

Circumdatin I, a New Ultraviolet-A Protecting Benzodiazepine Alkaloid from a Marine Isolate of the Fungus *Exophiala*

Dahai Zhang, Xiudong Yang, Jung Sook Kang, Hong Dae Choi, Byeng Wha Son

Received: September 21, 2007 / Accepted: January 7, 2008 © Japan Antibiotics Research Association

Abstract A new ultraviolet-A (UV-A) protecting benzodiazepine alkaloid, circumdatin I (1), along with the previously described circumdatin C (2) and circumdatin G (3), have been isolated from the mycelium of a marinederived fungus of the genus *Exophiala*. The structures of the three circumdatins were assigned on the basis of physicochemical evidence. The absolute stereochemistry of 1 was determined by comparison of optical rotation and CD experiments with those of 2 and 3.

Keywords marine-derived fungus, *Exophiala*, circumdatin, benzodiazepine alkaloid, ultraviolet-A (UV-A)

Marine-derived microorganisms such as bacteria, fungi, and cyanobacteria have proven to be a rich source of new biologically active secondary metabolites [1]. As part of our search for bioactive substances from marine-derived microorganism [2], the marine sponge-associated fungus was studied because the mycelium extract showed potent UV-A protecting activity. An assay-guided purification resulted in isolation of a new benzodiazepine alkaloid, circumdatin I (1), and two known circumdatins C (2) [3] and G (3) [4] from the marine isolate of fungus *Exophiala* sp. We report here on the isolation and structural elucidation of compounds $1\sim3$.

Fungal Isolation and Culture

The fungal strain was isolated from the surface of the marine sponge *Halichondria panicea* collected at Bogil Island, Jeonnam Province, Korea, in 2006 and identified as an *Exophiala* sp. (Family: Herpotrichiellaceae) on the basis of morphological evaluation and fatty acid methyl ester analysis (Korean Culture Center of Microorganism, Seoul, Korea, similarity index of 0.828). A voucher specimen is deposited at Pukyong National University with the code MFC353-1. The fungus was cultured (10 liters) for 3 weeks (static) at 29°C in SWS medium containing of soytone 0.1% and soluble starch 1.0% in seawater.

Extraction and Isolation

The cultured broth was filtered with through cheesecloth to give a mycelium cake. The freeze-dried mycelium cake was extracted with CH_2Cl_2 -MeOH (1:1) to afford crude extract (1.8 g), which was subjected to silica gel flash chromatography. Elution was performed with *n*-hexane-EtOAc (stepwise, $0 \sim 100\%$ EtOAc) to yield four fractions. Fractions 3 and 4, which were active in UV-A protecting assay, were separated by medium-pressure liquid chromatography (MPLC) (ODS) using a H₂O - MeOH gradient elution to afford crude compounds 1, 2, and 3, respectively. These were further purified by HPLC (YMC, ODS-A, 10×250 mm, 1.0 ml/minute) utilizing a 30 minutes gradient program of $50 \sim 100\%$ MeOH in H₂O to furnish 1 (6.5 mg), 2 (6.0 mg), and 3 (2.0 mg), respectively.

H. D. Choi: Department of Chemistry, Dongeui University, Busan 614-714, Korea

B. W. Son (Corresponding author), **D. Zhang, X. Yang:** Department of Chemistry, Pukyong National University, Busan 608-737, Korea, E-mail: sonbw@pknu.ac.kr

J. S. Kang: College of Dentistry, Pusan National University, Busan 602-739, Korea

1: a colorless solid; $[\alpha]_{D}^{20} = -236^{\circ}$ (*c* 0.2, MeOH); CD (MeOH) nm ($\Delta \varepsilon$) 302 (-2.2), 296 (-2.4), 254 (+7.5), 236 (-16.8), 214 (+29.7); UV (MeOH) λ_{max} nm (log ε) 210 (7.4), 240 (7.5), 283 (6.9), 329 (6.9), 340 (sh) (6.8); IR (KBr) v_{max} 3435, 1654, 1615 cm⁻¹; LR-EI-MS *m/z* 323 [M]⁺ (100), 281 (32), 252 (20), 224 (7), 189 (10), 162 (29), 121 (69), 106 (28), 92 (19), 63 (66). HR-EI-MS *m/z* 323.0907 [M]⁺ (calcd for C₁₇H₁₃N₃O₄, 323.0906). See Table 1 for NMR spectral data.

2 and **3** were isolated as a colorless solid, and showed spectral data virtually identical to those reported in the literature [3, 4].

Ultraviolet-A Protecting Assay

Samples to be tested were dissolved in MeOH, and the solution (200 μ l) was dispensed into wells of a 96-well microtiter tray. The absorbance of the sample solution was measured at 340 nm with microplate reader (Packard Co., Spectra CountTM). The ultraviolet-A protecting activity was expressed as ED₅₀, which is the concentration of the tested compound required to give a 50% increase of the absorbance from that of the blank solution [MeOH (200 μ l)]. ED₅₀'s were determined by linear regression of data plotted on a semi-log scale.

Structural Elucidation and Bioactivity

1 was isolated as a colorless solid which yielded a molecular formula of $C_{17}H_{13}N_3O_4$ by HR-EI-MS and ¹³C-NMR methods. The IR spectrum of **1** exhibited bands characteristic for hydroxyl (3435 cm⁻¹) and amide (1654 cm⁻¹) functionalities.

The ¹H- and ¹³C-NMR data for benzodiazepine **1**, including the results from DEPT, COSY and TOCSY experiments, showed the presence of 1,2,4-trisubstituted benzene, 1,2,3-trisubstituted benzene, 1,1-disubstituted ethane, and two amides groups (Table 1, Fig. 1).

Detailed analyses of the HMQC and HMBC spectra of **1** suggest the presence of 1,2,2-trisubstituted 3-methyl-7-hydroxy-1,4-benzodiazepine and 2,3-disubstituted 8-hydroxyquinazolin-4-one. The presence of 2,3-disubstituted 8-hydroxyquinazolin-4-one was further supported by UV spectral data [329 nm (log ε 6.9), 340 (sh) (6.8)].

The connectivities and assignments of the functional groups for 1, which led to the planar structure for this metabolite, were made by interpretation of HMBC NMR data (Fig. 1) and by the comparison of the NMR data with those of 2 [3] and 3 [4].

The NMR data of $1 \sim 3$ showed similar patterns, except

 Table 1
 NMR spectral data for circumdatin I (1)^a

Position	$\delta_{_{ m H}}$ (mult., J)	$\delta_{ m C}$ (mult.)
1	8.69 (d, 6.0)	
2		166.6 (s)
3		132.4 (s)
4	7.10 (d, 3.2)	114.1 (d)
5		157.2 (s)
6	7.00 (dd, 9.0, 3.2)	117.8 (d)
7	7.38 (d, 9.0)	130.0 (d)
8		124.5 (s)
10		161.2 (s)
11		121.8 (s)
12	7.58 (dd, 8.0, 1.0)	116.5 (d)
13	7.37 (dd, 8.0, 8.0)	127.8 (d)
14	7.27 (dd, 8.0, 1.0)	119.1 (d)
15		152.8 (s)
16		134.8 (s)
18		155.1 (s)
19	4.31 (dq, 6.5, 6.0)	49.6 (d)
20	1.56 (d, 6.5)	14.9 (q)

^a Recorded in DMSO-d₆ at 400 MHz (¹H) and 100 MHz (¹³C).



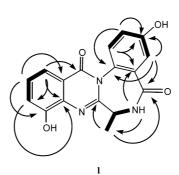


Fig. 1 Chemical structures of circumdatins I (1), C (2), and G (3). Structure of 1 elucidated by ¹H-¹H COSY (→) and HMBC (→) correlations.

for the appearance of a new oxygenated sp^2 -quaternary carbon in place of one sp^2 -methine of **2** [δ 7.76 (H-15), 128.8 (C-15)] and **3** [δ 7.57 (H-5), 128.6 (C-5)], respectively (Table 1).

Thus, **1** was characterized as 5,15-dihydroxycircumdatin F [5] on the basis of direct comparison of NMR data of **1** with those of **2** and **3**. The absolute stereochemistry of **1** was investigated using CD. The CD spectrum (MeOH) of **1** showed the following Cotton effects at 302 nm ($\Delta\varepsilon$, -2.2), 254 (+7.5), 236 (-16.8), 214 (+29.7), which were very similar to those of **2** [CD (MeOH): 307 nm ($\Delta\varepsilon$, -4.3), 263 (+8), 233 (-22) and 213 (+29)] [3]. Thus, the absolute configuration of asymmetric center for **1** was determined to be 19(*S*). This conclusion was further supported by the comparison of the optical rotations among **1**~3. The value of specific rotation of **1** ($[\alpha]_{D}^{20} - 236^{\circ}$) was in negative, the same phase as those of **2** ($[\alpha]_{D}^{20} - 75^{\circ}$) [3] and **3** ($[\alpha]_{D}^{20} - 221^{\circ}$) [4], implying that both compounds shared the same configuration.

2 and **3** were identified as the known compounds, circumdatins C and G, respectively, by comparison of their spectroscopic data to the published data [3, 4].

1∼3 were evaluated for UV-A protecting activity, and they exhibited an UV-A protecting activity with ED_{50} values of 98, 101, and 105 µM, respectively, which are more potent than the positive control, oxybenzone (ED_{50} , 350 µM), a currently used sunscreen agent.

Benzodiazepine alkaloids, for example, circumdatins A~H $[3 \sim 6]$. epi-aszonalenins A~C [7], benzodiazepinedione [8], and asperlicins [9], were widespread microbial products commonly found in nutrient rich cultures of both terrestrial and marine fungi, and exhibited interesting biological activities, treatment of gastrointestinal and CNS disorder [5, 9], inhibition of mitochondrial NADH oxidase [6], and psychoactive properties [7]. Circumdatins A~H have been previously isolated from Aspergillus ochraceus and have been suggested as good chemotaxonomic markers for this species [3~6].

Acknowledgements This work was supported by a grant from MarineBio21, Ministry of Maritime Affairs and Fisheries, Korea. Mass and CD spectral data were kindly provided by the Korea Basic Science Institute.

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