

SCIENTIFIC REPORTS



OPEN

Influence of leaf vein density and thickness on hydraulic conductance and photosynthesis in rice (*Oryza sativa* L.) during water stress

Muhammad Adnan Tabassum¹, Guanglong Zhu¹, Abdul Hafeez², Muhammad Atif Wahid³, Muhammad Shaban³ & Yong Li¹

Received: 06 April 2016

Accepted: 18 October 2016

Published: 16 November 2016

The leaf venation architecture is an ideal, highly structured and efficient irrigation system in plant leaves. Leaf vein density (LVD) and vein thickness are the two major properties of this system. Leaf laminae carry out photosynthesis to harvest the maximum biological yield. It is still unknown whether the LVD and/or leaf vein thickness determines the plant hydraulic conductance (K_{plant}) and leaf photosynthetic rate (A). To investigate this topic, the current study was conducted with two varieties under three PEG-induced water deficit stress (PEG-IWDS) levels. The results showed that PEG-IWDS significantly decreased A , stomatal conductance (g_s), and K_{plant} in both cultivars, though the IR-64 strain showed more severe decreases than the Hanyou-3 strain. PEG-IWDS significantly decreased the major vein thickness, while it had no significant effect on LVD. A , g_s , and K_{plant} were positively correlated with each other, and they were negatively correlated with LVD. A , g_s and K_{plant} were positively correlated with the inter-vein distance and major vein thickness. Therefore, the decreased photosynthesis and hydraulic conductance in rice plants under water deficit conditions are related to the decrease in the major vein thickness.

Photosynthesis is an important physiological process that is very sensitive to abiotic stresses^{1,2}. Diffusive (stomatal or mesophyll conductance) and biochemical impairments are considered two major responses that decrease photosynthesis under drought conditions^{3,4}. Stomatal conductance (g_s) is a fundamental process required for CO_2 acquisition and is regulated by stomatal opening and closing^{5,6}. A decreasing leaf turgor pressure and an increasing vapor pressure deficit (VPD) closes the stomata rapidly in response to water deficit condition⁷. Thus, stomatal limitation is a key cause of the decrease in A that occurs under water-limited conditions^{7,8}.

The water transportation capacity of plants is known as the plant hydraulic conductance^{9,10}, which is determined by the root, stem and leaf hydraulic conductance (K_{leaf})¹¹. The root contribution ranges from one-third to one-half of the internal plant resistances^{12,13}. The transpiration rate (E) or stomatal conductance exhibit significant and linear correlations with K_{plant} in a number of higher plants and rice plant^{14–18}. Therefore, the capacity of water transport system controls the plant growth as it maintains the hydraulic link between the roots and leaves¹⁹.

The leaf hydraulic architecture is the key location for gas exchange between the plant and its environment^{20,21}, and extra-vascular resistance imposes one-quarter or higher resistance ($\geq 30\%$) in K_{leaf} ^{22,23}. A decrease in K_{leaf} leads to stomatal closure, which reduces photosynthesis^{24,25}. Therefore, a strong correlation has been observed between g_s and K_{leaf} ^{22,23,26}. The leaf venation architecture is a perfect illustration of a highly efficient irrigation structure^{27,28}. Veins are made up of phloem and xylem vessels implanted in parenchyma, rarely in sclerenchyma, that are wrapped in bundle sheath cells. Leaf veins in monocots of the *Poaceae* family are divided into three categories (major, minor and transverse veins) in addition to the leaf midrib, and are different in sizes and

¹Ministry of Agriculture Key Laboratory of Crop Ecophysiology and Farming System in the Middle Reaches of the Yangtze River, College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, China.

²Cotton Physiology Lab for Efficient Production, College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, China. ³National Key Laboratory of Crop Genetic Improvement, College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, China. Correspondence and requests for materials should be addressed to Y.L. (email: liyong@mail.hzau.edu.cn)

| Varieties | Treatment | A ($\mu\text{mol m}^{-2}\text{s}^{-1}$) | g_s ($\text{mol m}^{-2}\text{s}^{-1}$) | C_i ($\mu\text{mol mol}^{-1}$) | E ($\text{mmol m}^{-2}\text{s}^{-1}$) | Ψ_{leaf} (MPa) | K_{plant} ($\text{mmol m}^{-2}\text{s}^{-1}\text{MPa}^{-1}$) |
|---------------|-------------|---|--|------------------------------------|---|----------------------------|---|
| Hanyou-3 | WWC | 20.2 ± 0.3a | 0.37 ± 0.04a | 236 ± 4 | 6.29 ± 0.86a | -1.48 ± 0.03bc | 4.26 ± 0.58a |
| | PEG-IWDS5% | 19.9 ± 0.3a | 0.27 ± 0.01ab | 258 ± 3 | 6.45 ± 0.31a | -1.37 ± 0.02a | 4.90 ± 0.24a |
| | PEG-IWDS10% | 14.9 ± 0.4b | 0.32 ± 0.00ab | 312 ± 4 | 4.07 ± 0.23ab | -1.37 ± 0.03ab | 3.41 ± 0.19ab |
| | PEG-IWDS15% | 14.6 ± 0.9b | 0.13 ± 0.03b | 277 ± 27 | 2.82 ± 0.04b | -1.70 ± 0.02c | 2.13 ± 0.03b |
| IR-64 | WWC | 20.8 ± 1.3a | 0.29 ± 0.04a | 258 ± 10 | 4.44 ± 0.32a | -1.57 ± 0.04ab | 2.83 ± 0.21a |
| | PEG-IWDS5% | 12.5 ± 0.4b | 0.21 ± 0.02ab | 263 ± 4 | 3.77 ± 0.49a | -1.48 ± 0.02a | 2.64 ± 0.35a |
| | PEG-IWDS10% | 12.4 ± 1.0b | 0.17 ± 0.02b | 260 ± 6 | 3.27 ± 0.46a | -1.44 ± 0.06ab | 2.98 ± 0.15a |
| | PEG-IWDS15% | 6.5 ± 0.6c | 0.15 ± 0.01b | 314 ± 3 | 1.30 ± 0.08b | -1.85 ± 0.02b | 0.88 ± 0.05b |
| ANOVA | | | | | | | |
| Treatment (T) | | ** | ** | ns | ** | ** | * |
| Variety (V) | | ns | ns | ns | ns | ns | * |
| T × V | | ** | ns | ns | ns | ns | ns |

Table 1. Effects of PEG-induced water deficit stress on photosynthesis (A), stomatal conductance (g_s), intercellular CO_2 concentration (C_i), transpiration rate (E) and leaf water potential (Ψ_{leaf}) of newly-developed leaves of two rice varieties at the vegetative stage. Water deficit stress was simulated by adding 5, 10 or 15% (W/V) PEG6000 to the nutrient solution. WWC, well-watered condition; PEG-IWDS, PEG-induced water deficit stress. The data are presented as the means ± SE with 3 replicates. ns, not significant; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. The data followed by the different letters of each variety within a single column are significant at $P < 0.05$ level.

functions^{29–33}. The major longitudinal veins run from the leaf lamina into the leaf sheath, while minor longitudinal veins mostly terminate at the junction of the leaf lamina and leaf sheath^{34–37}.

The leaf venation architecture has many functions, including mechanical support³⁸, sugars and hormone transportation³⁹, and replacement of water lost through E during photosynthetic processes²³. An enormous variation is found in the vein arrangement, size and density, and in the geometry of phloem and xylem vessels within the leaf vascular bundles. Thicker veins have a greater water transportation and sugar translocation capacity due to the greater number and/or size of the xylem and phloem vessels⁴⁰.

During the last two decades, numerous studies have been carried out to explore the relationship between K_{leaf} and leaf vein structure. The leaf vein length per unit leaf area is called as the vein length per unit area (VLA) or leaf vein density (LVD). Positive, negative^{41,42} and no correlations⁴³ have been found between LVD and K_{leaf} in these studies. In our previous study, a significant positive correlation between K_{leaf} , K_{plant} and LVD was observed in rice plants under well watered condition, but no relationship was observed between K_{leaf} and K_{plant} under drought stress, although K_{leaf} showed a positive correlation with LVD⁴⁴.

It is still unknown which vein property in rice crops is more closely related to the leaf photosynthetic rate and K_{plant} under drought conditions. The current study had the following objectives: (i) to elaborate the effects of PEG-induced water deficit stress (PEG-IWDS) on gas exchange parameters; (ii) to elaborate whether the LVD or leaf vein thickness is related to K_{plant} ; and (iii) to elaborate whether the LVD or leaf vein thickness is related to gas exchange parameters under PEG-IWDS.

Results

PEG-induced water deficit stress decreased the gas exchange parameters. More severe depression was observed in the IR-64 variety than in the Hanyou-3 variety (Table 1). IR-64 had a significant decrease in A under all PEG-IWDS conditions, while A was decreased non significantly under 5% PEG-IWDS in Hanyou-3. Under 15% PEG-IWDS, A was decreased by 68.7% in IR-64 compared with a smaller decrease of 27.8% in Hanyou-3. Hanyou-3 showed a significant decrease in g_s under 15% PEG-IWDS, and IR-64 revealed a significant decrease in g_s under both the 10% and 15% PEG-IWDS conditions. The intercellular CO_2 concentration (C_i) increased under all stress levels in both varieties, but a significant increase was observed in IR-64 under 15% PEG-IWDS. Hanyou-3 and IR-64 both showed a significant decrease in E under 15% PEG-IWDS, but a more severe decrease (70.7%) was observed in IR-64 than Hanyou-3 (55.2%). The decrease in leaf water potential (Ψ_{leaf}) was only significant in IR-64 under 15% PEG-IWDS. There was a positive relationship between A and g_s (Fig. 1a). K_{plant} showed a significant decrease in both varieties under 15% PEG-IWDS, although IR-64 showed a more severe decrease (68.8%) than Hanyou-3 (49.9%) (Table 1). A and g_s showed positive correlations with K_{plant} (Fig. 1b,c).

Leaf size was decreased under all PEG-IWADS conditions in Hanyou-3, but in IR-64, it was only significantly decreased under 15% PEG-IWDS (Table 2). Compared with IR-64, a more severe decrease in leaf size was observed in Hanyou-3 under all PEG-IWDS conditions. Leaf size showed positive correlation with the major, and minor vein thickness as well as with inter-vein distances (IVD), while it showed negative correlations with LVD and LVD_{minor} (data not shown). LVD and LVD_{minor} showed non-significant increases in both varieties under all PEG-IWADS conditions. Interestingly, IR-64 had a higher leaf vein density than Hanyou-3 under all treatment conditions. On the other hand, IVD decreased non-significantly under all treatment conditions in both varieties, and Hanyou-3 had a higher IVD than IR-64. LVD had a negative correlation with A and K_{plant} , but a non-significant relationship with g_s (Fig. 2). Similarly, LVD_{minor} had negative correlations with A and K_{plant} (Fig. 3a,c), but g_s was not significantly related to LVD_{minor} (Fig. 3b). IVD was positively correlated with A and K_{plant} (Fig. 3d,f) and was not related to g_s (Fig. 3e).

Major vein thickness decreased significantly in Hanyou-3 under 10 and 15% PEG-IWDS while a non-significant decrease was observed in IR-64 under all PEG-IWDS conditions (Table 3). Minor vein thickness

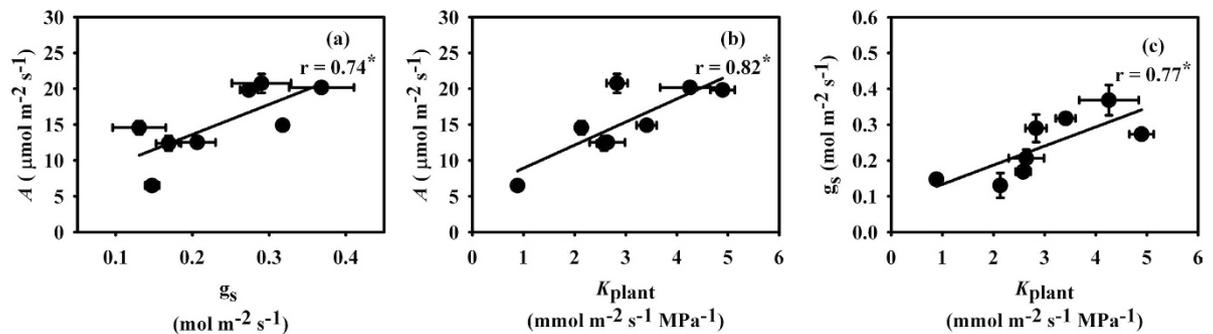


Figure 1. Relationships between photosynthesis (A) and stomatal conductance (g_s) (a) and plant hydraulic conductance (K_{plant}) (b) and relationship between g_s and K_{plant} (c). The data are presented as the mean values of 3 replicates. * $P < 0.05$.

| Varieties | Treatment | Single leaf area (cm ²) | LVD (no. mm ⁻¹) | LVD _{minor} (no. mm) | IVD (mm) |
|---------------|-------------|-------------------------------------|-----------------------------|-------------------------------|----------------|
| Hanyou-3 | WWC | 72.3 ± 1.1a | 3.71 ± 0.09a | 2.96 ± 0.10a | 0.270 ± 0.007a |
| | PEG-IWDS5% | 53.3 ± 2.8b | 3.87 ± 0.13a | 3.10 ± 0.12a | 0.260 ± 0.008a |
| | PEG-IWDS10% | 52.5 ± 1.6bc | 4.28 ± 0.10a | 3.49 ± 0.10a | 0.234 ± 0.006a |
| | PEG-IWDS15% | 40.0 ± 1.4c | 4.13 ± 0.11a | 3.29 ± 0.08a | 0.243 ± 0.006a |
| IR-64 | WWC | 25.9 ± 0.4a | 4.64 ± 0.07a | 3.67 ± 0.05a | 0.216 ± 0.003a |
| | PEG-IWDS5% | 21.7 ± 0.9ab | 4.71 ± 0.23a | 3.65 ± 0.23a | 0.215 ± 0.010a |
| | PEG-IWDS10% | 22.7 ± 1.2ab | 4.99 ± 0.16a | 3.88 ± 0.15a | 0.201 ± 0.006a |
| | PEG-IWDS15% | 19.4 ± 0.2b | 5.21 ± 0.17a | 4.08 ± 0.14a | 0.193 ± 0.006a |
| ANOVA | | | | | |
| Treatment (T) | | *** | ns | ns | ns |
| Variety (V) | | *** | ** | * | * |
| T × V | | * | ns | ns | ns |

Table 2. Effects of PEG-induced water deficit stress on the single leaf area, leaf vein density (LVD), minor leaf vein density (LVD_{minor}), and inter-vein distance (IVD) of newly developed leaves of two rice varieties at the vegetative stage. Water deficit stress was simulated by adding 5, 10 or 15% (W/V) PEG6000 to the nutrient solution. WWC, well-watered condition; PEG-IWDS, PEG-induced water deficit stress. The data are presented as the means ± SE with 3 replicates. ns, not significant; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. The data followed by the different letters of each variety within a single column are significant at $P < 0.05$ level.

decreased significantly in Hanyou-3 under 5% PEG-IWDS but non-significantly decreased under 10 and 15% PEG-IWDS. Moreover, the decrease in minor vein thickness was non-significant in IR-64 under all PEG-IWDS conditions. Major vein thickness showed a positive correlation with A , g_s and K_{plant} (Fig. 4). However, leaf minor vein thickness did not show any significant relationship with gas exchange or K_{plant} (data not shown).

Discussion

Stomatal closure in response to water deficit stress will limit photosynthesis by restricting CO₂ entry from the ambient environment into the intercellular air spaces of mesophyll cells^{45–48}. Moreover, decreased g_m and impaired biochemical processes are non-stomatal limitations to photosynthesis that occur under severe or long-term water deficit conditions^{49–51}. It is therefore logical that photosynthesis exhibited positive correlations with g_s or/and g_m in previous studies^{52–55}. In the current study, A was also positively correlated with g_s (Fig. 1a). Chaves *et al.*⁷ reported that increased VPD and reduced turgor potential are major causes of stomatal closure under water-limited conditions. However, it is the boundary layers (leaf and canopy), as well as the driving force (VPD), that determine E , while K_{plant} determines the water potential at that E ^{9,10}. Thus, a high K_{plant} can maintain a high g_s and the consequent A without leading to desiccation of the plant leaves^{14,56–59}. Linear correlations between K_{plant} and E or g_s were previously found in a number of higher plant species^{15–18}. K_{leaf} is a major component of K_{plant} ; the positive correlation between K_{plant} and gas exchange may be related to K_{leaf} . In the present study, the positive correlations between g_s , A and K_{plant} suggest that K_{plant} is one of the key regulators of photosynthesis (Fig. 1).

The environmental signals present before and during leaf development determines the vein traits, like other leaf traits including leaf size and stomatal density^{60,61}. Plasticity in vein traits was observed within the canopy and across environments for a given plant species. In this study, plasticity in leaf size, LVD, IVD and vein thickness were also observed under different PEG-IWDS conditions. Sack *et al.*⁶² suggested that LVD has a key influence on hydraulic conductance, g_s and A , and LVD is positively correlated with A . In the present study, A was negatively correlated with LVD and LVD_{minor} (Figs 2a and 3a) and positively correlated with IVD (Fig. 3d). This negative relationship between A and LVD is in accordance with negative relationships reported in angiosperms^{63–67}, but

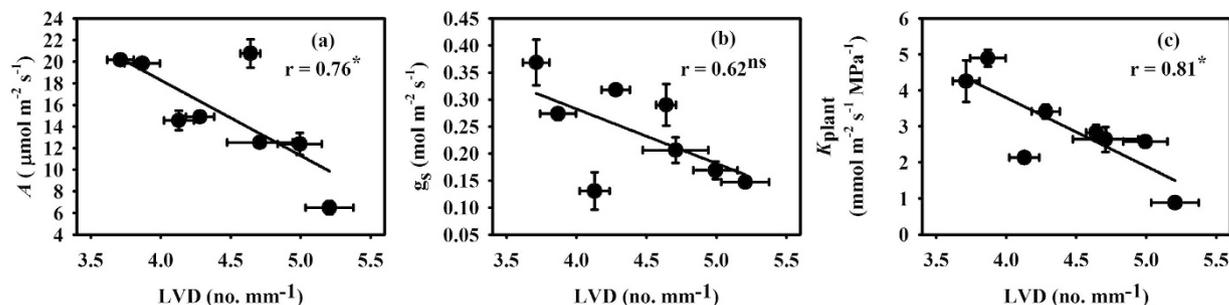


Figure 2. Relationship of photosynthesis (A) (a), stomatal conductance (g_s) (b) and plant hydraulic conductance (K_{plant}) (c) with leaf vein density (LVD). The data are presented as the mean values of 3 replicates. ns, not significant; $*P < 0.05$.

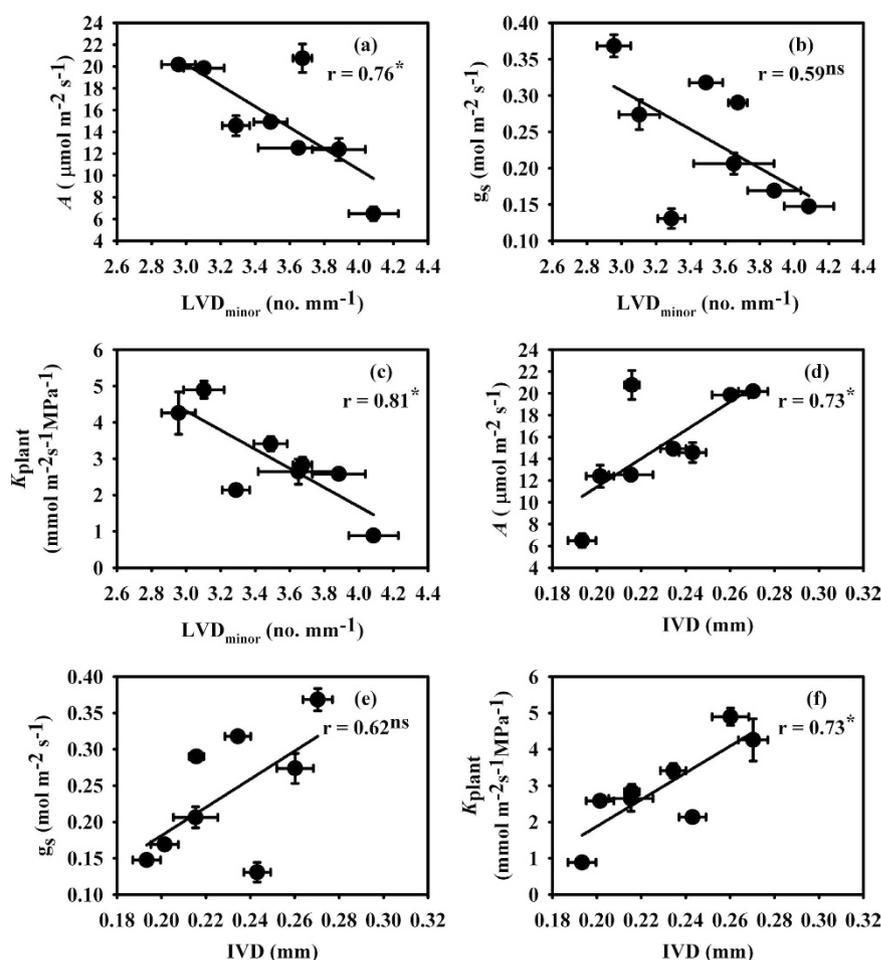


Figure 3. Relationships of photosynthesis (A) (a), stomatal conductance (g_s) (b) and plant hydraulic conductance (K_{plant}) (c) with minor leaf vein density ($\text{LVD}_{\text{minor}}$) and inter vein distance (IVD) (d–f). The data are presented as the mean values of 3 replicates. ns, not significant; $*P < 0.05$.

different from the study by Xiong *et al.*⁶⁸, who did not observe any relationship between g_s , A and LVD during studies of the *Oryza* genus under well-watered condition.

Rice leaves are small and have more highly lobed mesophyll cells than C_4 crop species⁶⁹. They also have a lower LVD than C_4 crops due to the higher number of mesophylls between veins. C_4 plants, such as *Setaria viridis* and sorghum, have seven veins per millimeter, but rice has fewer than six veins per millimeter⁷⁰. Although rice (C_3) and maize (C_4) both belong to the tropical-warm temperate grass family, rice has higher rates of photorespiration. This higher rate of photorespiration decreases the photosynthetic capacity by 30–35% at 30–35°C ambient temperature⁷¹, and drought conditions make this more severe, so rice does not attain the full potential photosynthesis like C_4 plants.

| Varieties | Treatment | Major vein thickness (mm) | Minor vein thickness (mm) |
|---------------|-------------|---------------------------|---------------------------|
| Hanyou-3 | WWC | 0.256 ± 0.006a | 0.120 ± 0.007a |
| | PEG-IWDS5% | 0.238 ± 0.003a | 0.093 ± 0.002b |
| | PEG-IWDS10% | 0.207 ± 0.006b | 0.100 ± 0.001ab |
| | PEG-IWDS15% | 0.172 ± 0.002c | 0.101 ± 0.001ab |
| IR-64 | WWC | 0.177 ± 0.006a | 0.098 ± 0.002a |
| | PEG-IWDS5% | 0.158 ± 0.004a | 0.092 ± 0.001a |
| | PEG-IWDS10% | 0.168 ± 0.003a | 0.092 ± 0.001a |
| | PEG-IWDS15% | 0.152 ± 0.002a | 0.096 ± 0.001a |
| ANOVA | | | |
| Treatment (T) | | *** | ns |
| Varieties (V) | | *** | ns |
| T × V | | * | ns |

Table 3. Effects of PEG-induced water deficit stress on the leaf major and minor vein thickness of newly developed leaves of two rice varieties at the vegetative stage. Water deficit stress was simulated by adding 5, 10 or 15% (W/V) PEG6000 to the nutrient solution. WWC, well-watered condition; PEG-IWDS, PEG-induced water deficit stress. The data are presented as the means ± SE with 3 replicates. ns, not significant; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. The data followed by the different letters of each variety within a single column are significant at $P < 0.05$ level.

K_{plant} was negatively correlated with LVD (Fig. 2c), while it had a positive correlation with IVD (Fig. 3f). Mesophyll cells are more numerous in C_3 than C_4 plants, which increases IVD in C_3 plants^{70,72} and reduces their K_{leaf} ^{23,43}. Smillie *et al.*⁷³ reported that IVD of rice plants is more dependent on cell size than cell number, which suggests that the lower IVD under water deficit conditions is mostly a result of more tightly packed, small mesophyll cells. The tightly packed mesophyll cells in smaller leaves under water deficit (Table 2) would produce more resistance in the apoplastic pathway for water transport in leaves, which would decrease K_{leaf} and the subsequent K_{plant} .

Vein size also decreases under drought stress in addition to leaf vein differentiation. Martre and Durand⁷⁴ reported that the vascular tissue is composed of xylem and phloem cells, and it carries out the transportation of different compounds. The flow rate of this transportation is determined by the size of the xylem and phloem cells. Decreases in the diameters of the xylem and phloem vessels were observed in *Ctenanthe setosa*, *Vigna unguiculata* and *Triticum aestivum* under water deficient conditions^{75–77}, likely because the thin xylem vessels provide protection from cavitation under water-limited conditions⁷⁸. In the present study, the major and minor vein thicknesses were also decreased under PEG-IWDS. The Hanyou-3 and IR-64 varieties showed more severe decreases in the major vein thickness (32.8% and 14.1%) than in the minor vein thickness (15.9% and 1.3%) under 15% PEG-IWDS (Table 3). The major, minor (longitudinal) and transverse veins have different sizes and functions^{29–31,33}. The major leaf veins are the main supply lines for receiving water directly from the roots via the stem and leaf sheath, as they run from the leaf blade into the sheath while minor veins terminate at the junction of the leaf blade and sheath^{34–37}. Water absorbed by the roots rises through the major veins from the leaf base to the leaf tips. After exiting the major veins, water reaches the minor veins via transverse veins and is finally distributed to mesophyll cells or is used for transpiration via stomata^{31–33}. Ocheltree *et al.*⁷⁹ suggested that g_s is strongly correlated with extra vascular resistance (outside large veins) under normal water regimes, while large vein resistance has a strong correlation with g_s under drought conditions. The current study suggests that the decreased major vein thickness that occurred under PEG-IWDS would increase the major vein resistance and restrict water uptake from the roots to leaves, and hence decreased K_{plant} and subsequently g_s and A .

Based on the present findings, we conclude that PEG-IWDS decreases K_{plant} , photosynthesis, leaf vein thickness and IVD, while it increases LVD and LVD_{minor} . LVD is negatively correlated with K_{plant} and photosynthesis, while major vein thickness is positively correlated with K_{plant} , g_s and A under PEG-IWDS condition in rice crops.

Materials and Methods

Plant materials. Two rice cultivars, Hanyou-3 and IR-64, were selected because they had different drought tolerances with regard to photosynthesis in previous study. Hanyou-3 is considered drought-tolerant, while IR-64 is considered drought-sensitive. Seeds were surface-sterilized for 90 minutes using 10% H_2O_2 , then washed with tap water to remove any residual H_2O_2 . The seeds were germinated on moist filter paper until the radical emerged in the laboratory, then they were transferred to a seedling tray with tap water under natural environmental conditions. Seedlings were supplied with 1/8th-strength Hoagland solution on the fifth day of germination to avoid nutrient deficiency. Seedlings were transplanted after fifteen days of germination. Each bucket contained 10.5 L Hoagland solution. Seedlings were transplanted using a split block design such that each bucket had four seedlings of each variety. This experiment had six replicates and four treatments: the well-watered condition (WWC) and 5%, 10% and 15% (w/v) PEG-IWDS. Treatments were applied when seedlings reached 40 days of age. The composition of the full strength nutrient solution was as follows: macronutrients ($mg\ l^{-1}$): 40 N as $(NH_4)_2SO_4$ and $Ca(NO_3)_2$, 10 P as KH_2PO_4 , 40 K as K_2SO_4 and KH_2PO_4 , and 40 Mg as $MgSO_4$; micronutrients ($mg\ l^{-1}$): 2.0 Fe as Fe-EDTA, 0.5 Mn as $MnCl_2 \cdot 4H_2O$, 0.05 Mo as $(NH_4)_6Mo_7O_{24} \cdot 4H_2O$, 0.2 B as H_3BO_3 , 0.01 Zn as $ZnSO_4 \cdot 7H_2O$, 0.01 Cu as $CuSO_4 \cdot 5H_2O$, 2.8 Si as $Na_2SiO_3 \cdot 9H_2O$. Dicyandiamide was added to the nutrient solution as a nitrification

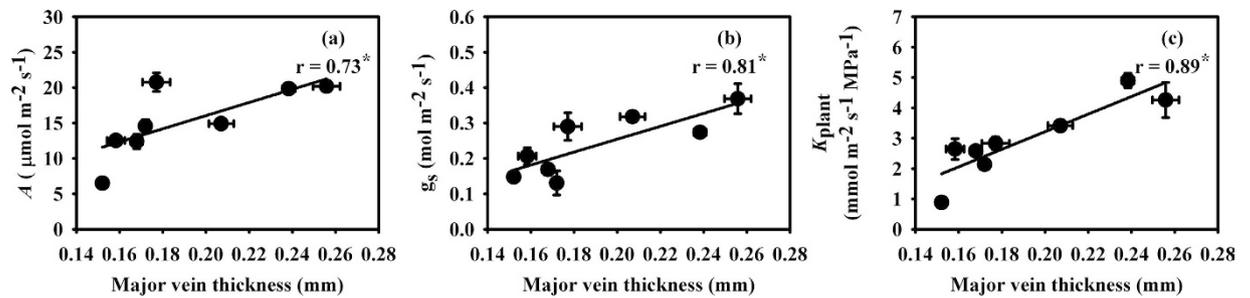


Figure 4. Relationships of photosynthesis (A) (a), stomatal conductance (g_s) (b) and plant hydraulic conductance (K_{plant}) (c) with leaf major vein thickness. The data are presented as the mean values of 4 replicates. * $P < 0.05$.

inhibitor. Solutions were changed every fifth day, and the pH was maintained at 5.50 ± 0.05 every day by adding 0.1 mol L^{-1} HCl or NaOH. The experiment was conducted under natural environmental conditions in Huazhong Agricultural University (114.37E, 30.48N) Wuhan, Hubei, China.

Gas exchange measurements. The gas exchanges were measured inside a growth chamber to avoid the fluctuations of the outdoor environment. The photosynthetic photon flux density (PPFD) was controlled to $1,000 \mu\text{mol m}^{-2} \text{s}^{-1}$ using T5 fluorescent lamps and halogen incandescent lamps fixed on a down and upward moving panel. There were three fans built in the roof of the growth chamber to avoid over-heating of the growth chamber, and the air temperature was set to $30/25^\circ\text{C}$ day/night with 11 h photoperiod. The relative humidity in the growth chamber was controlled at 65%.

Leaf area measurement. Three newly expanded leaves for each variety and replicate were detached, followed by leaf area measurement using a leaf area meter (Li-Cor 3000 C, Li-Cor, NE, USA).

Leaf vein density measurement. Rice leaf veins were divided into three categories based on their size (i.e., midrib, major and minor veins) to calculate the leaf vein density⁷³. One centimeter leaf sections were excised with a razor blade from the middle portion of newly-developed leaves after measuring the leaf width. These sections were immediately immersed in tap water and carried to a laboratory to observe all visible longitudinal leaf vein numbers. In the laboratory, all visible leaf veins (sum of the midrib, major and minor leaf veins) were counted under 40x magnification using a light microscope (SA3300, Beijing Tech Instrument Co., Ltd, Beijing, China). IVD was calculated by dividing the leaf width with the respective total longitudinal leaf vein numbers. LVD was calculated as total vein length per leaf area, and $\text{LVD}_{\text{minor}}$ was calculated as the total minor vein length per leaf area.

Measurement of plant hydraulic conductance. During the gas exchange measurements, newly and fully developed leaves were used to measure the day time leaf water potential using a WP4C Dewpoint Potential Meter (Decagon, Pullman, WA, USA). K_{plant} was calculated following the formula described by Brodribb and Holbrook⁸¹:

$$K_{\text{plant}} = E / (\Psi_{\text{solution}} - \Psi_{\text{leaf}}) \quad (1)$$

where Ψ_{solution} was 0 for WWC, and was -0.05 , -0.18 and -0.38 MPa, respectively, for the 5%, 10% and 15% PEG-IWDS.

Leaf vein thickness measurement. Minor vein thickness was measured for each side of the leaf (avoiding midribs) using a leaf thickness measuring instrument (YI-20030A, China Jiliang University), while major vein thickness was measured using a DTG03 digital thickness gauge (Digital Micrometers Ltd, Sheffield, UK).

Statistical analysis. One and two-way analyses of variance (ANOVA) were applied to assess the differences between treatments with Statistics 8.1 analytical software. Linear regression and correlation analysis were performed to test the possible correlations between the studied parameters using Sigma Plot 12 (SPSS Inc., Chicago, IL, USA).

References

- Hsiao, T. C. & Acevedo, E. Plant responses to water deficits, water-use efficiency, and drought resistance. *Agricultural meteorology* **14**, 59–84 (1974).
- Huang, B. Recent advances in drought and heat stress physiology of turfgrass — A review. *Acta Horticulturae* **661**, 185–192 (2004).
- Flexas, J., Bota, J., Loreto, F., Cornic, G. & Sharkey, T. Diffusive and metabolic limitations to photosynthesis under drought and salinity in C_3 plants. *Plant Biology* **6**, 269–279 (2004).
- Flexas, J. *et al.* Rapid variations of mesophyll conductance in response to changes in CO_2 concentration around leaves. *Plant, Cell & Environment* **30**, 1284–1298 (2007).
- Medici, L. O., Azevedo, R. A., Canellas, L. P., Machado, A. T. & Pimentel, C. Stomatal conductance of maize under water and nitrogen deficits. *Pesquisa Agropecuária Brasileira* **42**, 599–601 (2007).
- Dodd, I. C. Hormonal interactions and stomatal responses. *Journal of Plant Growth Regulation* **22**, 32–46 (2003).
- Chaves, M., Flexas, J. & Pinheiro, C. Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annals of Botany* **103**, 551–560 (2009).

8. Parida, A. K. & Das, A. B. Salt tolerance and salinity effects on plants: a review. *Ecotoxicology and Environmental Safety* **60**, 324–349 (2005).
9. Tyree, M. & Zimmermann, M. Xylem Structure and the Ascent of Sap, Springer. Berlin, Germany (2002).
10. Cowan, I. An electrical analogue of evaporation from, and flow of water in plants. *Planta* **106**, 221–226 (1972).
11. Martre, P. *et al.* Plasma membrane aquaporins play a significant role during recovery from water deficit. *Plant Physiology* **130**, 2101–2110 (2002).
12. Simonneau, T. & Inra, R. H. The use of tree root suckers to estimate root water potential. *Plant, Cell & Environment* **14**, 585–591 (1991).
13. Javot, H. & Maurel, C. The role of aquaporins in root water uptake. *Annals of Botany* **90**, 301–313 (2002).
14. Meinzer, F. & Grantz, D. Stomatal and hydraulic conductance in growing sugarcane: stomatal adjustment to water transport capacity. *Plant, Cell & Environment* **13**, 383–388 (1990).
15. Sperry, J. & Pockman, W. Limitation of transpiration by hydraulic conductance and xylem cavitation in *Betula occidentalis*. *Plant, Cell & Environment* **16**, 279–287 (1993).
16. Saliendra, N. Z., Sperry, J. S. & Comstock, J. P. Influence of leaf water status on stomatal response to humidity, hydraulic conductance, and soil drought in *Betula occidentalis*. *Planta* **196**, 357–366 (1995).
17. Meinzer, F. *et al.* Environmental and physiological regulation of transpiration in tropical forest gap species: the influence of boundary layer and hydraulic properties. *Oecologia* **101**, 514–522 (1995).
18. Hubbard, R., Ryan, M., Stiller, V. & Sperry, J. Stomatal conductance and photosynthesis vary linearly with plant hydraulic conductance in ponderosa pine. *Plant, Cell & Environment* **24**, 113–121 (2001).
19. Tyree, M. T. & Sperry, J. S. Vulnerability of xylem to cavitation and embolism. *Annual Review of Plant Biology* **40**, 19–36 (1989).
20. Whitehead, D. Regulation of stomatal conductance and transpiration in forest canopies. *Tree Physiology* **18**, 633–644 (1998).
21. Brodribb, T. J., Feild, T. S. & Jordan, G. J. Leaf maximum photosynthetic rate and venation are linked by hydraulics. *Plant Physiology* **144**, 1890–1898 (2007).
22. Sack, L., Cowan, P., Jaikumar, N. & Holbrook, N. The ‘hydrology’ of leaves: co-ordination of structure and function in temperate woody species. *Plant, Cell & Environment* **26**, 1343–1356 (2003).
23. Sack, L. & Holbrook, N. M. Leaf hydraulics. *Annual Review of Plant Biology* **57**, 361–381 (2006).
24. Sperry, J. S. Hydraulic constraints on plant gas exchange. *Agricultural and Forest Meteorology* **104**, 13–23 (2000).
25. Johnson, D. M., Woodruff, D. R., McCulloh, K. A. & Meinzer, F. C. Leaf hydraulic conductance, measured *in situ*, declines and recovers daily: leaf hydraulics, water potential and stomatal conductance in four temperate and three tropical tree species. *Tree Physiology* **29**, 879–887 (2009).
26. Brodribb, T. J., Holbrook, N. M., Zwieniecki, M. A. & Palma, B. Leaf hydraulic capacity in ferns, conifers and angiosperms: impacts on photosynthetic maxima. *New Phytologist* **165**, 839–846 (2005).
27. Pelletier, J. D. & Turcotte, D. L. Shapes of river networks and leaves: are they statistically similar? *Philosophical Transactions of the Royal Society of London B: Biological Sciences* **355**, 307–311 (2000).
28. Roth-Nebelsick, A., Uhl, D., Mosbrugger, V. & Kerp, H. Evolution and function of leaf venation architecture: a review. *Annals of Botany* **87**, 553–566 (2001).
29. Yamazaki, K. Studies on the connecting strand of the vascular system in rice leaves. *Proceedings of the Crop Science Society of Japan* **29**, 400–403 (1960).
30. Lush, W. Leaf structure and translocation of dry matter in a C₃ and a C₄ grass. *Planta* **130**, 235–244 (1976).
31. Altus, D. & Canny, M. Loading of assimilates in wheat leaves. I. The specialization of vein types for separate activities. *Functional Plant Biology* **9**, 571–581 (1982).
32. Altus, D. & Canny, M. Water pathways in wheat leaves. I. The division of fluxes between different vein types. *Functional Plant Biology* **12**, 173–181 (1985).
33. Altus, D., Canny, M. & Blackmann, D. Water pathways in wheat leaves. II. Water-conducting capacities and vessel diameters of different vein types, and the behaviour of the integrated vein network. *Functional Plant Biology* **12**, 183–199 (1985).
34. Chonan, N., Kawahara, H. & Matsuda, T. Morphology of vascular bundles of leaves in gramineous crops. I. Observations on vascular bundles of leaf blades, sheaths and internodes in rice plants. *Proceedings of the Crop Science Society of Japan* **42**, 425–432 (1974).
35. Colbert, J. T. & Evert, R. F. Leaf vasculature in sugarcane (*Saccharum officinarum* L.). *Planta* **156**, 136–151 (1982).
36. Dannenhoffer, J. M. & Evert, R. F. Development of the vascular system in the leaf of barley (*Hordeum vulgare* L.). *International Journal of Plant Sciences*, 143–157 (1994).
37. Russell, S. & Evert, R. Leaf vasculature in *Zea mays* L. *Planta* **164**, 448–458 (1985).
38. Niklas, K. J. Effects of vibration on mechanical properties and biomass allocation pattern of *capsella bursa-pastoris* (cruciferae). *Annals of Botany* **82**, 147–156 (1998).
39. Kehr, J. & Buhtz, A. Long distance transport and movement of RNA through the phloem. *Journal of Experimental Botany* **59**, 85–92 (2008).
40. Sack, L. & Scoffoni, C. Leaf venation: structure, function, development, evolution, ecology and applications in the past, present and future. *New Phytologist* **198**, 983–1000 (2013).
41. Sack, L. & Scoffoni, C. Measurement of leaf hydraulic conductance and stomatal conductance and their responses to irradiance and dehydration using the Evaporative Flux Method (EFM). *Journal of Visualized Experiments* **70**, e4179 (2012).
42. Nardini, A. & Salleo, S. Limitation of stomatal conductance by hydraulic traits: sensing or preventing xylem cavitation? *Trees* **15**, 14–24 (2000).
43. Flexas, J., Scoffoni, C., Gago, J. & Sack, L. Leaf mesophyll conductance and leaf hydraulic conductance: an introduction to their measurement and coordination. *Journal of Experimental Botany* **64**, 3965–3981 (2013).
44. Tabassum, M. A. *et al.* Rice (*Oryza sativa* L.) hydraulic conductivity links to leaf venation architecture under well-watered condition rather than PEG-induced water deficit. *Acta Physiologica Plantarum* **38**, 1–11 (2016).
45. Chaves, M. Effects of water deficits on carbon assimilation. *Journal of Experimental Botany* **42**, 1–16 (1991).
46. Cornic, G. Drought stress inhibits photosynthesis by decreasing stomatal aperture—not by affecting ATP synthesis. *Trends in Plant Science* **5**, 187–188 (2000).
47. Wilson, K. B., Baldocchi, D. D. & Hanson, P. J. Quantifying stomatal and non-stomatal limitations to carbon assimilation resulting from leaf aging and drought in mature deciduous tree species. *Tree Physiology* **20**, 787–797 (2000).
48. Chaves, M. M. *et al.* How plants cope with water stress in the field? Photosynthesis and growth. *Annals of Botany* **89**, 907–916 (2002).
49. Maroco, J. P., Rodrigues, M. L., Lopes, C. & Chaves, M. M. Limitations to leaf photosynthesis in field-grown grapevine under drought—metabolic and modelling approaches. *Functional Plant Biology* **29**, 451–459 (2002).
50. Bernacchi, C. J., Portis, A. R., Nakano, H., von Caemmerer, S. & Long, S. P. Temperature response of mesophyll conductance. Implications for the determination of Rubisco enzyme kinetics and for limitations to photosynthesis *in vivo*. *Plant Physiology* **130**, 1992–1998 (2002).
51. Flexas, J., Bota, J., Escalona, J. M., Sampol, B. & Medrano, H. Effects of drought on photosynthesis in grapevines under field conditions: an evaluation of stomatal and mesophyll limitations. *Functional Plant Biology* **29**, 461–471 (2002).
52. Escalona, J. M., Flexas, J. & Medrano, H. Stomatal and non-stomatal limitations of photosynthesis under water stress in field-grown grapevines. *Functional Plant Biology* **27**, 87–87 (2000).

53. Bota, J., Medrano, H. & Flexas, J. Is photosynthesis limited by decreased Rubisco activity and RuBP content under progressive water stress? *New Phytologist* **162**, 671–681 (2004).
54. Grassi, G. & Magnani, F. Stomatal, mesophyll conductance and biochemical limitations to photosynthesis as affected by drought and leaf ontogeny in ash and oak trees. *Plant, Cell & Environment* **28**, 834–849 (2005).
55. Galle, A. *et al.* The role of mesophyll conductance during water stress and recovery in tobacco (*Nicotiana sylvestris*): acclimation or limitation? *Journal of Experimental Botany* **60**, 2379–2390 (2009).
56. Küppers, M. Carbon relations and competition between woody species in a Central European hedgerow. *Oecologia* **64**, 344–354 (1984).
57. Nardini, A., Tyree, M. T. & Salleo, S. Xylem cavitation in the leaf of *Prunus laurocerasus* and its impact on leaf hydraulics. *Plant Physiology* **125**, 1700–1709 (2001).
58. Bhaskar, R. *et al.* Influences of whole plant water transport and drought on gas exchange within a chaparral community. *Ecological Society of America Annual Meeting Tucson, AZ* (2002).
59. Hirasawa, T., Ozawa, S., Taylaran, R. D. & Ookawa, T. Varietal differences in photosynthetic rates in rice plants, with special reference to the nitrogen content of leaves. *Plant Production Science* **13**, 53–57 (2010).
60. Zwieniecki, M., Boyce, C. & Holbrook, N. Hydraulic limitations imposed by crown placement determine final size and shape of *Quercus rubra* L. leaves. *Plant, Cell & Environment* **27**, 357–365 (2004).
61. Pantin, F., Simonneau, T. & Muller, B. Coming of leaf age: control of growth by hydraulics and metabolics during leaf ontogeny. *New Phytologist* **196**, 349–366 (2012).
62. Sack, L. *et al.* How do leaf veins influence the worldwide leaf economic spectrum? Review and synthesis. *Journal of Experimental Botany* **64**, 4053–4080 (2013).
63. Brodribb, T. J. & Holbrook, N. M. Declining hydraulic efficiency as transpiring leaves desiccate: two types of response. *Plant, Cell & Environment* **29**, 2205–2215 (2006).
64. Maherali, H., Moura, C. F., Caldeira, M. C., Willson, C. J. & Jackson, R. B. Functional coordination between leaf gas exchange and vulnerability to xylem cavitation in temperate forest trees. *Plant, Cell & Environment* **29**, 571–583 (2006).
65. Fichot, R. *et al.* Hydraulic efficiency and coordination with xylem resistance to cavitation, leaf function, and growth performance among eight unrelated *Populus deltoides* × *Populus nigra* hybrids. *Journal of Experimental Botany* **62**, 2093–2106 (2011).
66. Gleason, S. M. *et al.* Weak coordination among petiole, leaf, vein, and gas-exchange traits across Australian angiosperm species and its possible implications. *Ecology and Evolution* **6**, 267–278 (2016).
67. Walls, R. L. Angiosperm leaf vein patterns are linked to leaf functions in a global-scale data set. *American Journal of Botany* **98**, 244–253 (2011).
68. Xiong, D. *et al.* Leaf hydraulic conductance is coordinated with leaf morpho-anatomical traits and nitrogen status in the genus *Oryza*. *Journal of Experimental Botany* **66**, 741–748 (2015).
69. Burundukova, O., Zhuravlev, Y. N., Solopov, N. & P'yankov, V. A method for calculating the volume and surface area in rice mesophyll cells. *Russian Journal of Plant Physiology* **50**, 133–139 (2003).
70. Karki, S., Rizal, G. & Quick, W. P. Improvement of photosynthesis in rice (*Oryza sativa* L.) by inserting the C4 pathway. *Rice* **6**, 28 (2013).
71. Sage, R. Environmental and evolutionary preconditions for the origin and diversification of the C4 photosynthetic syndrome. *Plant Biology* **3**, 202–213 (2001).
72. Ueno, O., Kawano, Y., Wakayama, M. & Takeda, T. Leaf vascular systems in C₃ and C₄ grasses: a two-dimensional analysis. *Annals of Botany* **97**, 611–621 (2006).
73. Smillie, I., Pyke, K. & Murchie, E. Variation in vein density and mesophyll cell architecture in a rice deletion mutant population. *Journal of Experimental Botany* **63**, 4563–4570 (2012).
74. Martre, P. & Durand, J. L. Quantitative analysis of vasculature in the leaves of *Festuca arundinacea* (Poaceae): implications for axial water transport. *International Journal of Plant Sciences* **162**, 755–766 (2001).
75. Kutlu, N. *et al.* Changes in anatomical structure and levels of endogenous phytohormones during leaf rolling in *Ctenanthe setosa* under drought stress. *Turkish Journal of Biology* **33**, 115–122 (2009).
76. El-Afry, M. M., El-Nady, M. F., Abdelmonteleb, E. B. & Metwaly, M. M. S. Anatomical studies on drought-stressed wheat plants (*Triticum aestivum* L.) treated with some bacterial strains. *Acta Biologica Szegediensis* **56**, 165–174 (2012).
77. Farouk, S. & Amany, A. R. Improving growth and yield of cowpea by foliar application of chitosan under water stress. *Egyptian Journal of Biology* **14**, 14–16 (2012).
78. Jacobsen, A. L., Ewers, F. W., Pratt, R. B., Paddock, W. A. & Davis, S. D. Do xylem fibers affect vessel cavitation resistance? *Plant Physiology* **139**, 546–556 (2005).
79. Ocheltree, T. W., Nippert, J. B., Kirkham, M. B. & Prasad, P. V. V. Partitioning hydraulic resistance in *Sorghum bicolor* leaves reveals unique correlations with stomatal conductance during drought. *Functional Plant Biology* **41**, 25–36 (2014).
80. von Cammerer, S. & Farquhar, G. D. Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves. *Planta* **153**, 376–387 (1981).
81. Brodribb, T. & Holbrook, N. M. Changes in leaf hydraulic conductance during leaf shedding in seasonally dry tropical forest. *New Phytologist* **158**, 295–303 (2003).

Acknowledgements

This work was financially supported by the national key research and development program of China (2016YFD0300102), the National Natural Science Foundation of China (31301840), a Foundation for the Author of National Excellent Doctoral Dissertation of PR China (201465), and Fundamental Research Funds for the Central Universities (2662015PY031).

Author Contributions

Y.L. and M.A.T. conceived and designed the research. M.A.T. conducted the experiments and collected and analyzed data. M.A.T. wrote the manuscript. G.L.Z., A.H., M.A.W. and M.S. helped to collect data during the experiments and gave valuable comments. Y.L. revised the manuscript.

Additional Information

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Tabassum, M. A. *et al.* Influence of leaf vein density and thickness on hydraulic conductance and photosynthesis in rice (*Oryza sativa* L.) during water stress. *Sci. Rep.* **6**, 36894; doi: 10.1038/srep36894 (2016).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016