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DNA barcode for the identification of the sand fly *Lutzomyia longipalpis* plant feeding preferences in a tropical urban environment

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Little is known about the feeding behavior of hematophagous insects that require plant sugar to complete their life cycles. We studied plant feeding of *Lutzomyia longipalpis* sand flies, known vectors of *Leishmania infantum/chagasi* parasites, in a Brazilian city endemic with visceral leishmaniasis. The DNA barcode technique was applied to identify plant food source of wild-caught *L. longipalpis* using specific primers for a locus from the chloroplast genome, ribulose diphosphate carboxylase. DNA from all trees or shrubs within a 100-meter radius from the trap were collected to build a barcode reference library. While plants from the Anacardiaceae and Meliaceae families were the most abundant at the sampling site (25.4% and 12.7% of the local plant population, respectively), DNA from these plant families was found in few flies; in contrast, despite its low abundance (2.9%), DNA from the Fabaceae family was detected in 94.7% of the sand flies. The proportion of sand flies testing positive for DNA from a given plant family was not significantly associated with abundance, distance from the trap, or average crown expansion of plants from that family. The data suggest that there may indeed be a feeding preference of *L. longipalpis* for plants in the Fabaceae family.

Visceral leishmaniasis is a lethal vectorborne disease caused by protozoa from the genus *Leishmania*, and in recent decades, it has increasingly affected populations in urban settings of Brazil¹. The first major urban outbreak took place in Teresina, Piauí State, in the early 1980s after a massive planting of acacias. It has thus been hypothesized that the two events were connected.

The quantity and diversity of flora in a habitat are associated with the quality and availability of resources to insects². Plants supply insects with food, protection against predators and adequate shelter to lay their eggs^{3,4}. Accordingly, environmental changes related to plant types and abundance can modify the behavior of disease vectors⁵.

Interactions between sand flies, the vectors of *Leishmania* parasites, and vegetation have been considered^{6,7}. Costa specifically suggested a relation between acacias, which attract sand flies of the species *Lutzomyia longipalpis* and offer protection against predators, and vector proliferation in areas where these plants are abundant⁸. The relationship between acacias and visceral leishmaniasis vectors has been previously considered since one of the largest known epidemics of the disease was observed in Sudan among displaced people living in areas with abundant acacia trees⁹.

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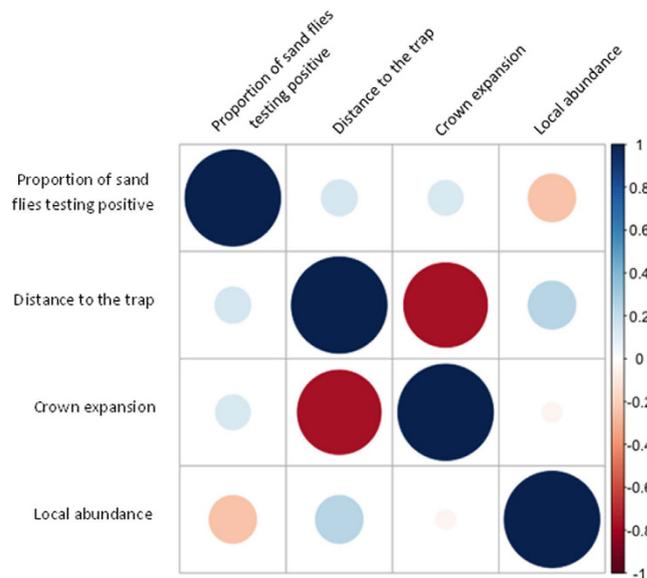


Figure 1. Correlations between the proportion of sand flies testing positive for DNA from a given plant family and plant characteristics. Red represents a negative correlation and blue represents a positive correlation. Larger circles represent stronger correlations. Relationships are presented for (A) proportion of sand flies testing positive for DNA from a given plant family, (B) average distance of plants from a given family to a trap, (C) average crown expansion of plants from a given family, and (D) local abundance of plants from a given family. A strong negative correlation was observed between the distance of plants from a given family to the trap and the average crown expansion of plants from that family.

Sand flies require sugar as their energy source^{10–12}. However, there is little information about specific sources of these sugars and about how sand flies are attracted to plants, especially under natural conditions^{7,13}.

To extend current knowledge about the ecology of insects and plants, more information about insect-plant interactions is necessary¹⁴. Direct analysis of the plant content in an insect's digestive tract is a precise approach to understanding feeding behavior¹⁵.

The use of the DNA barcode tool, a technique that is able to identify the food content in invertebrates, using short sequences obtained in conserved regions of the chloroplast, has been widely reported in the literature^{16–19}. Specifically, the ribulose diphosphate carboxylase (*rbcl*) gene has been successfully used to identify food source^{14,20,21}. This same locus was used by Junnila *et al.*²² to identify plant-derived food content in wild-caught *Phlebotomus papatasi* sand flies.

The DNA barcode approach can be used to identify relationships between presence of plant types and vectorial capacity or simply the likelihood of vector habitats in a particular area. One application of this technique, therefore, would be informing vector control measures, such as urban landscaping techniques, for leishmaniasis and other vectorborne diseases. Here, we describe a study designed to identify plant-based feeding preferences of *L. longipalpis* through the use of DNA barcode technology in a Brazilian city where visceral leishmaniasis is endemic.

Results

We used *rbcl* PCR to detect and identify plant DNA in the guts of 100 *L. longipalpis* captured during 5 days in the tropical urban environment of Teresina, Piauí State. Fifty-seven percent of the flies were found positive for plant DNA. In the vicinity of the trapping location, we identified 22 species of plants belonging to 14 families (Supplementary Table S1). DNA from the ubiquitous plant family Fabaceae was most prevalent and was identified in 94.7% (54/57) of sand flies in which plant DNA was detected. On average, each sand fly was positive for the DNA of 2.1 ± 1.4 plant families. To consider whether plant availability or proximity may have contributed to observed feeding trends among sand flies, relationships between plant characteristics and frequency of plant DNA detection in sand fly guts were assessed. The proportion of sand flies testing positive for DNA of a given plant family was not significantly correlated with local abundance of plants from that family, average distance of plants from the trap, or average crown expansion of plants in the family (Fig. 1). A statistically significant, negative correlation was found between the average distance between plants from a family and the trap and their average crown expansion (Pearson $r = -0.77$; $p = 0.002$) (Fig. 1 and Supplementary Table S1).

On each of the five trap days, DNA from four to eight of the plant families was detected in the flies captured and analyzed (Table 1). Although there was little variation in measurements of abiotic factors (*i.e.*, temperature, air velocity, relative humidity) across the sampling period, humidity was highest on the two days with lower temperatures (Day 1 and Day 4). DNA from 7.5 plant families on average was detected in sand flies captured on these two cooler, more humid days, while DNA from 5 plant families on average was detected in sand flies captured on the warmer, less humid days.

Trap day	Temperature (°C)	Air velocity (m/s)	Relative humidity	Number of sand flies caught and analyzed	Total number of plant families detected in the sand fly guts
1	23	0	92	12	7
2	26	1	78	12	6
3	26	0	83	12	5
4	24	1	92	12	8
5	26	0	81	9	4

Table 1. Abiotic factors and sampling frequency per day of trapping.

Plant Family	Abundance (% of local plant population)	Average Distance from Trap (meters)	Average Crown Expansion (meters)	N (%) Flies with DNA
Anacardiaceae	52 (25.4)	62.3	8.41	2 (3.5)
Bignoniaceae	2 (1.0)	50	9.27	2 (3.5)
Caricaceae	13 (6.3)	72.5	1.68	8 (14.0)
Fabaceae	6 (2.9)	62	8.43	54 (94.7)
Malpighiaceae	17 (8.3)	69.4	3.54	21 (36.8)
Meliaceae	26 (12.7)	62	4.14	1 (1.8)
Myrtaceae	5 (2.4)	60.8	3.97	7 (12.3)
Oxalidaceae	3 (1.5)	60	3.5	22 (38.6)
Rubiaceae	4 (2.0)	73.3	2.52	1 (1.8)
Annonaceae	18 (8.8)	53.4	2.58	0 (0.0)
Musaceae	13 (6.3)	73.1	2.9	0 (0.0)
Rutaceae	25 (12.2)	63.8	3.88	0 (0.0)
Sapindaceae	1 (0.5)	25	12.2	0 (0.0)
Poaceae**	20 (9.8)	14.4	1.09*	0 (0.0)

Table 2. Characteristics of plants by family and frequency of sand fly feeding. *Average leaf area expansion. **The only species belonging to the Family Poaceae in this study was *Zea mays* (maize), which does not make a crown.

Discussion

This study verified the feasibility of detecting plant DNA in the digestive tracts of *L. longipalpis* using a barcode approach. The successful application of this approach is consistent with previous studies investigating feeding behavior of *P. papatasi*²². Furthermore, detection of plant DNA in sand fly guts is biologically plausible as sand flies feed directly from the plant's tissue^{23,24} to obtain sugars that will be used as energy sources²⁵.

However, despite the success of the approach presented here in distinguishing *L. longipalpis* feeding sources, plant DNA was not detected in nearly half of the caught sand flies. This may be explained by factors such as DNA degradation by enzymes in the digestive tract—a process that occurs with blood DNA ingested by females during blood feeding^{26,27}—or due to sugar acquisition from honeydew excreted by aphids and coccidians^{28,29}.

At the sand fly trapping site, plant species belonging to 14 families were collected and identified. The Fabaceae family was the most frequently detected food source of *L. longipalpis* (Table 2) despite the low abundance (2.9% of local plants) and relatively high average distance from the trap of plants from this family. Additionally, DNA from plant families, specifically Anacardiaceae and Meliaceae, that were most represented in the area (*i.e.*, plants from these families had the highest abundance) was found in fewer insects than DNA from the Fabaceae family (Table 2 and Supplementary Table S1). Of further note, some plant families (*i.e.*, Rutaceae and Annonaceae) with abundance higher than that of Fabaceae were found in none of the sand flies (Table 2). This attests to the fact that sand flies have a feeding preference for certain plants or plant families.

The plant families Fabaceae, Anacardiaceae, Meliaceae, Rutaceae and Annonaceae have pantropical distributions. The Fabaceae family, commonly known as legumes, is considered to be the third largest angiosperm family³⁰. The species belonging to this family have carbohydrate heterogeneity, varying from simple sugar to complex heteropolysaccharides³¹. The Anacardiaceae family is known for having many fruit trees of high economic value, due to their wood and their production of substances used in industry or medicine³². The Meliaceae family includes trees producing meliacin, known for its insecticidal properties³³. Plants from the Rutaceae family have a large variety of secondary metabolites, as alkaloids, coumarins, flavonoids, limonoids, and volatile oils³⁴. Species belonging to the Annonaceae family produce bio compounds with medicinal, allelopathic or pesticide properties³⁵.

The attraction of sand flies to certain plants, as well as to certain honey odors, has been reported in different studies^{36–39}. According to Muller and Schlein¹² sand flies seeking sugar sources are primarily guided by attractive factors. These factors can be the carbohydrate composition of certain plants or plant families, high CO₂ emission which can be detected by the insect⁴⁰, or even the release of a phytochemical affecting the olfactory system^{41,42}.

Muller *et al.*¹³ who evaluated the *P. papatasi* attraction index for different plant species, found that phytochemicals serve as potential attractants for insects. Magalhaes-Junior *et al.*⁴³ additionally verified that volatile plant compounds, 1-octen-3-ol, 1-nonanol and 1-heptanol, can act as attractive factors for insects from the *L. longipalpis* species. It is worth noting that the compounds 1-octen-3-ol⁴⁴ and 1-nonanol⁴⁵ have been identified in different species belonging to the Fabaceae family.

Through an assessment of the relationship between abiotic factors and the number of families detected in sand flies per collection day, it was evidenced that more plant families were used as a feeding sources on those days with lower temperatures and higher relative humidity. However, no reports were found in the literature about how these factors might influence the dynamics of sugar acquisition in these insects. Currently, it is known that abiotic factors such as temperature and relative humidity influence the population dynamics in the different sand fly species^{46–48} including *L. longipalpis*⁴⁹.

Accordingly, our study suggested there is indeed a feeding preference by *L. longipalpis* for the Fabaceae family, represented in this study by specimens of the species *Albizia niopoides*, *Anadenanthera macrocarpa*, *Cenostigma macrophyllum* and *Tamarindus indica*. At the same time, the data suggests that *L. longipalpis* had less preference for plants belonging to the families Anacardiaceae, Meliaceae, Rutacea or Annonaceae.

Upon further verification of *L. longipalpis* plant feeding preferences, it will be necessary to determine whether sand fly abundance varies with the presence or absence of plants from certain families. If so, the combined findings may have implications for controlling urban epidemics of vector borne disease. In particular, removal of preferred sugar sources for sand flies could reduce their prevalence and thus reduce density-dependent transmission of the pathogens they harbor.

It is also recommended that new studies relating the feeding interaction between sand flies and plants focus not only in the identification of plant species or plant families serving as a carbohydrate source, but also in the identification of the chemical components that lead to the insect preference for certain food source. Additionally, it is necessary to understand the role of each abiotic factor in this feeding process, and this way, sand fly population control measures may be applied.

Materials and Methods

Field collection of phlebotomine sand flies. Insect and plant collection were conducted in a region of the city of Teresina (05°07'700"S/42°46'426"W), capital of Piauí State, in Brazil's Northeast. The sand flies were captured using a CDC light trap, deployed in an animal shelter outside of a household. The trap remained at the site for five consecutive nights working 12 hours per night (6 P.M. to 6 A.M.) in January, 2015. The captured sand flies were taken to the laboratory for identification and processing. Only *L. longipalpis* sand flies were separated randomly from the sample of sand flies collected every trapping night. In order to avoid the contamination of the samples during the DNA extraction process, every plant fragment was removed from the sand fly bodies. Previously collected insects were submerged in a solution of 0.5% hypochlorite with 0.01 ml/ml Triton X-100 detergent, agitated gently for 1 min with forceps, and then rinsed in double distilled water (ddH₂O) for 1 min as proposed by Matheson *et al.*¹⁵. After this procedure, the insects were stored at –80 °C for further DNA extraction. All the techniques used for plant identification had been previously tested using DNA directly from plants (Lima *et al.*, submitted). For every capture day, values of temperature, relative humidity and wind speed were obtained in collaboration with the National Meteorology Institute (INMET).

Field collection of plants. The botanical material (leaves and flowers) was collected under the criteria that plants should be, at the most, 100 meters apart from the sand fly trap, which is within the interval of dispersal of the sand fly *Lu. longipalpis*⁵⁰. In total, 22 plant species were collected and subsequently identified at the Graziela Barroso Herbarium at the Federal University of Piauí. For genetic analysis, species were labeled and stored at –80 °C until DNA extraction. Information on species included in the present study is provided in Supplementary Table S1.

Distances of each plant specimen from the CDC traps were recorded using a Garmin Dakota 10 GPS device (Garmin, USA) and the average expansion of the crown was measured according to methods proposed by Blozan⁵¹.

DNA extraction, PCR and sequencing. During the DNA extraction process, sand flies were put in 1.5 mL test tubes and 300 µl of CTAB lysis buffer (50 mL 2x CTAB + 50 mL ddH₂O + 200 mL mercaptoethanol (0.2%)) was added. The samples were macerated with an Argos Pellet Mixer (Argos, USA), and 10 µl K proteinase (Invitrogen, USA) was added to each tube (concentration 10 mg/mL). Immediately after, the samples were incubated for two hours in a 60 °C hot bath. Extraction was then conducted using the PureLink Genomic DNA Mini Kit (Invitrogen, USA) according to the manufacturer's instructions. The plant genomic DNA extraction was carried out using the kit BIOPUR Extração Mini Spin Planta (Biometrix, Brazil). The DNA was extracted and analyzed in 1% agarose gel under UV light and subsequently was quantified by spectrophotometer NanoDrop™ 2000 (ThermoScientific, USA) and the fluorometer Qubit® 2.0 (Invitrogen, USA).

For each separated DNA sample, a segment of chloroplast gene *rbcL* was amplified by PCR using the PCR Master Mix (Promega, USA) according to the manufacturer's instructions for a 50 µL final volume, containing 1 µL of each primer (*rbcLaF* and *rbcLaR*⁵²), and 5 µL DNA template. The reactions took place in a T100 (Bio-Rad, USA) thermal cycler. The thermal cycle conditions have been described by Bafeel *et al.*⁵². PCR products were visualized in 1% agarose gel under UV light and purified using ExoSap-IT (Affymetrix, USA) for the sand fly samples and PureLink PCR Purification Kit (Invitrogen, USA) for the plants. Further, this material was quantified by using fluorometer Qubit® 2.0 (Invitrogen, USA) and purity verified in spectrophotometer NanoDrop™ 2000 (ThermoScientific, USA). Sequencing was carried out in an ABI 3500 automatic sequencer (Applied Biosystems, USA).

Only sequences with a PHRED score above 30 were used in the analysis. Contig assembly was carried out using CodonCode Aligner (CodonCode Corporation). Local alignments were done using BLAST⁵³.

Assembly of DNA barcode library. DNA sequences obtained from each primer were deposited in Genbank database, linked to the National Center for Biotechnology Information (NCBI) (Supplementary Table S1). The deposited sequences were set up for starting a reference library of plant species from the city of Teresina, and from other tropical cities.

Identification of the plant DNA in sand flies. Each sequence of plant DNA found in the insects was compared to a local plant library using the BLASTN. Only matches with highest score and, at least, 90% identity were considered for the sand flies' plant feeding preference⁵⁴.

Statistical Methods. Descriptive statistics were summarized for individual plant characteristics and abiotic factors (*i.e.*, temperature, relative humidity and wind speed) across the five trap days. Correlations between the average distance of the plant family from the trap, the average crown expansion of plants from a given family, the proportion of plants in the area from a given family, and the proportion of sand flies testing positive for DNA from a given plant family were investigated for the entire sample of sand flies.

All statistical analyses were performed using R version 3.2.1 (R Foundation for Statistical Computing, Vienna, Austria).

References

- Costa, C. H. N., Pereira, H. F. & Araújo, M. A. Epidemia de leishmaniose visceral no Estado do Piauí, Brasil, 1980–1986. *Rev Saude Publica* **24**, 361–372 (1990).
- Costa, F. V., Siqueira Neves, F., Oliveira Silva, J. & Fagundes, M. Relationship between plant development, tannin concentration and insects associated with *Copaifera langsdorffii* (Fabaceae). *Arthropod-Plant Interactions* **5**, 9–18 (2010).
- Gawande, N. D., Subashini, S., Murugan, M. & Subbarayalu, M. Molecular screening of insecticides with sigma glutathione S-transferases (GST) in cotton aphid *Aphis gossypii* using docking. *Bioinformation* **10**, 679–683 (2014).
- Mello, M. O. & Silva-Filho, M. C. Plant-insect interactions: an evolutionary arms race between two distinct defense mechanisms. *Braz J Plant Physiol* **14**, 71–81 (2002).
- Conley, A. K., Watling, J. I. & Orrock, J. L. Invasive plant alters ability to predict disease vector distribution. *Ecol Appl* **21**, 329–334 (2011).
- Rutledge, L. C. & Ellenwood, D. A. Production of Phlebotomine sand flies on the open forest floor in Panama: phytologic and edaphic relations. *Environ Entomol* **4**, 83–89 (1975).
- Cameron, M. M., Pessoa, F. A. C., Vasconcelos, A. W. & Ward, R. D. Sugar meal sources for the phlebotomine sandfly *Lutzomyia longipalpis* in Ceará State, Brazil. *Med Vet Entomol* **9**, 263–272 (1995).
- Costa, C. H. N. Characterization and speculations on the urbanization of visceral leishmaniasis in Brazil. *Cad Saude Publica* **24**, 2959–2963 (2008).
- Elnaiem, D. E. *et al.* Factors affecting variations in exposure to infections by *Leishmania donovani* in eastern Sudan. *East Mediterr Health J* **9**, 827–836 (2003).
- Jacobson, R. L., Studentsky, L. & Schlein, Y. Glycolytic and chitinolytic activities of *Phlebotomus papatasi* (Diptera: Psychodidae) from diverse ecological habitats. *Folia Parasit* **54**, 301–309 (2007).
- Mascari, T. M. & Foil, L. D. Laboratory Evaluation of the Efficacy of Fluorescent Biomarkers for Sugar-Feeding Sand Flies (Diptera: Psychodidae). *J Med Entomol* **47**, 664–669 (2010).
- Muller, G. C. & Schlein, Y. Different methods of using attractive sugar baits (ATSB) for the control of *Phlebotomus papatasi*. *J Vector Ecol* **36** Suppl 1, S64–70 (2011).
- Muller, G. C., Revay, E. E. & Schlein, Y. Relative attraction of the sand fly *Phlebotomus papatasi* to local flowering plants in the Dead Sea region. *J Vector Ecol* **36** Suppl 1, S187–194 (2011).
- Gravendeel, B., Eurlings, M. & Heijerman, T. Use of DNA barcoding for host plant identification. *Entomologische Berichten* **69**, 30–35 (2009).
- Matheson, C. D. *et al.* A PCR method for detection of plant meals from the guts of insects. *Organisms Diversity & Evolution* **7**, 294–303 (2008).
- Jurado-Rivera, J. A., Vogler, A. P., Reid, C. A. M., Petitpierre, E. & Gómez-Zurita, J. DNA barcoding insect–host plant associations. *Proc Biol Sci* **276**, 639–648 (2009).
- Pumarino, L., Alomar, O. & Agustí, N. Development of specific ITS markers for plant DNA identification within herbivorous insects. *B Entomol Res* **101**, 271–276 (2011).
- Staudacher, K., Wallinger, C., Schallhart, N. & Traugott, M. Detecting ingested plant DNA in soil-living insect larvae. *Soil biology & biochemistry* **43**, 346–350 (2011).
- Wallinger, C. *et al.* The effect of plant identity and the level of plant decay on molecular gut content analysis in a herbivorous soil insect. *Mol Ecol Resour* **13**, 75–83 (2013).
- García-Robledo, C., Erickson, D. L., Staines, C. L., Erwin, T. L. & Kress, W. J. Tropical Plant–Herbivore Networks: Reconstructing Species Interactions Using DNA Barcodes. *Plos One* **8**, e52967, doi: 10.1371/journal.pone.0052967 (2013).
- Kajtoch, L. A. DNA metabarcoding study of a polyphagous beetle dietary diversity: the utility of barcodes and sequencing techniques. *Folia Biologica* **62**, 223–234 (2014).
- Junnila, A., Muller, G. C. & Schlein, Y. Identification of plant tissues from the gut of *Phlebotomus papatasi* by DNA analysis. *Acta Trop* **117**, 14–18 (2011).
- Warburg, A. & Faiman, R. Research priorities for the control of phlebotomine sand flies. *Journal of vector ecology: journal of the Society for Vector Ecology* **36** Suppl 1, S10–16 (2011).
- Stone, C. M. & Foster, W. A. In *Ecology of parasite-vector interactions* 35–79 (Wageningen Academic Publishers, 2013).
- Cavalcante, R. R., Pereira, M. H., Freitas, J. M. & Gontijo Nde, F. Ingestion of saliva during carbohydrate feeding by *Lutzomyia longipalpis* (Diptera; Psychodidae). *Memorias do Instituto Oswaldo Cruz* **101**, 85–87 (2006).
- Sant'Anna, M. R. *et al.* Blood meal identification and parasite detection in laboratory-fed and field-captured *Lutzomyia longipalpis* by PCR using FTA databasing paper. *Acta Trop* **107**, 230–237 (2008).
- Valinsky, L., Ettinger, G., Bar-Gal, G. K. & Orshan, L. Molecular identification of bloodmeals from sand flies and mosquitoes collected in Israel. *J Med Entomol* **51**, 678–685 (2014).

28. Souza, N. A. *et al.* The influence of sugars and amino acids on the blood-feeding behaviour, oviposition and longevity of laboratory colony of *Lutzomyia longipalpis* (Lutz & Neiva, 1912) (Diptera: Psychodidae, Phlebotominae). *Memorias do Instituto Oswaldo Cruz* **90**, 751–757 (1995).
29. Schlein, Y. & Jacobson, R. L. Sugar meals and longevity of the sandfly *Phlebotomus papatasi* in an arid focus of *Leishmania major* in the Jordan Valley. *Med Vet Entomol* **13**, 65–71 (1999).
30. Andrade, A. L. P., Miotto, S. T. S. & Santos, É. P. d. A subfamília Faboideae (Fabaceae Lindl.) no Parque Estadual do Guartelá, Paraná, Brasil. *Hoehnea* **36**, 737–768 (2009).
31. Aguilera, J. M. & Stanley, D. W. A review of textural defects in cooked reconstituted legumes—The influence of storage and processing. *J Food Proc Preserv* **9**, 145–169 (1985).
32. Nadia, T. d. L., Machado, I. C. & Lopes, A. V. Polinização de *Spondias tuberosa* Arruda (Anacardiaceae) e análise da partilha de polinizadores com *Ziziphus joazeiro* Mart. (Rhamnaceae), espécies frutíferas e endêmicas da caatinga. *Brazilian Journal of Botany* **30**, 89–100 (2007).
33. Pereira Júnior, O. L. *et al.* Atividade biológica de um novo triterpeno isolado de *Guarea cannata* (Meliaceae). *Acta Amazonica* **23**, 173–176 (1993).
34. Supabphol, R. & Tangjitjareonkun, J. Chemical constituents and biological activities of *Zanthoxylum limonella* (Rutaceae): A review. *Trop J Pharm Res* **13**, 2119–2130 (2014).
35. Lemos, E. E. P. d. A produção de anonáceas no Brasil. *Revista Brasileira de Fruticultura* **36**, 77–85 (2014).
36. Schlein, Y. & Warburg, A. Phytophagy and the feeding cycle of *Phlebotomus papatasi* (Diptera: Psychodidae) under experimental conditions. *J Med Entomol* **23**, 11–15 (1986).
37. Schlein, Y. & Yuval, B. Leishmaniasis in the Jordan Valley IV. Attraction of *Phlebotomus papatasi* (Diptera: Psychodidae) to plants in the field. *J Med Entomol* **24**, 87–90 (1987).
38. Junnila, A., Muller, G. C. & Schlein, Y. Attraction of *Phlebotomus papatasi* to common fruit in the field. *J Vector Ecol* **36** Suppl 1, S206–211 (2011).
39. Wasserberg, G., Kirsch, P. & Rowton, E. D. Orientation of colonized sand flies *Phlebotomus papatasi*, *P. duboscqi*, and *Lutzomyia longipalpis* (Diptera: Psychodidae) to diverse honeys using a 3-chamber in-line olfactometer. *J Vector Ecol* **39**, 94–102 (2014).
40. Schlein, Y. & Jacobson, R. L. High nocturnal CO₂ emanation guides the sand fly *Phlebotomus papatasi* to sugar-rich plants. *Physiol Entomol* **33**, 353–359 (2008).
41. Foster, W. A. Phytochemicals as population sampling lures. *J Am Mosq Control Assoc* **24**, 138–146 (2008).
42. Machado, V. E. *et al.* Attraction of the sand fly *Nyssomyia neivai* (Diptera: Psychodidae) to chemical compounds in a wind tunnel. *Parasit Vectors* **8**, 1–4 (2015).
43. Magalhaes-Junior, J. *et al.* A laboratory evaluation of alcohols as attractants for the sandfly *Lutzomyia longipalpis* (Diptera: Psychodidae). *Parasit Vectors* **7** (2014).
44. Ogunwande, I. A. *et al.* Essential oils from the leaves of six medicinal plants of Nigeria. *Nat Prod Commun* **8**, 243–248 (2013).
45. Cakmak, Y. S., Aktumsek, A., Duran, A. & Çetin, O. Antioxidant Activity and Biochemical Screening of two *Glycyrrhiza* L. Species. *Br J Pharm Res* **11**, 2231–2919 (2016).
46. Marcondes, C. B., Santos-Neto, L. G. & Lozovei, A. L. Ecology of phlebotomine sandflies (Diptera, Psychodidae) in Brazilian Atlantic Forest. *Rev Soc Bras Med Trop* **34**, 255–260 (2001).
47. Barata, R. A. *et al.* Phlebotomine sandflies (Diptera: Psychodidae) in Governador Valadares, a transmission area for American tegumentary leishmaniasis in State of Minas Gerais, Brazil. *Rev Soc Bras Med Trop* **44**, 136–139 (2011).
48. Prudhomme, J. *et al.* Ecology and spatiotemporal dynamics of sandflies in the Mediterranean Languedoc region (Roquedur area, Gard, France). *Parasit Vectors* **8**, 642, doi: 10.1186/s13071-015-1250-2 (2015).
49. Ximenes, M. d. F. d. M. *et al.* Effect of Abiotic Factors on Seasonal Population Dynamics of *Lutzomyia longipalpis* (Diptera: Psychodidae) in Northeastern Brazil. *J Med Entomol* **43**, 990–995 (2006).
50. Morrison, A. C., Ferro, C., Morales, A., Tesh, R. B. & Wilson, M. L. Dispersal of the Sand Fly *Lutzomyia-Longipalpis* (Diptera, Psychodidae) at an Endemic Focus of Visceral Leishmaniasis in Colombia. *J Med Entomol* **30**, 427–435 (1993).
51. Blozan, W. Tree Measuring Guidelines of the Eastern Native Tree Society. *Native Tree Society BBS - Exploration of Trees and Forests*. Available at: http://www.nativetreesociety.org/measure/tree_measuring_guidelines.htm (Accessed: 6th January 2015) (2008).
52. Bafeel, S. O. *et al.* Comparative evaluation of PCR success with universal primers of maturase K (matK) and ribulose-1, 5-bisphosphate carboxylase oxygenase large subunit (rbcL) for barcoding of some arid plants. *Plant Omics* **4**, 195–198 (2011).
53. Altschul, S. F., Gish, W., Miller, W., Myers, E. W. & Lipman, D. J. Basic local alignment search tool. *J Mol Biol* **215**, 403–410 (1990).
54. van Velzen, R., Weitschek, E., Felici, G. & Bakker, F. T. DNA barcoding of recently diverged species: relative performance of matching methods. *Plos One* **7**, e30490, doi: 10.1371/journal.pone.0030490 (2012).

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Author Contributions

L.L., V.S. and C.C. designed the study; L.L., M.F., V.B. and L.S. analyzed the data; L.L. and M.M. collected materials; L.L., M.M., O.K., I.A. and A.W. performed the laboratory work; L.L., M.F., L.S. and C.C. wrote the manuscript. All authors read and approve the final manuscript.

Additional Information

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