

The C-terminal module IV of connective tissue growth factor is a novel immune modulator of the Th17 response

Raquel Rodrigues-Díez¹, Raúl R Rodrigues-Díez¹, Sandra Rayego-Mateos¹, Beatriz Suarez-Alvarez¹, Carolina Lavoz¹, Luiz Stark Aroeira², Elsa Sánchez-López³, Macarena Orejudo¹, Matilde Alique¹, Carlos Lopez-Larrea⁴, Alberto Ortiz⁵, Jesús Egido⁶ and Marta Ruiz-Ortega¹

Connective tissue growth factor (CTGF/CCN2) is a matricellular protein susceptible to proteolytic degradation. CCN2 levels have been suggested as a potential risk biomarker in several chronic diseases. In body fluids, CCN2 full-length and its degradation fragments can be found; however, their *in vivo* effects are far from being elucidated. CCN2 was described as a profibrotic mediator, but this concept is changing to a proinflammatory cytokine. *In vitro*, CCN2 full-length and its C-terminal module IV (CCN2(IV)) exert proinflammatory properties. Emerging evidence suggest that Th17 cells, and its effector cytokine IL-17A, participate in chronic inflammatory diseases. Our aim was to explore whether CCN2(IV) could regulate the Th17 response. *In vitro*, stimulation of human naive CD4⁺ T lymphocytes with CCN2(IV) resulted in differentiation to Th17 phenotype. The *in vivo* effects of CCN2(IV) were studied in C57BL/6 mice. Intraperitoneal administration of recombinant CCN2(IV) did not change serum IL-17A levels, but caused an activation of the Th17 response in the kidney, characterized by interstitial infiltration of Th17 (IL17A⁺/CD4⁺) cells and upregulation of proinflammatory mediators. In CCN2(IV)-injected mice, elevated renal levels of Th17-related factors (IL-17A, IL-6, STAT3 and ROR γ t) were found, whereas Th1/Th2 cytokines or Treg-related factors (TGF- β and Foxp-3) were not modified. Treatment with an anti-IL-17A neutralizing antibody diminished CCN2(IV)-induced renal inflammation. Our findings unveil that the C-terminal module of CCN2 induces the Th17 differentiation of human Th17 cells and causes a renal Th17 inflammatory response. Furthermore, these data bear out that IL-17A targeting is a promising tool for chronic inflammatory diseases, including renal pathologies.

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Connective tissue growth factor (CTGF/CCN2) is a cysteine-rich secreted matricellular protein that exerts many biological functions depending on the cellular context.¹ CCN2 is a 36–38 kDa monomeric protein composed of 349 residues. This protein has a modular architecture comprising four functional modules: an amino-terminal insulin-like growth-factor-binding domain (IGFB), the cysteine-rich domain (also called von Willebrand type c domain), a thrombospondin type 1 repeat domain, and the C-terminal heparin-binding domain (Module 4) also known as a carboxy-terminal cystine knot domain (namely here CCN2(IV)).² Between modules

2 and 3 is the ‘hinge region’ that is cleaved by most proteases, including matrix metalloproteases (MMPs) 1, 2, 3, 7, 9, and 13, elastase, and plasmin, the latter two of which also cleave the individual modules to yield four fragments.³ In particular, MMP2 clips CCN2 *in vitro* to a 10-kDa fragment and that both full-length and clipped CCN2 were recovered from fibroblast cell culture supernatants.¹ In biological fluids, CCN2 can be found in various forms: full-length molecule, fragments including N-terminal and C-terminal halves cleaved in the ‘hinge region,’ as well as the individual 10–12 kDa that corresponds to CCN2(IV).^{1,4}

¹Cellular Biology in Renal Diseases Laboratory, Instituto de Investigación Sanitaria Fundación Jiménez Díaz, Universidad Autónoma Madrid, Madrid, Spain; ²Servicio de Nefrología, Hospital Universitario La Paz- IdiPAZ, Madrid, Spain; ³Translational Oncology Unit, Instituto de Investigaciones Biomédicas ‘Alberto Sols,’ CSIC, Madrid, Spain; ⁴Department of Immunology, Hospital Universitario Central de Asturias, Oviedo, Spain; ⁵Dialysis, Fundación Jiménez Díaz, Madrid, Spain and ⁶Renal Research Laboratory, Fundación Jiménez Díaz, Madrid, Spain

Correspondence: Dr M Ruiz-Ortega, PhD, Cellular Biology in Renal Diseases Laboratory, Instituto de Investigación Fundación Jiménez Díaz, Universidad Autónoma Madrid, Avda. Reyes Católicos, 2, 28040 Madrid, Spain. E-mail: mruizo@fdj.es

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These variations produce a subfamily of secreted proteins with distinct functions. However, the biological *in vivo* effects of CCN2 and its fragments are still unknown. Several independent studies have evaluated CCN2 levels in the urine and/or serum in several chronic kidney diseases. Some groups have found elevated levels of the full-length CCN2,^{5,6} or the N-terminal,^{6,7} or the C-terminal^{8–10} CCN2 fragments determined by ELISA, using antibodies that recognized each part of the molecule. Based on these data, CCN2 has been proposed as a risk biomarker of human diabetic nephropathy and other forms of chronic kidney disease,^{2,5–10} and for cardiac dysfunction in patients exhibiting myocardial fibrosis and chronic heart failure.¹¹

CCN2 has been found overexpressed in almost all human disorders characterized by excessive scarring and fibrosis.^{2,12} The concept of CCN2 as a downstream mediator of profibrotic factors, including transforming growth factor- β (TGF- β) and Angiotensin II, is the chief operating paradigm in the field.^{13,14} Therapeutic approaches that selectively block endogenous CCN2 activity have proven beneficial effects in fibrotic-related diseases, including experimental lung, liver, vascular, and renal diseases, and some authors have suggested that CCN2 could be a therapeutic target for fibrosis.^{12–16} However, cardiac CCN2 overexpression conferred cardioprotection in Angiotensin II-infused mice and in ischemia–reperfusion injury,^{17,18} showing that CCN2 exerts protective effects in some pathological settings. These data suggest that before using CCN2 blockers in humans, it is necessary to fully understand the *in vivo* biological functions of this protein and its fragments.

Although many authors have proposed that CCN2 overproduction plays a major role in pathways that lead to fibrosis,¹² the direct contribution of CCN2 to fibrosis *in vivo* is still unclear. In contrast, *in vitro* studies support the novel concept of CCN2 as a proinflammatory cytokine.¹⁹ CCN2 is a chemotactic factor for immune cells,²⁰ has promoted cell adhesion and migration, and has upregulated proinflammatory factor production, including cytokines, chemokines, and adhesion molecules.¹⁹ We have previously demonstrated that systemic administration of CCN2(IV) elicited the recruitment of inflammatory cells in the murine kidney after 24 h, suggesting that CCN2(IV) could induce acute inflammation *in vivo*,²¹ but there are no data on sustained inflammation. Several studies have investigated the biological functions of CCN2 modules. *In vitro* studies using recombinant proteins have shown that the full-length CCN2 and its C-terminal module IV (named here CCN2(IV)) increased matrix production, caused epithelial-to-mesenchymal transition of tubular epithelial cells, and induced endothelial cell migration and proliferation, whereas the N-terminal domain was not effective in these studies.^{22–24} In human renal fibroblasts, the N-terminal domain binds IGF-I and enhances the induction of matrix components.^{25–27} Data from several laboratories support an important role for CCN2(IV) in the regulation of several

process induced by other proteins, including mitosis, angiogenic activity, and heparin binding.^{4,28,29} These data show the complexity of this molecule.

The classical view of the immune response regulation based on Th1/Th2 paradigm of T helper (Th) cells has been enlarged by the discovery of novel subsets, including proinflammatory Th17 (CD4⁺/IL-17A⁺) cells and immunosuppressive CD4⁺/CD25^{high}/Foxp3⁺ regulatory T cells (Tregs).^{30,31} Growing evidence suggests that IL-17A, the Th17 effector cytokine, besides participating in immune-mediated diseases, is also involved in chronic inflammatory diseases such as atherosclerosis, and hypertension.^{30–33} Recent studies suggest that blockade of IL-17A is a promising tool for chronic human inflammatory diseases, as observed in rheumatoid arthritis, uveitis, and psoriasis.^{34–36} Our aim was to explore whether the CCN2(IV) could regulate the Th17 response and therefore contribute to persistent inflammation. First, we evaluated the direct effect of CCN2(IV) in human CD4⁺ T lymphocyte differentiation. Second, we investigated whether CCN2(IV) could regulate Th17 response *in vivo* using a model of systemic CCN2(IV) administration into mice.

MATERIALS AND METHODS

Human T-Cell Isolation

Human peripheral blood mononuclear cells (PBMCs) were obtained from peripheral blood of healthy donors (Blood Transfusion Center, Oviedo, Spain) after written informed consent and approval in accordance with the Declaration of Helsinki Principles. PBMCs were isolated by Ficoll-Hypaque density gradient centrifugation (Lymphoprep, Oslo, Norway). Naive CD4⁺ T cells were subsequently isolated from PBMCs by negative selection using naive CD4⁺ T cell isolation Kit II (Miltenyi Biotec GmbH, Bergisch Gladbach, Germany) according to the manufacturer's instructions. The purity of isolated cells was always >95%.

In Vitro Th17 Polarization Cell

Naive T-cell differentiation toward Th17 effector cells was performed as following. Isolated naive CD4⁺ T cells were cultured at a concentration of 5×10^5 cells/ml in complete RPMI-1640 medium supplemented with 10% heat-inactivated fetal calf serum (FBS) and 100 U/ml streptomycin–penicillin solution at 37 °C and 5% CO₂ in 48-well plates. For the experiments, cells were stimulated in Th17 cell polarizing medium containing 50 U/ml interleukin-2 (IL-2), plate-bound anti-CD3 (3 μ g/ml) and anti-CD28 (3 μ g/ml), and neutralizing anti-IFN- γ and anti-IL4 monoclonal antibodies (5 μ g/ml each, eBiosciences, San Diego, CA, USA), and different stimuli were added (1 ng/ml TGF- β , 20 ng/ml IL-6, 50 ng/ml CCN2(IV); supplied by Peprotech, NJ, USA). After 2 days, cells were restimulated with Th17 cell polarizing medium and stimuli, adding additionally 10 μ g/ml IL-23 to each well. After 5 days, the Th17 phenotype was analyzed by flow cytometry.

Design of the Experimental Model

Studies were performed in adult male C57BL/6 mice (9–12 weeks old, 20 g; obtained from Harlan Interfauna Ibérica, S.A., Barcelona, Spain) and maintained at the local animal facilities, with free access to food and water, normal light/dark cycles, and under special pathogen-free conditions. C57BL/6 mice received a single intraperitoneal injection of recombinant CCN2(IV) (corresponding to human CCN2 module 4, a protein of 98A.A. residues and 11 kDa; purchased from Peprotech), at the dose of 2.5 ng/g of body weight, dissolved in saline, as described previously.²¹ The recombinant CCN2(IV) present endotoxin levels of <0.01, and the quality and purity was also confirmed by MALDI-TOF (data not shown). Mice were studied after 5, 10, and 15 days ($n = 10$ mice in the 10-day group and 5 mice in the other groups). A control group with same volume of saline injection was also studied ($n = 10$ mice). To discard that CCN2(IV) inflammatory reaction was caused by trans-xeno differences in human/murine protein, the IgM/IgG ratio was evaluated in mice serum (no differences between groups were found, data not shown).

The model of unilateral ureteral obstruction (UO) was studied in male C57BL/6 mice. The model was performed under isoflurane-induced anesthesia; the left ureter was ligated with silk (4/0) at two locations and cut between ligatures to prevent urinary tract infection (obstructed kidney), as described previously.³⁷ A control group of sham-operated mice was also studied ($n = 8$ mice per group).

For IL-17A neutralization studies, mice were injected with 100 μ g/mouse of anti-IL-17A neutralizing antibody or isotype K1 IgG control (eBiosciences, $n = 5$ mice per group), based on previous studies,³⁸ and starting 24 h before CCN2(IV) injection and every 72 h thereafter until the time of killing at 10 days.

Mice were killed under anesthesia (Ketamine-HCl/Xylazine-HCl) and then the kidneys were perfused *in situ* with cold saline before removal. One kidney from each mouse was fixed in buffered formalin, embedded in paraffin, and used for immunohistochemistry. The other kidney was snap-frozen in liquid nitrogen for gene and protein studies. All the procedures on animals were performed according to the European Community and Instituto de Investigación Sanitaria Fundación Jiménez Díaz Animal Research Ethical Committee guidelines.

Flow Cytometry

For analysis of intracellular cytokine production, cultured human CD4⁺ T cells were stimulated with PMA (50 ng/ml) and ionomycin (1 μ g/ml; both from Sigma-Aldrich) in the presence of monensin (eBiosciences) for 6 h. Following surface staining with APC-labeled anti-CD4 and PerCP-Cy5.5-labeled anti-CD3 (Biolegend, San Diego, CA, USA), cells were fixed and permeabilized using Fix and Perm reagent (eBiosciences). Furthermore, intracellular cytokine staining was performed with an anti-IL17A-PE antibody

(Biolegend). Isotype controls were used to define markers settings. In blood samples from mice models, circulating levels of T lymphocytes were evaluated. Cell surface staining was performed using FITC-labeled anti-CD3 and PE-labeled anti-CD4 (BD Pharmingen, San Diego, CA, USA). After cell surface staining, factor X-linked forkhead/winged helix (Foxp3) was stained using Foxp3 staining kit (BD Pharmingen) according to the manufacture's instructions. Samples were acquired on a BD FACScaliburTM Cytometer using the CellQuest Pro Software package (Becton Dickinson, Franklin Lakes, NJ, USA). The specific fluorescence intensity was quantified as the mean fluorescence intensity (MFI) calculated by subtracting the background of isotype-matched control staining from the total fluorescence.

Renal Histology and Immunohistochemistry

Paraffin-embedded sections were stained using standard histology procedures. Immunostaining was carried out in 3 μ m thick tissue sections that were deparaffinized and antigen retrieved using the PT Link system (Dako Diagnósticos S.A, Barcelona, Spain) with Sodium Citrate Buffer (10 mM) adjusted to pH 6 or pH 9 depending on the immunohistochemical marker. Immunohistochemical staining was performed using the Dako Autostainer (Dako). Briefly, endogenous peroxidase was blocked and then sections were incubated for 30–20 min at room temperature with the following primary antibodies: mouse anti-CD4 (Dako, IS649) and rabbit anti-CD3 (Dako, IS503) ready to use, rat anti F4/80 (1/5000; Serotec, MCAP497), and rabbit anti-IL-17A (1:250; Abcam: ab9565). After washing, slides were treated with the EnVisionTM DuoFLEX Doublestain System using 3,3'-diaminobenzidine as chromogen. For F4/80 staining, a rabbit anti-rat IgG antibody was used as linker before EnVision treatment. Sections were counterstained with Carazzi's hematoxylin. The total number of positive stained cells was quantified in five randomly chosen fields ($\times 200$) using Image-Pro Plus software. Data are expressed as positive stained area *vs* total analyzed area. Samples from each animal were examined in a blind manner.

Double immunofluorescence staining for CD4 and IL-17A was performed in OCT (Optimal Cutting Temperature, Sakura) embedded frozen sections. Before fixation (PFA 4% 10 min at room temperature), slides were incubated for 1 h at room temperature with 4% of bovine serum albumin (BSA) and serum in 1 \times phosphate-buffered saline (PBS) to eliminate nonspecific protein binding sites. Primary antibody rabbit anti-IL-17A (1:250; Abcam: ab9565) was incubated overnight at 4 °C. After washing, slides were treated with the corresponding anti-IgG Alexa488-conjugated secondary antibody (1:300). To determinate T-cell colocalization, antibody to CD4 was incubated overnight followed by an Alexa633-conjugated secondary antibody (1:300). Nuclei were contrasted with DAPI. Negative controls were incubated with a nonspecific immunoglobulin of the same isotype as the primary antibody and without primary antibody.

Protein Studies

Kidney extracts were lysed in lysis buffer (50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 2 mM EDTA, 2 mM EGTA, 0.2% Triton X-100, 0.3% NP-40, 100 μ M phenylmethylsulphonyl-fluoride, 1 mM dithiothreitol, 100 μ M Na₃VO₄, and 1 mM protease-inhibitor cocktail (Sigma)). Nuclear proteins were isolated as described previously.²¹ Protein concentration was determined by the BCA method (Pierce). Tissue protein extracts (30 μ g/lane) were separated on 8–12% polyacrylamide-SDS gels under reducing conditions. Samples were then transferred onto PVDF membranes (Bio-Rad, Alcobendas, Spain), blocked with 5% nonfat dry milk, in TBS with 0.05% Tween-20, and incubated overnight at 4 °C with the primary antibodies, subsequently incubated with peroxidase-conjugated IgG (Amersham), and developed by ECL chemiluminescence (GE Healthcare, Buckinghamshire, UK). Autoradiographs were scanned using the Gel Doc™ EZ imager and analyzed with the Image Lab 3.0 software (Bio-Rad).

The primary antibodies used were: Affinity Purified anti-mouse ROR γ t (1:1000; eBiosciences: 14-6981), Affinity Purified anti-mouse/human/rat Foxp3 (1:1000; eBiosciences: 14-4774), rabbit anti-phospho-Stat3 (Tyr705, 1:1000; Cell Signaling; #9131), and p65 NF- κ B subunit (C-20, 1:1000; Santa Cruz; sc-372). The efficacy of protein loading and transfer to membranes was assessed by incubation with mouse anti-GAPDH antibody (1:5000; Chemicon, MAB374). In renal protein extracts, cytokine levels were analyzed by ELISA (100 ng/ml). Interferon- γ (IFN- γ), IL-4, IL-10, IL-17A, and TGF- β protein levels were assayed by an ELISA kit from eBiosciences; IL-6, monocyte chemotactic protein-1 (MCP-1), and RANTES (Regulated upon Activation, Normal T-cell Expressed, and Secreted) from BD Bioscience. Cytokine levels were quantified by comparison with a standard curve and data were expressed as fold change over the mean of value of control mice levels.

Gene Expression Studies

Total RNA was isolated from renal samples with Trizol (Invitrogen, Groningen, The Netherlands). cDNA was synthesized using the High capacity cDNA Archive Kit (Applied Biosystems, Foster City, CA, USA) using 2 μ g of total RNA primed with random hexamer primers, following the manufacturer's instructions. Multiplex RT-PCR was performed using fluorogenic (FAM) primers designed by Assay-on-Demand™ gene expression products (Applied Biosystems): MCP-1: Mm00441242_m1, RANTES: Mm01302428_m1, monocyte chemotactic protein-1 receptor 2 (CCR2): Mm-99999051-gH, CXCL1 (GRO- α): Mm04207460_m1, α -smooth muscle actin (α -SMA): Mm01546133_m1, Fibronectin: Mm01256734_m1, Type I Pro-Collagen: Mm00483888_m1, Vimentin: Mm00449208_m5. Data were normalized to eukaryotic ribosomal 18s rRNA (4310893E) (VIC). IL-17A gene expression (sense 5'-GGACTC TCCACCGCAATGA-3' and antisense 5'-GACCAGGATCTCT TGCTGGA-3') was detected using SYBR Green Master Mix

(Applied Biosystems) and normalized with eukaryotic ribosomal 18s (sense 5'-CAGCTTTGCAACCATACTCCC-3', and antisense 5'-CCGTCGTAGTTCCGACCATAA-3'). The mRNA copy numbers were calculated for each sample by the instrument software using Ct value ('arithmetic fit point analysis for the lightcycler'). Results were expressed in copy numbers, calculated relative to control mice, after normalization against 18s.

Statistical Analysis

Statistical analysis was done using the SPSS statistical software (version 11.0, Chicago, IL, USA). After Kolmogorov–Smirnov test, which determined the nonnormal sample distribution of the data, differences between groups were assessed by Mann–Whitney *U*-test. Differences were considered significant when $P < 0.05$.

RESULTS

CCN2(IV) Induced Th17 Differentiation of Human Naive CD4⁺ T Cells

Depending on cytokine environment, human naive CD4⁺ T cells can differentiate into different Th effector subsets. IL-6 is an indispensable factor that generates Th17 cells from naive T cells³¹ through activation of the retinoic acid-related orphan receptor- γ t (ROR γ t) and signal transducer and activator of transcription 3 (STAT3).^{39,40} *In vitro* stimulation of isolated human naive CD4⁺ T cells, in Th17 cell polarizing conditions as described in the Materials and Methods section, with IL-6 or CCN2(IV) for 5 days induced a significant increase in the number of IL-17A⁺-producing CD4⁺ T cells (Th17 cells, determined by flow cytometry) compared with unstimulated cells in the same cultured conditions (Figures 1a and b). In CCN2(IV)-treated CD4⁺ T cells, elevated levels of phosphorylated STAT-3 were found (Figure 1c), showing that CCN2(IV) regulates this molecular mechanism involved in Th17 differentiation.

CCN2(IV) *In Vivo* Administration in Mice Did Not Modulate Circulating IL-17A Levels

In murine models of inflammatory diseases, including multiple sclerosis, inflammatory bowel disease, and arthritis, serum IL-17A levels are elevated, and the Th17 cell population is expanded and plays a pathogenic role;³⁰ therefore, we first evaluated whether systemic administration of CCN2(IV) in mice could regulate circulating IL-17A levels. However, in CCN2(IV)-injected mice, plasma IL-17A levels were not modified at any time point studied (5, 10, and 15 days; Table 1), and remained at similar levels to control mice. Moreover, the total number of circulating CD4⁺ cells were not changed between groups (evaluated flow cytometry, data not shown). Our data suggest that CCN2(IV) *in vivo* did not induce an expansion of systemic murine Th17 cells, the main cells responsible for circulating IL-17A levels, at least in the conditions of our study.

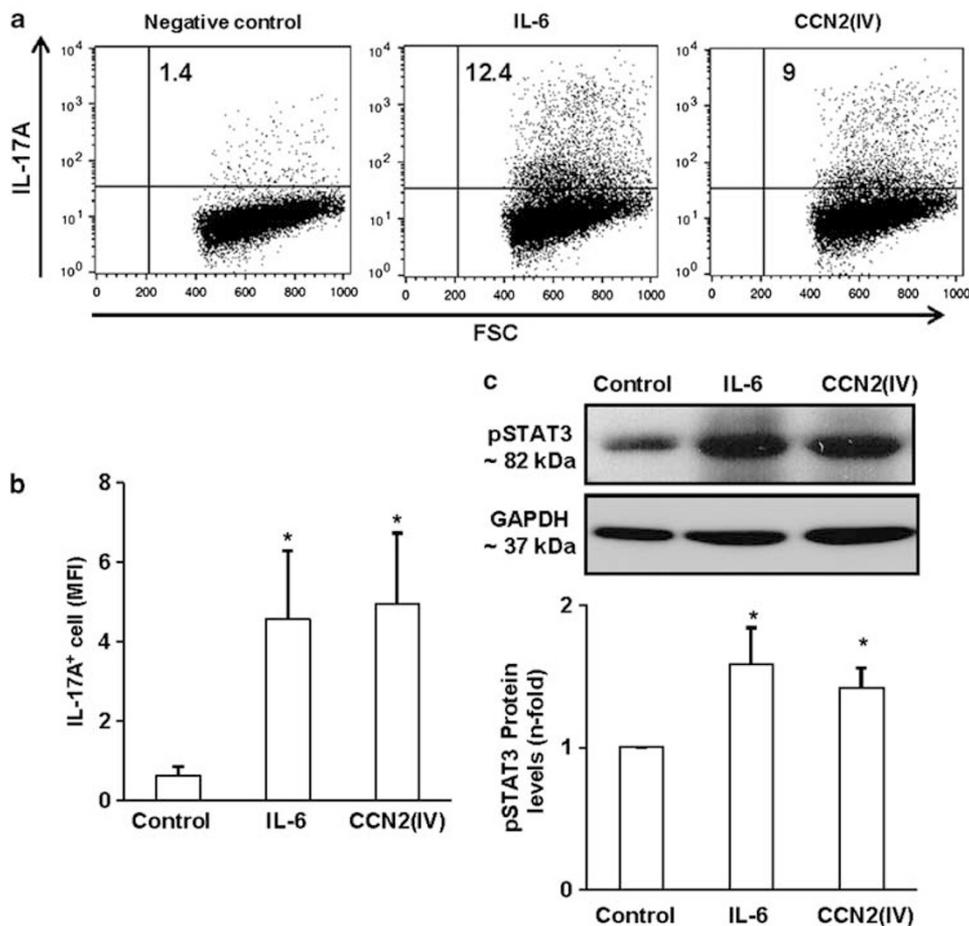


Figure 1 CCN2(IV) *in vitro* induced Th17 differentiation of human naive CD4⁺ T cells. Naive CD4⁺ T cells were isolated from PBMCs. 5 × 10⁵ cells/ml were incubated in Th17 cell polarizing medium (containing IL-2, anti-CD3, anti-CD28, neutralizing anti-IFN- γ , and anti-IL4) in the presence or absence of different stimuli (20 ng/ml IL-6 or 50 ng/ml CCN2(IV)) for 5 days. Intracellular IL-17A staining was evaluated by flow cytometry and expressed as mean fluorescence intensity (MFI) calculated by subtracting the background of isotype-matched control staining from the total fluorescence. (a) Dot plots show a representative experiment of the induced IL-17A expression on gated CD4⁺ T cells after stimulation with IL-6 or CCN2(IV) cytokines, and without stimulation. (b) The percentage of CD4⁺ T cells expressing IL-17A cells (mean \pm s.e.m. of six experiments done) is shown. (c) Activation of STAT-3 was evaluated by levels of phosphorylated STAT3 in cellular extracts. A representative western blot experiment and data as mean \pm s.e.m. as fold change over control of four experiments. **P* < 0.05 vs control.

Table 1 IL-17A serum levels in CCN2(IV)-injected mice

group	IL-17A serum levels	<i>P</i> -value vs control
Control	32.5 \pm 8.4	
CCN2(IV), 5 days	30.5 \pm 2.0	1
CCN2(IV), 10 days	26.7 \pm 4.1	0.905
CCN2(IV), 15 days	34.5 \pm 7.2	0.841

C57BL/6 mice received a single i.p. injection of 2.5 ng/g body weight recombinant CCN2(IV) and were killed after 5, 10, or 15 days (*n* = 5, 10, and 5 mice per group, respectively) compared with control group (i.p. vehicle (saline), *n* = 10). Data are expressed in pg/ml as mean \pm s.e.m. of 5 to 10 mice per group.

CCN2(IV) Induced an Activation of the Th17 Response in the Kidney

Next, we evaluated whether CCN2(IV) could modulate the Th17 response at tissue levels. CCN2(IV) *in vivo* induced an inflammatory response in the murine kidney after 24 h,²¹ but there were no data at longer times. The involvement of a specific Th response activation can be determined by the evaluation of Th hallmark cytokines (Th1, Th2, Th17, and Treg cell-related cytokines: IFN- γ , IL-4, IL-17A, and TGF- β 1/IL-10, respectively). We have previously observed that after 24 h, CCN2(IV) increased renal levels of IFN- γ and IL-4, downregulated IL-10, and did not change IL-17A renal levels.²¹ After 5 days, a slight upregulation of renal levels of all

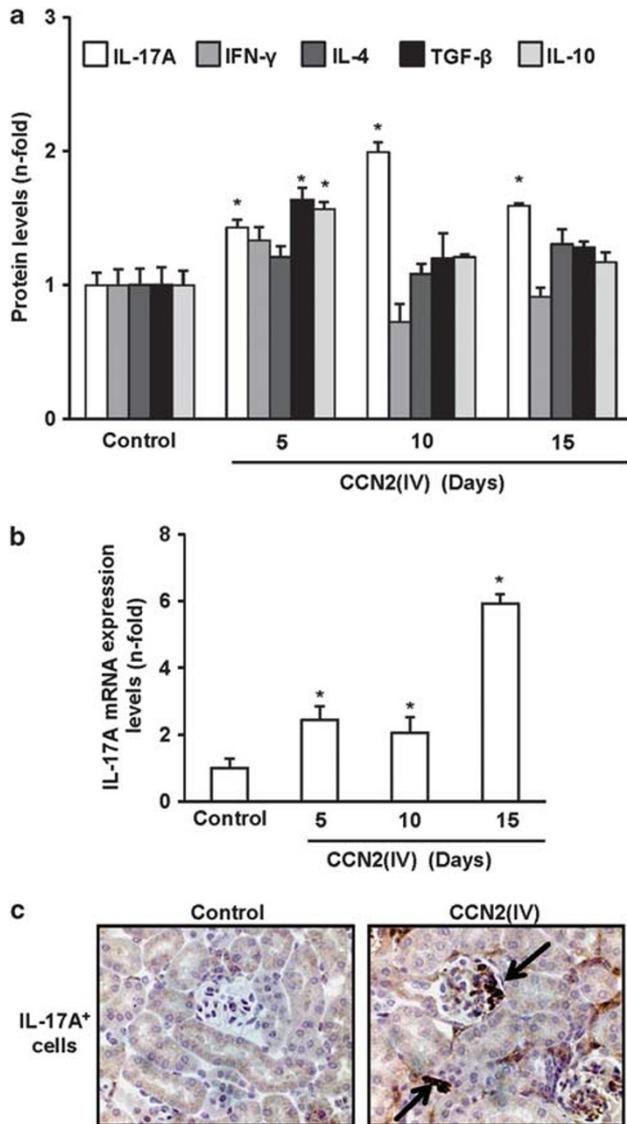


Figure 2 CCN2(IV) *in vivo* upregulated the Th17 hallmark cytokine IL-17A in the kidney, whereas it did not change renal levels of Th1, Th2, or Treg-related cytokines. C57BL/6 mice received a single i.p. injection of 2.5 ng/g body weight recombinant CCN2(IV) or vehicle (saline) and were killed after 5, 10, or 15 days ($n = 5, 10, \text{ and } 5$ mice per group, respectively) compared with control group (vehicle; $n = 10$). (a) Renal protein levels of IL-17A, IFN- γ , IL-4, IL-10, and TGF- β (Th17, Th1, Th2, Treg, and hallmark cytokines) analyzed by ELISA and expressed as fold change over control of mean \pm s.e.m. (10 to 5 animals per group). * $P < 0.05$ vs control. (b) Total kidney RNA was isolated and IL-17A mRNA expression was evaluated by real-time PCR. Results are expressed as fold change over control of mean \pm s.e.m. (5 to 10 animals per group). * $P < 0.05$ vs control. (c) Immunolocalization of IL-17A in the kidney of CCN2(IV)-treated mice. (a) IL-17A immunostaining in control and CCN2(IV)-treated mice, studied after 10 days, as a representative image of 5 mice per group, analyzed in independent experiments. Arrows mark positive IL-17A-expressing cells ($\times 400$ magnification).

cytokines was observed in CCN2(IV)-injected mice (Figure 2a). Importantly, only IL-17A remained elevated at 10 and 15 days of CCN2(IV) administration (Figure 2a), as

confirmed by real-time PCR (Figure 2b). These findings show that CCN2(IV) promotes a dynamic Th differentiation process in the kidney. Moreover, positive IL-17A staining was found in glomerular and tubulointerstitial areas of CCN2(IV)-injected mice after 10 days (immunohistochemistry), whereas no IL-17A expression was observed in control mice kidneys (Figure 2c). Our data clearly demonstrated that CCN2(IV) *in vivo* increased renal production of IL-17A, whereas Th1/Th2/Treg-related cytokines were not changed.

In the present study, at all time points evaluated (5, 10, and 15 days), CCN2(IV)-treated mice had a significant increase in the number of F4/80⁺ cells (that mainly recognizes monocytes/macrophages) and CD3⁺ and CD4⁺ T lymphocytes, compared with controls (Figures 3a and b). To detect Th17 cells, double immunostaining using antibodies against IL-17A and CD4 was performed (Figure 3c). After 10 days of CCN2(IV) administration, IL-17A⁺/CD4⁺ cells were observed, demonstrating the presence of Th17 cells in the kidney in response to CCN2(IV). However, some IL17A⁺/CD4⁻ cells were also found, suggesting that other cells besides Th17, probably tubular epithelial cells as described previously,⁴¹ can produce IL-17A.

The regulation of inflammatory cell infiltration in the kidney is controlled by the local production of chemokines such as MCP-1 and RANTES, the main factors involved in the recruitment of macrophages and T lymphocytes, respectively.⁴² Among the intracellular signaling systems involved in the pathogenesis of renal inflammation, the activation of the NF- κ B pathway has special interest.⁴³ In CCN2(IV)-treated mice, elevated renal mRNA and protein levels of MCP-1 and RANTES were found at all time points studied (real-time PCR and ELISA; Figures 4a and b). Moreover, in renal nuclear protein extracts of CCN2(IV)-treated mice, elevated levels of p65 NF- κ B subunit were observed as compared with controls (Figure 4c), demonstrating renal NF- κ B activation.

In contrast, CCN2(IV) administration in mice did not induce any significant change in renal function or induction of proteinuria as compared with controls (data not shown). The evaluation of profibrotic-related genes showed a transient upregulation of α -SMA (a marker of activated fibroblasts) and some extracellular matrix proteins, including fibronectin and procollagen, after 5 days of CCN2(IV) injection, which were downregulated thereafter (Figure 5). Moreover, there were no changes in renal morphology and collagen content (determined by Masson Trichrome, not shown). Our experimental data show that CCN2(IV) *in vivo* induced a sustained inflammatory response in the kidney, characterized by local production of IL-17A, but not fibrosis or kidney dysfunction.

IL-17A Blockade Inhibited CCN2(IV)-Induced Renal Inflammatory Response

To identify the role of IL-17A in the inflammatory response caused by CCN2(IV) at 10 days, a neutralizing IL-17A

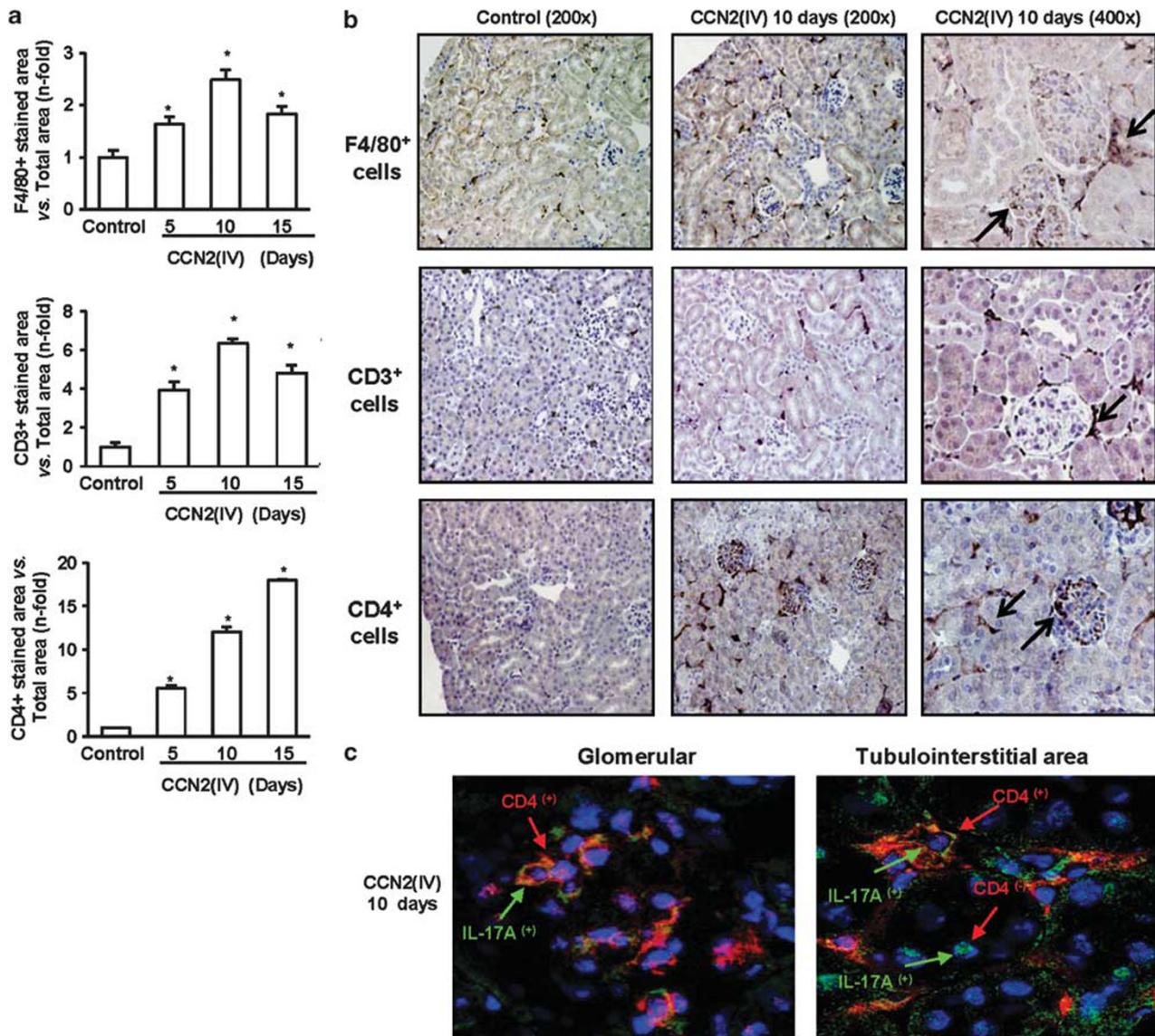


Figure 3 CCN2(IV) *in vivo* caused a sustained inflammatory cell infiltration in the kidney. C57BL/6 mice received a single i.p. injection of 2.5 ng/g body weight recombinant CCN2(IV) or vehicle (saline) and were killed after 5, 10, or 15 days ($n=5, 10,$ and 5 mice per group, respectively) compared with control group ($n=10$). The inflammatory cell infiltration was characterized in paraffin-embedded renal sections by immunohistochemistry with anti-F4/80 (marker mainly of monocytes/macrophages), anti-CD3 (lymphocytes T marker), and anti-CD4 (effectors lymphocytes T marker) antibodies. (a) The immunohistochemistry quantification expressed as mean \pm s.e.m. of fold change as compared with controls. $*P<0.01$ vs control group. (b) A representative animal from each group ($\times 200$ magnifications). Arrows indicate infiltrating cells in detail ($\times 400$ magnifications). (c) Confocal analysis of CCN2(IV)-treated renal samples showing double immunostaining of IL-17A⁺/CD4⁺ cells (Th17 cells), both in glomeruli and tubulointerstitial areas. Some IL-17A⁺/CD4⁻ cells were observed in tubulointerstitial areas. Nuclei are in blue (DAPI staining; $\times 600$ magnification).

antibody was used. In CCN2(IV)-injected mice, IL-17A neutralization markedly decreased the number of infiltrating immune cells (macrophages and CD3⁺ and CD4⁺ T lymphocytes) compared with IgG-treated mice (Figures 6a and b). Blockade of IL-17A significantly diminished renal CCN2(IV)-induced overexpression of MCP-1, RANTES, MCP-1 receptor (CCR2), and CXCL1 (Figure 6c). These data clearly demonstrate the involvement of IL-17A in CCN2(IV)-mediated renal inflammation.

CCN2(IV) Upregulated Th17-Related Factors in the Kidney

Next, we explored whether CCN2(IV) regulates the upstream mechanisms involved in Th17 cell activation in the kidney. In CCN2(IV)-injected mice, renal IL-6 and ROR γ t protein levels were significantly increased compared with control mice (Figures 7a and b). Activation of STAT3, evaluated by increased levels of phosphorylated STAT3, was observed at 10 days after CCN2(IV) administration (Figure 7b).

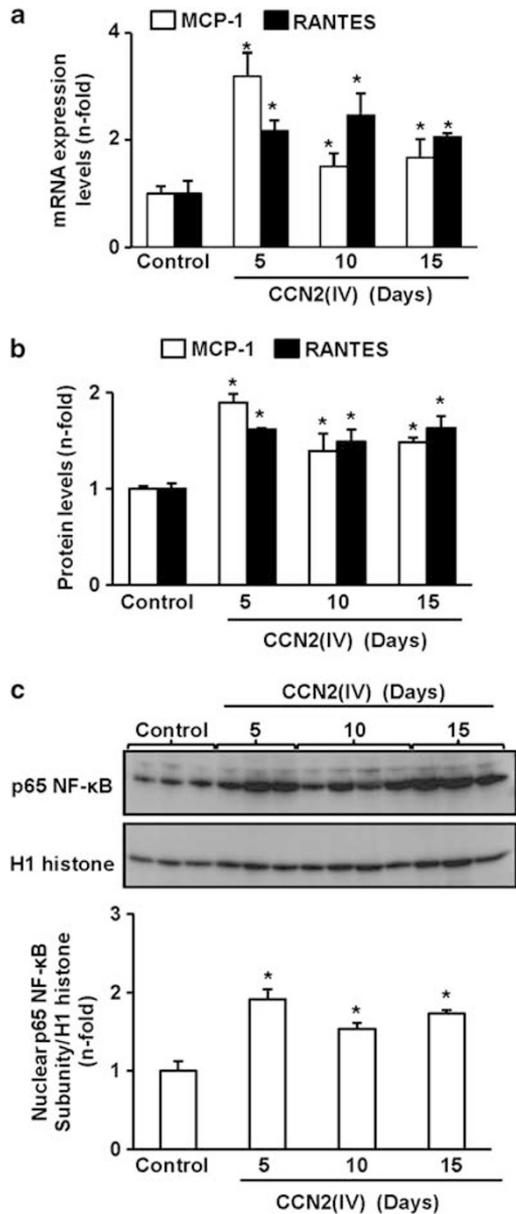


Figure 4 CCN2(IV) increased proinflammatory chemokine expression and activated NF- κ B in the kidney. In renal samples from control and CCN2(IV)-injected mice for 5, 10, and 15 days, gene expression of MCP-1 and RANTES was evaluated by real-time PCR (a) and protein levels by ELISA (b). Activation of renal NF- κ B was evaluated determining the nuclear levels of p65 NF- κ B subunit by western blot (c). Data are expressed as *n*-fold increase over control as mean \pm s.e.m. of 5–10 animals per group. **P* < 0.05 vs control.

We have further evaluated whether CCN2(IV) could regulate Th17-related factors in cultured renal cells. Previous studies have demonstrated that CCN2(IV) increased IL-16 production in renal cells.²¹ In cultured renal cells, incubation with CCN2(IV) increased STAT3 phosphorylation, with a maximal response after 10 min of stimulation (Figure 7c).

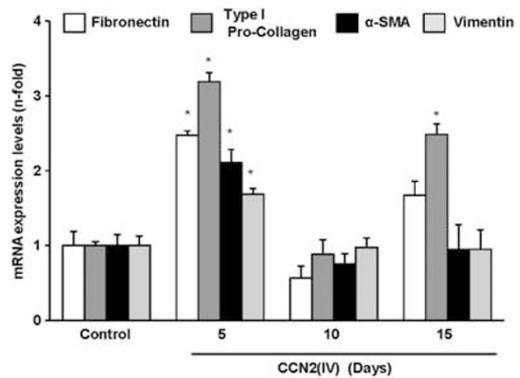


Figure 5 CCN2(IV) induced a transient upregulation of matrix-related proteins in the kidney. In renal samples from control and CCN2(IV)-injected C57BL/6 mice for 5, 10, and 15 days, gene expression of fibronectin, type I collagen, α -SMA, and vimentin was evaluated by real-time PCR. Data are expressed as *n*-fold increase over control as mean \pm s.e.m. of 5–10 animals per group. **P* < 0.05 vs control.

CCN2(IV) did not Regulate Treg-Related Transcription Factors in the Kidney

Th17/Treg differentiation from naive CD4⁺ cells is a process closely regulated that depends on cytokine environment and activation of specific transcription factors. Treg cell differentiation is regulated through the activation of Foxp3.⁴⁴ In CCN2(IV)-injected mice, Foxp3 renal levels were not elevated compared with controls at all times studied (Figure 8a). Moreover, frequency of the CD4⁺/Foxp3⁺ Treg circulating cells, assessed by flow cytometry, were not modified in response to CCN2(IV) administration (Figure 8b).

Elevated Renal Levels of IL-17A in Nonimmune Experimental Models of Renal Damage Characterized by CCN2 Overexpression

Previous studies have shown the involvement of Th17 cells in nonimmune inflammatory experimental renal injury, such as the model of UUO.⁴⁵ In this model we have previously described elevated renal levels of CCN2 in the obstructed kidneys.³⁷ Now, we have observed that obstructed kidneys also presented elevated levels of IL-17A as compared with controls (Figure 9), showing an association between CCN2 and IL-17A upregulation in experimental renal damage.

DISCUSSION

Our *in vitro* and *in vivo* studies support the novel concept of CCN2(IV) as a proinflammatory cytokine that is able to modulate the Th17 immune response. Our findings expand the known profibrogenic CCN2 signaling pathways to a true cytokine with an active role in sustained renal inflammation.

Our studies carried out on human CD4⁺ T lymphocytes showed that CCN2(IV) drives Th17 differentiation. The differentiation of naive CD4⁺ T cells into different Th effector subsets is a process closely regulated by specific cytokines and transcription factors.^{30,31,39,40} Stimulation of

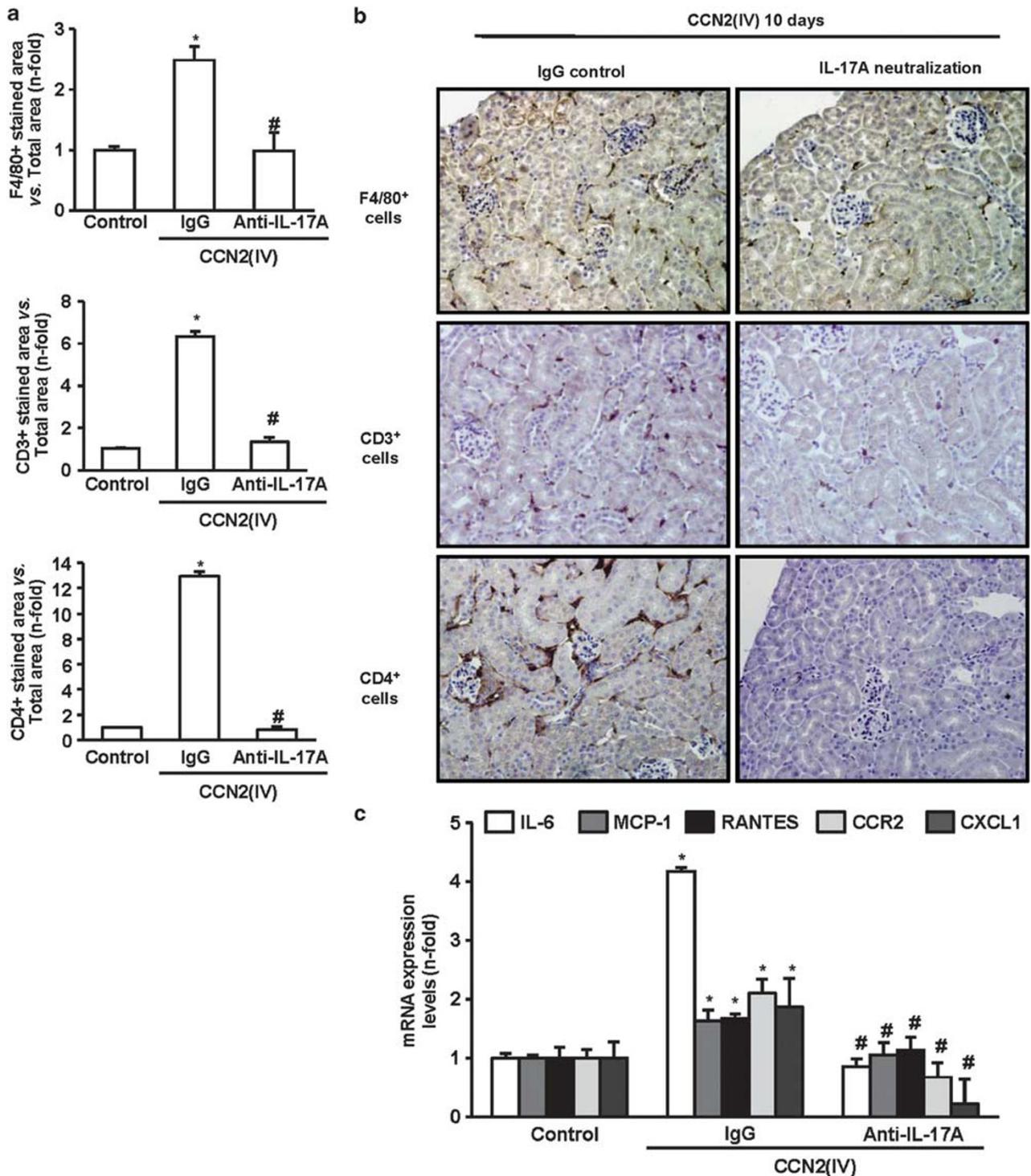


Figure 6 IL-17A blockade inhibited CCN2(IV)-induced renal inflammatory responses. To block IL-17A responses, mice were treated with an IL-17A neutralizing antibody or its corresponding IgG isotype as control ($n = 5$ mice per group) 24 h before CCN2(IV) injection and every 72 h thereafter until the time of killing at day 10. IL-17A neutralization diminished renal inflammatory cell infiltration in CCN2(IV)-injected mice. **(a)** Quantification of F4/80-, CD3-, or CD4-positive cells, expressed as mean \pm s.e.m. fold change of positive staining vs total area over control. **(b)** Representative immunohistochemistry from each group ($\times 200$ magnification). **(c)** IL-17A neutralization decreased CCN2(IV)-induced renal chemokine expression. IL-6, MCP-1, RANTES, CCR2, and CXCL1 mRNA expression as mean \pm s.e.m. analyzed by real-time PCR. * $P < 0.05$ vs control; # $P < 0.05$ vs CCN2(IV)-IgG.

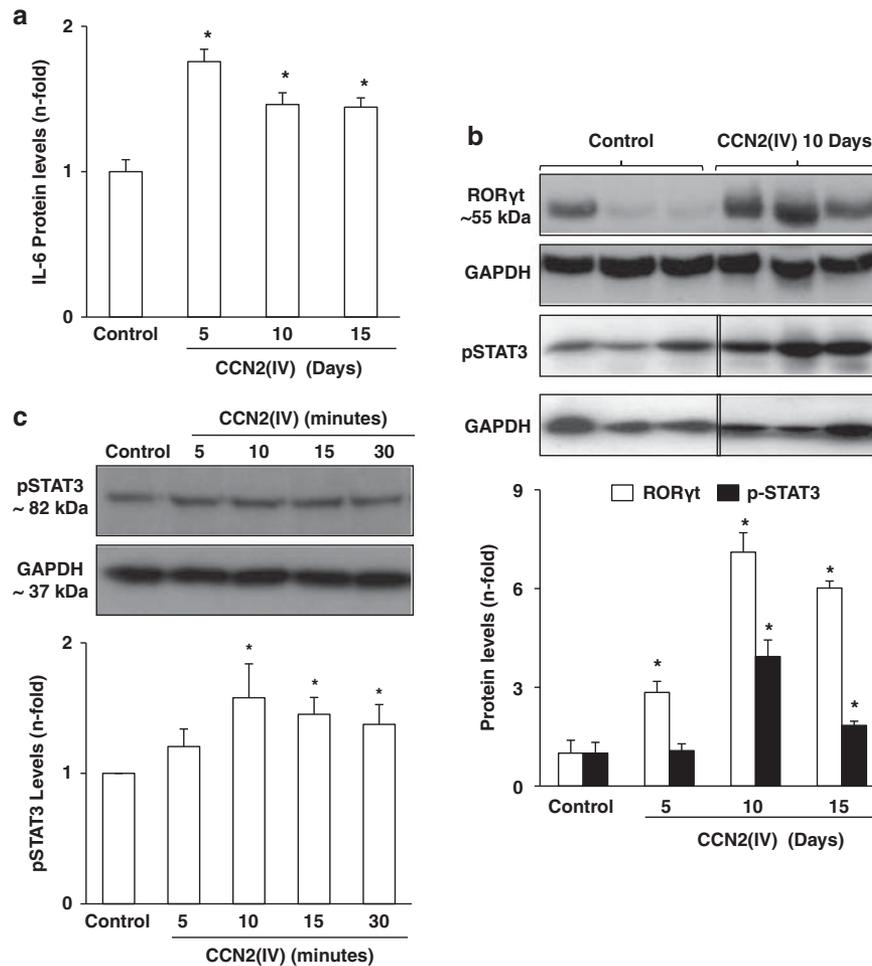


Figure 7 CCN2(IV) activated Th17-related factors in mice kidney. Renal protein levels of control and CCN2(IV)-injected mice for 5, 10, and 15 days were isolated. (a) Renal IL-6 levels evaluated by ELISA. The renal levels of Th17-related transcription factors ROR γ t and phosphorylated STAT3 (p-STAT3) were analyzed by western blot. (b) Quantification of protein levels of 5 to 10 mice per group, and a representative western blot of control and CCN2(IV)-treated mice killed at day 10. Data are expressed as mean \pm s.e.m. as fold change over control. * $P < 0.05$ vs control. (c) Mice tubular epithelial cells were incubated with 10 ng/ml CCN2(IV) until 30 min and the phosphorylated levels of STAT-3 were evaluated by western blot. (c) A representative experiment of four experiments performed is shown, and data are expressed mean \pm s.e.m. * $P < 0.05$ vs control.

isolated human naive CD4⁺ T cells with CCN2(IV) induced a Th17 cell polarization, characterized by increased IL-17A⁺/CD4⁺ T cells, similar to that observed with IL-6, a known inducer of Th17 differentiation. Th1 cell differentiation is controlled by T-bet and STAT4, Th2 is GATA-3/STAT6 dependent, Treg is Foxp-3 mediated, and Th17 depends on STAT3 activation and subsequent ROR γ t induction.^{38–40} The importance of Th17-related factors have been demonstrated using knockout mice, where STAT3-null cells fail to express ROR γ t,³⁹ and lack of ROR γ t results in profound Th17 deficiency.⁴⁰ *In vitro* CCN2(IV) activated STAT-3 in isolated human naive CD4⁺ T cells and in cultured renal cells. *In vivo* CCN2(IV) upregulated Th17-related factors in the kidney, including IL-6 production and activation of STAT3 and ROR γ t, demonstrating that this molecular mechanism is operating in CCN2(IV)-induced renal damage and it could underlie the observed activation of the local Th17 response.

In contrast, no changes in renal levels of Th1/Th2 cytokines or Treg-related factors such as TGF- β and Foxp-3 were found, suggesting that Th1/Th2/Treg responses were not modulated by CCN2(IV), at least in the murine kidney.

Recent studies have shown the importance of Th17 cells, and its hallmark cytokine IL-17A, in immune-mediated glomerulonephritis, including experimental antimyeloperoxidase glomerulonephritis, crescentic glomerulonephritis, and lupus nephritis.^{46,47} The direct involvement of Th17 cells in renal damage has been demonstrated by an experimental study in mice, showing that injection of Th17 cells resulted in developed albuminuria after 3 days, glomerular neutrophil infiltration, and increased renal CXCL1 mRNA.⁴⁸ In the kidney of CCN2(IV)-treated mice, infiltrating Th17 cells were detected by double CD4⁺/IL-17A⁺ staining, suggesting that Th17 cells could contribute to renal damage by CCN2(IV).

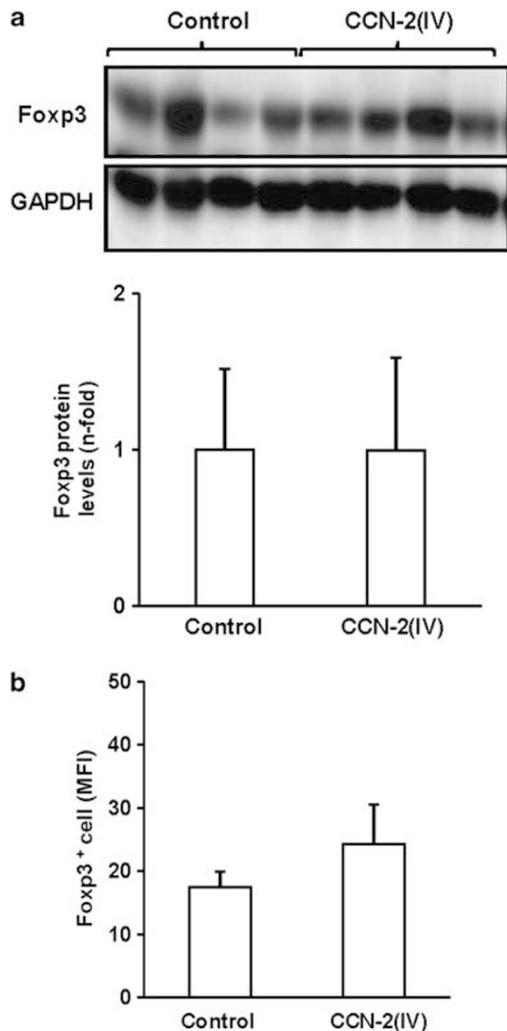


Figure 8 CCN2(IV) did not regulate Treg-related transcription factors. Foxp3 renal levels were analyzed by western blot. **(a)** Data as ratio of renal Foxp3/GAPDH protein as mean \pm s.e.m. of 10 mice per group and a representative western blot experiment. Treg (CD4⁺ Foxp3⁺) cells were analyzed in plasma by flow cytometry. **(b)** The percentage of CD4⁺ T cells expressing Foxp3 cells (mean \pm s.e.m. of 10 mice per group). * $P < 0.05$ vs control.

Besides Th17 cells, IL-17A can be produced by other cells, including CD8⁺ T cells, $\gamma\delta$ T cells, natural killer cells, neutrophils, and some resident cells.⁴⁹ In the model of ischemia-reperfusion, neutrophils, but not Th17 cells, are the main sources of IL-17A and contribute to renal injury by natural killer T activation and IL-12/IFN- γ production.⁵⁰ In glomeruli of patients with lupus nephritis, double-negative (CD4, CD8) T cells are a major source of IL-17A,⁵¹ and in renal allograft rejection, positive staining for IL-17A has been detected in tubular cells,⁴¹ as we have observed here in CCN2(IV)-treated mice, suggesting that renal cells could produce this cytokine and contribute to extend the damage.

The biological actions of IL-17A are predominantly proinflammatory.⁴⁹ There are several pathways by which this

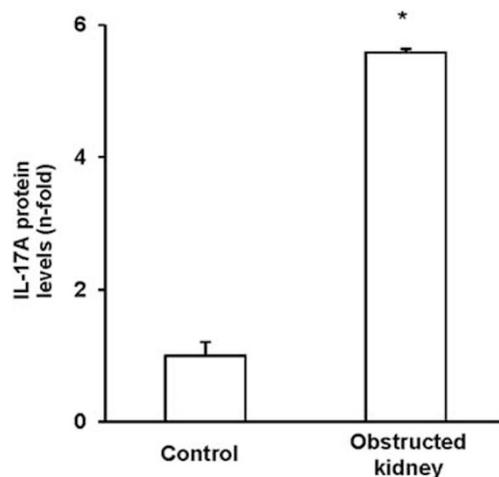


Figure 9 Upregulation of renal levels of IL-17A in the model of unilateral ureteral obstruction in mice. IL-17A renal levels were evaluated by ELISA and expressed as data as mean \pm s.e.m. of 6 animals per group. * $P < 0.05$ vs control mice.

cytokine might affect local inflammation. In tubular cells, mesangial cells, vascular smooth muscle cells, endothelial cells, and fibroblasts, IL-17A induces the release of cytokines and chemokines such as IL-16, MCP-1, RANTES, CXCL1, and CXCL8 that recruit different leukocyte subsets leading to target organ injury.⁵² IL-17A can also directly promote monocyte chemotaxis.⁵³ Our *in vitro* and *in vivo* data clearly show that CCN2(IV) induces IL-17A production. In CCN2(IV)-injected mice, renal overexpression of IL-17A was associated with the presence of inflammatory infiltrating cells and upregulation of chemokines that were reduced in IL-17A neutralization experiments, clearly demonstrating the involvement of IL-17A in CCN2(IV)-mediated renal inflammation. Studies in knockout mice have shown that the absence of IL-17A resulted in less glomerular crescent formation and improved renal function in experimental nephrotoxic nephritis.⁵⁴ Several IL-17A blockade strategies, including neutralizing antibodies or receptor gene targeting, ameliorated several experimental nonimmune diseases.^{32–36} Preliminary data in rheumatoid arthritis and uveitis,³⁴ and more recently, data from two phase 2 clinical trials, in which patients with psoriasis were treated with specific targeting antibodies for IL-17A or its receptor,^{35,36} have shown beneficial effects in humans. CCN2 has been involved in several chronic diseases, including skin disorders, cardiac hypertrophy, atherosclerosis, pulmonary hypertension, and lung and liver diseases,¹² where IL-17A is a key player.^{32,33,55–58} In the nonimmune murine model of UUU, we have observed elevated renal levels of IL-17A and infiltrating inflammatory cells associated with renal CCN2 overexpression. Although future studies are needed, our data showing that IL-17A blockade ameliorated CCN2-induced renal inflammation support the concept of

IL-17A-neutralizing antibody as a promising tool for chronic inflammatory diseases, including chronic kidney diseases.

CCN2 overexpression has been described in human fibrotic diseases, and it is produced temporally and spatially in close proximity to fibrotic areas.^{2,12} Antagonists of CCN2 have proven effective in blocking profibrogenic CCN2 signaling pathways *in vitro* and have yielded promising data with respect to preventing experimental fibrosis.¹⁴ Regarding renal diseases, knockdown of CCN2 gene expression with antisense gene transfer into rat kidney ameliorates tubulointerstitial fibrosis in obstructive nephropathy¹⁴ and in experimental mice diabetes in C57BL/6 mice.¹⁶ CCN2 is known to function downstream of TGF- β , driving extracellular matrix accumulation and fibrosis.^{2,12} Indeed, in cultured renal cells, CCN2 blockade diminished matrix proteins synthesis induced by TGF- β and Angiotensin II.⁵⁹ Although initial studies in fibroblasts, and other cultured cells, showed that recombinant CCN2, both the C-terminal and the full molecule, induce extracellular matrix production,^{2,7,14} several *in vivo* studies have shown that CCN2 alone is not sufficient to cause ongoing fibrotic changes. In a model of a skin fibrosis, injection of CCN2 alone did not induce fibrosis and only the simultaneous application of TGF- β 1 caused persistent fibrosis.⁶⁰ CCN2 overexpression in rat lungs only caused a temporal induction of procollagen gene expression and transient matrix accumulation, but it is not sufficient to induce progressive fibrosis.⁶¹ Podocyte-specific CCN2-transgenic mice (in C57BL/6 background) exhibited no glomerular abnormalities, proteinuria, or matrix accumulation, although they were more susceptible to streptozotocin-induced diabetes.⁶² Our *in vivo* data showed a similar response in the kidney. Systemic CCN2(IV) administration into C57BL/6 mice increased profibrotic genes renal expression at day 5, and decreasing thereafter, and no collagen accumulation was observed.

Our findings expand the known profibrogenic CCN2 signaling pathways to a true cytokine with an active role in sustained renal inflammation mediated by Th17 cells. Different strategies for blocking CCN2 activity have proven beneficial effects in fibrotic-related pathologies, including renal diseases,¹² whereas CCN2 overexpression induces cardioprotection.^{17,18} Our results set out that IL17A neutralization block inflammatory events caused by the C-terminal module of CCN2 in the kidney, and suggest that IL-17A targeting could be a better therapeutic option than CCN2 blockers for human use in inflammatory diseases.

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DISCLOSURE/CONFLICT OF INTEREST

The authors declare no conflict of interest.

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