

SCIENTIFIC REPORTS



OPEN

D-form KLKLLLLLKLK-NH₂ peptide exerts higher antimicrobial properties than its L-form counterpart via an association with bacterial cell wall components

Takayuki Manabe & Kiyoshi Kawasaki

Received: 01 November 2016

Accepted: 20 January 2017

Published: 06 March 2017

The antimicrobial peptide KLKLLLLLKLK-NH₂ was developed based on sapsin B, and synthesized using D-amino acids. Biochemical properties of the D-form and L-form KLKLLLLLKLK-NH₂ peptides were compared. In order to limit the effects due to bacterial resistance to proteolysis, antimicrobial activities of the peptides were evaluated after short-term exposure to bacteria. D-form KLKLLLLLKLK-NH₂ exhibited higher antimicrobial activities than L-form KLKLLLLLKLK-NH₂ against bacteria, including *Staphylococcus aureus* and *Escherichia coli*. In contrast, both the D-form and L-form of other antimicrobial peptides, including Mastoparan M and Temporin A, exhibited similar antimicrobial activities. Both the D-form KLKLLLLLKLK-NH₂ and L-form KLKLLLLLKLK-NH₂ peptides preferentially disrupted *S. aureus*-mimetic liposomes over mammalian-mimetic liposomes. Furthermore, the D-form KLKLLLLLKLK-NH₂ increased the membrane permeability of *S. aureus* more than the L-form KLKLLLLLKLK-NH₂. Thus suggesting that the enhanced antimicrobial activity of the D-form was likely due to its interaction with bacterial cell wall components. *S. aureus* peptidoglycan preferentially inhibited the antimicrobial activity of the D-form KLKLLLLLKLK-NH₂ relative to the L-form. Furthermore, the D-form KLKLLLLLKLK-NH₂ showed higher affinity for *S. aureus* peptidoglycan than the L-form. Taken together, these results indicate that the D-form KLKLLLLLKLK-NH₂ peptide has higher antimicrobial activity than the L-form via a specific association with bacterial cell wall components, including peptidoglycan.

Cationic antimicrobial peptides act as innate immune agents in multicellular organisms including mammals and insects (reviewed in ref. 1). Antimicrobial peptides specifically bind to negatively-charged bacterial surface molecules, such as the endotoxin lipopolysaccharide^{2–4}, and disrupt the bacterial membrane leading to cell lysis and/or cell death. The Shai-Matsuzaki-Huang model is a well-known machinery that explains the activity of antimicrobial peptides^{1,5–7}. Antimicrobial peptides have attracted great interest as a novel class of antibiotics because they exhibit broad-spectrum antimicrobial activities and they are not likely to induce resistance. Unfortunately, the susceptibility of antimicrobial peptides to proteases limits their use in pharmaceutical applications. To overcome this limitation, antimicrobial peptides may be synthesized from D-amino acids instead of L-amino acids, which makes them resistant to proteolytic degradation.

Both D-form and L-form antimicrobial peptides showed similar antimicrobial activity^{8–12}. However, D-form antimicrobial peptides were more stable and retained antimicrobial activity for extended periods of time compared to L-form antimicrobial peptides in the presence of NaCl, CaCl₂, or human serum albumin at physiological concentrations¹¹, and D-form antimicrobial peptides were resistant to enzymatic degradation¹⁰. Furthermore, a D-form 13-mer antimicrobial peptide, but not the L-form 13-mer antimicrobial peptide, reduced endotoxin-induced lethality in mice⁸. The D-form-specific effect was explained by protease resistance *in vivo*^{9,12}. Furthermore, antimicrobial peptides were thought to exert their effects without specific target receptors which require close contact based on the structure of the peptides^{10,12}.

Faculty of Pharmaceutical Sciences, Doshisha Women's College, Kyotanabe, Kyoto 610-0395, Japan. Correspondence and requests for materials should be addressed to K.K. (email: kkawasak@dwc.doshisha.ac.jp)

	MIC ($\mu\text{g/ml}$)	
	D-form	L-form
<i>S. aureus</i>	1	16
<i>E. coli</i>	8	16
<i>C. albicans</i>	32	64

Table 1. Minimum Inhibitory Concentrations (MICs) of D-form and L-form KLKLLLLLKLK-NH₂ against *S. aureus*, *E. coli*, and *C. albicans*.

	MIC ($\mu\text{g/ml}$)	
	D-form	L-form
KLKLLLLLKLK-NH ₂	4	>128
FIKRIARLLRKIF-NH ₂	4	4
INLKAIAALAKKLL-NH ₂	16	16
FLPLIGRVLSGIL-NH ₂	8	8

Table 2. Minimum Inhibitory Concentrations (MICs) of D-form and L-form antimicrobial peptides against *S. aureus*.

Sapecin B is an antimicrobial peptide that was originally isolated from the culture medium of an embryonic cell line, NIH-Sape-4, derived from *Sarcophaga peregrina* (flesh fly). It displays potent activity against Gram-positive bacteria¹³. Two other related proteins, sapecin and sapecin C, were also isolated from culture medium of NIH-Sape-4^{13–15}. Sapecin B has significant sequence similarity to a scorpion venom toxin, charybdotoxin^{13,16}. Structural comparison of sapecin B and charybdotoxin identified the undecapeptide RSLCLLHCRLK-NH₂, which corresponds to amino acid residues 7 to 17 of sapecin B with C-terminal amidation^{16,17}. The peptide fragment RSLCLLHCRLK-NH₂ showed significant antimicrobial activity, suggesting that this region is responsible for the antimicrobial activity of the peptide¹⁷. The undecapeptide KLKLLLLLKLK-NH₂ was developed by modifying the primary structure of RSLCLLHCRLK-NH₂. In addition to its activity against Gram-positive bacteria, Gram-negative bacteria, and fungi¹⁸, KLKLLLLLKLK-NH₂ has been shown to enhance mammalian immune responses via undefined molecular mechanisms^{19–21}. The antimicrobial activity of the D-form KLKLLLLLKLK-NH₂, which was synthesized using D-amino acids, persisted longer than the L-form because of its resistance to proteolytic degradation¹⁸.

In this study, we examined the antimicrobial properties of D-form KLKLLLLLKLK-NH₂. D-form KLKLLLLLKLK-NH₂ displays higher antimicrobial activity against bacteria than its L-form; however, this elevated activity could not be explained by resistance to proteolytic degradation. It is important to note that other D-form antimicrobial peptides did not show higher antimicrobial activity than their L-form counterparts. Furthermore, D-form KLKLLLLLKLK-NH₂ showed higher affinity for bacterial cell wall components, such as peptidoglycan, than its L-form. Thus, the enhanced antimicrobial activity of the D-form KLKLLLLLKLK-NH₂ relative to its L-form is due to direct interactions with bacterial cell surface components.

Results

MICs of D-form KLKLLLLLKLK-NH₂ were lower than those of L-form KLKLLLLLKLK-NH₂.

Previously, D-form KLKLLLLLKLK-NH₂ was shown to persist longer in bacterial culture medium and it showed higher antimicrobial activity to *Staphylococcus aureus* than the L-form¹⁸. In order to further examine the antimicrobial properties of D-form KLKLLLLLKLK-NH₂, we determined the MICs of the peptides against *S. aureus*, *Escherichia coli*, and *Candida albicans*. MICs of D-form KLKLLLLLKLK-NH₂ were lower than those of its L-form, especially against *S. aureus* where the MIC of the D-form was 16-fold lower than the L-form (Table 1). We determined minimum inhibitory concentrations (MICs) of other antimicrobial peptides, including KLKLLLLLKLK-NH₂, a derivative of KLKLLLLLKLK-NH₂¹⁸, FIKRIARLLRKIF-NH₂ (Kn2–7) derived from *Buthus martensii* scorpion venom²², INLKAIAALAKKLL-NH₂ (Mastoparan M) derived from *hornet venom*²³, and FLPLIGRVLSGIL-NH₂ (Temporin A) derived from *Rana temporaria* reference²⁴ against *S. aureus*. All of these peptides are expected to form a helical structure similar to KLKLLLLLKLK-NH₂^{16,17,22–24}. MIC of D-form KLKLLLLLKLK-NH₂ against *S. aureus* is more than 32-fold lower than that of the L-form (Table 2). In contrast, the MIC of D-forms and L-forms of Mastoparan M, Kn2–7, and Temporin A against *S. aureus* (Table 2) were similar. These observations indicate that KLKLLLLLKLK-NH₂ and its related peptide KLKLLLLLKLK-NH₂ are unique because these D-form peptides display lower MICs against *S. aureus* than their L-forms.

D-form KLKLLLLLKLK-NH₂ displayed higher antimicrobial activity than L-form KLKLLLLLKLK-NH₂.

In order to further explore the antimicrobial properties of D-form KLKLLLLLKLK-NH₂, we incubated microorganisms with the peptides for 10 min to minimize the effect of proteolytic degradation. D-form KLKLLLLLKLK-NH₂ showed higher antimicrobial activity to *S. aureus*, *Enterococcus hirae*, *E. coli*, and *Salmonella enterica* serovar Typhimurium than L-form KLKLLLLLKLK-NH₂ (Fig. 1a–d). In contrast, D-form and L-form KLKLLLLLKLK-NH₂ showed similar antimicrobial activity against *C. albicans* (Fig. 1e). Under the experimental conditions in which we examined antimicrobial activity against *S. aureus*, specific degradation of the L-form

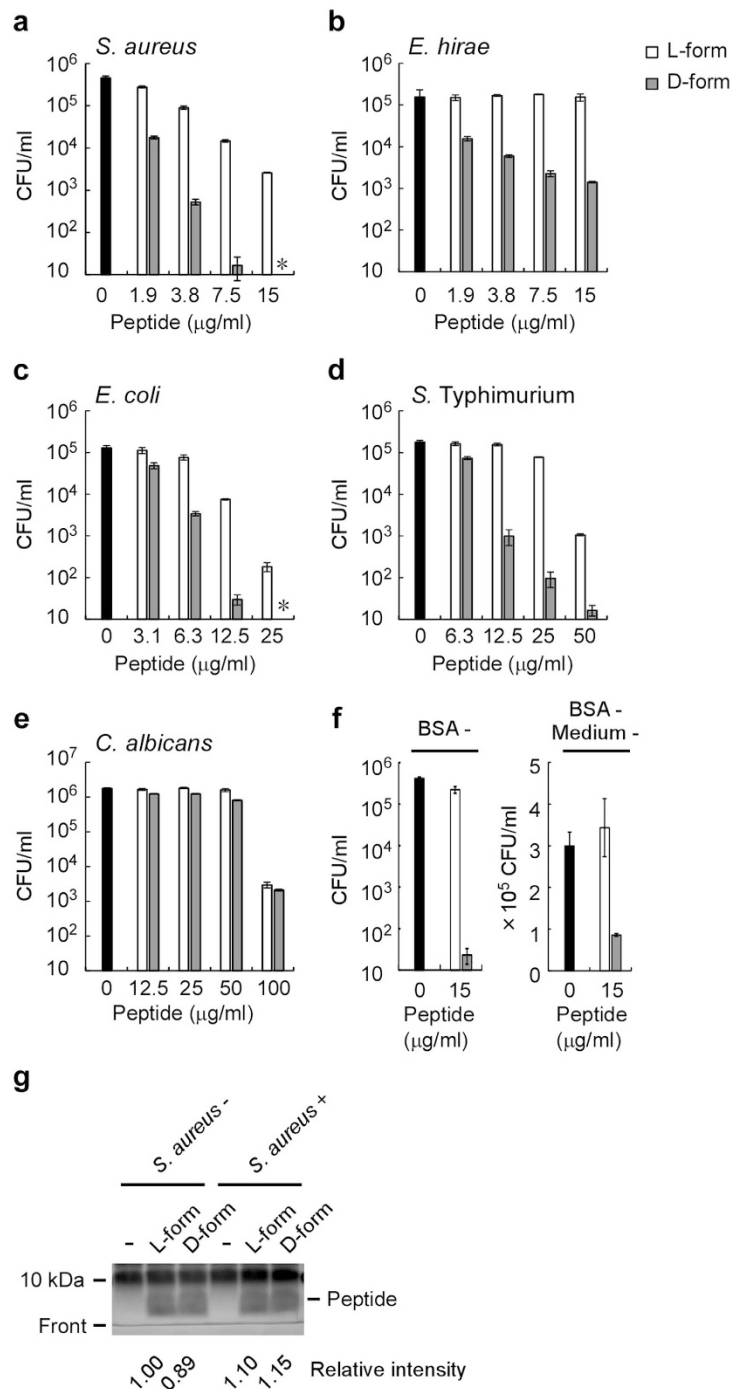


Figure 1. Antimicrobial activities of D-form KLKLLLLLKLK-NH₂ to bacteria were higher than those of the L-form counterpart. Antimicrobial activities of D-form KLKLLLLLKLK-NH₂ (D-form) and L-form KLKLLLLLKLK-NH₂ (L-form) against *S. aureus* (a,f), *E. hirae* (b), *E. coli* (c), *S. Typhimurium* (d), and *C. albicans* (e) were examined. (f) Antimicrobial activities were examined in the absence of bovine serum albumin (BSA-) or in the absence of both bovine serum albumin and culture medium (BSA-, medium-). Gray bars and white bars represent CFUs in assay mixtures treated with the indicated concentrations of D-form and L-form peptides, respectively. Black bars represent CFUs in assay mixtures without peptide. The error bars represent the mean ± standard deviations from triplicate plates. Asterisk (*) indicates no bacteria were detected (<10 CFU/ml). Concentrations of dimethyl sulfoxide in the assay mixtures were 0.15% (a,b and f) and 1.0% (c-e). (g) *S. aureus* (*S. aureus*-) was treated without (-) or with 15 μg/ml D-form KLKLLLLLKLK-NH₂ (D-form) or L-form KLKLLLLLKLK-NH₂ (L-form) as performed in (a). Control assay mixtures without *S. aureus* (*S. aureus*-) were also prepared. The assay mixtures (10 μl) were subjected to SDS-polyacrylamide gel electrophoresis using Any kD™ precast polyacrylamide gel. Proteins, including KLKLLLLLKLK-NH₂ (molecular weight 1321.8), were visualized with Ruby protein gel stain. The full-length gel was shown in supplementary figure 1. Relative intensities of peptide signals were measured using Multi Gauge ver.3 (FUJIFILM), and the values normalized with that of L-form peptide (*S. aureus*-) were indicated.

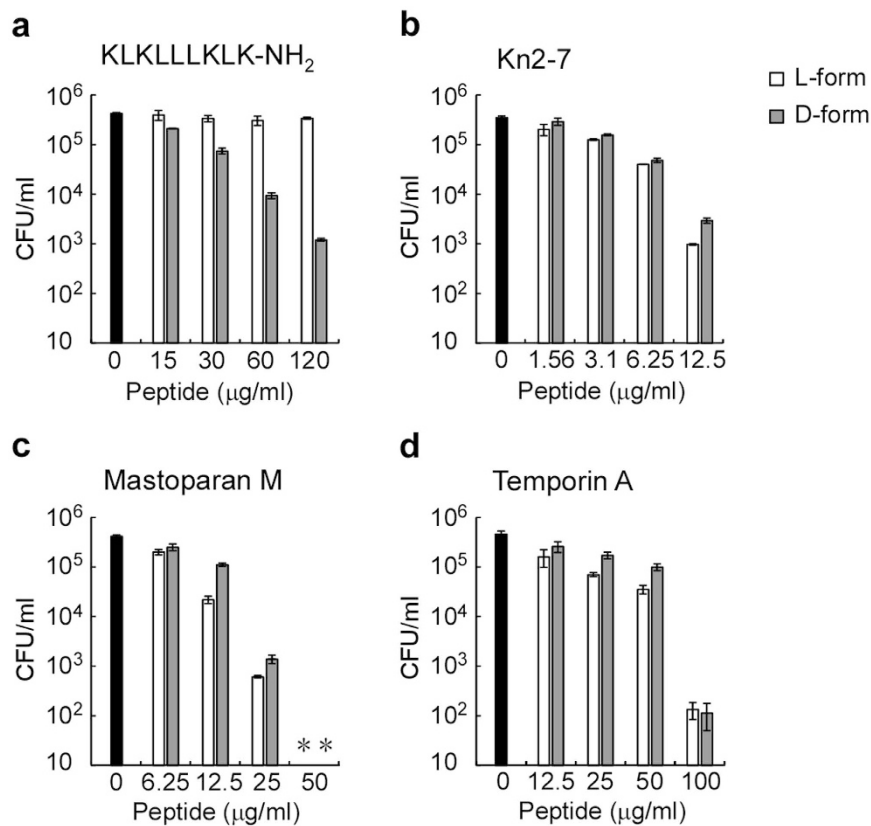


Figure 2. Similar antimicrobial activities of D-forms and L-forms of Kn2-7, Mastoparan M, and Temporin A. Antimicrobial activities of KLKLLLLKLNH₂ (a), Kn2-7 (b), Mastoparan M (c), and Temporin A (d) against *S. aureus* were examined. Gray bars and white bars represent CFUs in assay mixtures treated with indicated concentrations of D-form and L-form peptides, respectively. Black bars represent CFUs in assay mixtures treated without peptide. The error bars represent the mean \pm standard deviations from triplicate plates. Asterisk (*) indicates bacteria were not detected (less than 10 CFU/ml). Concentrations of dimethyl sulfoxide in the assay mixtures were 0.5%.

peptide was not observed (Fig. 1g). This observation suggests that the higher antimicrobial activity of D-form KLKLLLLKLNH₂ was not due to its resistance to proteolytic degradation. In addition, in order to exclude the possibility that bovine serum albumin or some components from culture medium specifically affect the antimicrobial activity of KLKLLLLKLNH₂, we performed experiments without culture medium and/or bovine serum albumin in the assay mixture. D-form KLKLLLLKLNH₂ also showed higher antimicrobial activity to *S. aureus* than L-form KLKLLLLKLNH₂ in the absence of culture medium and/or bovine serum albumin (Fig. 1f). It is noteworthy that antimicrobial activity of both L-form and D-form peptide in the absence of culture medium and bovine serum albumin were lower than those in our standard assay condition (Fig. 1a and f). The antimicrobial activity of D-form KLKLLLLKLNH₂ was also higher than that of its L-form counterpart (Fig. 2a). In contrast, the D-forms and L-forms of Kn2-7, Mastoparan M, and Temporin A peptides displayed similar antimicrobial activities against *S. aureus* (Fig. 2b–d). These results indicate that KLKLLLLKLNH₂ and its derivative KLKLLLLKLNH₂ are unique in that their D-forms have antimicrobial activities than their L-forms.

D-form KLKLLLLKLNH₂ increased bacterial membrane permeability. Cationic antimicrobial peptides bind to the negatively charged bacterial surface and penetrate into the bacterial membrane. Therefore, their effects on bacterial membrane permeability closely correlate with antimicrobial activity. Effects of KLKLLLLKLNH₂ and Mastoparan M on membrane permeability of *S. aureus* were monitored by ethidium bromide influx rates. As shown in Fig. 3a, both D-form KLKLLLLKLNH₂ (20 μ g/ml) and L-form KLKLLLLKLNH₂ (20 μ g/ml) increased ethidium bromide influx rates; however, the rates were higher in response to D-form KLKLLLLKLNH₂ than the L-form KLKLLLLKLNH₂. In contrast, D-form and L-form Mastoparan M (20 μ g/ml) increased ethidium bromide influx rates to a similar extent (20 μ g/ml) (Fig. 3b). These observations are consistent with the findings that the antimicrobial activity of D-form KLKLLLLKLNH₂ against *S. aureus* was higher than that of its L-form KLKLLLLKLNH₂ (Fig. 1a). However, that antimicrobial activity of D-form Mastoparan M against *S. aureus* was similar with that of its L-form (Fig. 2c).

***S. aureus* peptidoglycan and *E. coli* lipopolysaccharide preferentially inhibited the antimicrobial activity of D-form KLKLLLLKLNH₂.** Most cationic antimicrobial peptides interact with bacterial membranes. Previously, sapeicin was shown to have a high affinity for cardiolipin²⁵. This observation encouraged

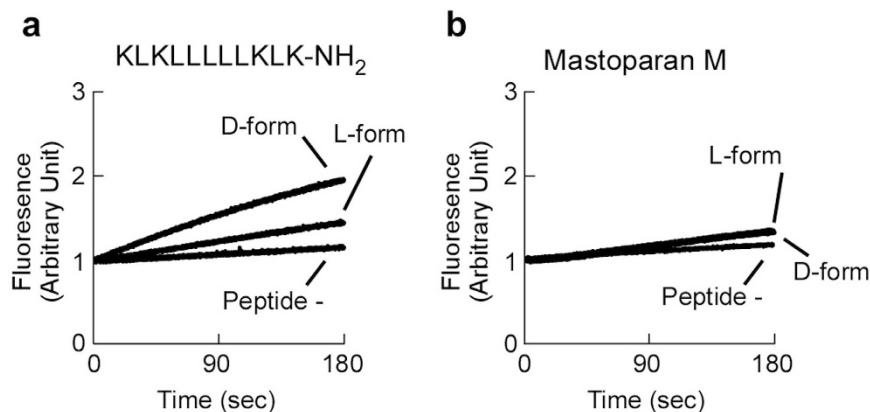


Figure 3. Effect of D-form KLKLLLLLKLK-NH₂ on *S. aureus* membrane permeability was higher L-form KLKLLLLLKLK-NH₂. (a) *S. aureus* was treated with 20 µg/ml of D-form KLKLLLLLKLK-NH₂, 20 µg/ml of L-form KLKLLLLLKLK-NH₂, or without peptide (peptide-). (b) *S. aureus* was treated with 20 µg/ml D-form Mastoparan M, 20 µg/ml L-form Mastoparan M, or without peptide (peptide-). Ethidium influx was monitored by fluorescence for 180 sec after the addition of ethidium bromide to the bacteria/peptide suspension.

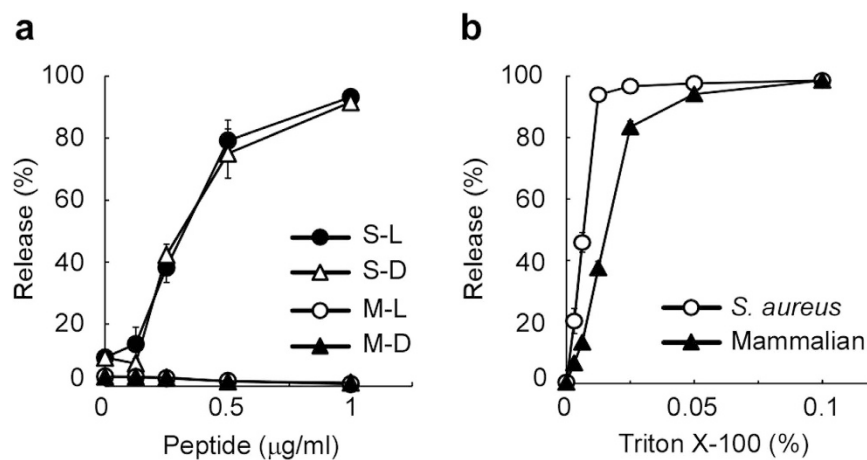


Figure 4. D-form and L-form KLKLLLLLKLK-NH₂ disrupt *S. aureus*-mimetic liposomes. (a) *S. aureus*-type and mammalian-type liposomes with calcein were exposed to D-form and L-form KLKLLLLLKLK-NH₂; S-L, *S. aureus*-mimetic liposomes treated with L-form peptide; S-D, *S. aureus*-mimetic liposomes treated with D-form peptide; M-L, mammalian-mimetic liposomes treated with L-form peptide; and M-D, mammalian-mimetic liposomes treated with D-form peptide. (b) *S. aureus*-type (*S. aureus*) and mammalian-type (Mammalian) liposomes containing calcein were exposed to Triton X-100. The amount of calcein that leaked from the liposomes was measured using a spectrofluorophotometer and normalized to determine the % release relative to 0.1% Triton X-100. The error bars represent the mean \pm standard deviations from triplicate assays.

us to examine whether D-form KLKLLLLLKLK-NH₂ specifically disrupts liposomes that mimic the cellular membrane of *S. aureus*. Both D-form and L-form KLKLLLLLKLK-NH₂ released calcein from *S. aureus*-mimetic liposomes^{17,26}, which consisted of phosphatidylglycerol and cardiolipin (Fig. 4a). On the other hand, neither D-form nor L-form KLKLLLLLKLK-NH₂ was able to release calcein from mammalian-mimetic liposomes²⁷ that consisted of phosphatidylcholine, phosphatidylethanolamine, and cholesterol (Fig. 4a). Mammalian-mimetic liposomes demonstrated similar sensitivity to Triton X-100 as *S. aureus*-mimetic liposomes, excluding the possibility that mammalian-mimetic liposomes are resistant to chemical treatments (Fig. 4b). These observations indicate that both D-form and L-form KLKLLLLLKLK-NH₂ preferentially disrupt *S. aureus*-mimetic liposomes, which likely contributes to the antimicrobial activity of KLKLLLLLKLK-NH₂. Thus, the ability to disrupt *S. aureus*-mimetic liposomes is not the cause of higher antimicrobial activity of D-form KLKLLLLLKLK-NH₂ relative to its L-form.

To identify a specific target of D-form KLKLLLLLKLK-NH₂, we analyzed whether bacterial cell wall components were able to inhibit the antimicrobial activities. A comparison of the antimicrobial activities of D-form and L-form KLKLLLLLKLK-NH₂ revealed that 1.9 µg/ml of D-form and 7.5 µg/ml of L-form displayed similar antimicrobial activity to *S. aureus*. The antimicrobial effect of D-form KLKLLLLLKLK-NH₂ was almost inhibited by 40 µg/ml of *S. aureus* peptidoglycan, but the same concentration failed to abrogate the antimicrobial

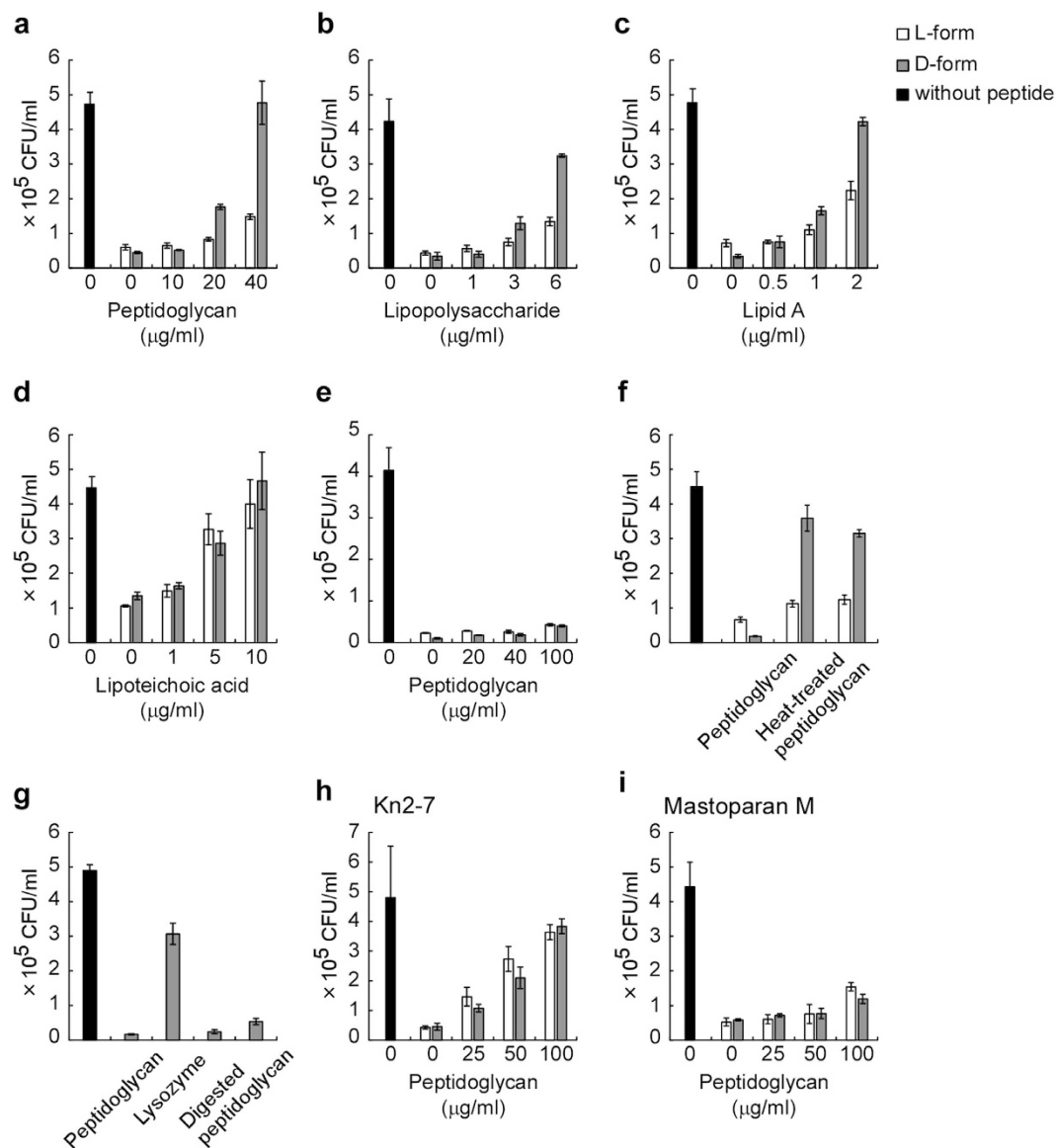


Figure 5. *S. aureus* peptidoglycan and *E. coli* lipopolysaccharide preferentially inhibit the antimicrobial activity of D-form KKLKLLLLLKLK-NH₂. Antimicrobial activities of D-form KKLKLLLLLKLK-NH₂ (1.9 $\mu\text{g/ml}$) and L-form KKLKLLLLLKLK-NH₂ (7.5 $\mu\text{g/ml}$) against *S. aureus* were examined in the presence of the indicated concentrations of peptidoglycan from *S. aureus* (a), lipopolysaccharide from *E. coli* (b), lipid A (c), lipoteichoic acid from *S. aureus* (d), and peptidoglycan from *E. coli* (e). (f) Antimicrobial activities of D-form KKLKLLLLLKLK-NH₂ (1.9 $\mu\text{g/ml}$) and L-form KKLKLLLLLKLK-NH₂ (7.5 $\mu\text{g/ml}$) against *S. aureus* were examined in the absence or presence of peptidoglycan (40 $\mu\text{g/ml}$) or heat-treated peptidoglycan (40 $\mu\text{g/ml}$) from *S. aureus*. (g) Antimicrobial activities of D-form KKLKLLLLLKLK-NH₂ (2.0 $\mu\text{g/ml}$) against *S. aureus* were examined in the absence or presence of 40 $\mu\text{g/ml}$ of peptidoglycan treated with lysozyme (digested peptidoglycan), 40 $\mu\text{g/ml}$ of peptidoglycan treated without lysozyme (peptidoglycan), or control buffer treated with lysozyme (lysozyme). Antimicrobial activities of D-form and L-form peptides of Kn2-7 (6.25 $\mu\text{g/ml}$) (h) or Mastoparan M (8 $\mu\text{g/ml}$) (i) against *S. aureus* were examined in the presence of the indicated concentrations of peptidoglycan from *S. aureus*. Gray bars and white bars represent CFUs in assay mixtures treated with D-form and L-form peptides, respectively. Black bars represent CFU in assay mixtures treated without peptide. The error bars represent the mean \pm standard deviations from triplicate plates. Concentrations of dimethyl sulfoxide in the assay mixtures were 0.15%.

activity of L-form (Fig. 5a). These observations highlight the potential for a specific interaction between D-form KKLKLLLLLKLK-NH₂ and peptidoglycan. In order to exclude the possibility that some contaminants, such as proteases, in peptidoglycan samples might affect the inhibitory effects, heat-treated peptidoglycan was used for the analysis. As shown in Fig. 5f, heat-treated peptidoglycan showed similar inhibitory effects on the antimicrobial activities with those of untreated peptidoglycan. To further confirm that peptidoglycan is a specific target of D-form KKLKLLLLLKLK-NH₂, the antimicrobial effects were investigated in the presence of lysozyme-digested

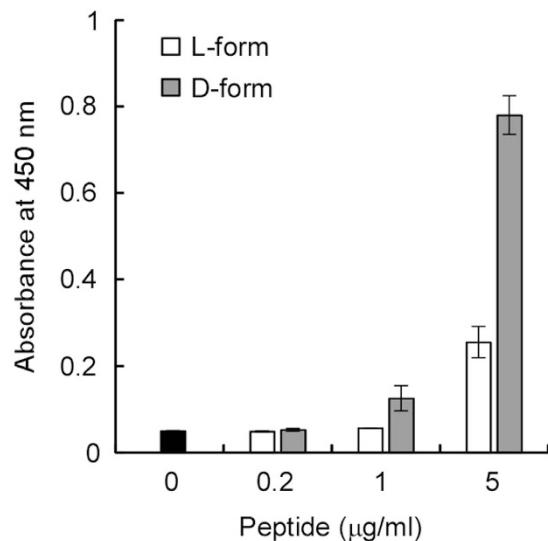


Figure 6. D-form KLKLLLLLKLK-NH₂ showed higher affinity for peptidoglycan than L-form KLKLLLLLKLK-NH₂. Peptidoglycan-coated wells were treated without (black bar) or with the indicated concentrations of biotin-labeled D-form KLKLLLLLKLK-NH₂ (gray bar) or biotin-labeled L-form KLKLLLLLKLK-NH₂ (white bar). Binding of biotin-labeled peptide to peptidoglycan was determined based on avidin-labeled peroxidase activity. The mean values with standard deviations of triplicate assays are shown.

peptidoglycans (Fig. 5g). The D-form did not show an inhibitory effect on antimicrobial activity. The antimicrobial activity of D-form KLKLLLLLKLK-NH₂ was preferentially inhibited by lipopolysaccharide prepared from *E. coli* (Fig. 5b). Furthermore, antimicrobial activity of D-form KLKLLLLLKLK-NH₂ was also preferentially inhibited by synthetic *E. coli* lipid A, a membrane anchor region of lipopolysaccharide (Fig. 5c). In contrast, lipoteichoic acid prepared from *S. aureus* inhibited the antimicrobial effect of both D-form and L-form peptides similarly, indicating that the inhibitory effect was not specific for the D-form peptide (Fig. 5d). Peptidoglycan prepared from *E. coli* had a weak inhibitory effect on the antimicrobial activity of D-form and L-form peptides (Fig. 5e). Taken together, these observations indicate that some cell surface components, such as *S. aureus* peptidoglycan, preferentially associate with D-form KLKLLLLLKLK-NH₂ rather than its L-form. Moreover, this preferential association accounts for higher antimicrobial activity of D-form KLKLLLLLKLK-NH₂ than that of the L-form.

In addition, inhibitory effects of peptidoglycan against Kn2-7 and Mastoparan M were examined. As shown in Fig. 5h, peptidoglycan shows significant inhibitory effects against the antimicrobial activities of both D-forms and L-forms of Kn2-7. Furthermore, peptidoglycan shows weak inhibitory effects to antimicrobial activities of Mastoparan M, and the inhibitory effect was not specific for the D-form peptide (Fig. 5i).

D-form KLKLLLLLKLK-NH₂ showed higher affinity for *S. aureus* peptidoglycan than L-form KLKLLLLLKLK-NH₂. The inhibitory effect of *S. aureus* peptidoglycan on the antimicrobial activity of D-form KLKLLLLLKLK-NH₂ suggested a specific interaction between these two molecules. To determine whether there was a direct association, direct binding between KLKLLLLLKLK-NH₂ and *S. aureus* peptidoglycan was examined. Biotin-labeled D-form or L-form KLKLLLLLKLK-NH₂ was added to multi-well plates that were coated with immobilized *S. aureus* peptidoglycan. Binding of biotin-labeled peptides to the D-form or L-form KLKLLLLLKLK-NH₂ was quantified using avidin-labeled peroxidase. As shown in Fig. 6, D-form KLKLLLLLKLK-NH₂ has a higher affinity for *S. aureus* peptidoglycan than the L-form counterpart.

Discussion

Incorporation of D-amino acids into antimicrobial peptides has been shown to improve their therapeutic efficacy; however, little is known about how the underlying mechanisms make them distinct from their L-form counterparts (reviewed in ref. 28). In this study we found that D-form KLKLLLLLKLK-NH₂ showed higher antimicrobial activity against both Gram-positive and Gram-negative bacteria, including *S. aureus* and *E. coli*, relative to its L-form counterpart. Moreover, the enhanced antimicrobial activity of the D-form was not due to its resistance to proteolytic degradation. D-form KLKLLLLLKLK-NH₂ showed higher affinity for *S. aureus* peptidoglycan than the L-form counterpart. Peptidoglycan and lipopolysaccharide prepared from *S. aureus* and *E. coli*, respectively, selectively inhibited the antimicrobial activities of D-form KLKLLLLLKLK-NH₂. Thus, specific interactions between D-form peptides and components of the bacterial cell wall may contribute to its elevated antimicrobial activity.

Cationic antimicrobial peptides target the negatively charged cell surface of microorganisms. In some cases, D-forms of naturally-occurring antimicrobial peptides have antimicrobial activities similar to those of L-form counterparts, and it is believed that the interaction between antimicrobial peptide and microbial cell surface is not due to specific, close interactions^{10,12}. This general notion is consistent with our observations of similar antimicrobial activities of the D-forms and L-forms of Mastoparan M, Kn2-7, and Temporin A. In addition,

D-form KLKLLLLLKLK-NH₂ showed similar activity to disrupt *S. aureus*-mimetic liposomes when compared to the L-form. These observations indicate that the interaction between antimicrobial peptides and anionic bacterial-type liposomes does not require close contact based on the structure, but charge-based interactions are important for antimicrobial activities. In contrast to the previous studies, our results showed that D-form KLKLLLLLKLK-NH₂ had a higher affinity for some cell surface compounds than its L-form counterpart, and the affinity of the D-form for bacterial surface components contributed to its antimicrobial activity. Our observations indicated that some specific, close contact between antimicrobial peptides and bacterial cell surface components increase antimicrobial activities in addition to charge-based contact. Peptidoglycan is consisted of sugars and peptides, and they are chiral components. The chiral portions of peptidoglycan might be involved in the association of D-form KLKLLLLLKLK-NH₂. It is noteworthy that high affinity of D-form KLKLLLLLKLK-NH₂ to cell surface components including peptidoglycan does not necessary indicate direct targeting. There might be mechanisms to facilitate peptide transfer to the plasma membrane, which determine the effective concentration.

Comparison of the D-4Leu and L-4Leu antimicrobial peptides revealed that the D-form had a greater tendency to bind to the biofilm exopolysaccharide alginate²⁹. This current study of KLKLLLLLKLK-NH₂ largely recapitulated these findings. To date, the molecular basis for the close interaction of D-form peptides with bacterial cell surface components remains unknown; however, the importance of precise structures of the bacterial molecules involved in these interactions has been shown. Antimicrobial activities of D-form KLKLLLLLKLK-NH₂ were preferentially inhibited by *S. aureus* peptidoglycan but not by *E. coli* peptidoglycan. This difference is likely based on the structural differences between *S. aureus* peptidoglycan and *E. coli* peptidoglycan.

Based on our observations, replacement of all L-amino acids with D-amino acids in an antimicrobial peptide may introduce structural changes that are beneficial for antimicrobial activity. It is important to note that not all antimicrobial peptides have distinct activities based on whether they are expressed as a D-form or L-form, and the number of these peptides may be fairly low. Future studies should focus on elucidating the specific interactions of the D-form modification with bacteria as well as the molecular basis underlying this this phenomenon. This will aid in the development of peptide therapeutics.

Methods

Reagents and antimicrobial peptides. Dimethyl sulfoxide, bovine serum albumin (fraction V), cardiolipin, L- α -phosphatidyl-DL-glycerol, peptidoglycan purified from *S. aureus*, lysozyme, and lipoteichoic acid purified from *S. aureus* were purchased from Sigma-Aldrich. Cholesterol, 2-dioleoyl-*sn*-glycero-3-phosphocholine, and 2-dioleoyl-*sn*-glycero-3-phosphoethanolamine were purchased from Avanti Polar Lipids Inc. Peptidoglycan purified from *E. coli* was purchased from InvivoGen. Calcein was purchased from Dojindo. Triton X-100 was purchased from Thermo Fisher Scientific. Lipopolysaccharide purified from *E. coli* 0111:B4 was purchased from List Biological Laboratories, Inc. Synthetic lipid A was purchased from Peptide Institute Inc. Ruby protein gel stain and Any kDTM precast polyacrylamide gels were purchased from Bio-Rad.

Antimicrobial peptides and biotin-labeled antimicrobial peptides were commercially synthesized by Hayashi Kasei, Thermo Fisher Scientific, and the Toray Research Center. C-terminals of the synthetic peptides were modified by amidation. All peptides were initially suspended in dimethyl sulfoxide.

Culture of bacteria and fungi. *S. aureus* (NBRC100910), *E. coli* W3110 (NBRC12713), *E. hirae* (NBRC3181), *C. albicans* (NBRC1385), and *S. Typhimurium* 14028s (ATCC14028)² were used in this study. *S. aureus*, *E. coli*, and *S. Typhimurium* were grown at 37 °C with aeration in Muller-Hinton II medium (BD Biosciences). *E. hirae* was grown at 37 °C in LB medium (Nacalai Tesque, Inc.) containing 0.5% glucose (w/v). *C. albicans* was grown at room temperature with aeration in YM medium (BD Biosciences). All experiments were conducted using bacterial cells and *C. albicans* in the logarithmic-phase of growth.

Determination of MIC. Bacterial suspensions in Muller-Hinton II medium were adjusted to an optical density of 550 nm (OD₅₅₀) = 0.0011. *C. albicans* suspensions in YM medium were adjusted to OD₆₅₀ = 0.033. Peptides were serially diluted in 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride, 0.2% bovine serum albumin, and 2.56% dimethyl sulfoxide. The peptide solution (100 μ l) was mixed with 100 μ l of bacteria or *C. albicans* suspensions. Bacterial cultures were incubated for one day at 37 °C. *C. albicans* cultures were incubated for two days at room temperature. Cell growth was monitored optically and the MIC was determined.

Assay for antimicrobial activity. Bacteria and *C. albicans* were suspended in growth medium. Peptides suspended in dimethyl sulfoxide were serially diluted in 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride, 0.2% bovine serum albumin as described previously¹³. Concentrations of dimethyl sulfoxide in the assay mixtures are indicated in the figure legends. In order to examine the effects of bovine serum albumin on the assay, 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride, was used for the dilution of peptides. Peptide solution (500 μ l) was added to 500 μ l of bacteria suspensions and then the mixture was incubated at 37 °C for 10 min. In order to examine the effects of culture medium components on the assay, bacteria suspension was prepared with 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride. Alternatively, 500 μ l of peptide solution was added to 500 μ l of *C. albicans* suspensions and the mixture was incubated at room temperature for 10 min. The inhibitory effects of the bacterial components were analyzed by incubating 450 μ l of *S. aureus* suspension with 500 μ l of peptide solution plus 50 μ l of inhibitor samples at 37 °C for 10 min. Then, the peptide/bacteria suspensions were diluted and plated onto LB agar, LB agar containing 0.5% glucose, or YM agar. After cultivation of the plates, colony forming units (CFU) in the peptide/bacteria suspension were calculated based on the average of triplicate plates.

Assay for membrane permeability. To examine membrane permeability, ethidium influx rates were examined as previously described^{30,31}. *S. aureus* suspension cultures were adjusted to an OD₆₀₀ of 0.4 in 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride and 0.2% bovine serum albumin. Then, peptide in dimethyl sulfoxide (8 μ l) or dimethyl sulfoxide alone (8 μ l) was added to 2 ml of *S. aureus* suspensions. At 30 sec after the addition of peptide, ethidium bromide was added to a final concentration of 5 μ g/ml, and fluorescence of the ethidium-nucleic acid complex was monitored using a RF-5300PC spectrofluorometer (Shimadzu). Excitation and emission wavelengths were 545 nm with 5 nm slits and 600 nm with 10 nm slits, respectively.

Preparation of liposomes containing calcein and assay for resistance to antimicrobial peptides. Liposomes were prepared as previously described with some modifications^{18,32}. *S. aureus*-mimetic liposomes^{17,26} and mammalian-mimetic liposomes²⁷ were prepared by combining phospholipid mixtures (cardiolipin: L- α -phosphatidyl-DL-glycerol = 1:3 (mol/mol)) and 2-dioleoyl-*sn*-glycero-3-phosphocholine: 1,2-dioleoyl-*sn*-glycero-3-phosphoethanolamine:Cholesterol = 4:2:3 (mol/mol/mol)), respectively. A 50 mM calcein solution was prepared by mixing 100 mg calcein with 3.1 ml of solution A (0.3 ml of 1 M Tris-HCl (pH 7.4), 0.9 ml of 5 M sodium hydroxide, and 13.8 ml of water). The liposomes were prepared by adding 1 ml of calcein solution to the dried lipid mixtures, followed by vortexing for 10 min at room temperature. The liposome solution (1 ml) was centrifuged at 3000 \times g for 10 min, and the precipitate was resuspended in 1 ml of 20 mM Tris-HCl (pH 7.4) containing 150 mM sodium chloride. The centrifugation step was repeated two more times. Finally, the precipitate was suspended in 100 μ l of 20 mM Tris-HCl (pH 7.4) containing 150 mM sodium chloride.

Liposome suspensions were prepared by diluting of 1 μ l of liposomes into 40 ml of 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride. Peptides were serially diluted in 10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride and 1% dimethyl sulfoxide. Peptide samples (20 μ l) were added to 2 ml of liposome suspension, and the mixtures were incubated at room temperature for 10 min. Calcein leakage from the liposomes was examined using a RF-5300PC spectrofluorometer. Excitation and emission wavelengths were 490 nm and 520 nm (with a 5 nm slit width), respectively³².

Digestion and heat-inactivation of peptidoglycan. Peptidoglycan (120 μ g) prepared from *S. aureus* was added to 1 mg/ml of lysozyme in phosphate buffered saline (150 μ l)³³. Peptidoglycan without lysozyme and lysozyme without peptidoglycan were also prepared as controls. Samples were incubated overnight at 37 °C, and then incubated at 100 °C for 15 min to inactivate lysozyme. For heat-inactivation of peptidoglycan, 600 μ g of peptidoglycan suspended in water (300 μ l) was incubated at 100 °C for 15 min. The samples were sonicated for 10 sec at setting 1 using a Branson sonifier model S-150D. These samples were used as inhibitor samples for antimicrobial activity assays.

Peptidoglycan-binding assay. Peptidoglycan-binding assays were performed as previously described with some modifications^{34–36}. Peptidoglycan from *S. aureus* (100 μ g/ml) was suspended in 0.2% trifluoroacetic acid and sonicated twice for 10 sec at setting 1 using a Branson sonifier model S-150D. The peptidoglycan suspension (50 μ l) was used to coat the wells of a flat bottom 96-well microplate (Thermo Fisher Scientific). The plate was incubated at room temperature until the water evaporated. The plate was placed at 60 °C for 1 h to dry out completely, and then blocked with 200 μ l of 5 mg/ml bovine serum albumin in binding buffer (10 mM phosphate buffer (pH 6.0) containing 130 mM sodium chloride, 0.05% Tween 20, and 0.01% trifluoroacetic acid) at 37 °C for 2 h. The plate was washed four times with 200 μ l of binding buffer. Biotin-labeled peptides in 100 μ l of binding buffer containing 0.5% dimethyl sulfoxide were added to the wells and incubated at 37 °C for 2 h. Detection of biotin-labeled peptides was performed using Vectastain ABC reagent (Vector Laboratories) according to manufacturer's instructions. The wells were washed four times with binding buffer, then 100 μ l of avidin-labeled peroxidase was added to each well, and the plate was incubated at 37 °C for 1 h. The wells were washed again as described above. After washing, 100 μ l of 3, 3', 5, 5'-tetramethylbenzidine substrate was added and the plate was incubated at room temperature. After 10 min, the reaction was stopped by the addition of 100 μ l of 0.5 M sulfuric acid. Absorbance was measured at 450 nm.

References

- Zasloff, M. Antimicrobial peptides of multicellular organisms. *Nature* **415**, 389–395 (2002).
- Gunn, J. S. *et al.* PmrA-PmrB-regulated genes necessary for 4-aminoarabinose lipid A modification and polymyxin resistance. *Mol Microbiol* **27**, 1171–1182 (1998).
- Kawasaki, K., China, K. & Nishijima, M. Release of the lipopolysaccharide deacylase PagL from latency compensates for a lack of lipopolysaccharide aminoarabinose modification-dependent resistance to the antimicrobial peptide polymyxin B in *Salmonella enterica*. *Journal of bacteriology* **189**, 4911–4919 (2007).
- Vaara, M. & Viljanen, P. Binding of polymyxin B nonapeptide to gram-negative bacteria. *Antimicrobial agents and chemotherapy* **27**, 548–554 (1985).
- Matsuzaki, K. Why and how are peptide-lipid interactions utilized for self-defense? Magainins and tachyplesins as archetypes. *Biochimica et biophysica acta* **1462**, 1–10 (1999).
- Huang, H. W. Molecular mechanism of antimicrobial peptides: the origin of cooperativity. *Biochimica et biophysica acta* **1758**, 1292–1302 (2006).
- Shai, Y. Mechanism of the binding, insertion and destabilization of phospholipid bilayer membranes by alpha-helical antimicrobial and cell non-selective membrane-lytic peptides. *Biochimica et biophysica acta* **1462**, 55–70 (1999).
- Santamaria, C. *et al.* Bactericidal and antitoxic properties of short cationic peptides derived from a snake venom Lys49 phospholipase A2. *Antimicrobial agents and chemotherapy* **49**, 1340–1345 (2005).
- Hamamoto, K., Kida, Y., Zhang, Y., Shimizu, T. & Kuwano, K. Antimicrobial activity and stability to proteolysis of small linear cationic peptides with D-amino acid substitutions. *Microbiology and immunology* **46**, 741–749 (2002).
- Wade, D. *et al.* All-D amino acid-containing channel-forming antibiotic peptides. *Proceedings of the National Academy of Sciences of the United States of America* **87**, 4761–4765 (1990).
- Huang, J. *et al.* Inhibitory effects and mechanisms of physiological conditions on the activity of enantiomeric forms of an alpha-helical antibacterial peptide against bacteria. *Peptides* **32**, 1488–1495 (2011).

12. Chen, Y. *et al.* Comparison of biophysical and biologic properties of alpha-helical enantiomeric antimicrobial peptides. *Chemical biology & drug design* **67**, 162–173 (2006).
13. Yamada, K. & Natori, S. Purification, sequence and antibacterial activity of two novel sapecin homologues from *Sarcophaga embryonica* cells: similarity of sapecin B to charybdotoxin. *The Biochemical journal* **291** (Pt 1), 275–279 (1993).
14. Matsuyama, K. & Natori, S. Purification of three antibacterial proteins from the culture medium of NIH-Sape-4, an embryonic cell line of *Sarcophaga peregrina*. *The Journal of biological chemistry* **263**, 17112–17116 (1988).
15. Matsuyama, K. & Natori, S. Molecular cloning of cDNA for sapecin and unique expression of the sapecin gene during the development of *Sarcophaga peregrina*. *The Journal of biological chemistry* **263**, 17117–17121 (1988).
16. Bontems, F., Roumestand, C., Gilquin, B., Menez, A. & Toma, F. Refined structure of charybdotoxin: common motifs in scorpion toxins and insect defensins. *Science* **254**, 1521–1523 (1991).
17. Yamada, K. & Natori, S. Characterization of the antimicrobial peptide derived from sapecin B, an antibacterial protein of *Sarcophaga peregrina* (flesh fly). *The Biochemical journal* **298** Pt 3, 623–628 (1994).
18. Alvarez-Bravo, J., Kurata, S. & Natori, S. Novel synthetic antimicrobial peptides effective against methicillin-resistant *Staphylococcus aureus*. *The Biochemical journal* **302** (Pt 2), 535–538 (1994).
19. Okuyama-Nishida, Y. *et al.* Prevention of death in bacterium-infected mice by a synthetic antimicrobial peptide, L5, through activation of host immunity. *Antimicrobial agents and chemotherapy* **53**, 2510–2516 (2009).
20. Fritz, J. H. *et al.* The artificial antimicrobial peptide KLKLLLLLKLK induces predominantly a TH2-type immune response to co-injected antigens. *Vaccine* **22**, 3274–3284 (2004).
21. Schellack, C. *et al.* IC31, a novel adjuvant signaling via TLR9, induces potent cellular and humoral immune responses. *Vaccine* **24**, 5461–5472 (2006).
22. Cao, L. *et al.* Antibacterial activity and mechanism of a scorpion venom peptide derivative *in vitro* and *in vivo*. *PLoS one* **7**, e40135 (2012).
23. Li, M. L., Liao, R. W., Qiu, J. W., Wang, Z. J. & Wu, T. M. Antimicrobial activity of synthetic all-D mastoparan M. *International journal of antimicrobial agents* **13**, 203–208 (2000).
24. Mangoni, M. L. *et al.* Structure-function relationships of temporins, small antimicrobial peptides from amphibian skin. *European journal of biochemistry/FEBS* **267**, 1447–1454 (2000).
25. Matsuyama, K. & Natori, S. Mode of action of sapecin, a novel antibacterial protein of *Sarcophaga peregrina* (flesh fly). *Journal of biochemistry* **108**, 128–132 (1990).
26. Zhao, C. *et al.* RL-37, an alpha-helical antimicrobial peptide of the rhesus monkey. *Antimicrobial agents and chemotherapy* **45**, 2695–2702 (2001).
27. Won, A., Ruscito, A. & Ianoul, A. Imaging the membrane lytic activity of bioactive peptide latarcin 2a. *Biochimica et biophysica acta* **1818**, 3072–3080 (2012).
28. Li, H. *et al.* Roles of d-Amino Acids on the Bioactivity of Host Defense Peptides. *International journal of molecular sciences* **17** (2016).
29. Yin, L. M., Lee, S., Mak, J. S., Helmy, A. S. & Deber, C. M. Differential binding of L- vs. D-isomers of cationic antimicrobial peptides to the biofilm exopolysaccharide alginate. *Protein and peptide letters* **20**, 843–847 (2013).
30. Murata, T., Tseng, W., Guina, T., Miller, S. I. & Nikaido, H. PhoPQ-mediated regulation produces a more robust permeability barrier in the outer membrane of *Salmonella enterica* serovar typhimurium. *Journal of bacteriology* **189**, 7213–7222 (2007).
31. Kawasaki, K. & Manabe, T. Latency of the lipid A deacylase PagL is involved in producing a robust permeation barrier in the outer membrane of *Salmonella enterica*. *Journal of bacteriology* **192**, 5837–5840 (2010).
32. Yoneyama, F. *et al.* Peptide-lipid huge toroidal pore, a new antimicrobial mechanism mediated by a lactococcal bacteriocin, lactacin Q. *Antimicrobial agents and chemotherapy* **53**, 3211–3217 (2009).
33. Baba, K., Okada, M., Kawano, T., Komano, H. & Natori, S. Purification of sarcotoxin III, a new antibacterial protein of *Sarcophaga peregrina*. *Journal of biochemistry* **102**, 69–74 (1987).
34. Senyurek, I. *et al.* Dermcidin-derived peptides show a different mode of action than the cathelicidin LL-37 against *Staphylococcus aureus*. *Antimicrobial agents and chemotherapy* **53**, 2499–2509 (2009).
35. Udompetcharaporn, A. *et al.* Identification and characterization of a QM protein as a possible peptidoglycan recognition protein (PGRP) from the giant tiger shrimp *Penaeus monodon*. *Developmental and comparative immunology* **46**, 146–154 (2014).
36. Shi, X. Z., Zhao, X. F. & Wang, J. X. A new type antimicrobial peptide astacidin functions in antibacterial immune response in red swamp crayfish *Procambarus clarkii*. *Developmental and comparative immunology* **43**, 121–128 (2014).

Acknowledgements

We thank Dr. Shunji Natori for his encouragement to this work. This work was supported in part by JSPS KAKENHI Grant number 26460179.

Author Contributions

T.M. performed experiments and wrote the manuscript. K.K. conducted the research and wrote the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Manabe, T. and Kawasaki, K. D-form KLKLLLLLKLK-NH₂ peptide exerts higher antimicrobial properties than its L-form counterpart via an association with bacterial cell wall components. *Sci. Rep.* **7**, 43384; doi: 10.1038/srep43384 (2017).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2017