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ORIGINAL ARTICLE

No evidence for attenuated stress-induced extrastriatal dopamine signaling in psychotic disorder

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Stress is an important risk factor in the etiology of psychotic disorder. Preclinical work has shown that stress primarily increases dopamine (DA) transmission in the frontal cortex. Given that DA-mediated hypofrontality is hypothesized to be a cardinal feature of psychotic disorder, stress-related extrastriatal DA release may be altered in psychotic disorder. Here we quantified for the first time stress-induced extrastriatal DA release and the spatial extent of extrastriatal DA release in individuals with non-affective psychotic disorder (NAPD). Twelve healthy volunteers (HV) and 12 matched drug-free NAPD patients underwent a single infusion [¹⁸F] fallypride positron emission tomography scan during which they completed the control and stress condition of the Montreal Imaging Stress Task. HV and NAPD did not differ in stress-induced [¹⁸F]fallypride displacement and the spatial extent of stress-induced [¹⁸F]fallypride displacement in medial prefrontal cortex (mPFC) and temporal cortex (TC). In the whole sample, the spatial extent of stress-induced subjective stress. Psychotic symptoms during the scan or negative, positive and general subscales of the Positive and Negative Syndrome Scale were not associated with stress-induced [¹⁸F]fallypride displacement nor the spatial extent of stress-induced [¹⁸F]fallypride displacement in NAPD. Our results do not offer evidence for altered stress-induced extrastriatal DA signaling in NAPD, nor altered functional relevance. The implications of these findings for the role of the DA system in NAPD and stress processing are discussed.

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INTRODUCTION

In the last decade, significant progress has been made in understanding the role of the dopamine (DA) system in the human stress response.^{1–3} Evidence has emerged showing that, at least in part, the stress response is facilitated by DA release in the striatum^{1,2,4} and prefrontal cortex (PFC).^{5,6} Dopaminergic (DAergic) involvement in the stress response is particularly relevant for psychiatric disorders such as psychotic disorder,⁷ as evidence suggests that stress has an important role in the onset of psychotic symptoms^{8,9} and DAergic abnormalities are a hallmark feature of psychotic disorder.¹⁰ Investigating stress-related DAergic activity in the context of psychotic disorder could thus provide new insights into the pathogenesis of the disorder.

Stress-induced DAergic activity in humans has been studied *in vivo* with positron emission tomography (PET), hinging on competition between radioligand binding and endogenous DA release.¹¹ In these studies, DA release was assessed during a psychosocial evaluation paradigm² (for metabolic stress, see Adler *et al.*¹², Brunelin *et al.*¹³). Although psychosocial stress in healthy volunteers (HV) produced modest and variable changes in striatal DA release, ^{1,2,4,14} the same stressor reliably increased DA release in the (associative) striatum of individuals across the psychosis continuum.^{1,2,4} Importantly, this suggests that the putative

association between stress and psychotic disorder may be moderated by the DA system.

Preclinical work, however, has revealed that short-lived stressors consistently and primarily increase DAergic activity in the PFC analog of the rodent. ^{15,16} Moreover, selective destruction of frontal DA neurons increases stress-related DA transmission in mesolimbic regions, ^{16,17} hinting at a key regulatory role for PFC DA transmission in the stress response. Because DA-mediated hypofrontality is hypothesized to be a cardinal feature of psychotic disorder, ^{18,19} this preclinical work indirectly suggests that the well-documented link between stress and psychotic disorder^{20,21} may be underlain by cortical DA function. More specifically, decreased cortical DA function may constitute a neurochemical feature of vulnerability to psychotic disorder and underlie increased behavioral stress sensitivity. ²¹

In the only two human studies currently available, psychosocial stress in HV increased medial PFC (mPFC) DA release⁶ and increased the area (that is, spatial extent) of mPFC DA release⁵ assessed with high-affinity $D_{2/3}$ binding ligand [18 F]fallypride. 22 In an add-on sample of first-degree relatives of patients with psychotic disorder, Lataster *et al.* 3 showed that the spatial extent of stress-induced mPFC DA release decreased as a function of increased subjective stress. Although this latter finding hints at stress-related DA-mediated hypofrontality in the psychosis

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Table 1. Sample demographics							
	HV	NAPD	Statistics (P-value, test statistic)				
Gender			(1, 0 ^a)				
Male	8	8					
Female	4	4					
Age	48.08 (9.94)	44.67 (11.24)	$(0.44, -0.79^{b})$				
Education ^c	5.83 (1.4)	5.33 (1.44)	(0.4, 0.86 ^a)				
Smoking			(0.38, 0.54 ^a)				
Nonsmoker	11	10					
Smoker	1	2					
Cannabis lifetime ^d	0.23 (0.83)	0.67 (1.23)	(0.31, 1.04)				
Other drugs lifetime ^{d,e}	0 (0)	0.01 (0.04)	(0.31, 1.04)				
Injected radioligand (MBq)	189.83 (8.2)	187.92 (10.86)	$(0.4, -0.85^{b})$				
Specific activity (GBq)	2611.42 (872.96)	2146.25 (1198.6)	(0.98, -0.03 ^b)				
Current symptoms ^f	_	11.83 (3.93)	_				
Years off AP	_	7.09 (4.96)	_				
Cumulative haloperidol equivalents ⁹	_	4303.07 (12 280.64)	_				

Abbreviations: AP, antipsychotics; HV, healthy volunteer; NAPD, non-affective psychotic disorder. ^aChi² test. ^bT-test. ^cHighest finished education, scored on a scale ranging from 1 (primary school) to 8 (Masters degree). ^dLifetime use scored on a scale ranging from 1 (one to five times) to 8 (> 100 times). ^eStimulants, sedatives, opiates, cocaine, psychedelics, XTC, MDMA, PCP and inhalants subscales. ^fPositive subscale of the Positive and Negative Syndrome Subscale (PANSS). ^gCumulative haloperidol equivalents were calculated by converting the weekly antipsychotics dose to haloperidol equivalents and multiplying it by the number of weeks the antipsychotics were taken.

continuum, investigating stress-induced PFC DAergic activity in established psychotic disorder could further elucidate the role of this mechanism in the pathogenesis of the illness. To these aims, we investigated the effect of psychosocial stress on extrastriatal DA signaling in a sample of HV and medication-free individuals with a diagnosis of non-affective psychotic disorder (NAPD) using [18F]fallypride PET.

However, measuring extrastriatal DAergic activity remains methodologically challenging; the density of extrastriatal D₂ receptors is 2–8% compared with the striatum.²³ Radioligands with suboptimal affinity and selectivity to investigate DAergic activity in extrastriatal areas may yield low signal-to-noise ratio, thus limiting quantification. 11,24 Although [18F]fallypride has been used to quantify DA release in cortical regions due to its high affinity and specificity, the effects of amphetamine on extrastriatal DA release quantified using [¹⁸F]fallypride have not been uniformly consistent.^{25–29} While this has been attributed to the radioligand's inherent signal-to-noise ratio, 26,28 within-subject variation introduced by two-day scanning protocols, with control and experimental scan on separate days, may also constitute a source of measurement error, particularly in the context of subtle changes in neurotransmitter activity. To minimize within-subject variation, we utilized a validated single infusion [18F]fallypride paradigm, which circumvents subtraction and yields a model fit approach.3,30,31

In the current study, medication-free NAPD and HV underwent a well-validated experimental psychosocial stress paradigm, the Montreal Imaging Stress Task (MIST).² All the participants completed a MIST control and stress condition in a single [18F] fallypride session. Subjective stress responses, psychotic symptoms and plasma cortisol levels were assessed throughout each condition. Conform previous work, we first investigated stressinduced [18F]fallypride displacement and the spatial extent of stress-induced [18F]fallypride displacement in mPFC,5,6 after which we explored other extrastriatal regions. It was expected that both outcome parameters of DA signaling would be positively associated with the subjective stress response in HV. Consistent with the notion of DA-mediated hypofrontality, we expected that NAPD would show less stress-induced extrastriatal [18F]fallypride displacement and a decrease in the spatial extent of stressinduced extrastriatal [18F]fallypride displacement, compared with HV.

MATERIALS AND METHODS

Sample

The sample consisted of 12 HV (unrelated to Lataster $et~al.^5$) and 12 NAPD matched on age, gender and education (Table 1). All NAPD were diagnosed with a non-affective psychotic disorder (Supplementary Table 1). Four included NAPD were antipsychotics naive. Except for one NAPD, the remaining group was treated with antipsychotics for < 2 years. At the time of scanning, NAPD were off antipsychotics for at least 1 year (Table 1), were not exposed to mood stabilizers, were off antidepressants (total n=5) for longer than 1 year and did not take benzodiazepines on the day of the scan (Supplementary Table 1). NAPD showed relatively low acute psychotic symptom scores (Table 1), but did not meet the criteria for remission according to the Positive and Negative Syndrome Scale (PANSS) criteria (less than a score of 3 on all relevant items according to van Os $et~al.^{32}$). HV were matched to NAPD with a past of minimal illicit drug use (Table 1).

Participants were recruited through regional and national media and, additionally, NAPD were recruited through local mental health services. The RWTH Aachen University ethics committee approved the study. PET approval was granted by the national authority for radiation protection in humans in Germany (Bundesamt für Strahlenschutz, BfS). Written informed consent was obtained before participation. Inclusion criteria independent of group: (i) age 18-60 years (ii) able to provide informed consent. Exclusion criteria independent of group: (i) current/past use of illicit drugs according to the Composite International Diagnostic Interview (World Health Organization, 1990) (lifetime: >15 times cannabis, >5 times other drugs; illicit drug use in the past year), (ii) foreign bodies precluding a magnetic resonance imaging (MRI) scan, (iii) neurological disease, (iv) pregnancy. NAPD-specific inclusion criterion: diagnosis of non-affective psychotic disorder according to the Diagnostic and Statistical Manual of Mental Disorders (DSM-IV) criteria. HV-specific exclusion criteria: lifetime history of psychiatric illness according to DSM-IV criteria and lifetime neuroleptic use. On the day of scanning, a urine screening was performed to exclude current drug use and pregnancy.

Psychosocial stress paradigm

Psychosocial stress was induced using the MIST.² The MIST is a mental arithmetic task with an evaluative psychosocial component and has been prescribed in detail before.^{1,2,5,33} Psychosocial feedback during the MIST was scripted. All participants were exposed to identical feedback by an investigator who was previously unknown to them. Time and difficulty were automatically adjusted during the experimental condition using a computer algorithm preventing users from exceeding 60–70% correct answers. The MIST training version was practised for 15 min at least 2 h before scan. Participants completed 10 6-min blocks of MIST control and experimental version. Control and experimental sessions were separated by a break (Figure 1).



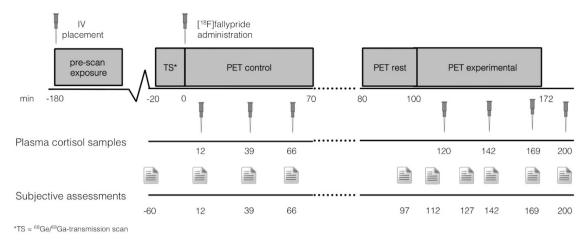


Figure 1. Graphical overview of the single infusion design. Following the transmission scan, the radioligand was injected after which participants always performed the control block of the MIST for 70 min. After a 10-min break, participants were repositioned using the scanner coordinate system and reference skin marks. At 100 min post injection, participants performed the MIST experimental condition for 70 min. Plasma cortisol samples were collected in intervals ranging from 22 to 54 min. MIST, Montreal Imaging Stress Task; PET, positron emission tomography.

Behavioral and physiological assessments

PANSS positive, negative and general symptoms³⁴ were assessed by a trained researcher before the scan. Subjective stress and psychotic symptoms were briefly assessed pre-scan (n = 1), during each PET part (n=8) and post scan (n=1) (Figure 1). Subjective stress responses were assessed using seven-point Likert Scale items: 'I feel relaxed' (reversed), 'I feel judged' and 'I do not live up to expectations', on the basis of previous work ($\alpha = 0.69$).^{5,33} Psychotic symptoms (positive only) during the scan were assessed using the following items: 'I hear voices', 'I see things' and 'I feel suspicious' (α = 0.7). Plasma cortisol samples were also collected throughout each PET part (n=6) and post scan (n=1); Figure 1). Plasma cortisol levels were determined using a radio immunoassay.³⁵

Image acquisition and analyses

MRI scan. T1-weighted MRI scans were acquired on a 1.5T Philips (Philips Medical Systems, Herrsching, Germany) machine with TE = 4.59 ms, TR = 30 ms, matrix dimensions = 256×256 , slice thickness = 2 mm, slice number = 176. During the data acquisition phase, this scanner was replaced by a Siemens 3T scanner (Siemens Healthcare, Munich, Germany). Remaining scans (37.5%) were collected using the Magnetization Prepared Rapid Acquisition Gradient-Echo sequence, with TE = 2.52 ms, TR = 1900 ms, matrix dimensions = 256 × 256, slice thickness = 1 mm, slice number = 176. A similar proportion of HV (5/12) and NAPD (4/12) MRI scans were obtained on the second machine.

Radioligand preparation. The radiosynthesis of [18F]fallypride was a highyield modification of the synthesis method for [18F]desmethoxyfallypride, described in detail previously. 36,37

PET acquisition. All PET measurements were performed in the supine position in a quiet environment. Head position was fixed using a vacuum plastic mould to limit the head movement.³⁸ The scans were performed in three-dimensional mode on a Siemens ECAT EXACT HR+ scanner (Siemens-CTY, Knoxville, TN, USA). Sixty-three slices of 2.425 mm slice thickness (pixel $size = 2 \text{ mm} \times 2 \text{ mm}$) were reconstructed per time frame by filtered back projection (Hamm filter) after Fourier rebinning into two-dimensional sinograms. Data sets were corrected for random coincidences, scatter radiation and attenuation (10 min ⁶⁸Ge/⁶⁸Ga-transmission scan). The image matrix was 128×128. The PET data were smoothed (4 mm FWHM), realigned, co-registered (transformation matrix based on first 10 realigned frames) (PMOD v3.1 (PMOD Technologies, Zurich, Switzerland)) and normalized (SPM 8, Wellcome Trust, London, UK). For every participant, an attenuation-corrected average image of the first 15 min was created. These frames were chosen because of their minimal amount of movement and subsequent high signal-to-noise ratio.³⁹ The remaining frames were realigned to the 15-min mean image using squared difference sum (dissimilarity function) and trilinear interpolation as rigid matching settings in PMOD v3.1 and inspected frame by frame. To quantify the remaining

discrepancy between mean frame and other frames, individual data sets X, Y, Z and pitch, roll, yaw parameters were exported from SPM 8 (realign option with trilinear interpolation). HV and NAPD did not differ in movement parameters (data upon request) and total sample movement parameters were low (X, Y, Z movement all < 5 mm and pitch, roll, yaw all < 5°).

Data were collected in two segments, a control and experimental part, in a single session with single bolus administration.^{3,33} The PET acquisition protocol is visualized in Figure 1. Dynamic frames were collected every 60 s for the first 6 min, after which they were collected every 120 s for the remainder of the emission scan, in accordance with previous work.³ Break frames typically consisted of frame 39-42 and were discarded before preprocessing.

PET analysis. Time-activity curves were obtained for the cerebellum (reference region) and temporal and frontal regions. Two masks were created: one containing cerebellum only and another containing all regions (results section). Regions were based on Brodmann definitions, identical to previous work. 5,33 Using the Automated Anatomical Labeling mask provided by PMOD v3.1, hippocampus and amygdala were located for all participants. Using the PMOD v3.1 crop and tailor functions, hippocampus and amygdala were drawn and inspected slice by slice to ensure mask coverage. All masks were custom-tailored to the individual's MRI, transferred to co-registered PET data in PMOD v3.1 and visually inspected for fit by two independent raters. Given that striatal and extrastriatal regions differ in time to reach pseudo equilibrium, stress-induced [18F]fallypride displacement in striatal regions was not investigated; these values could not be reliably investigated with the current design, which was optimized to detect extrastriatal DA signaling.3

PET data were analyzed using a modified simplified reference tissue model, 40 in accordance with previous work. 3,30,31,33,41–44 Stress-induced [18F]fallypride displacement, reflecting DA release, was quantified using time-activity curve plots and receptor kinetic parameters. The statistically significant change in radioligand displacement was calculated for every region of interest (ROI) as the Z-value of γ (γ /std(γ)). ^{33,41} Here, γ is considered an additional time-varying parameter in the simplified reference tissue model estimating the amplitude of ligand displacement at start of the experimental condition in a single scan session (based on the assumption that changes in competition between DA release and radioligand competition are reflected in the estimation of γ^{31}). Given that this design does not assume a physiological steady state, it is suitable to investigate time-varying changes in DA concentrations. The Z-value of γ as a proxy of stimulus-induced changes in DA release is highly correlated with BP_{ND} (binding potential relative to non-displaceable radioligand)^{33,41} and has been validated using [18F]fallypride.43

y was calculated over an exponential decay function $h(t) = \exp(-\tau(t-T))$, where t = measurement time, T = time of experimental condition initiation and τ controls the rate at which activation effects die away (dissipation



Table 2. Extrastriatal stre	Table 2. Extrastriatal stress-induced [¹8F]fallypride displacement in HV and NAPD	splacement in HV and NAPD						
Region	HV stress-induced $l^{18}F$] fallypride displacement mean $Z(\gamma)^3$ (s.d.)	NAPD stress-induced [¹⁸ F] fallypride displacement mean Z(y) ^a (s.d.)	P-value group diff. Z(y)	T-value group diff. Z(y)	HV spatial extent of stress-induced ${}^{l}^{B}F$ fallypride displacement mean ${}^{\infty}^{b}$ (s.d.)	NAPD spatial extent of stress- induced [¹⁸ F]fallypride displacement mean n% ^b (s.d.)	P-value group diff. spatial extent	T-value group diff. spatial extent
Frontal lobe mPFC ^c	2.68 (3.99)	1.59 (2.98)	0.46	-0.76	24.92 (12.16)	19.12 (10.67)	0.23	- 1.24
Temporal lobe Temporal CTX ^{c,d,e} Hippocampus ^d Parahippocampal gyrus ^d Amyddala ^d	12.88 (4.02) - 0.42 (1.08) - 0.59 (82) - 0.17 (0.13)	13.79 (3.02) - 0.39 (0.74) - 0.72 (0.47) - 0.48 (0.83)	0.54 0.95 0.65	3.21 0.06 -0.45	8.96 (6.49) < 1% 1.34 (1.83)	7.34 (7.05) 1.8 (3.1) 1.08 (1.18) < 1%	0.27 0.3 0.68	- 1.14 1.06 - 0.42 - 0.81
VIII) gadara	(51.5)). 	;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;	(00:0)	0/1/	7.0	2

of total in tracer displacement and spatial extent of Abbreviations: CTX, cortex; diff., difference; HV, healthy volunteer; mPFC, medial prefrontal cortex; NAPD, non-affective psychotic disorder. "Stress-induced [18Ffallypride displacement (Z(y)). "Percentage ^cSignificant stress-induced increase temporal ²Temporal cortex (inferior and superior interest (ROI) showing significant stress-induced [18F]fallypride displacement (Z-value of γ). >4/12. with Cook's distance removed oę in region displacement in voxels

rate, set to τ =0.03 min⁻¹),^{3,31,43} yielding a γ variate estimation interval peaking at 11 min after experimental condition onset, with the peak dissipating to 10% in 69 min.

Because previous work has demonstrated that psychological paradigms not only affect the intensity (amount) of DA release, but also the area affected, 3,33,45 the spatial extent of [18 F]fallypride displacement was calculated as the percentage of voxels in an ROI showing significant radioligand displacement (quantified as γ) after correction (p(/number of total voxels)). This approach requires that voxel T-values in a given ROI are homogenously distributed for groups of interest (HV, NAPD); this assumption was tested by calculating the decrease in number of active voxels (that is, significant γ values) when increasing the T-value by 1 (tested for multiple T-values) in all ROIs and comparing this between groups (data upon request). High correlations (up to T=0.87) between ROI ligand displacement and the spatial extent of ligand displacement (in voxels) were observed, suggesting that the area of DA release increases with DA release.

Analyses

Similar to previously published work investigating stress-induced [18F] fallypride displacement⁶ and the spatial extent stress-induced [18F] fallypride displacement,³ the total sample consisted of 12 matched HV and NAPD. *A priori* power analyses indicated a power of 0.82 to detect a group difference which is comparable to previous work using [18F] fallypride.³

Multilevel regression models with subject as the within level were applied to investigate increases in subjective stress and (positive) psychotic symptoms from control to experimental condition. Difference scores (stress-control condition) for subjective stress/symptoms were calculated for follow-up analyses. The area under the curve 46 was calculated for plasma cortisol levels (nmol I $^{-1}$). The area under the curve or nmol I $^{-1}$ cortisol difference values were used for all cortisol analyses. Regions with mean BP $_{\rm ND}$ < 0.5 in HV were not taken into account to prevent a low signal-to-noise ratio.

To replicate previous findings, we first investigated stress-induced mPFC [¹⁸F]fallypride displacement and the spatial extent of stress-induced mPFC [¹⁸F]fallypride displacement in HV. This was followed by an attempt to discover additional extrastriatal regions involved in stress processing in HV (Table 2 for all identified regions). For these purposes, *t*-tests (spatial extent/radioligand displacement >0) were performed. The same procedure was repeated for NAPD; no additional regions were identified in NAPD. Next, group differences (HV vs NAPD) in stress-induced radioligand displacement and its spatial extent were investigated in regions showing significant stress-induced radioligand displacement (using analysis of variance).

Follow-up analyses were performed using stress-induced increases in subjective stress/psychotic symptoms, symptom scores on PANSS subscales (positive, negative, general)³⁴ and the amount of years off antipsychotics (day of scan—last day of antipsychotics use/365) as outcome variables. The α was set to the conventional threshold of P=0.05. Given the matched nature of the samples, covariates were not included in group comparisons. When analyzing single groups, age and gender were entered as nuisance covariates.

RESULTS

Demographics, behavioral and physiological assessments

Groups did not differ on demographic variables (Table 1; all not significant). Recreational illicit drug use ceased long before the scan and no included participants reported current drug use (years since last use (M=17.83, s.d.=7.52)). Antipsychotics naive NAPD (n=4) and antipsychotics-free (currently non-medicated >1 year) participants did not differ in their PANSS score on the positive subscale (t(1,23)=0.25, P=0.81). Subjective stress during the scan increased from control to experimental condition (b=0.63, z(188)=6.07, P<0.0001), regardless of group (b=-0.24, z(1188)=-1.14, P=0.26). NAPD increased in positive psychotic symptoms from control to stress condition (b=0.21, z(95)=2.79, P=0.005). Subjective stress in the whole sample (b=-1.24, z(116)=-7.93, P<0.001) and positive psychotic symptoms in NAPD (b=-0.26, z(58)=-2.21, P=0.03) significantly decreased following a debriefing session 15 min after the scan finished.



Cortisol $(nmol \, I^{-1})$ decreased as a function of time in HV (b = -0.34, t(64) = -2.87, P = 004), but not in NAPD (b = -0.02, z)(66) = -0.11, P = 0.91).

There were no differences in area under the curve cortisol between conditions (t(18) = 1.65, P = 0.12), nor were there group differences (b = 474.42, t(1,9) = 0.21, P = 0.83) in area under the curve cortisol difference scores or an association with subjective stress (b = 671.38, t(18) = 0.43, P = 0.67).

Stress-induced [18F]fallypride displacement: main effects and group differences

The average HV BP_{ND} calculated over the whole paradigm using the simplified reference tissue model⁴⁰ in the mPFC (M=0.51.s.d. = 0.2), temporal cortex (TC; M = 0.63, s.d. = 0.16), hippocampus (M = 1.56, s.d. = 0.88), parahippocampal gyrus (M = 0.66, s.d. = 0.18)and amygdala (M = 4.13, s.d. = 1.56) was higher than 0.5 These regions were therefore included in the mask. No additional regions with mean $\mathrm{BP}_{\mathrm{ND}}>0.5$ were identified in NAPD.

In the mPFC and TC, a significant stress-induced increase in radioligand displacement and the spatial extent of radioligand displacement could be observed in HV and NAPD separately (P < 0.05), but not in the hippocampus, parahippocampal gyrus or amygdala (P > 0.05). No group differences in stress-induced radioligand displacement were observed in a priori selected ROI, the mPFC (Table 2), nor when looking at the dorso-mPFC (b = -0.05, t(1,23) = -0.12, P = 0.91) or ventro-mPFC (b = -0.09, t)(1,23) = -0.23, P = 0.82) subregions separately. Moreover, no group differences in stress-induced radioligand displacement were observed in the TC (Table 2).

Similarly, no group differences were observed in the spatial extent of stress-induced radioligand displacement in the mPFC (Table 2), dorso-mPFC (b = -3.11, t(1,23) = -0.55, P = 0.59), ventromPFC (b = -6.86, t(1,23) = -1.3, P = 0.21) or TC (Table 2) (Figures 2 and 3).

Stress-induced [18F]fallypride displacement: follow-up analyses In the whole sample, stress-induced radioligand displacement in mPFC (F(23) = 0.11, P = 0.74) or TC (F(23) = 0.88, P = 0.36) was not associated with subjective stress. The association between the spatial extent of stress-induced mPFC radioligand displacement and subjective stress in the whole sample did not reach significance (F(23) = 1.71, P = 0.2). When looking at mPFC subregions, the association between subjective stress and the spatial extent of stress-induced radioligand displacement in ventro-mPFC (F(23) = 2.48, P = 0.09) and dorso-mPFC (F(23) = 0.15, P = 0.87) was not significant (Figure 4). Further investigation revealed a significant positive association between subjective stress and the spatial extent of stress-induced radioligand displacement in right ventro-mPFC (F(23) = 4, P = 0.03; Figure 4), but not left ventro-mPFC (F(23) = 0.83, P = 0.45). Subjective stress was not associated with the spatial extent of stress-induced radioligand displacement in TC (F(23) = 0.63, P = 0.54; Figure 4).

The spatial extent of stress-induced radioligand displacement (b = 1.13, t(7) = 7.75, P = 0.001), but not stress-induced radioligand displacement (b = -0.22, t(7) = -2, P = 0.12), in ventromPFC was positively associated with duration of antipsychoticsfree period.

Psychotic symptoms during the scan in NAPD were not associated with stress-induced radioligand displacement in mPFC (b=1.25, t(11)=-0.13, P=0.9) or TC (b=-1.42, t(11)=-0.67,P = 0.53), or the spatial extent of stress-induced radioligand displacement in mPFC (b=-3.68, t(11)=0.51, P=0.62) or TC (b = -3.24, t(11) = -0.63, P = 0.55). PANSS positive, negative or general symptoms in NAPD were also not associated with stressinduced radioligand displacement or the spatial extent of stressinduced radioligand displacement in mPFC or TC (Table 3).

Adding years off antipsychotics as a covariate did not change results. Moreover, antipsychotics-naive NAPD and antipsychotics-free participants did not differ in stress-induced radioligand displacement or the spatial extent of stress-induced radioligand displacement in any of the identified regions (data not

Finally, cumulative haloperidol equivalents (antipsychotics in the past) were not associated with stress-induced radioligand displacement in mPFC (b < 0.01, t(11) = -0.47, P = 0.65) or TC (b < 0.01, t(11) = 1.05, P = 0.32), or the spatial extent of stressinduced tracer displacement in mPFC (b < -0.01, t(11) = -0.61, P = 0.55) or TC (b < -0.01, t(11) = -0.37, P = 0.72).

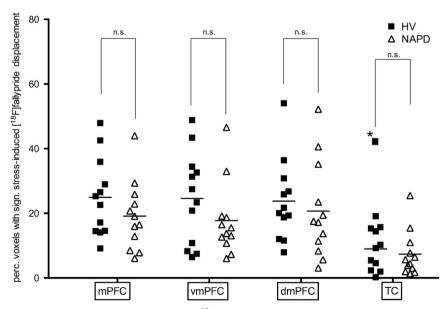


Figure 2. Group averages for the spatial extent of stress-induced [¹⁸F]fallypride displacement. NAPD did not significantly differ from HV in the spatial extent of stress-induced [¹⁸F]fallypride displacement in any (sub)region. Ventro-mPFC (vmPFC) and dorso-mPFC (dmPFC) are mPFC subregions. *, outlier (Cook's distance >4 per n), removed from mean. Not significant (NS) at P = 0.05. HV, healthy volunteer; mPFC, medial prefrontal cortex; NAPD, non-affective psychotic disorder; TC, temporal cortex.

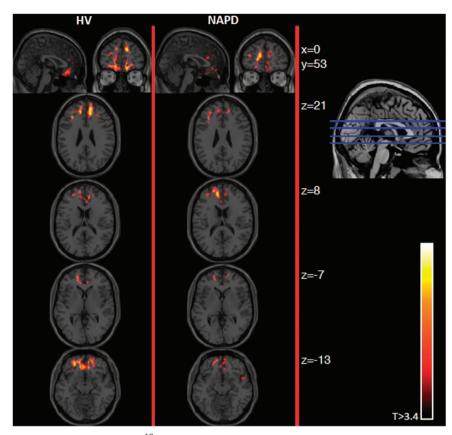


Figure 3. Parametric maps showing stress-induced [18 F]fallypride displacement in mPFC. Graphical representation showing stress-induced [18 F]fallypride in HV and NAPD in coronal (top row, left images), sagittal (top row, right images) and axial view (columns). Coronal image and Montreal Neurological Institute (MNI) z coordinates on the right depict the axial slice position. Starting position (top) was x = 0, y = 53, z = 21 (MNI). Mean t-maps per group show the stress-induced [18 F]fallypride displacement throughout the mPFC. Individual t-maps were generated using displacement parameter γ ($t = \gamma/\text{sd}(\gamma)$) and were averaged across all participants per group. Images are thresholded at 3.4 for visualization purposes. HV, healthy volunteer; mPFC, medial prefrontal cortex; NAPD, non-affective psychotic disorder.

DISCUSSION

Using [¹⁸F]fallypride PET, the effect of psychosocial stress on extrastriatal DA signaling was investigated in HV and NAPD. In accordance with previous work, extrastriatal DA release⁶ and the spatial extent (area/size of DA release in voxels) of DA release^{3,31} served as primary outcome measures of stress-related DA signaling. We showed that psychosocial stress increases extrastriatal DA signaling in HV: both DA release and the spatial extent of DA release increased in mPFC and TC. Moreover, we did not find evidence for altered stress-induced extrastriatal DA signaling in NAPD. This is based on the observations that (i) psychological stress increased DA signaling to a similar extent in HV and NAPD, (ii) subjective stress and the spatial extent of stress-induced DA release were similarly associated in HV and NAPD and (iii) stress-related DA signaling was not associated with positive, negative or general symptom scales of the PANSS in NAPD.³⁴

BP_{ND} values in frontal and temporal areas were in ranges comparable to previous studies^{6,28} although inter-individual variability was observed in the hippocampus and amygdala, which may be the result of the inherent small size of these structures. The observation that stress increased mPFC DA signaling in HV confirm previous data.^{5,6} In addition, increases in DA signaling in TC were observed. Although stress-induced TC DA signaling in humans has not been reported before, it is consistent with functional magnetic resonance imaging studies using the MIST,^{47,48} suggesting that these effects might be, in part, DAergic.

Contrary to expectations, differences in stress-induced frontal and temporal DA signaling between HV and NAPD were not observed. In combination with the absence of a correlation between measures of stress-induced DA signaling and psychotic symptoms (during scan or assessed with PANSS), these results could suggest that stress-related extrastriatal DA signaling is unaffected in NAPD. Here, we offer four explanations.

First, these results seemingly contrast with the hypothesis of DA-mediated hypofrontality in psychosis. ^{18,19} However, the concept of hypofrontality is often assessed indirectly (for example, cerebral blood flow) and in the context of cognitive performance, ^{18,49,50} not stress. Little *in vivo* evidence exists for D ^{2/3}-mediated hypofrontality in psychotic disorder ¹⁰ and a positive association between amphetamine-induced PFC DA release measured with [¹⁸F]fallpyride and schizotypal personality traits²⁹ may even suggest increased cortical DA transmission in psychotic disorder. Although inconsistent, ^{51–53} changes at the D₁ receptor have been observed in schizophrenia. Moreover, experimental animal work suggests an important role for PFC D₁ receptors in the stress response ⁵⁴ and a D₁, but not D₂, agonist can restore stress-related DAergic PFC–striatum interactions. ⁵⁵ Altogether, this could indicate that, although DA transmission at D_{2/3} during stress may be unaltered in psychotic disorder, activity at the D₁ may be abnormal.

A second viable explanation may be that the absence of differences between HV and NAPD could be explained by the relatively low amount of acute psychotic symptoms (PANSS score; Table 1). This would, however, go against evidence that increased stress sensitivity is present in those at risk for psychotic disorder, for non-acute psychotic disorder and even remitted psychotic disorder. In addition, stress-induced increases in psychotic symptoms during the scan confirmed increased stress sensitivity

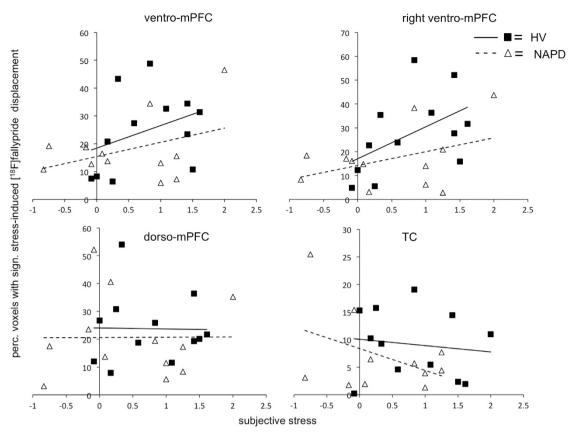


Figure 4. Association between the spatial extent of stress-induced [18]fallypride displacement and subjective stress in the whole sample. Subjective stress and the spatial extent of stress-induced radioligand displacement were associated in ventro-mPFC (trend, P = 0.06) and, more specifically, right ventro-mPFC (P = 0.02) in the whole sample, but not in dorso-mPFC (P = 0.93) and TC (P = 0.33). For visualization purposes, HV and NAPD were depicted separately. HV, healthy volunteer; mPFC, medial prefrontal cortex; NAPD, non-affective psychotic disorder; TC, temporal cortex.

Table 3. Associations between stress-induced [¹⁸F]fallypride displacement and psychotic symptoms on the Positive and Negative Syndrome Subscale (PANSS) in NAPD

	Association between stress-induced [¹⁸ F]fallypride displacement (Ζ(γ)) and PANSS symptoms			Association between spatial extent of stress-induced [¹⁸ F] fallypride displacement (% voxels) and PANSS symptoms				
	Coefficient	95% CI	T-value	P-value	Coefficient	95% CI	T-value	P-value
Positive subscale								
mPFC	0.16	-0.36 to 0.69	0.72	0.5	0.88	-1.09 to 2.84	1.03	0.33
Temporal CTX	-0.05	-0.68 to 0.56	-0.22	0.84	0.9	-0.42 to 2.21	1.57	0.16
Negative subscale								
mPFC	0.29	-0.95 to 1.53	0.54	0.61	-0.45	- 5.26 to 4.37	-0.21	0.84
Temporal CTX	-0.7	-2.02 to 0.62	- 1.22	0.26	0.49	-2.97 to 3.94	0.33	0.75
General subscale								
mPFC	0.05	-0.58 to 0.52	-0.12	0.91	0.26	- 1.83 to 2.34	0.28	0.78
Temporal CTX	-0.16	-0.77 to 0.45	- 0.6	0.57	0.48	-0.98 to 1.94	0.76	0.47

Abbreviations: CI, confidence interval; CTX, cortex; HV, healthy volunteer; mPFC, medial prefrontal cortex; NAPD, non-affective psychotic disorder.

in our sample of NAPD. We recently reported a negative correlation between the spatial extent of mPFC DA release and subjective stress/subclinical psychotic symptoms in healthy first-degree relatives of individuals with psychotic disorder. This could suggest functional cortical DAergic alterations in the stress response in some, but not all, individuals across the psychosis continuum. One way to investigate whether stress-related PFC DA signaling is dependent on illness phase is the addition of a group of acutely psychotic NAPD.

A third explanation may be that the use of [¹⁸F]fallypride has contributed to the absence of group differences. Amphetamine-induced PFC DA release quantified with fallypride has mostly yielded negative results. ^{25,26,28} However, three separate studies using the MIST^{3,6} (including the current one), as well as a study investigating response inhibition, ⁵⁹ have reported PFC DAergic activity measured with fallypride. While the reason for this discrepancy between stimulant- and task-based studies using [¹⁸F]fallypride is unclear, it may be related to the different



mechanisms of action task- and stimulant-induced DA release in the cortex. Whereas psychological tasks elicit increased DA synthesis and release, corresponding with increased cell firing, 60 stimulants increase extracellular DA release through DA and noradrenaline transporter blockade 61 and decrease overall cell firing. 62 A replication study with higher affinity radioligands such as FLB 457 (refs. 26, 63) could be useful to assess the suitability of fallypride to detect task-induced cortical DA release, as has been done recently for stimulants. 26

A final explanation could be that the sample displayed abnormalities in cortical neurotransmission not directly related to the DA system. This assumption is based on the observation that cognitive and negative symptoms in NAPD were not associated with stress-related DA signaling. One potential candidate neurotransmitter system could be glutamate. Glutamate transmission in the cortex has an essential role in stress processing, ⁶⁴ and cognitive and negative symptoms of schizophrenia have been associated with altered frontal glutamate activity, ⁶⁵ but not always consistently so. ⁶⁶ Thus, alterations in cortical glutamate transmission could potentially account for negative and cognitive symptoms in the sample of NAPD while also explaining their increased stress sensitivity to the task.

Although there may be multiple explanations for the absence of differences between NAPD and HV, stress-induced mPFC DA release⁶ and the spatial extent of mPFC DA release³ are associated with physiological and behavioral parameters. This suggests that PFC DAergic processing has a functional role in the stress response, which is potentially unaltered in NAPD. This was reflected in the correlation between the subjective stress response and spatial extent of ventro-mPFC DA release in the current study. However, an association between subjective stress and ventromPFC DA release was not observed. Although high correlations were observed between the spatial extent of DA release and DA release, this may indicate that increases in subjective stress are associated with a greater area of DA release without altering the amount of DA released. This could be interpreted as a compensatory processing mechanism, where increased resources are necessary to obtain the same result.

The spatial extent of ventro-mPFC DA release in response to stress increased as NAPD were longer off antipsychotics. Two possible explanations exist for this association. First, as NAPD are longer off antipsychotics, their DAergic stress response may progressively approximate that of HV. This is in line with an association between D₁ receptor density and drug-free interval⁶⁷ and could suggest that DA receptor density may normalize following prolonged exposure to antipsychotics. The association between time off antipsychotics and the spatial extent of stress-related DA release may reflect gradual homeostatic downregulation of PFC D_{2/3} receptors, previously upregulated through extended antipsychotics blockade, although such upregulations are dependent on mode of antipsychotics administration. 68,69

An alternative explanation may be that as acute psychotic symptoms decrease, DAergic abnormalities normalize. This is in line with work showing that striatal DA function of remitted schizophrenia patients⁷⁰ and antipsychotics-treated schizophrenia patients⁷¹ is more similar to HV. However, this explanation goes against alterations in stress sensitivity that persist beyond acute psychotic disorder⁵⁸ and the observation that the MIST increased psychotic symptoms in NAPD. Here, again, an acutely psychotic group of NAPD could be of added value.

Strengths and limitations

The current findings need to be interpreted in light of strengths, limitations and sample size.

Strengths of the study include minimal past drug use in the sample, thereby excluding substance-induced NAPD and associated confounds in the DA system. Given that, in particular,

cannabis use is associated with psychotic symptoms⁷² and DA function,^{73–75} this may have increased our sensitivity to investigate stress-related DA function. Moreover, the single infusion paradigm limited within-subjection variation, further decreasing measurement error. Finally, the direction and location of task effects in HV were similar to a previous study using an identical design, which suggest a degree of stability.⁵

Some limitations of the study need to be addressed. A general limitation is that the single infusion protocol with fallypride used in the current study has not been associated with measures directly related to DA activity, hence use of the term 'DA signaling'. Moreover, striatal DA signaling could not be reliably investigated; actual and simulated data³¹ indicate that the current design would produce unreliable estimates for the striatum, given the slow time course of radioligand binding. Future [¹⁸F]fallypride studies could increase scan duration or, in the case of a single infusion paradigm, prolong the control condition to investigate striatal and extrastriatal DA signaling simultaneously.

Because of model assumptions and to limit stress exposure to the scanning period, the task order was fixed to control-experimental, similar to previous work.^{5,14} Although this may have introduced order effects, a recent study demonstrated stress-induced DA release independent of the order of conditions.⁶ This makes it unlikely that order effects had a major effect on our outcome measures.

In addition, benzamide binding is affected by cerebral blood flow. Flowever, in response to behavioral challenges and in low-binding areas, regional cerebral blood flow effects are rather small and are not expected to explain the presented results. Other studies with a single infusion paradigm have discussed this issue in greater detail. 5,30,33,45

In the absence of a task-induced effect on plasma cortisol levels, our results could reflect socially desirable behavior or increased effort in the stress condition. The association between subjective stress and the spatial extent of ventro-mPFC DA release does, however, suggest an effect of the stressor. This is also confirmed from by data from one HV who was scanned in a control-control sequence (data upon request); changes in subjective stress or [¹⁸F] fallypride displacement were not observed.

Rather, the absence of cortisol effects may be related to time of day; a significant association between sampling time and cortisol (nmol I⁻¹) in HV was observed. Both the current study as well as another recent study who failed to find an effect of the MIST on cortisol levels⁶ collected PET data in the afternoon. In contrast, in a previous study, we did find an effect of the MIST on cortisol levels, but PET data were collected around noon. Future studies may, therefore, want to include physiological stress parameters that are less sensitive to time of day than cortisol.

Another observation was that MIST effects on the spatial extent of stress-induced ventro-mPFC DA release were smaller than previous work using an identical acquisition protocol (~25% here vs ~50%).³ This may be related to different versions of the task; the current study used an auto-adjust version (set to 70% correct responses), whereas a manually calibrated task (aiming at 90% correct response) was used previously. Task differences may have affected the perceived stressfulness of the paradigm and, correspondingly, DAergic processing. Moreover, image preprocessing software, scanner type and head fixation procedures may further explain these between-study differences.

Some limitations related to the sample also need to be addressed. Although NAPD were off antipsychotics for longer than 1 year, past antipsychotic use may have affected DA receptor density and thus masked subtle illness-related effects on stress-induced DA signaling. Although this is a limitation we acknowledge, repeating the analyses with time off antipsychotics as a covariate did not affect the results described in this manuscript. A sample of neuroleptic-naive participants could be valuable in detecting alterations in the extrastriatal DAergic stress response, if

present, associated with NAPD. In addition, the NAPD sample included five individuals with brief psychotic disorder as their main diagnosis (Supplementary Table 1); low-grade residual symptoms in these individuals may have limited the power to detect associations between stress-related DA signaling and psychotic symptoms. Finally, post hoc power calculations indicated that group differences with effect sizes (Cohen's d) up until 0.5 may have been overlooked. To detect small-to-moderate group differences, replication with larger sample sizes is essential.

CONCLUSIONS

Preclinical 16,17 and human studies have previously shown that stress affects DAergic activity in frontal cortical areas. The preliminary evidence presented here does not suggest altered extrastriatal DA signaling in the context of stress in NAPD. While we have demonstrated that frontal DA signaling is functionally relevant in the stress response, it is not clear how this is related to the putative link between stress and psychotic disorder. Follow-up studies in acutely psychotic and neuroleptic-naive NAPD could provide new insights into the role of stress-related extrastriatal DAergic processing in NAPD.

CONFLICT OF INTEREST

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REFERENCES

- 1 Mizrahi R, Addington J, Rusjan PM, Suridjan I, Ng A, Boileau I et al. Increased stress-induced dopamine release in psychosis. Biol Psychiatry 2012; 71: 561–567.
- 2 Pruessner JC, Champagne F, Meaney MJ, Dagher A. Dopamine release in response to a psychological stress in humans and its relationship to early life maternal care: a positron emission tomography study using [11C]raclopride. J Neurosci 2004; 24: 2825–2831.
- 3 Lataster J, Collip D, Ceccarini J, Hernaus D, Haas D, Booij L *et al.* Familial liability to psychosis is associated with attenuated dopamine stress signaling in ventromedial prefrontal cortex. *Schizophr Bull* 2014; **40**: 66–77.
- 4 Soliman A, O'Driscoll GA, Pruessner J, Holahan AL, Boileau I, Gagnon D *et al.* Stress-induced dopamine release in humans at risk of psychosis: a [11C]raclopride PET study. *Neuropsychopharmacology* 2008; **33**: 2033–2041.
- 5 Lataster J, Collip D, Ceccarini J, Haas D, Booij L, van Os J et al. Psychosocial stress is associated with in vivo dopamine release in human ventromedial prefrontal cortex: a positron emission tomography study using [(1)(8)F]fallypride. Neuroimage 2011; 58: 1081–1089.
- 6 Nagano-Saito A, Dagher A, Booij L, Gravel P, Welfeld K, Casey KF et al. Stress-induced dopamine release in human medial prefrontal cortex--18F-fallypride/PET study in healthy volunteers. Synapse 2013; 67: 821–830.
- 7 Walker EF, Diforio D. Schizophrenia: a neural diathesis-stress model. *Psychol Rev* 1997; **104**: 667–685.
- 8 Kelleher I, Keeley H, Corcoran P, Ramsay H, Wasserman C, Carli V *et al.* Childhood trauma and psychosis in a prospective cohort study: cause, effect, and directionality. *Am J Psychiatry* 2013; **170**: 734–741.
- 9 Lataster J, Myin-Germeys I, Lieb R, Wittchen HU, van Os J. Adversity and psychosis: a 10-year prospective study investigating synergism between early and recent adversity in psychosis. Acta Psychiatr Scand 2012; 125: 388–399.
- 10 Howes OD, Kapur S. The dopamine hypothesis of schizophrenia: version III--the final common pathway. Schizophr Bull 2009; 35: 549–562.

- 11 Egerton A, Mehta MA, Montgomery AJ, Lappin JM, Howes OD, Reeves SJ et al. The dopaminergic basis of human behaviors: a review of molecular imaging studies. Neurosci Biobehav Rev 2009: 33: 1109–1132.
- 12 Adler CM, Elman I, Weisenfeld N, Kestler L, Pickar D, Breier A. Effects of acute metabolic stress on striatal dopamine release in healthy volunteers. *Neu-ropsychopharmacology* 2000; 22: 545–550.
- 13 Brunelin J, d'Amato T, Van Os J, Costes N, Suaud Chagny MF, Saoud M. Increased left striatal dopamine transmission in unaffected siblings of schizophrenia patients in response to acute metabolic stress. *Psychiatry Res* 2010; 181: 130–135.
- 14 Montgomery AJ, Mehta MA, Grasby PM. Is psychological stress in man associated with increased striatal dopamine levels?: A [11C]raclopride PET study. Synapse 2006; 60: 124–131.
- 15 Deutch AY, Tam SY, Roth RH. Footshock and conditioned stress increase 3,4-dihydroxyphenylacetic acid (DOPAC) in the ventral tegmental area but not substantia nigra. Brain Res 1985: 333: 143–146.
- 16 Abercrombie ED, Keefe KA, DiFrischia DS, Zigmond MJ. Differential effect of stress on in vivo dopamine release in striatum, nucleus accumbens, and medial frontal cortex. J Neurochem 1989; 52: 1655–1658.
- 17 King D, Zigmond MJ, Finlay JM. Effects of dopamine depletion in the medial prefrontal cortex on the stress-induced increase in extracellular dopamine in the nucleus accumbens core and shell. *Neuroscience* 1997; **77**: 141–153.
- 18 Davis KL, Kahn RS, Ko G, Davidson M. Dopamine in schizophrenia: a review and reconceptualization. *Am J Psychiatry* 1991; **148**: 1474–1486.
- 19 Weinberger DR, Berman KF, Illowsky BP. Physiological dysfunction of dorsolateral prefrontal cortex in schizophrenia. III. A new cohort and evidence for a monoaminergic mechanism. Arch Gen Psychiatry 1988; 45: 609–615.
- 20 Howes OD, Murray RM. Schizophrenia: an integrated sociodevelopmental-cognitive model. *Lancet* 2014; 383: 1677–1687.
- 21 Myin-Germeys I, van Os J. Stress-reactivity in psychosis: evidence for an affective pathway to psychosis. *Clin Psychol Rev* 2007; **27**: 409–424.
- 22 Mukherjee J, Christian BT, Dunigan KA, Shi B, Narayanan TK, Satter M et al. Brain imaging of 18F-fallypride in normal volunteers: blood analysis, distribution, testretest studies, and preliminary assessment of sensitivity to aging effects on dopamine D-2/D-3 receptors. Synapse 2002; 46: 170–188.
- 23 Suhara T, Sudo Y, Okauchi T, Maeda J, Kawabe K, Suzuki K et al. Extrastriatal dopamine D2 receptor density and affinity in the human brain measured by 3D PET. Int J Neuropsychopharmacol 1999; 2: 73–82.
- 24 Hirvonen J, Aalto S, Lumme V, Nagren K, Kajander J, Vilkman H et al. Measurement of striatal and thalamic dopamine D2 receptor binding with 11C-raclopride. Nucl Med Commun 2003; 24: 1207–1214.
- 25 Cropley VL, Innis RB, Nathan PJ, Brown AK, Sangare JL, Lerner A *et al.* Small effect of dopamine release and no effect of dopamine depletion on [18F]fallypride binding in healthy humans. *Synapse* 2008; **62**: 399–408.
- 26 Narendran R, Frankle WG, Mason NS, Rabiner EA, Gunn RN, Searle GE et al. Positron emission tomography imaging of amphetamine-induced dopamine release in the human cortex: a comparative evaluation of the high affinity dopamine D2/3 radiotracers [11C]FLB 457 and [11C]fallypride. Synapse 2009; 63: 447–461.
- 27 Riccardi P, Li R, Ansari MS, Zald D, Park S, Dawant B *et al.* Amphetamine-induced displacement of [18F] fallypride in striatum and extrastriatal regions in humans. *Neuropsychopharmacology* 2006; **31**: 1016–1026.
- 28 Slifstein M, Kegeles LS, Xu X, Thompson JL, Urban N, Castrillon J et al. Striatal and extrastriatal dopamine release measured with PET and [(18)F] fallypride. Synapse 2010; 64: 350–362.
- 29 Woodward ND, Cowan RL, Park S, Ansari MS, Baldwin RM, Li R et al. Correlation of individual differences in schizotypal personality traits with amphetamine-induced dopamine release in striatal and extrastriatal brain regions. Am J Psychiatry 2011; 168: 418–426.
- 30 Alpert NM, Badgaiyan RD, Livni E, Fischman AJ. A novel method for noninvasive detection of neuromodulatory changes in specific neurotransmitter systems. *Neuroimage* 2003; 19: 1049–1060.
- 31 Ceccarini J, Vrieze E, Koole M, Muylle T, Bormans G, Claes S et al. Optimized in vivo detection of dopamine release using 18F-fallypride PET. J Nucl Med 2012; 53: 1565–1572.
- 32 van Os J, Burns T, Cavallaro R, Leucht S, Peuskens J, Helldin L et al. Standardized remission criteria in schizophrenia. Acta Psychiatr Scand 2006; **113**: 91–95.
- 33 Hernaus D, Collip D, Lataster J, Ceccarini J, Kenis G, Booij L *et al.* COMT Val158Met genotype selectively alters prefrontal [18F]fallypride displacement and subjective feelings of stress in response to a psychosocial stress challenge. *PLoS One* 2013; **8**:
- 34 Kay SR, Fiszbein A, Opler LA. The positive and negative syndrome scale (PANSS) for schizophrenia. *Schizophr Bull* 1987; **13**: 261–276.
- 35 Dressendorfer RA, Kirschbaum C, Rohde W, Stahl F, Strasburger CJ. Synthesis of a Cortisol Biotin Conjugate and Evaluation as a Tracer in an Immunoassay for Salivary Cortisol Measurement. J Steroid Biochem 1992; 43: 683–692.



- 36 Grunder G, Siessmeier T, Piel M, Vernaleken I, Buchholz HG, Zhou Y et al. Quantification of D2-like dopamine receptors in the human brain with 18F-desmethoxyfallypride. J Nucl Med 2003; 44: 109–116.
- 37 Stark D, Piel M, Hubner H, Gmeiner P, Grunder G, Rosch F. In vitro affinities of various halogenated benzamide derivatives as potential radioligands for noninvasive quantification of D(2)-like dopamine receptors. Bioorg Med Chem 2007; 15: 6819–6829.
- 38 Green MV, Seidel J, Stein SD, Tedder TE, Kempner KM, Kertzman C et al. Head movement in normal subjects during simulated PET brain imaging with and without head restraint. J Nucl Med 1994: 35: 1538–1546.
- 39 Mawlawi O, Martinez D, Slifstein M, Broft A, Chatterjee R, Hwang DR et al. Imaging human mesolimbic dopamine transmission with positron emission tomography: I. Accuracy and precision of D(2) receptor parameter measurements in ventral striatum. J Cereb Blood Flow Metab 2001; 21: 1034–1057.
- 40 Innis RB, Cunningham VJ, Delforge J, Fujita M, Gjedde A, Gunn RN et al. Consensus nomenclature for in vivo imaging of reversibly binding radioligands. *J Cereb Blood Flow Metab* 2007; **27**: 1533–1539.
- 41 Backman L, Nyberg L, Soveri A, Johansson J, Andersson M, Dahlin E et al. Effects of working-memory training on striatal dopamine release. *Science* 2011; **333**: 718.
- 42 Badgaiyan RD, Fischman AJ, Alpert NM. Dopamine release during human emotional processing. *Neuroimage* 2009; **47**: 2041–2045.
- 43 Christian BT, Lehrer DS, Shi B, Narayanan TK, Strohmeyer PS, Buchsbaum MS et al. Measuring dopamine neuromodulation in the thalamus: using [F-18]fallypride PET to study dopamine release during a spatial attention task. Neuroimage 2006; 31: 139–152.
- 44 Badgaiyan RD. Detection of dopamine neurotransmission in "real time". Front Neurosci 2013: 7: 125.
- 45 Ceccarini J, Koole M, Van Laere K. Reply: Task- versus amphetamine-induced displacement of high-affinity D2/3 receptor ligands. J Nucl Med 2013; 54: 1849–1850.
- 46 Pruessner JC, Kirschbaum C, Meinlschmid G, Hellhammer DH. Two formulas for computation of the area under the curve represent measures of total hormone concentration versus time-dependent change. *Psychoneuroendocrinology* 2003; 28: 916–931
- 47 Dedovic K, Rexroth M, Wolff E, Duchesne A, Scherling C, Beaudry T et al. Neural correlates of processing stressful information: an event-related fMRI study. Brain Res 2009: 1293: 49–60.
- 48 Wang J, Rao H, Wetmore GS, Furlan PM, Korczykowski M, Dinges DF et al. Perfusion functional MRI reveals cerebral blood flow pattern under psychological stress. Proc Natl Acad Sci USA 2005; 102: 17804–17809.
- 49 Fusar-Poli P, Howes OD, Allen P, Broome M, Valli I, Asselin MC et al. Abnormal frontostriatal interactions in people with prodromal signs of psychosis: a multimodal imaging study. Arch Gen Psychiatry 2010; 67: 683–691.
- 50 Meyer-Lindenberg A, Miletich RS, Kohn PD, Esposito G, Carson RE, Quarantelli M et al. Reduced prefrontal activity predicts exaggerated striatal dopaminergic function in schizophrenia. Nat Neurosci 2002: 5: 267–271.
- 51 Abi-Dargham A, Moore H. Prefrontal DA transmission at D1 receptors and the pathology of schizophrenia. *Neuroscientist* 2003; **9**: 404–416.
- 52 Karlsson P, Farde L, Halldin C, Sedvall G. PET study of D(1) dopamine receptor binding in neuroleptic-naive patients with schizophrenia. Am J Psychiatry 2002; 159: 761–767.
- 53 Okubo Y, Suhara T, Suzuki K, Kobayashi K, Inoue O, Terasaki O et al. Decreased prefrontal dopamine D1 receptors in schizophrenia revealed by PET. Nature 1997; 385: 634–636.
- 54 Arnsten AF. Prefrontal cortical network connections: key site of vulnerability in stress and schizophrenia. *Int J Dev Neurosci* 2011; **29**: 215–223.
- 55 Scornaiencki R, Cantrup R, Rushlow WJ, Rajakumar N. Prefrontal cortical D1 dopamine receptors modulate subcortical D2 dopamine receptor-mediated stress responsiveness. Int J Neuropsychopharmacol 2009: 12: 1195–1208.
- 56 Collip D, Nicolson NA, Lardinois M, Lataster T, van Os J, Myin-Germeys I et al. Daily cortisol, stress reactivity and psychotic experiences in individuals at above average genetic risk for psychosis. Psychol Med 2011; 41: 2305–2315.
- 57 Myin-Germeys I, van Os J, Schwartz JE, Stone AA, Delespaul PA. Emotional reactivity to daily life stress in psychosis. Arch Gen Psychiatry 2001; 58: 1137–1144.
- 58 Myin-Germeys I, Delespaul P, van Os J. Behavioural sensitization to daily life stress in psychosis. *Psychol Med* 2005; 35: 733–741.
- 59 Albrecht DS, Kareken DA, Christian BT, Dzemidzic M, Yoder KK. Cortical dopamine release during a behavioral response inhibition task. Synapse 2014; 68: 266–274.

- 60 Dugast C, Suaud-Chagny MF, Gonon F. Continuous in vivo monitoring of evoked dopamine release in the rat nucleus accumbens by amperometry. Neuroscience 1994: 62: 647–654.
- 61 Bymaster FP, Katner JS, Nelson DL, Hemrick-Luecke SK, Threlkeld PG, Heiligenstein JH et al. Atomoxetine increases extracellular levels of norepinephrine and dopamine in prefrontal cortex of rat: a potential mechanism for efficacy in attention deficit/hyperactivity disorder. Neuropsychopharmacology 2002; 27: 699–711.
- 62 Bunney BS, Walters JR, Roth RH, Aghajanian GK. Dopaminergic neurons: effect of antipsychotic drugs and amphetamine on single cell activity. J Pharmacol Exp Ther 1973; 185: 560–571.
- 63 Narendran R, Jedema HP, Lopresti BJ, Mason NS, Gurnsey K, Ruszkiewicz J et al. Imaging dopamine transmission in the frontal cortex: a simultaneous microdialysis and [11C]FLB 457 PET study. Mol Psychiatry 2014; 19: 302–310.
- 64 Moghaddam B. Stress activation of glutamate neurotransmission in the prefrontal cortex: implications for dopamine-associated psychiatric disorders. *Biol Psychiatry* 2002; **51**: 775–787.
- 65 Merritt K, McGuire P, Egerton A. Relationship between glutamate dysfunction and symptoms and cognitive function in psychosis. Front Psychiatry 2013; 4: 151.
- 66 Poels EM, Kegeles LS, Kantrowitz JT, Slifstein M, Javitt DC, Lieberman JA et al. Imaging glutamate in schizophrenia: review of findings and implications for drug discovery. Mol Psychiatry 2014; 19: 20–29.
- 67 Abi-Dargham A, Xu X, Thompson JL, Gil R, Kegeles LS, Urban N et al. Increased prefrontal cortical D(1) receptors in drug naive patients with schizophrenia: a PET study with [(1)(1)C]NNC112. J Psychopharmacol 2012; 26: 794–805.
- 68 Chen JF, Aloyo VJ, Weiss B. Continuous treatment with the D2 dopamine receptor agonist quinpirole decreases D2 dopamine receptors, D2 dopamine receptor messenger RNA and proenkephalin messenger RNA, and increases mu opioid receptors in mouse striatum. Neuroscience 1993; 54: 669–680.
- 69 Samaha AN, Reckless GE, Seeman P, Diwan M, Nobrega JN, Kapur S. Less is more: antipsychotic drug effects are greater with transient rather than continuous delivery. *Biol Psychiatry* 2008; **64**: 145–152.
- 70 Laruelle M, Abi-Dargham A, Gil R, Kegeles L, Innis R. Increased dopamine transmission in schizophrenia: relationship to illness phases. *Biol Psychiatry* 1999; 46: 56–72
- 71 Demjaha A, Egerton A, Murray RM, Kapur S, Howes OD, Stone JM et al. Antipsychotic treatment resistance in schizophrenia associated with elevated glutamate levels but normal dopamine function. *Biol Psychiatry* 2014; **75**: e11–e13.
- 72 Kuepper R, van Os J, Lieb R, Wittchen HU, Hofler M, Henquet C. Continued cannabis use and risk of incidence and persistence of psychotic symptoms: 10 year follow-up cohort study. BMJ 2011; 342: d738.
- 73 Kuepper R, Ceccarini J, Lataster J, van Os J, van Kroonenburgh M, van Gerven JM et al. Delta-9-tetrahydrocannabinol-induced dopamine release as a function of psychosis risk: 18F-fallypride positron emission tomography study. *PLoS One* 2013; 8: e70378
- 74 Mizrahi R, Kenk M, Suridjan I, Boileau I, George TP, McKenzie K et al. Stress-induced dopamine response in subjects at clinical high risk for schizophrenia with and without concurrent cannabis use. Neuropsychopharmacology 2013; 39: 1479-1489
- 75 Stokes PR, Egerton A, Watson B, Reid A, Breen G, Lingford-Hughes A et al. Significant decreases in frontal and temporal [11C]-raclopride binding after THC challenge. Neuroimage 2010; 52: 1521–1527.
- 76 Dagher A, Gunn R, Lockwood G, Cunningham V, Grasby P, Brooks D. Quantitative functional brain imaging with positron emission tomography. Academic Press: San Diego, CA, USA, 1998.
- 77 Cumming P, Xiong G, la Fougere C, Rominger A, Bartenstein P, Buchholz HG *et al.*Surrogate markers for cerebral blood flow correlate with [(1)(8)F]-fallypride binding potential at dopamine D(2/3) receptors in human striatum. *Synapse* 2013; 67: 199–203.



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