

# ILLUMINATING THE BRAIN

Systems neuroscientists are pushing aside their electrophysiology rigs to make room for the tools of 'optogenetics'. **Lizzie Buchen** reports from a field in the process of reinvention.

**T**he centrifuge tube was the first that neuroscientist Philip Sabes had held in his hand for 15 years. The small, polypropylene container, no larger than a AAA battery, held a few drops of liquid at its base. It looked like water but, Sabes had been told by his collaborators, it contained a high concentration of viruses — and he had to get them into the brain of a monkey. “Honestly, I really felt like I didn’t know what I was doing,” says Sabes, of his work last November at the Keck Center for Integrative Neuroscience at the University of California, San Francisco (UCSF). “I basically knew nothing about molecular biology. This was way outside my area of expertise.”

Sabes’s training was in physics, machine learning and human perception, and his lab has been working with humans and non-human primates to develop models of how the brain turns perceptions into actions; for example, seeing a fly and swatting it away. He’s not alone in his molecular-biology naivety at the Keck Center — there is no cell-culture facility, no PCR machine and no bench-top centrifuge. The centre’s one ice machine spits out large cubes instead of the crushed ice routinely used for chilling reagents — it was ordered by mistake, and no one has cared enough to fix the situation. Sabes and his colleagues have

had no need for such apparatus. Researchers in their field of ‘systems neuroscience’ try to understand how networks of neurons process sensations and control behaviours such as learning and decision-making. And up to this point, much of their progress has been made using electrophysiology, stimulating and recording from the brains of animals as they perform a task or develop a new skill.

Now though, advances in a five-year-old field called optogenetics are convincing these scientists to crack open molecular-biology textbooks. Using a hybrid of genetics, virology and optics, the techniques involved enable researchers to instantaneously activate or silence specific groups of neurons within circuits with a precision that electrophysiology and other standard methods do not allow. Systems neuroscientists have longed for such an advance, which allows them their first real opportunity to pick apart the labyrinthine jumble of cell types in a circuit and test what each one does. “It has revolutionized my approach to science,” says Antonello Bonci, a neurophysiologist at the UCSF Ernest Gallo Clinic and Research Center in Emeryville who began using the technique in 2007. “It can clarify unequivocally the role of specific classes of cells, and solve controversies that have been going on for many, many years.” Among the

clarifications sought is the precise function of ‘place’ cells, hippocampal neurons that fire only when an animal finds itself in a specific location; another is the function of complex activity patterns observed when an animal is paying attention or executing a movement.

## A field’s evolution

The transition phase isn’t easy. Optogenetic tools were first used in cell cultures and mice, which are amenable to genetic manipulation. Now systems neuroscientists must adapt them to function in organisms they traditionally study such as rats, birds and primates. With the technical challenge comes a personal one, as researchers leave their experimental comfort zone for a new field.

Most, however, anticipate that any discomfort will be worthwhile. “This is God’s gift to neurophysiologists,” says Robert Desimone, director of the McGovern Institute for Brain Research at the Massachusetts Institute of Technology (MIT) in Cambridge, who has been using electrophysiology for more than 30 years. “Molecular techniques were always beyond us, so this is our first opportunity. It’s a revolution. But we’re catching up to the revolution that had been going on for the rest of the world.”

Optogenetics started out in 2005, when a team at Stanford University in California led by

Karl Deisseroth and his then-postdoc Ed Boyden inserted a light-sensitive channel from green algae, called channelrhodopsin-2 (ChR2), into neurons growing in a dish. Exposed to a pulse of blue light, the channels opened and a flood of positive ions poured into the neurons, making them fire<sup>1</sup>. Within a year, 30 labs had contacted Deisseroth to ask for the technology. By March 2010, Deisseroth had sent protocols or genetic constructs to more than 500 labs around the world, and Boyden, now at MIT, had sent them to in excess of 250.

The technique has been refined greatly in the past five years, but the basic steps are the same (see 'Six steps to optogenetics'). First, researchers create a genetic construct containing the ChR2 gene or another 'opsin' gene, along with genetic elements that control its expression — for example, a specific 'promoter' sequence. Then, they package up the construct in a virus. When the virus is injected into an animal's brain, it widely infects neurons and delivers the construct, but the opsin is expressed in only a subgroup of cells with the necessary machinery to activate its promoter. The opsin proteins sit in the membrane of those neurons, and researchers trigger them with light of a specific wavelength, typically delivered through an optic fibre threaded through an animal's skull. Deisseroth and Boyden have discovered or engineered many other opsins that allow neurons to be manipulated in different ways, including neural silencers.

The technique is popular, but complex. The fastest adopters were those who work with cells grown *in vitro* and animals such as flies, worms and mice, for which they could take advantage of established genetic tools and well-characterized animal lines. "The number of tangible results are still of the order of half a dozen or so. But we're on the rising phase of the exponential," says Karel Svoboda, a neuroscientist at the Howard Hughes Medical Institute's Janelia Farm research campus in Ashburn, Virginia, who last year started mapping mouse cortical circuits using optogenetics<sup>2</sup>.

From the start, these developments sent quivers through the systems-neuroscience community. "When I first saw this stuff, at a meeting where Karl was presenting some of the earliest results, I thought, 'Oh God, finally,'" says Loren Frank, another neuroscientist at the Keck Center. "And then I thought, oh great. Here I was, I'd gotten pretty good at the stuff I thought I needed to do, and now I have to learn an entirely new field."

Frank — who says he has "always had a bit of molecular-biology envy" — began

collaborating with Deisseroth to use optogenetics on rats just over three years ago, for his studies of place cells and hippocampal circuits. Silencing or activating specific cells could help show whether or how they help an animal explore a new location or recognize a familiar one. "If you want to actually understand the system, you have to start trying to get control of the system and actually dissect the circuit," says Frank. Standard techniques have not allowed that type of dissection. Electrodes inserted into an animal's brain typically stimulate hundreds of thousands of cells; using lesions or drugs also hits circuits like a hammer. Optogenetics could be a scalpel, turning particular neurons in a circuit on or off within milliseconds.

### From brain to behaviour

By April 2009, Frank was making progress in rats, achieving ChR2 expression in a discrete set of hippocampal neurons and getting them to fire. And down the hall, Sabes was ready to try the technique in primates as part of his efforts to understand circuits involved in hand-eye coordination. "I knew optogenetics was on the horizon, and it was potentially exciting, but realistically, I just didn't know anything about this stuff," he says.

Sabes and two songbird researchers at the Keck Center, Michael Brainard and Allison Doupe, teamed up with Frank, Deisseroth and another colleague, Linda Wilbrecht, a neuroscientist at the Gallo Center. The group, one of the first big collaborations aiming to apply optogenetics to rats, birds and primates, received a US\$1.6-million National Institutes of Health grant in September 2009 through the financial stimulus. The neuroscientists describe what they'd like to do — in Sabes's case, for example, alter patterns of activity that occur in the parietal lobe when an animal reaches for something, and work out which patterns are important for planning, initiating or adjusting the movement. Wilbrecht's lab leads efforts to generate the appropriate constructs and select the best viruses, and Deisseroth tries to build new viral and optogenetic tools that they need.

One huge advantage for mouse researchers has been the ready-made bank of animal lines engineered to express an enzyme called Cre recombinase in subsets of neurons, such as all dopaminergic ones, which they can use in combination with specially designed genetic constructs to achieve the cellular specificity they want. In other animals, a promoter must be found that will only be turned on in dopaminergic neurons. Some promoters and viruses that work in mice do not work in rats or

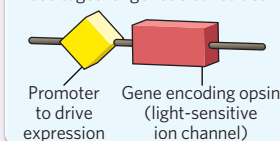
**"Optogenetics can solve controversies that have been going on for many, many years."**

## SIX STEPS TO OPTOGENETICS

With optogenetic techniques, researchers can modulate the activity of targeted neurons using light.

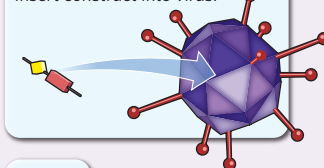
### STEP 1

Piece together genetic construct.



### STEP 2

Insert construct into virus.



### STEP 3

Inject virus into animal brain; opsin is expressed in targeted neurons.



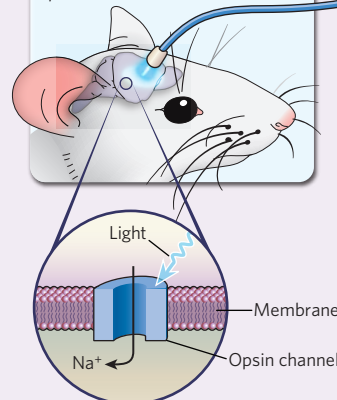
### STEP 4

Insert 'optrode', fibre-optic cable plus electrode.



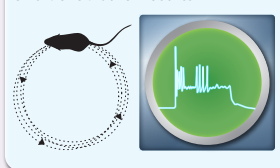
### STEP 5

Laser light of specific wavelength opens ion channel in neurons.



### STEP 6

Record electrophysiological and behavioural results.



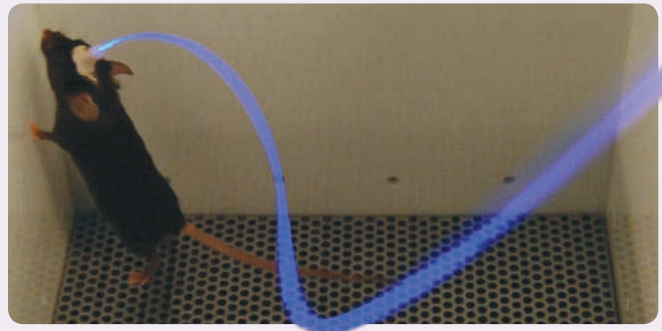
## Opto school

Nuné Lemaire stares down at a small black mouse that is sitting in a white plastic tub. The graduate student from the Massachusetts Institute of Technology in Cambridge reaches over to a heavy metal box, from which a thin, clear wire travels to a device mounted on the mouse's head. She flips a switch and the top of the mouse's skull glows blue. The animal begins walking quickly in anticlockwise circles until Lemaire flips the laser off. On cue, the mouse stops walking, brushes its whiskers and resumes sniffing.

The trick, although striking, is nothing new. The laser was activating neurons — made sensitive to blue light by genetic engineering — on the right side of the brain, compelling the animal

to walk in leftward circles. But it was new to Lemaire — she had implanted the headmount just a few hours before. She and a handful of others were taking part in the first four-day workshop on optogenetics, in March this year, at Karl Deisseroth's lab at Stanford University in California.

"Once they get the hands-on experience, they can be up and running in a couple weeks," says Deisseroth, who set up the course partly because he has been inundated with requests from people who want to learn the complex methods involved. The course — funded by Stanford — includes a lecture, hands-on experience and access to Deisseroth lab members and their years of expertise. "It was



Light delivered by an implanted fibre-optic cable activates neurons.

P. ANIKEVA/K. DEISSEROTH

questions, questions all the time, during lunch, during dinner, during all the experiments," says Lemaire. "We asked what kind of cement they're using, how they make sure the fibre doesn't get twisted." Deisseroth is building an

Optogenetics Innovation Laboratory for running workshops. Proper teaching is crucial, he says, "because of the unusual combination of skills required. Much more is needed than simply sending clones around". **L.B.**

primates, meaning that researchers have to start from scratch. Progress has been faster in rats because the animals are relatively easy to breed and are similar enough to mice for methods to be transferable. In March 2009, Deisseroth was the first to publish a rat optogenetic study, in which he manipulated circuit components in a rat model of Parkinson's disease to work out which parts might account for the relief afforded by deep-brain stimulation<sup>3</sup>.

An additional complication for primate researchers is that primate brains are larger than those of rodents, making it difficult to ensure the injected virus and the activating light penetrate deep enough. And troubleshooting in monkeys will take much longer than in rats, given the long lifetime and high value — experimental, financial and ethical — of the animals. But in April 2009, Desimone and his colleagues worked with Boyden to publish the first experiment showing that viruses can be used to insert opsin channels and control neural activity in a macaque<sup>4</sup>.

By November 2009, Sabes could be found holding his tube of virus, trying to get a similar primate system going in his lab. This was a pilot experiment: he was simply trying to see if he could get some of the virus into neurons at all. After bending and breaking a few needles, he successfully injected the tube's contents into a monkey's brain. He and two graduate students then hacked together an 'optrode' by gluing together a fibre-optic cable and an electrode. They fed the wires through a head fixture normally used for electrophysiology, and rigged half of a syringe as a support. "It felt very much like garage-development days, cutting stuff up to see what works," says Sabes.

In January, with the optrode inserted into the region where the virus had been injected

two months earlier, Sabes flipped on a blue laser and watched a screen showing electrical readout from the optrode. Yellow waveforms flashed across the screen, paired with a flurry of clicking noises: spikes of neural activity.

### Experts on hand

Like Sabes, the few other electrophysiologists starting to tackle primate optogenetics are keeping experts in the technique close by to avoid making a novice's mistakes (see 'Opto school'). "It's critical that at least the initial phase is collaborative," says Krishna Shenoy, a primate electrophysiologist at Stanford University who has been working closely with Deisseroth for two years to explore how neurons in the brain's motor cortex control arm movements.

Boyden's paper is still the only optogenetics publication on primates, and the technique is some way from generating new discoveries. It's still not known whether the opsins will be expressed consistently for the year or two required to train monkeys in sophisticated behaviours and then record from neurons repeatedly. "If it only expresses at the right level for a few months and then dies off or expresses too much, it's just untenable," says Shenoy.

All these questions take time and money to answer, and not every lab has an appetite for the work. "It's probably going to have a big impact, and I'm definitely interested," says Tirin Moore, a neurobiologist at Stanford. "But I'm less interested in pioneering the approach in monkeys and more interested in using the tools once it's clear that they work."

Meanwhile, Boyden and Deisseroth are hammering out some of the problems. Last month, Deisseroth reported a system in mice that could make detailed knowledge of promoters

unnecessary: two viruses containing separate genetic constructs are injected into two connected brain regions. Only neurons that run between the two regions will receive doses of both constructs, which then interact to express the opsin<sup>5</sup>. Boyden is designing a light source that would weigh a fraction of a gram so that animals can walk around freely rather than being tethered to a fibre-optic cable.

"Soon enough, this is going to be standard technology," says Sabes. "The hardware will be there, the viruses will be there, there will be a handful of constructs that everyone agrees works reasonably well." That aim is likely to be furthered by a two-year, \$14.9-million grant from the US Defense Advanced Research Projects Agency in Arlington, Virginia, that Sabes, Deisseroth and six other labs, led by Shenoy, won in April. The team will attempt to use primate optogenetics to explore brain repair after injury, including possible light-based neural prosthetics, devices that might stimulate appropriate patterns of activity in the surviving neurons.

For now, Sabes is still trying to analyse the results of his first monkey experiment. He has sent the animal's brain to a friend competent in histology, who will slice it, paper thin, onto slides. Then Sabes will have to grapple with a fluorescent microscope for the first time, trying to work out where and how well the opsin was actually expressed.

"I'm sure it's not that hard," says Sabes, "but I've never done it".

**Lizzie Buchen is a freelance writer based in San Francisco.**

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**"It felt like garage-development days, cutting stuff up to see what works."**

## Q&amp;A

# Strong medicine for French research

The medical-research adviser to France's president aims to shift power and money to universities.

**Arnold Munnich** has been scientific adviser for medical research and health to French president Nicolas Sarkozy since 2007. A paediatric geneticist at University Paris Descartes, he heads the Jean Frézal Centre of Medical Genetics at the Necker Hospital for Sick Children in Paris, an institute run jointly with INSERM, the national biomedical research agency.

## How did you come to be chosen as a presidential adviser?

It was serendipitous. Back in 2000, I wrote a provocative article in the newspaper *Le Monde*, criticizing the hype around the sequencing of the human genome. As a clinical geneticist on the front line of treating children with genetic diseases, I knew that the genome is ultimately just a long list of nucleotides, and that we still have a long way to go to understand it — that this was not going to change the life of the patient immediately.

Sarkozy, who was at that time mayor of Neuilly-sur-Seine in the western suburbs of Paris, happened to read the article and invited me to a breakfast meeting. He was extremely concerned about the ethical issues and health effects of genetics research, and also its possible negative consequences. I invited him to spend a day with us at the lab and hospital, which he did. Subsequently, we kept in touch and, during his presidential campaign in 2007, he occasionally asked me for advice. After the election he called me and said, "Arnold, do you want to be my adviser for medical research?" I told him, "Are you kidding? You know my life, and how busy I am already. I have no political ambitions, and am very happy in my job here." But he insisted that I could do more. So ever since, although I've kept the same hours at the hospital, I now work at the Élysée Palace from seven to nine in the morning and from six to nine in the evening.

## What is your main role as adviser?

I mostly focus on reforms of the funding and organization of French biomedical research. France historically had weak universities, and to remedy this the government created large national research agencies before and after the Second World War. These controlled every aspect of research — they

designed the programmes, funded them, carried them out and evaluated them.

But this centralized monolithic structure, inherited from the past, has its limitations. Sarkozy has decided that the French scientific system will instead pursue the model that has been so successful in other highly developed nations, with powerful research universities at its centre. One of his first actions after being elected in 2007 was to pass a law giving the universities autonomy from centralized state control. They can now manage their own land and buildings, entirely handle their own budgets and have some flexibility to set researchers' salaries as they see fit, instead of being tied to rigid national salary scales — this gives them more openness and freedom to retain, or to attract, the best researchers.

## Are the universities ready for this change?

The crucial task is to identify and support the most competitive research universities. Not all teaching universities can also be first-class research universities, so we are encouraging consolidation and mergers.

In 2008, we launched 'Operation Campus', a €5-billion (US\$6.2-billion) project to restructure and refurbish a select few universities. And as part of the €35-billion package of the *grand emprunt*, or 'big loan', announced by the president last December to boost the country's long-term competitiveness (see *Nature* 462, 838; 2009), €11 billion will go to the universities. There will be competition for the money, with five to ten of the best campuses receiving billion-euro endowments.

## Where does this leave the research agencies?

They now pilot national strategy and fund research, much like research councils. But universities have become the main operators, in charge of organizing their own research strategies. That has been accompanied by a marked shift away from the dominance of recurrent research funding, where the agencies funded labs on a rolling basis, to one where individual labs compete nationally for funding on the basis of grant proposals



for projects — with the major player here being the National Research Agency, based in Paris, which was created in 2007 and has a budget of €850 million this year. Independent evaluation has also been added outside of the agencies, with the creation in 2007 of the Agency for Evaluation of Research and Higher Education in Paris.

## An evaluation of INSERM in 2008 recommended a fusion of life-science research across the agencies. How far has this come?

The panel said that INSERM should be the lead agency for biomedical research, and fold in or absorb much of the life-science research done elsewhere, including that at specialized agencies for AIDS, cancer and other research. In 2009, we created the National Alliance for Life and Health Sciences, an association of eight bodies including INSERM, the National Centre for Scientific Research, the Pasteur Institute and universities that will coordinate research on specific themes across all these agencies. Eventually we would like to see even further consolidation.

## What about clinical research, where France is far weaker than the United Kingdom, for example?

The *grand emprunt* will help here. France has 32 university teaching hospitals, but they are more focused on teaching and care and less on research than their UK or US counterparts. We aim to upgrade five university teaching hospitals to centres of excellence with a focus on research. A long-standing bad habit in France has been to seek to spread funding evenly, but this only reduces effectiveness. So we want to fund just a few centres of excellence, those that are most open to reforming their practices to improve research. Overall, reforms of research are moving fast — faster perhaps than mentalities. ■

Interview by Declan Butler.

### Correction

The News Feature 'Illuminating the brain' (*Nature* 465, 26–28; 2010) contained an incorrect statement regarding the experimental work of Philip Sabes. In fact, no needles were broken during the primate experiment.