

# SCIENTIFIC REPORTS



OPEN

## The wheat R2R3-MYB transcription factor TaRIM1 participates in resistance response against the pathogen *Rhizoctonia cerealis* infection through regulating defense genes

Received: 08 March 2016

Accepted: 08 June 2016

Published: 01 July 2016

Tianlei Shan, Wei Rong, Huijun Xu, Lipu Du, Xin Liu &amp; Zengyan Zhang

The necrotrophic fungus *Rhizoctonia cerealis* is a major pathogen of sharp eyespot that is a devastating disease of wheat (*Triticum aestivum*). Little is known about roles of MYB genes in wheat defense response to *R. cerealis*. In this study, *TaRIM1*, a *R. cerealis*-induced wheat MYB gene, was identified by transcriptome analysis, then cloned from resistant wheat CI12633, and its function and preliminary mechanism were studied. Sequence analysis showed that *TaRIM1* encodes a R2R3-MYB transcription factor with transcription-activation activity. The molecular-biological assays revealed that the TaRIM1 protein localizes to nuclear and can bind to five MYB-binding site *cis*-elements. Functional dissection results showed that following *R. cerealis* inoculation, *TaRIM1* silencing impaired the resistance of wheat CI12633, whereas *TaRIM1* overexpression significantly increased resistance of transgenic wheat compared with susceptible recipient. TaRIM1 positively regulated the expression of five defense genes (*Defensin*, *PR10*, *PR17c*, *nsLTP1*, and *chitinase1*) possibly through binding to MYB-binding sites in their promoters. These results suggest that the R2R3-MYB transcription factor TaRIM1 positively regulates resistance response to *R. cerealis* infection through modulating the expression of a range of defense genes, and that *TaRIM1* is a candidate gene to improve sharp eyespot resistance in wheat.

Common wheat (*Triticum aestivum* L.) is one of most important staple crops in the world and plays a fundamental role in food security. Wheat production often is negatively affected by various fungal diseases. The sharp eyespot disease, mainly caused by the soil-borne fungus *Rhizoctonia cerealis* van der Hoeven (teleomorph: *Ceratobasidium cereale* D. Murray & L.L. Burpee), is a devastating disease of wheat production in the world<sup>1–3</sup>. The sharp eyespot symptoms in wheat include dark-bordered lesions on stem bases and based sheaths of adult-plants<sup>3</sup>. The sharp eyespot disease can destroy the transport tissues in stems of plants, and cause pre- and post-emergence damping off and premature spike senescence or ripening (white heads), leading to yield losses of ~10–30%. The environmentally safe and efficient way to protect wheat from sharp eyespot is to breed resistant wheat varieties. However, the resistances in partially-resistant wheat lines are controlled by multiple quantitative loci<sup>1,4</sup>. Currently, breeding sharp eyespot-resistant wheat cultivars using traditional methods is difficult since none of sharp eyespot-immune wheat cultivars/lines is available. Thus, to improve wheat resistance to *R. cerealis*, it is necessary to identify important genes in the defense response and clarify their defense roles.

In order to defend attack of pathogens, plants have evolved comprehensive and complicated defense system. The transcription factor families play an important role in the plants responses against different stresses. The myeloblastosis (MYB) transcription factors are characterized by a DNA-binding MYB domain. The MYB domain is composed of approximately 52 amino acid residues that adopt a helix-turn-helix conformation that intercalates

The National Key Facility for Crop Gene Resources and Genetic Improvement, Institute of Crop Science, Chinese Academy of Agricultural Sciences, Beijing 100081, China. Correspondence and requests for materials should be addressed to Z.Z. (email: zhangzengyan@caas.cn)

into the major groove of DNA<sup>5–6</sup>. Since the first plant MYB gene *COLORED1* (*Cl*) being involved in anthocyanin biosynthesis was identified in maize<sup>7</sup>, a large number of MYB proteins have been identified in different plant species. Based on the number of adjacent MYB repeats, MYB proteins can be divided into 4 classes: 1R-MYB, 2R-MYB (R2R3-MYB), 3R-MYB and 4R-MYB<sup>6</sup>. Numerous MYB proteins have been implicated in diverse biological processes, including cell cycle regulation, cell wall biosynthesis, development and reproduction, and defense responses to abiotic and biotic stresses<sup>6,8–14</sup>. To date, most of the identified MYB genes belong to R2R3-MYB subfamily, and a number of R2R3-MYB transcriptional factors have been evidenced to play different important roles in various plant species<sup>6,15–21</sup>. For example, in *Arabidopsis*, *BOS1* (*BOTRYTIS-SUSCEPTIBLE1*), an R2R3-MYB gene (AtMYB108), is required for restricting the spread of 2 necrotrophic pathogens *Botrytis cinerea* and *Alternaria brassicicola*, and involved in the tolerance to osmotic and oxidative stresses<sup>22</sup>. Overexpression of an *Arabidopsis* R2R3-MYB AtMYB96 could enhance tolerance to drought stress<sup>23</sup> and increase resistance to bacteria pathogens<sup>24</sup>. In barley, the MYB transcription factor HvMYB6 functions as positive regulator of basal and MLA-mediated immunity responses to *Blumeria graminis*<sup>25</sup>. Ectopic expression of the wheat MYB gene *TaMYB33* that was induced by NaCl and PEG stresses increased salt and drought tolerance in *Arabidopsis* plants<sup>17</sup>. The ectopic expression of *TaMYB73* improved salt tolerance of transgenic *Arabidopsis* plants<sup>15</sup>. Overexpression of the wheat pathogen-induced MYB gene *TaPIMP1* in transgenic wheat could significantly enhance resistance to the fungal pathogen *Bipolaris sorokiniana* and drought stresses<sup>26</sup>. Ectopic expression of a *Thinopyrum intermedium* MYB gene *TiMYB2R-1* could significantly increase resistance of transgenic wheat lines to take-all caused by *Gaeumannomyces graminis*<sup>27</sup>. Silencing of a wheat R2R3-MYB gene *TaMYB4* in wheat impaired the resistance to *Puccinia striiformis* f. sp. *tritici*<sup>5</sup>. However, none of MYB genes being involved in defense response to *R. cerealis* infection has been reported yet.

In this study, we identified and functional characterized a *R. cerealis*-induced MYB gene in wheat, named *TaRIM1*. Toward *R. cerealis* infection, the gene expression goes higher level. The sequence analysis and bio-molecular assays proved that *TaRIM1* protein is a R2R3-type MYB transcription factor. It is localized in the nucleus and can bind to MYB binding site *cis*-elements. Through generation of *TaRIM1*-silencing and overexpression wheat plants and assessment of their defense responses following *R. cerealis* inoculation, the functional dissection results indicated that *TaRIM1* positively modulated wheat defense response to *R. cerealis*. Further investigation suggested that *TaRIM1* might activate the expression of a range of defense-related genes, resulting in enhanced resistance to *R. cerealis* infection.

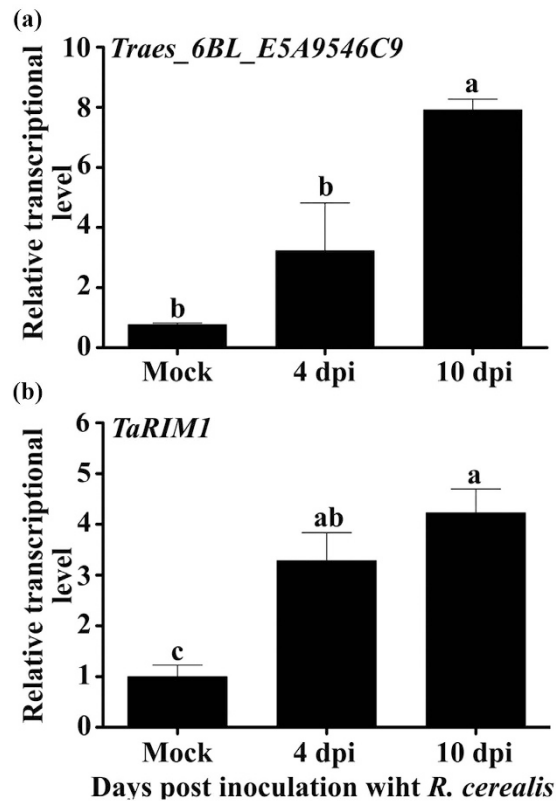
## Results

**Identification and cloned sequence of *TaRIM1* induced by *R. cerealis* infection.** To identify wheat genes being involved in defense response to *R. cerealis*, we performed transcriptomic analysis through Deep RNA-seq on 3 resistance lines of the recombinant inbred lines (RILs, being derived from the cross of sharp eyespot-resistant wheat line Shanhongmai and sharp eyespot-susceptible wheat cultivar Wenmai 6) at 4 and 10 d post inoculation (dpi) with *R. cerealis* high-virulence strain WK207 (Unpublished). Among the up-regulated sequences, the expression of the sequence with no. Traes\_6BL\_E5A9546C9, being homologous to the wheat MYB gene *TaMYB33* sequence, was up-regulated in the resistant wheat lines after *R. cerealis* inoculation. It showed a 4.18-fold at 4 dpi or a 10.23-fold at 10 dpi transcriptional increase than the mocked (Fig. 1a). Quantitative RT-PCR (qRT-PCR) analysis showed that the transcriptional levels of this gene were induced after *R. cerealis* inoculation (Fig. 1b), and the expression tendency by experimental qRT-PCR was in agreement with the RNA-Seq data. This gene was designated as *TaRIM1* and was suggested to be involved in wheat defense response to *R. cerealis* infection.

The full-length cDNA sequence (with 1028 bp, Fig. 2a, NCBI accession no. KU864997) of *TaRIM1* was obtained from *R. cerealis*-infected stem cDNA of the resistance wheat line CI12633 by RACE and nest RT-PCR. It includes the complete ORF with 732-bp, 5'-untranslated region (UTR) with 98-bp, and 3'-UTR with 198-bp. The genomic DNA sequence of *TaRIM1* was amplified from CI12633 genomic DNA. The comparison of the genomic and cDNA sequences indicated that no intron existed in genomic transcription unit of *TaRIM1*, at least in the amplification region. The deduced protein *TaRIM1* contains 243 amino acids with a predicted molecular weight of 26.61 KD and predicted PI of 6.09. As shown in Fig. 2a, the *TaRIM1* protein sequence possesses two conserved MYB DNA-binding domains [one (R2) located at amino acids 13–63 and another (R3) at amino acids 66–114], a putative nuclear localization signal (NLS, located at amino acids 110–136), and an acidic region (amino acids 138–191) possibly acting as a transcription activation domain<sup>28</sup>. The reconstructed phylogenetic tree analysis showed that this protein *TaRIM1* was clustered into the R2R3-MYB subfamily (Fig. 2b). Thus, *TaRIM1* most likely is a R2R3-MYB transcription factor with transcription-activation activity.

***TaRIM1* localizes to the nucleus and binds to MYB-binding site *cis*-elements.** As the *TaRIM1* protein sequence contains the NLS sequence (Fig. 2a), the p35S:*TaRIM1*-GFP (green fluorescent protein) fusion expressing-vector was prepared for investigating the subcellular localization of *TaRIM1*. The p35S:*TaRIM1*-GFP and control p35S:GFP construct DNAs were separately introduced into and transiently expressed in both wheat mesophyll protoplasts and onion epidermal cells. Confocal imaging of the transient expression showed that *TaRIM1*-GFP localized in the nucleus in both the wheat mesophyll protoplasts and onion epidermal cells (Fig. 3a,b), whereas the fluorescence of the control GFP was distributed throughout the cell (Fig. 3a,b). These results indicated that the expressing *TaRIM1* protein localizes in the nuclear.

The amino acid sequence of *TaRIM1* contains R2 and R3 DNA-binding domains. To investigate if *TaRIM1* binds to MYB-binding site (MBS) *cis*-elements, the glutathione *S*-transferase (GST)-*TaRIM1* recombinant protein was prepared, expressed, and purified. Electrophoretic mobility shift assay (EMSA) was used to examine the DNA binding ability of *TaRIM1* with MBS. Here, the tested 5 MBS *cis*-elements include ACI, MBS1-Bz, MBS1-w, RT1, and St1R that were bound by known functional R2R3-MYB, StMYB1R-1 and OsMYB3R-2 transcription

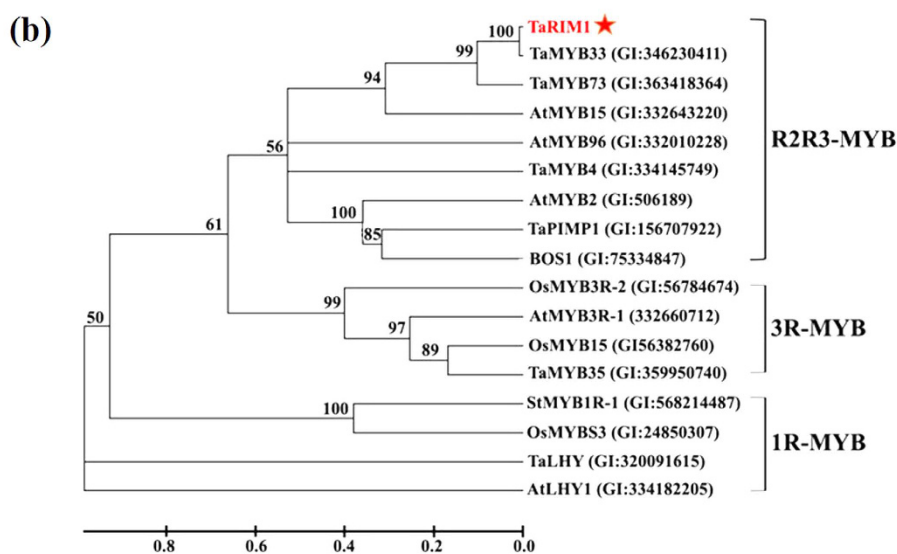
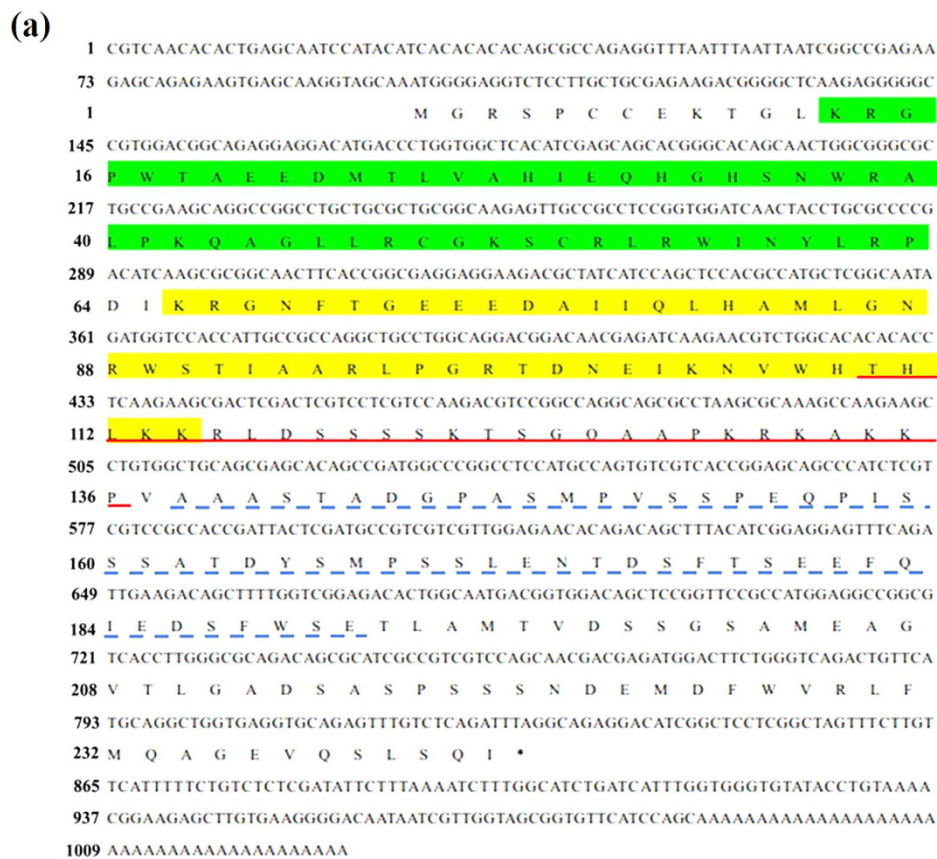


**Figure 1.** Transcriptional analyses of *TaRIM1* in *Rhizoctonia cerealis*-inoculated wheat. The RNAs are isolated from pooled stems of 3 resistant lines in recombinant inbred lines of cross of *R. cerealis*-resistant wheat Shanhongmai and susceptible wheat cultivar Wenmai 6 at 4, or 10 days post inoculation (dpi) with *R. cerealis* strain WK207 or mocked ones. (a) RNA-Seq data of the sequence *Traes\_6BL\_E5A9546C9* being corresponding to *TaRIM1*. (b) qRT-PCR verification of the transcription of *TaRIM1*. The transcriptional level in the mocked one is set to 1. The transcriptional levels with different letters are significantly different from each other based on statistically significant analysis on the results of three replications (*t*-test,  $**P < 0.01$ ).

factors<sup>26</sup> (Fig. 4a, Table S1). The EMSA results showed that the GST-*TaRIM1* protein could bind to all the tested 5 MBS probes, especially showing the strongest binding to ACI, but not bind to the GCC-box *cis*-element that is specifically bound by ERF transcription factors, whereas GST failed to bind with the MBS *cis*-element ACI and the GCC-box *cis*-element (Fig. 4b). These data proved that the protein *TaRIM1* can bind to these five MBS elements.

**Silencing of *TaRIM1* impairs wheat resistance to *R. cerealis*.** Virus-induced gene silencing (VIGS) is an efficient reverse-genetic tool for rapidly analyzing functions of genes in plants. Barley stripe mosaic virus (BSMV)-based VIGS is extensively used for investigating functions of interest genes in barley and wheat<sup>29–31</sup>. To explore whether *TaRIM1* plays an important role in wheat resistance response against *R. cerealis*, we used BSMV-based VIGS method to down-regulate transcriptional levels of *TaRIM1* in the resistant wheat line CI12633. At 15 dpi with the virus, the transcript of BSMV coat protein (*cp*) gene was readily detected (Fig. 5a), suggesting that BSMV successfully infected these wheat plants. Importantly, the transcript levels of *TaRIM1* were significantly reduced in CI12633 plants infected by BSMV:*TaRIM1* compared to BSMV:GFP infected CI12633 plants (control plants) (Fig. 5a,b), suggesting that *TaRIM1* transcript was successfully down-regulated in BSMV:*TaRIM1* infected plants, hereafter *TaRIM1*-silenced plants represented BSMV:*TaRIM1* infected ones. The *TaRIM1*-silenced and BSMV:GFP infected CI12633 plants were further inoculated with *R. cerealis*. Subsequently, the infection types (ITs) by the fungus were evaluated. At 45 dpi with *R. cerealis*, *TaRIM1*-silenced CI12633 plants showed more susceptible to the sharp eyespot disease caused by *R. cerealis* (ITs: ~2.8–3.8; Fig. 5c), whereas BSMV:GFP infected CI12633 plants showed more resistance of sharp eyespot (IT: 1.2, Fig. 5c). These results suggested the down-regulation of *TaRIM1* compromised the resistance to *R. cerealis* in CI12633, and that *TaRIM1* is required for host resistance response to *R. cerealis*.

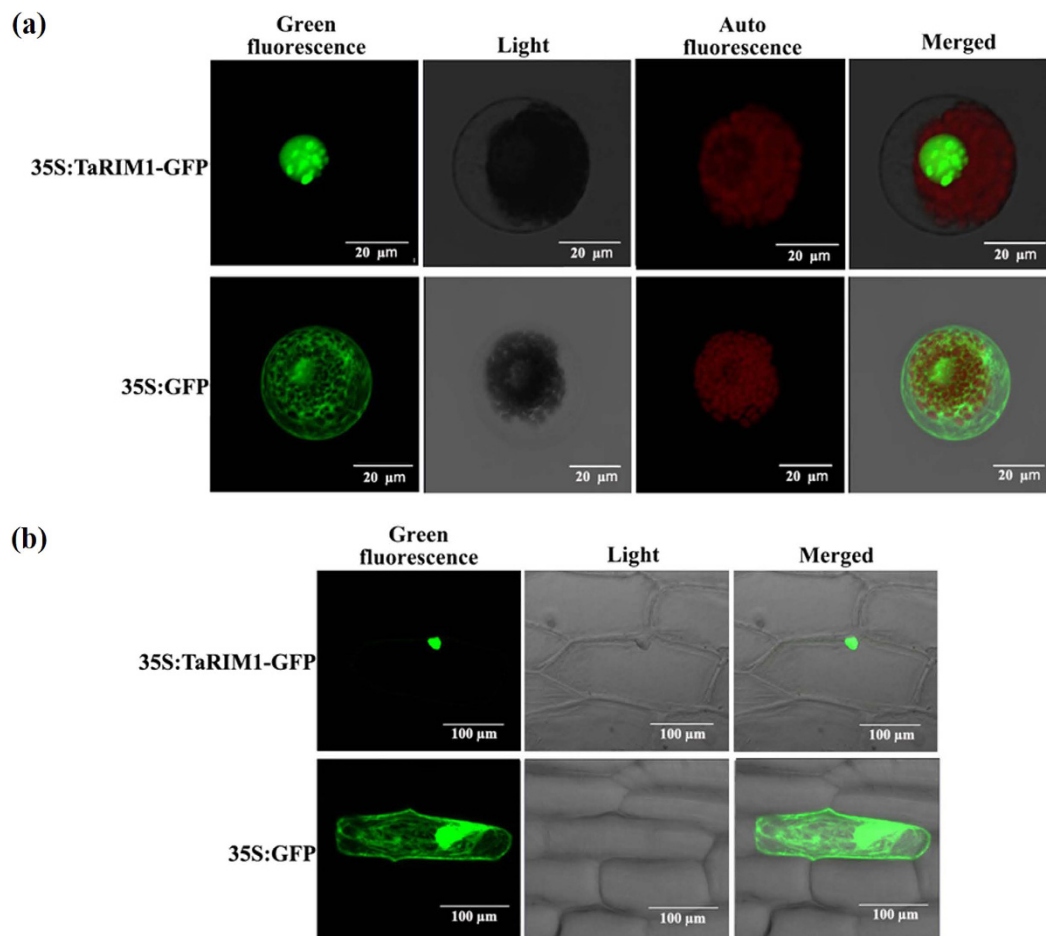
**Overexpression of *TaRIM1* enhances wheat resistance to *R. cerealis*.** To further investigate the defense role of *TaRIM1* in wheat, we constructed the *TaRIM1* overexpression vector pUbi:myc-*TaRIM1* (Fig. 6a), in which the expression of the fused protein gene *myc-TaRIM1* of a *c-myc* epitope tag and *TaRIM1* was driven by a maize ubiquitin (*Ubi*) promoter and terminated by the terminator of the *Agrobacterium tumefaciens* nopaline synthase gene (*Tnos*) in a modified monocot transformation vector pAHC25<sup>31–32</sup>. The pUbi:myc-*TaRIM1* vector



**Figure 2. The sequence and phylogenetic tree analyses of TaRIM1.** (a) The nucleotide sequence and deduced amino acid sequence of TaRIM1. Green part represents R2 domain, yellow space indicates R3 domain, the nucleus localization signal is marked by red lines, and blue dotted line represents the acidic region. (b) Phylogenetic analysis of the TaRIM1 protein. The phylogenetic tree including TaRIM1 protein and 16 known-function MYB proteins is constructed using neighbor-joining phylogeny of MEGA 5.0, and is showed in bootstrapped manner.

DNA was bombarded by gene gun into immature embryos of the spring wheat cultivar Yangmai 16 for generating transgenic wheat plants. The presence of *TaRIM1* transgene cassette was detected by the desired PCR product (374 bp) using the primer pairs specific to *TaRIM1-Tnos* transgene (Fig. 6b). Based on results of PCR detection in 3 successive generations of  $T_0$ - $T_2$ , five stably transgenic lines (MO1-MO5) containing *Ubi:myc-TaRIM1* transgene were selected. qRT-PCR assays showed that the transcriptional levels of *TaRIM1* in these five *TaRIM1*-overexpressing transgenic lines were significantly elevated compared with non-transformed (wild type,



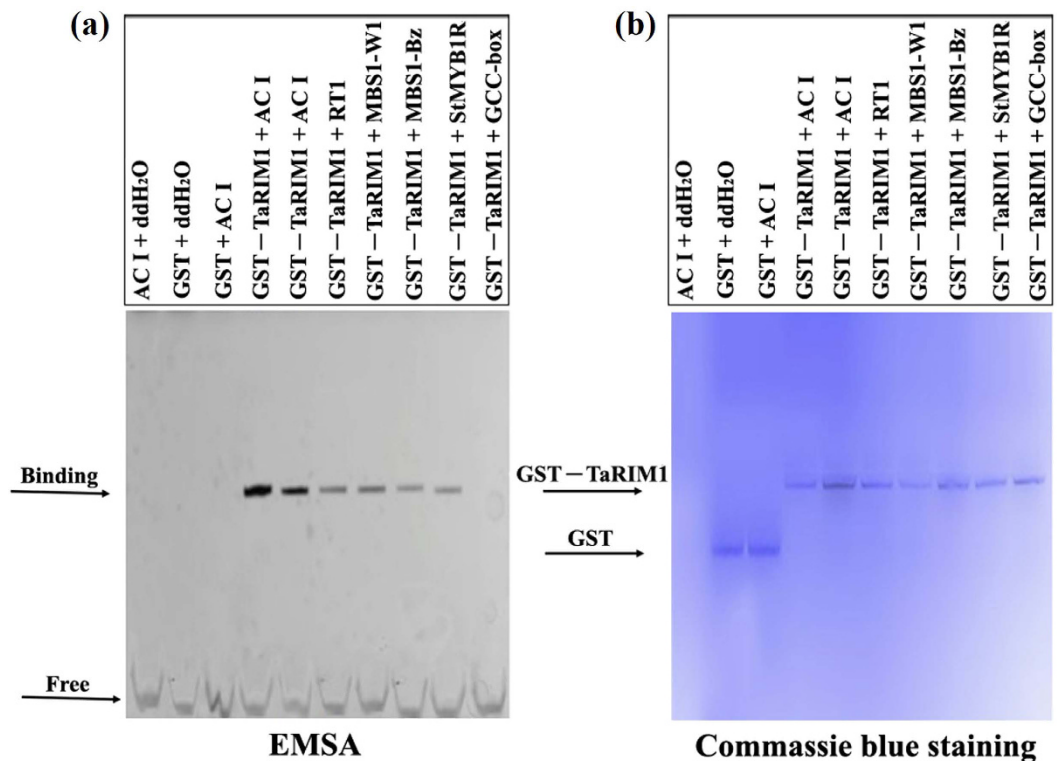


**Figure 3. Subcellular localization of the TaRIM1 protein in either the wheat mesophyll protoplasts or onion epidermal cells.** The 35S:TaRIM1-GFP and 35S:GFP constructs were separately introduced into and transiently expressed in both the wheat mesophyll protoplasts (a) and onion epidermal cells (b). The transient expressing green-fluorescence proteins (GFPs) were observed and photographed under 488 nm in Confocal Laser Scanning Microscopy (Zeiss LSM 700, Germany). Auto-fluorescence of the wheat chloroplast was observed and photographed under 639 nm in Confocal Laser Scanning Microscopy (Zeiss LSM 700, Germany).

WT) recipient wheat Yangmai 16 (Fig. 6c). Western blotting analysis indicated that the introduced *myc-TaRIM1* gene was translated into the *myc-TaRIM1* protein in these 5 overexpressing transgenic lines (MO1-MO5), but not in WT Yangmai 16 (Fig. 6d). Following inoculation with *R. cerealis* for ~47 d, all the 5 *TaRIM1*-overexpressing wheat lines in successive two ( $T_1$ - $T_2$ ) generations showed significantly enhanced-resistance to sharp eyespot compared with susceptible WT wheat Yangmai 16 (Table 1). These results indicated that TaRIM1 positively contributed to wheat defense response to *R. cerealis* infection.

**TaRIM1 positively regulates the expression of defense genes in wheat.** To explore the putative mechanism of *TaRIM1* in the resistance response to *R. cerealis* infection, we analyzed the transcript levels of 5 wheat defense genes by qRT-PCR in *TaRIM1*-silencing and overexpression wheat plants as well their controls after the pathogen inoculation. The examined genes include *Defensin* (NCBI accession no. CA630387), *PR10* (NCBI accession no. CA613496), *PR17c* (NCBI accession no. TA65181), *nsLTP1* (NCBI accession no. TC411506), and *chitinase1* (*Chit1*, NCBI accession no. CA665185). As shown in Fig. 7, the transcription levels of all the 5 defense genes were significantly decreased in susceptible *TaRIM1*-silenced wheat plants than that in BSMV:GFP infected control plants. These results suggested that the expression level of *TaRIM1* was correlated with the transcriptional levels of these defense genes. As expected, the transcription levels of these 5 defense genes were significantly elevated in resistant *TaRIM1*-overexpressing transgenic wheat lines compared to those in susceptible WT wheat Yangmai 16 plants (Fig. 8). These results indicated that TaRIM1 positively regulated, most likely activated, the expression of these defense genes in wheat.

MYB proteins can regulate the expression of defense- and stress-related genes following by interaction with MBS *cis*-elements. Our above EMSA results showed that the TaRIM1 protein could bind to 5 MBS motif sequences (Fig. 4). Furthermore, to address how TaRIM1 activates the afore-tested 5 defense genes, we obtained the promoters' sequences of these 5 defense genes from International Wheat Genome Sequencing Consortium (<http://www.wheatgenome.org/>), and then searched MBS *cis*-elements in -2000 to -1 bp promoter sequences



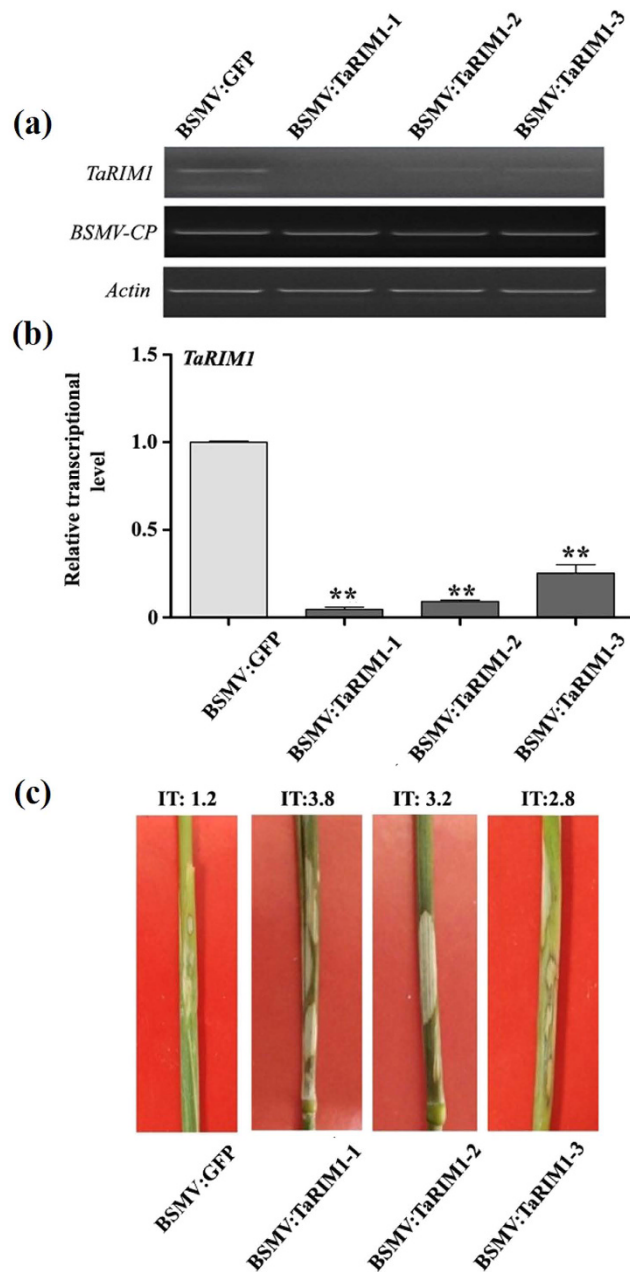
**Figure 4. Electrophoretic mobility shift assay on binding activity of TaRIM1 to MBS *cis*-element.** MBS indicates MYB-binding site. (a) The retarded band of binding of GST-TaRIM1 fusion protein to MBS probe and free probe are indicated by arrows. (b) Commaie blue staining shows the position of GST protein and GST-TaRIM1 fusion protein by arrows.

upstream of ATG of all of these genes using PLACE (<https://sogo.dna.affrc.go.jp/cgi-bin/sogo.cgi?lang=en&pj=640&action=page&page=newplace>) and the tested 5 MBS motif sequences. As shown in Table 2, the promoters of *Defensin* and *PR10* contain 4 MBS motif sequences, including ACI, MBS1, RT1, and St1R, respectively. The promoter of *Chit1* contains 3 MBS motif sequences, including ACI, MBS1, and RT1. The promoter of *PR17c* and *nsLTP1* contain 2 MBS *cis*-elements, including MBS1 and St1R. These data implied that TaRIM1 may interact with these MBS *cis*-elements in the promoters of these defense genes.

## Discussion

The sharp eyespot disease, caused primarily by the necrotrophic fungal pathogen *R. cerealis*, seriously limits the wheat production worldwide. The wheat defense response to *R. cerealis* is complicated and involves expression changes of a series of defense-related genes<sup>31,33</sup>. Identification of important genes in wheat defense response to *R. cerealis* is critical for developing wheat varieties with resistance to sharp eyespot. In plants, MYB transcription factors play important roles in development and defense responses against abiotic and biotic stresses. Many MYB genes are induced after various stress stimuli. For example, the Arabidopsis R2R3-MYB gene *BOS1* showed significant induction after *B. cinerea*, *bos1* mutant displayed more sensitivity to 2 necrotrophic pathogens, osmotic and oxidative stresses than WT plants<sup>22</sup>. In wheat, the R2R3-MYB gene *TaPIMP1* was induced after *B. sorokiniana* infection and drought stress, and positively regulated resistance responses to *B. sorokiniana* and drought stresses<sup>26</sup>. *TaLHY*, a wheat 1R-MYB gene, was induced by the infection of stripe rust pathogen strain CYR32. VIGS-based functional analysis suggest that *TaLHY* may positively participate in wheat defense response to the biotrophic fungal pathogen CYR32<sup>13</sup>. However, no MYB gene being involved in resistance to the necrotrophic pathogen *R. cerealis* has been identified yet.

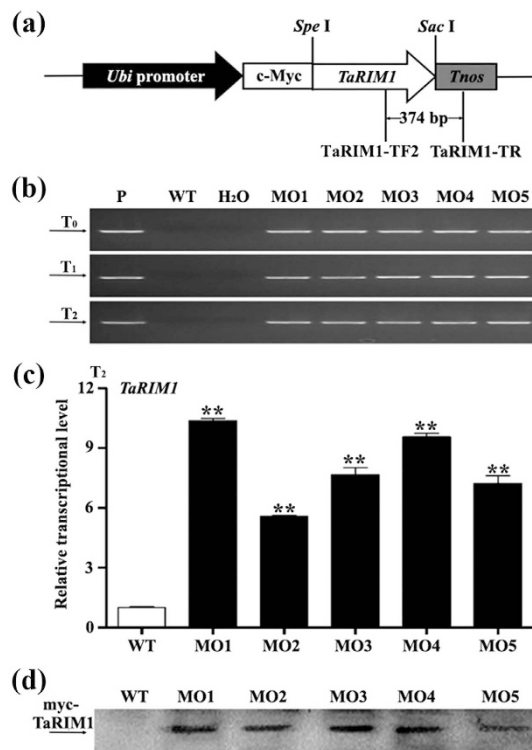
In this study, through RNA-Seq-based transcriptomic analyses, *TaRIM1*, a wheat MYB gene induced by *R. cerealis* in wheat, was identified, and cloned from *R. cerealis*-resistant wheat CI12633. In fact, some stress-induced genes play important roles in defense responses to the stresses, whereas other stress-induced genes do not. To explore the functional role of *TaRIM1* in wheat defense response to *R. cerealis*, we generated the *TaRIM1*-silencing and overexpression wheat plants. Through molecular characterization and assessment of resistance responses of *TaRIM1*-silencing and overexpression as well their control wheat plants after inoculation with the fungal pathogen, the functional dissection results displayed that silencing of *TaRIM1* did obviously impair resistance to *R. cerealis* in CI12633, *TaRIM1*-overexpression wheat lines exhibited significantly enhanced-resistance compared with susceptible WT recipient Yangmai16. These data suggest that *TaRIM1* is required for defense response against *R. cerealis* in wheat, at least for the resistant wheat line CI12633, and positively contributes to wheat defense response to *R. cerealis* infection. Moreover, the transgenic wheat produced in this study will provide potential wheat germplasm for enhancing resistance to sharp eyespot disease. To our



**Figure 5. VIGS-based functional analysis of TaRIM1 in response of wheat to *Rhizoctonia cerealis*.** (a) RT-PCR analysis of the transcription levels of *TaRIM1* and the barley stripe mosaic virus (BSMV) *CP* gene in wheat plants infected by BSMV:GFP or BSMV:TaRIM1 for 15 d. (b) qRT-PCR analysis of relative transcript level of *TaRIM1* in stems of wheat CI12633 plants infected by BSMV:TaRIM1 or BSMV:GFP at 45 dpi with *R. cerealis* strain WK207. The relative transcript level of *TaRIM1* in BSMV:TaRIM1-infected wheat plants is relative to that in BSMV:GFP-infected plants. Statistically significant differences between BSMV:GFP-infected and BSMV:TaRIM1-infected (*TaRIM1*-silencing) CI12633 plants were determined based on three replications using Student's *t*-test (\*\* $P < 0.01$ ). (c) Typical sharp eyespot phenotypes of *TaRIM1*-silencing CI12633 plants and BSMV:GFP-infected CI12633 plants after inoculation with *R. cerealis* strain WK207 for 45 days. IT indicates infection type of wheat plants to *R. cerealis* infection.

knowledge, TaRIM1 is the first reported member of MYB family positively participating in resistance response to *R. cerealis*.

In this report, Blast and phylogenetic analyses showed that the deduced protein TaRIM1 is a member of the R2R3-MYB subfamily. The sequence of the TaRIM1 protein possesses R2 and R3 MYB DNA-binding domains, a NLS and a putative transcription-activation domain, implying that the TaRIM1 protein is an activator-type R2R3-MYB transcription factor. Our biochemical and molecular-biological experiment results reveal that TaRIM1 is localized in the nuclear and can bind to all the test 5 MBS *cis*-elements. These biochemical properties



**Figure 6.** The transformation vector and molecular characterization of *TaRIM1*-overexpressing transgenic wheat plants. (a) The *TaRIM1*-overexpression transformation vector pUbi:myc-*TaRIM1*. The arrow indicates the fragment amplified in the PCR detection of the transgene. (b) PCR patterns of  $T_0$ - $T_2$  plants. P: indicates pUbi:myc-*TaRIM1* plasmid as the positive control; MO1-MO5 indicate 5 *TaRIM1* transgenic wheat lines. WT indicates non-transformation wheat (recipient) Yangmai 16. (c) qRT-PCR analysis of *TaRIM1* transcriptional levels in these 5 transgenic wheat lines. The relative transcript level of *TaRIM1* in these transgenic lines is relative to that in the WT plants. Three biological replicates each line were averaged and statistically treated (*t*-test;  $**P < 0.01$ ). Bars indicate standard error of the mean. (d) Western blot pattern of these 5 *TaRIM1*-overexpressing transgenic lines and WT Yangmai 16 using an anti-myc antibody.

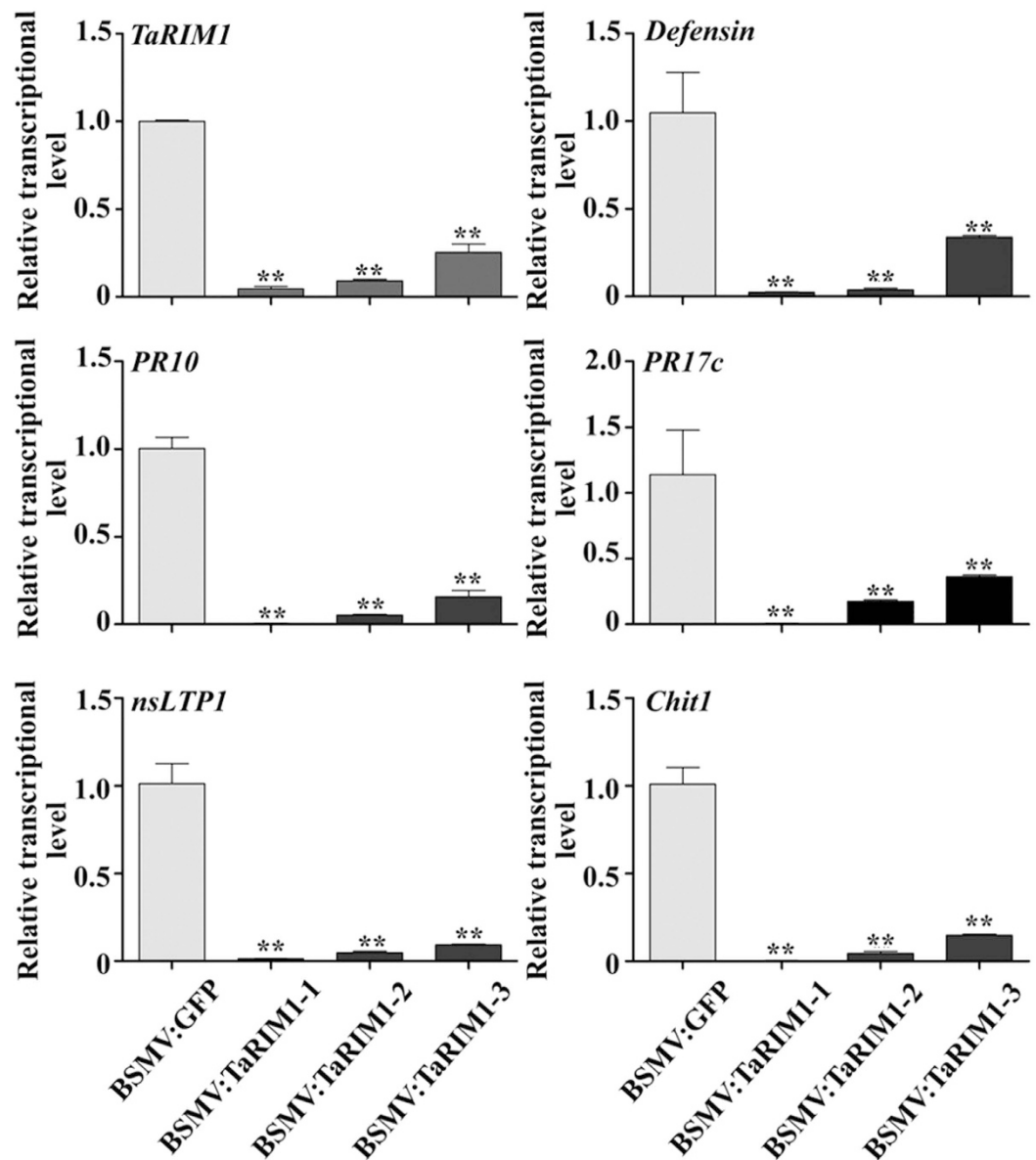
Lines	IT in $T_1$	DI in $T_1$	IT in $T_2$	DI in $T_2$
MO1	1.43**	28.60**	1.00**	20.00**
MO2	1.00**	20.00**	1.00**	20.00**
MO3	1.00**	20.00**	1.00**	20.00**
MO4	1.80**	36.00**	1.67**	33.40**
MO5	1.58**	31.60**	1.00**	20.00**
WT	2.75	55.00	2.42	48.40

**Table 1.** Sharp eyespot responses of *TaRIM1*-overexpression and wild-type wheat lines<sup>a</sup>. <sup>a</sup>MO1 to MO5 represent 5 *TaRIM1*-overexpression transgenic wheat lines, respectively. IT represents infection type. DI indicates disease index of sharp eyespot.  $T_1$  indicates  $T_1$  generation.  $T_2$  indicates  $T_2$  generation. Response value each line is the mean of at least 10 plants in each generation. Statistically significant differences between each line of *TaRIM1*-overexpression wheat (MO1 to MO5) and WT (non-transformation) recipient wheat Yangmai 16 were determined using Student's *t*-test ( $**P < 0.01$ ).

are in agreement with the sequence characteristics of *TaRIM1*, and are necessary for the active function of MYB transcription factors<sup>5</sup>.

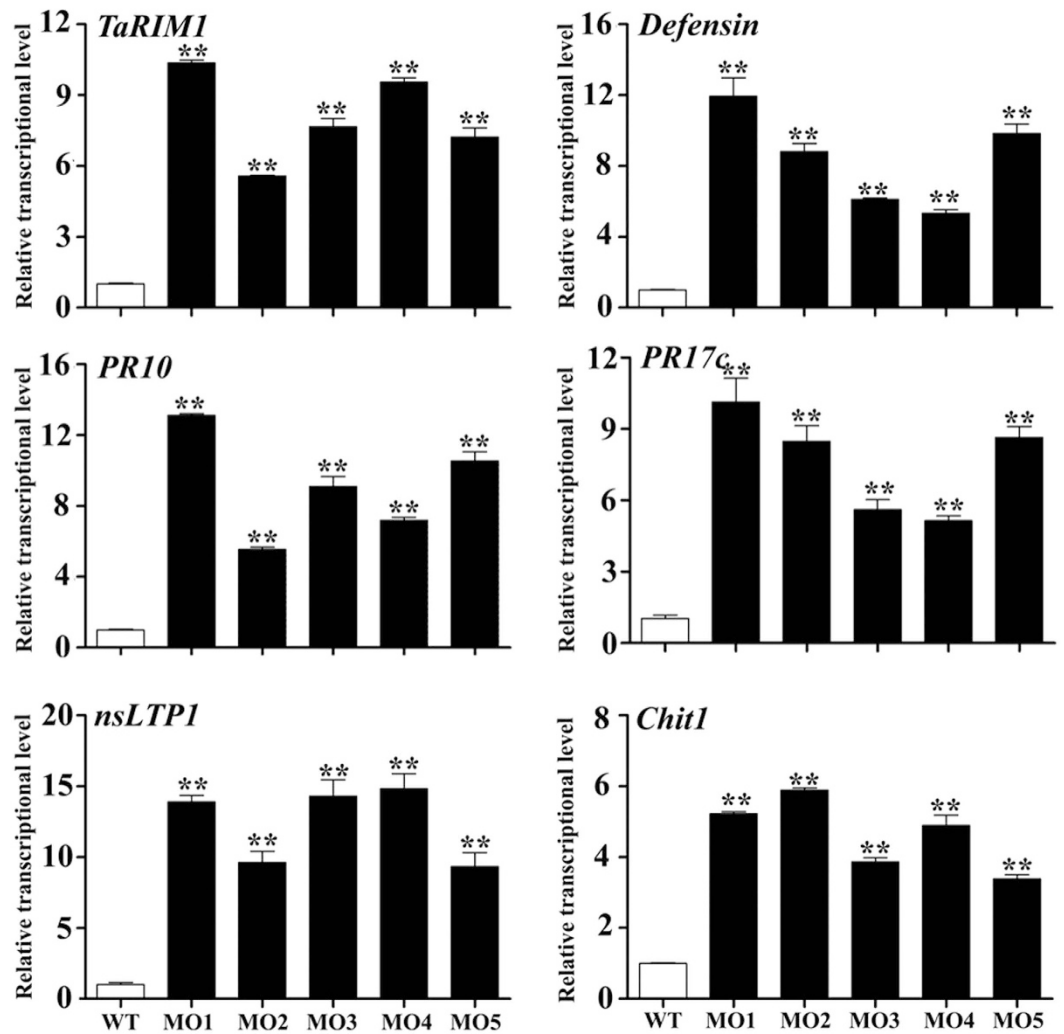
In plants, activator-type transcription factors have been implicated in defense responses through activating the expression of defense-related genes<sup>10,26–28,33–35</sup>. Defense genes positively contribute to resistance to pathogens in plants<sup>24,26</sup>. For example, transgenic wheat plants overexpressing a barley *chitinase* gene or a radish defensin gene *RsAFP2* or a wheat *LTP* gene showed enhanced-resistance to fungal pathogens<sup>36–38</sup>. To explore the putative mechanism of *TaRIM1* in wheat resistance response, we analyzed the transcriptional levels of 5 wheat defense genes, including *Defensin*, *PR10*, *PR17c*, *nsLTP1*, and *Chit1*, in *TaRIM1*-silencing and *TaRIM1*-overexpression wheat plants and their control plants. The results showed that the expression levels of these 5 defense genes were lower in *TaRIM1*-silenced wheat than in the control plants, whereas the transcriptional levels of the 5 genes in





**Figure 7.** qRT-PCR analysis of the relative transcript level of *TaRIM1* and five defense genes in *TaRIM1*-silencing and BSMV:GFP-infected CI12633 plants inoculated with *R. cerealis* for 45 d. The relative transcript level of the tested genes in *TaRIM1*-silencing wheat CI12633 plants is relative to that in BSMV:GFP-infected CI12633 plants. The examined genes include *Defensin* (NCBI accession no. CA630387), *PR10* (NCBI accession no. CA613496), *PR17c* (NCBI accession no. TA65181), *nsLTP1* (NCBI accession no. TC411506), and *chitinase1* (*Chit1*, NCBI accession no. CA665185). Statistically significant differences between *TaRIM1*-silencing wheat plants and BSMV:GFP-infected plants were determined based on three replications using Student's *t*-test (\*\* $P < 0.01$ ).

*TaRIM1*-overexpression wheat plants were elevated compared with non-transformed recipient wheat, suggesting that *TaRIM1* positively regulates the transcriptional levels of the above-tested 5 defense genes. These results suggest that overexpression of *TaRIM1* up-regulates, most likely activates, the expression of a subset of, at least the above-tested 5, defense-related genes. Many papers document that transcription factors firstly bind to specific DNA sequences (*cis*-acting elements) in target genes, and then modulate the transcription levels of these genes<sup>10,26–28,33–35,39</sup>. To address how *TaRIM1* up-regulates the expression of the 5 afore-tested defense genes, we analyzed the promoter sequences of the examined 5 defense genes in wheat. The promoter sequences of these 5 defense genes, including *Defensin*, *PR10*, *PR17c*, *nsLTP1*, and *Chit1*, contain 2–4 kinds of MBS *cis*-acting elements. Our EMSA results prove that *TaRIM1* indeed binds to these MBS *cis*-acting elements (Fig. 4). Thus, we deduce that the *TaRIM1* transcription factor interacts with the promoters of these defense genes and then activates the expression of these genes. Consequently, the expression change of a range of wheat defense-related genes



**Figure 8.** qRT-PCR analysis of the expression of *TaRIM1* and five defense genes in *TaRIM1*-overexpressing transgenic and WT lines inoculated with *R. cerealis* for 47 d. The relative transcript level of the tested genes in transgenic lines is relative to that in the WT plants. The examined genes include *Defensin* (NCBI accession no. CA630387), *PR10* (NCBI accession no. CA613496), *PR17c* (NCBI accession no. TA65181), *nsLTP1* (NCBI accession no. TC411506), and *chitinase1* (*Chit1*, NCBI accession no. CA665185). Statistically significant differences between transgenic lines with WT were determined based on three replications using Student's *t*-test (\*\* $P < 0.01$ ).

regulated by *TaRIM1* probably results in enhanced resistance in transgenic wheat to sharp eyespot caused by *R. cerealis* infection.

In conclusion, *TaRIM1*, a wheat MYB gene induced by *R. cerealis* infection, was identified and its functional role was dissected. *TaRIM1* encodes an activator-type R2R3-MYB transcription factor *TaRIM1*. *TaRIM1* positively contributes to wheat resistance response to *R. cerealis* infection through regulation of the expression of a range of defense-related genes. *TaRIM1* is a candidate gene for breeding wheat varieties with resistance to sharp eyespot caused by *R. cerealis* infection. This study provides novel insight into characteristics and functional roles of the MYB members in plants' defense responses.

## Methods

**Plant and fungal materials, and treatments.** *R. cerealis*-resistant wheat line CI12633 was used in cloning and VIGS analysis. The spring wheat cultivar Yangmai16 displaying susceptible was used as transformation recipient. Three resistant lines of RILs (cross of resistant wheat cultivar Shanhongmai and highly-susceptible wheat cultivar Wenmai 6), provided by Prof. Jizeng Jia in our Institute, were used in RNA-Sequencing.

The fungus *R. cerealis* Jiangsu-prevailing strain R0301 and North-China high-virulence strain WK207 were provided by Profs Huaigu Chen and Shibin Cai (Jiangsu Academy of Agricultural Sciences, China), and Prof. Jinfen Yu (Shandong Agricultural University), respectively.

Wheat plants were grown in a 15 h light (~22 °C)/9 h dark (~10 °C) regime. The treatments were conducted according to the protocol by Zhu *et al.*<sup>31</sup>.

Gene Name	Accession number	Site Name	Position	Sequence
<i>Defensin</i>	CA630387	MBS1	-765 to -760	CTGTTA
		RT1	-715 to -711	AACGG
		St1R	-591 to -586	GGATAA
		AC1	-373 to -368(-)	CCAACC
<i>PR10</i>	CA613496	St1R	-1777 to -1773	GGATA
		MBS1	-1608 to -1602(-)	CGGTTG
		RT1	-141 to -137(-)	AACGG
		AC1	-811 to -804	CACCTAAC
<i>PR17c</i>	TA65181	St1R	-58 to -54	GGATA
<i>nsLTP1</i>	TC411506	MBS1	-86 to -81(-)	CTGTTA
<i>Chit1</i>	CA665185	RT1	-893 to -888	AACGG
		AC1	-772 to -767(-)	CCTACC
		MBS1	-741 to -736	CGGTTG
		MBS1	-249 to -244(-)	CTGTTA

**Table 2.** MBS sequences in promoters of defense genes regulated by *TaRIM1*.

**RNA extraction and cDNA synthesis.** Total RNA was extracted using TRIZOL reagent (Qiagen, China) and subjected to DNase I (Takara, Japan) digestion and purification. The first-strand cDNA was synthesized using 2- $\mu$ g purified RNA, AMV reverse transcriptase and AP primer (5'-GGCCACGCGTCGACTAGTACTT TTTTTTTTTTTTTT-3') and Oligo dT primer according to the manual (Takara, Japan).

**Cloning of cDNA and genomic DNA full-length sequences of *TaRIM1*.** After inoculation with *R. cerealis* WK207 for 4 or 10 d, RNAs extracted from the infected base stems and sheaths of three resistant lines of the RILs were deeply sequenced. The further transcriptomic analyses were performed by Bioinformatics to identify up-regulated genes (Data unpublished). Among them, one with no. Traes\_6BL\_E5A9546C9 is homologous to the *TaMYB33* sequence, thus this corresponding gene was named *TaRIM1*.

To obtain the *R. cerealis*-resistance-related sequence of *TaRIM1*, used as the template to the full-length cDNA of *TaRIM1* was obtained from infected stems' cDNA of resistant wheat CI12633 by two rounds of 3'RACE reactions. In 1<sup>st</sup> round, the primers TaRIM1F-F1 (5'-GCAGCATTTACCTTCGGAC-3') and AUAP (5'-GGCCACGCGTCGACTAGTAC-3') were used. The primers TaRIM1F-F2 (5'-CGTCAACA CACTGAGCAATC-3') and AUAP (5'-GGCCACGCGTCGACTAGTAC-3') were used in 2<sup>nd</sup> round. Genomic sequence of *TaRIM1* was obtained from CI12633 genomic DNA by 2 round PCR amplifications and 2 pairs of primers (TaRIM1F-F1: 5'-GCAG CATTTACCTTCGGAC-3', TaRIM1F-R1: 5'-AACGATTATGTTCCTTCACA-3', TaRIM1F-F2: 5'-CGTCAACACACTGAGCAATC-3', and TaRIM1F-R2: 5'-AAACTA GCCGAGGAGCCG-3'). The purified PCR products were cloned into the pMD-18T vector (Takara) and sequenced.

Sequence blast was performed online using the website (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). Protein sequence analysis was performed using SMART (<http://smart.embl-heidelberg.de/>). The NLS region was predicted by cNLS Mapper ([http://nls-mapper.iab.keio.ac.jp/cgi-bin/NLS\\_Mapper\\_form.cgi](http://nls-mapper.iab.keio.ac.jp/cgi-bin/NLS_Mapper_form.cgi)). Phylogenetic tree analysis was performed using MEGA 5.0.

**Subcellular localization of *TaRIM1* in onion epidermal cells.** The *TaRIM1* coding region lacking the stop codon was amplified using primers TaRIM1-GFP-Hind III-F (5'-AGTAAAGCTTATGGGGAGGTC TCCTTGC-3', underline represents the *Hind* III restriction site) and TaRIM1-GFP-BamHI-R (5'-GCGGGATCCA ATCTGAGACAAACTCTG-3', underline represents the *Bam*H I restriction site). The PCR products was digested with restriction enzymes *Hind* III and *Bam*H I, then sub-cloned in-frame into 5'-terminus of GFP coding region in the p35S:GFP vector, resulting in the TaRIM1-GFP fusion construct p35S:TaRIM1-GFP.

The p35S:TaRIM1-GFP or p35S:GFP alone construct was separately introduced into wheat leaf protoplasts via the PEG-mediated transfection method following the protocol of Yoo *et al.*<sup>40</sup> or transformed into onion epidermal cell by particle bombardment following Zhang *et al.*<sup>41</sup>. After incubation at 25 °C for 12 h, GFP signals were then observed and photographed using Confocal Laser Scanning Microscopy (Zeiss LSM 700, Germany).

**VIGS-based functional analysis of *TaRIM1*.** The specific fragment (no. 576–836 nt) of *TaRIM1* was amplified from CI12633 cDNA by primers TaRIM1-NheIF (5'-ACAGCTAGCTCGTCCGCCACCGATTACT-3', underline represents *Nhe* I restriction site) and TaRIM1-NheIR (5'-GGCGCTAGCCTC TGCTAAATCTGAG ACAAC-3' underline represents *Nhe* I restriction site). PCR products were digested with *Nhe* I, then ligated into the BSMV- $\gamma$  vector, resulting in recombinant vector BSMV- $\gamma$ -TaRIM1.

BSMV:TaRIM1 and control virus BSMV:GFP viruses were used to inoculate the CI12633 using protocol following Zhu *et al.*<sup>31</sup>. At 14 d after infection, the fourth leaves of the inoculated seedlings were collected to monitor BSMV infection, the transcription of the BSMV coat protein (*CP*) gene with the specific primers (BSMV-CPF: 5'-TGACTGCTAAGGGTGGAGGA-3', BSMV-CPR: 5'-CGGTTGAACATCACGAAG AGT-3') and RT-PCR was used to evaluate if BSMV inoculates wheat plants. At 20 days after BSMV infection, BSMV infected plants

were inoculated with the small toothpick fragments harboring the well-developed mycelia of *R. cerealis* WK 207<sup>34</sup>. At 45 dpi with *R. cerealis*, ITs were scored, and sharp eyespot symptoms were photographed.

**EMSA on TaRIM1 binding activity to MBS *Cis*-elements.** The coding sequence of *TaRIM1* with *EcoR* I and *Xho* I restriction sites was amplified with the primers (GF: 5'-GATGAATTCAGAGGGGGCCGTGGACG-3', underline represents *EcoR* I restriction site. GR: 5'-CTACTCGAGCTGGCCGACGTCTTGGGA-3' underline represents *Xho* I restriction site) and then sub-cloned in-frame into 3'-terminus of GST in pGEX-4T-1 vector. The resulting GST-TaRIM1 recombinant protein was expressed in *Escherichia coli* BL21 cells after induction with 0.3 mM isopropyl- $\beta$ -D-thiogalactopyranose, and purified using MicroSpin module (GE Amersham).

The primers of 6 *cis*-element probes including 5 MBS and GCC-box were synthesized as the sequences in Table S1<sup>26</sup>.

Following a modified EMSA protocol<sup>42</sup>, each probe was mixed with ~2  $\mu$ g of recombinant GST-TaRIM1 or GST in a binding buffer. Each reaction mixture was incubated on ice for 6 h and loaded onto 8% polyacrylamide gel. After electrophoresis was performed at 100 V for 30 min, the gels were stained with ethidium bromide for visualization of the DNA bands.

**Generation and PCR detection of *TaRIM1*-overexpressing transgenic wheat.** ORF of *TaRIM1* was amplified with the primers (TVF: 5'-CAAACTAGTATGGGGAGG TCTCCTTGC-3', underline represents *Spe* I restriction site, TVR: 5'-CGTGAGCTCCT AAATCTGAGACAAACT-3', underline represents *Sac* I restriction site) to sub-clone in-frame with a *c-myc* epitope tag in a modified pAHC25-myc vector<sup>31,32</sup>, resulting in *TaRIM1*-overexpression transformation vector pUbi:myc-TaRIM1 (Fig. 6a). In predicted transformed plants, the expression of the fused protein gene *myc-TaRIM1* of a *c-myc* epitope tag and *TaRIM1* was driven by a maize ubiquitin (*Ubi*) promoter and terminated by the terminator of the *Agrobacterium tumefaciens* nopaline synthase gene (*Tnos*).

The resulting transformation vector pUbi:myc-TaRIM1 plasmid was transformed into 1,200 immature embryos of wheat Yangmai 16 by biolistic bombardment following Chen *et al.*<sup>34</sup>.

The overexpressing transgene was detected by a PCR product (374 bp) specific to the transgene using specific primers (TaRIM1-TF1: 5'-CGGACAACGAGATCAA GAAC-3', TaRIM1-TF2: 5'-AGCTTTACATCGGA GGAGTTTCAG-3' locating in the *TaRIM1* coding region, TaRIM1-TR: 5'-AAAACCCATCTCATAAATAACG-3' locating in *Tnos*) with 2 round of nested amplifications.

**Western blot analysis of myc-TaRIM1 protein in transgenic wheat.** The protein expression of introduced c-myc-TaRIM1 was evaluated by Western blotting. Total proteins were extracted from 0.4 g of ground base stem powders. About 10  $\mu$ g of total soluble proteins were separated on 12% sodium dodecyl sulfate polyacrylamide gels and transferred to PVDF membrane. The Western blots were incubated with a 2000-fold dilution of c-myc antibody (TransGen Biotech, China) at 4 °C for 12 h, and then with 1000-fold dilution of secondary antibody conjugated to horseradish peroxidase (TransGen Biotech, China) at ~25 °C for 1 h. The expressing myc-TaRIM1 proteins were visualized using the Pro-light HRP Chemiluminescent Kit (TIANGEN Biotech, China).

**qRT-PCR analyses of *TaRIM1* and defense genes.** qRT-PCR technique was used to examine the relative transcriptional levels of *TaRIM1* (*TaRIM1*-QF: 5'-CGACTCGTCCTCGTCCAAG-3', *TaRIM1*-QR: 5'-CGACGACG GCATCGAGTAAT-3') and five defense-marker genes in *TaRIM1*-silencing and overexpression wheat plants as well as their control wheat plants. These defense genes include *Defensin* (*Defensin*-Q-F: 5'-ATGTCCCGTGCCTTTTGCTA-3', *Defensin*-Q-R: 5'-CCAAACTACCGAGTCCCCG-3'), *PR10* (*PR10*-Q-F: 5'-CGTGGAGGTAAACGA TGAG-3', *PR10*-Q-R: 5'-GCTAAGTGTCCGGGGTAAT-3'), *PR17c* (*PR17c*-Q-F: 5'-ACGACATCACGGCGAGGT-3', *PR17c*-Q-R: 5'-CACGGGAAAGAGAGGATGA-3'), *nsLTP1* (*nsLTP1*-Q-F: 5'-ATGCGGGTTGGCGTGAAG-3', *nsLTP1*-Q-R: 5'-TGT TGCGGTGGTAGGTTGTTG-3') and *chitinase1* (*Chit1*-Q-F: 5'-ATGCTCTGGGACC GATACTT-3', *Chit1*-Q-R: 5'-AGCCTCACTTTGTTCTCGTTT-3'). qRT-PCR was performed (95 °C for 5 min, 41 cycles of 95 °C for 15 s and 60 °C for 31 s) using SYBR Green I Master Mix (Takara) using ABI PRISM 7300 detective system according to the manufacturer's instruction. The relative transcript levels were calculated using the  $2^{-\Delta\Delta CT}$  method<sup>43</sup>, where the wheat *actin* gene (NCBI accession no. BE425627) with the primers ActA: (5'-CACTGGAATGGTCAAGGCTG-3') and ActB (5'-CTCCATGTC ATCCAGTTG-3') was used as the internal reference.

**Assessment on wheat response to *R. cerealis*.** At least 10 plants for each wheat line were inoculated with developed mycelia of *R. cerealis* following the protocol of Wei *et al.*<sup>44</sup>. In T<sub>1</sub> generation, *R. cerealis* WK207 was used to inoculate plants, whereas *R. cerealis* isolate R0301 was used to inoculate T<sub>2</sub> and WT plants. ITs and disease index for wheat line were evaluated at ~47 dpi according to Wei *et al.*<sup>44</sup>.

## References

- Chen, J. *et al.* Mapping of QTL conferring resistance to sharp eyespot (*Rhizoctonia cerealis*) in bread wheat at the adult plant growth stage. *Theor. Appl. Genet.* **126**, 2865–2878 (2013).
- Hamada, M. S., Yin, Y. N., Chen, H. G. & Ma, Z. H. The escalating threat of *Rhizoctonia cerealis*, the causal agent of sharp eyespot in wheat. *Pest Manag. Sci.* **67**, 1411–1419 (2011).
- Lemańczyk, G. & Kwaśna, H. Effects of sharp eyespot (*Rhizoctonia cerealis*) on yield and grain quality of winter wheat. *Eur. J. Plant Pathol.* **135**, 187–200 (2013).
- Cai, S. B. *et al.* Germplasm development and mapping of resistance to sharp eyespot (*Rhizoctonia cerealis*) in wheat. *Scientia Agricultura Sinica*. **39**, 928–934 (2006). (Chinese with English abstract).
- Al-Attala, M. N. *et al.* A novel *TaMYB4* transcription factor involved in the defence response against *Puccinia striiformis* f. sp. *tritici* and abiotic stresses. *Plant Mol. Biol.* **84**, 589–603 (2014).



6. Dubos, C. *et al.* MYB transcription factors in Arabidopsis. *Trends Plant Sci.* **15**, 573–581 (2010).
7. Paz-Ares, J., Ghosal, D., Wienand, U., Peterson, P. A. & Saedler, H. The regulatory *c1* locus of *Zea mays* encodes a protein with homology to *myb* proto-oncogene products and with structural similarities to transcriptional activators. *EMBO J.* **6**, 3553–3558 (1987).
8. Abe, H. *et al.* Arabidopsis AtMYC2 (bHLH) and AtMYB2 (MYB) function as transcriptional activators in abscisic acid signaling. *Plant Cell.* **15**, 63–78 (2003).
9. Dai, X. Y. *et al.* Overexpression of an R1R2R3 MYB gene, *OsMYB3R-2*, increases tolerance to freezing, drought, and salt stress in transgenic Arabidopsis. *Plant Physiol.* **143**, 1739–1751 (2007).
10. Ma, Q. B. *et al.* Enhanced tolerance to chilling stress in *OsMYB3R-2* transgenic rice is mediated by alteration in cell cycle and ectopic expression of stress genes. *Plant Physiol.* **150**, 244–256 (2009).
11. Millar, A. A. & Gubler, F. The Arabidopsis *GAMYB-like* genes, *MYB33* and *MYB65*, are microRNA-regulated genes that redundantly facilitate anther development. *Plant Cell.* **17**, 705–721 (2005).
12. Su, C. F. *et al.* A novel MYB3-dependent pathway confers cold tolerance in rice. *Plant Physiol.* **153**, 145–158 (2010).
13. Zhang, Z. J. *et al.* *TaLHY*, a 1R-MYB Transcription Factor, Plays an Important Role in Disease Resistance against Stripe Rust Fungus and Ear Heading in Wheat. *PLoS One.* **10**, e0127723 (2015).
14. Zhou, J. L., Lee, C. H., Zhong, R. Q. & Ye, Z. H. MYB58 and MYB63 are transcriptional activators of the lignin biosynthetic pathway during secondary cell wall formation in Arabidopsis. *Plant Cell.* **21**, 248–266 (2009).
15. He, Y. N. *et al.* Ectopic expression of a wheat MYB transcription factor gene, *TaMYB73*, improves salinity stress tolerance in Arabidopsis thaliana. *J. Exp. Bot.* **63**, 1511–1522 (2012).
16. Mao, X. G. *et al.* Transgenic expression of *TaMYB2A* confers enhanced tolerance to multiple abiotic stresses in Arabidopsis. *Funct. Integr. Genomics.* **11**, 445–465 (2011).
17. Qin, Y. X. *et al.* Over-expression of *TaMYB33* encoding a novel wheat MYB transcription factor increases salt and drought tolerance in Arabidopsis. *Mol. Biol. Rep.* **39**, 7183–7192 (2012).
18. Stracke, R., Werber, M. & Weisshaar, B. The *R2R3-MYB* gene family in Arabidopsis thaliana. *Curr Opin Plant Biol.* **4**, 447–456 (2001).
19. Xue, G. P., Kooiker, M., Drenth, J. & McIntyre, C. L. *TaMYB13* is a transcriptional activator of fructosyltransferase genes involved in  $\beta$ -2, 6-linked fructan synthesis in wheat. *Plant J.* **68**, 857–870 (2011).
20. Zhang, L. C. *et al.* A wheat R2R3-MYB gene, *TaMYB30-B*, improves drought stress tolerance in transgenic Arabidopsis. *J. Exp. Bot.* **63**, 5873–5885 (2012).
21. Zhong, R. Q., Lee, C. H., Zhou, J. L., McCarthy, R. L. & Ye, Z. H. A battery of transcription factors involved in the regulation of secondary cell wall biosynthesis in Arabidopsis. *Plant Cell.* **20**, 2763–2782 (2008).
22. Mengiste, T., Chen, X., Salmeron, J. & Dietrich, R. The *BOTRYTIS SUSCEPTIBLE1* gene encodes an R2R3MYB transcription factor protein that is required for biotic and abiotic stress responses in Arabidopsis. *Plant Cell.* **15**, 2551–2565 (2003).
23. Seo, P. J. *et al.* The MYB96 transcription factor mediates abscisic acid signaling during drought stress response in Arabidopsis. *Plant Physiol.* **151**, 275–289 (2009).
24. Seo, P. J. & Park, C. M. MYB96-mediated abscisic acid signals induce pathogen resistance response by promoting salicylic acid biosynthesis in Arabidopsis. *New Phytol.* **186**, 471–483 (2010).
25. Chang, C. *et al.* Barley MLA immune receptors directly interfere with antagonistically acting transcription factors to initiate disease resistance signaling. *Plant Cell.* **25**, 1158–1173 (2013).
26. Zhang, Z. Y. *et al.* An R2R3 MYB transcription factor in wheat, *TaPIMP1*, mediates host resistance to *Bipolaris sorokiniana* and drought stresses through regulation of defense- and stress-related genes. *New Phytol.* **196**, 1155–1170 (2012).
27. Liu, X. *et al.* Transgenic wheat expressing *Thinopyrum intermedium* MYB transcription factor *TiMYB2R-1* shows enhanced resistance to the take-all disease. *J. Exp. Bot.* **64**, 2243–2253 (2013).
28. Fujimoto, S. Y., Ohta, M., Usui, A., Shinshi, H. & Ohme-Takagi, M. Arabidopsis ethylene-responsive element binding factors act as transcriptional activators or repressors of GCC box mediated gene expression. *Plant Cell.* **12**, 393–404 (2000).
29. Holzberg, S., Brosio, P., Gross, C. & Pogue, G. P. Barley stripe mosaic virus-induced gene silencing in a monocot plant. *Plant J.* **30**, 315–327 (2002).
30. Scofield, S. R., Huang, L., Brandt, A. S. & Gill, B. S. Development of a virus-induced gene-silencing system for hexaploid wheat and its use in functional analysis of the *Lr21*-mediated leaf rust resistance pathway. *Plant Physiol.* **138**, 2165–2173 (2005).
31. Zhu, X. L. *et al.* The wheat AGC kinase *TaAGC1* is a positive contributor to host resistance to the necrotrophic pathogen *Rhizoctonia cerealis*. *J. Exp. Bot.* **66**, 6591–6603 (2015).
32. Christensen, A. H. & Quail, P. H. Ubiquitin promoter-based vectors for high-level expression of selectable and/or screenable marker genes in monocotyledonous plants. *Transgenic Res.* **5**, 213–218 (1996).
33. Zhu, X. L. *et al.* The wheat ethylene response factor transcription factor pathogen-induced ERF1 mediates host responses to both the necrotrophic pathogen *Rhizoctonia cerealis* and freezing stresses. *Plant Physiol.* **164**, 1499–1514 (2014).
34. Chen, L. *et al.* Overexpression of *TiERF1* enhances resistance to sharp eyespot in transgenic wheat. *J. Exp. Bot.* **59**, 4195–4204 (2008).
35. Zhang, H. *et al.* Tomato stress-responsive factor *TSRF1* interacts with ethylene responsive element GCC box and regulates pathogen resistance to *Ralstonia solanacearum*. *Plant Mol. Biol.* **55**, 825–834 (2004).
36. Li, Z. *et al.* Expression of a radish defensin in transgenic wheat confers increased resistance to *Fusarium graminearum* and *Rhizoctonia cerealis*. *Funct. Integr. Genomics.* **11**, 63–70 (2011).
37. Shin, S. *et al.* Transgenic wheat expressing a barley class II chitinase gene has enhanced resistance against *Fusarium graminearum*. *J. Exp. Bot.* **59**, 2371–2378 (2008).
38. Zhu, X. L. *et al.* Overexpression of wheat lipid transfer protein gene *TaLTP5* increases resistances to *Cochliobolus sativus* and *Fusarium graminearum* in transgenic wheat. *Funct. Integr. Genomics.* **12**, 481–488 (2012).
39. Cheng, H. Q. *et al.* The cotton MYB108 forms a positive feedback regulation loop with CML11 and participates in the defense response against *Verticillium dahliae* infection. *J. Exp. Bot.* **67**, 1935–1950 (2016).
40. Yoo, S. D., Cho, Y. H. & Sheen, J. Arabidopsis mesophyll protoplasts: a versatile cell system for transient gene expression analysis. *Nat. Protoc.* **2**, 1565–1572 (2007).
41. Zhang, Z. Y. *et al.* A novel ERF transcription activator in wheat and its induction kinetics after pathogen and hormone treatments. *J. Exp. Bot.* **58**, 2993–3003 (2007).
42. Dong, N. *et al.* Overexpression of *TaPIEP1*, a pathogen-induced ERF gene of wheat, confers host-enhanced resistance to fungal pathogen *Bipolaris sorokiniana*. *Funct. Integr. Genomics.* **10**, 215–226 (2010).
43. Livak, K. J. & Schmittgen, T. D. Analysis of relative gene expression data using real-time quantitative PCR and the  $2^{-\Delta\Delta CT}$  Method. *Methods.* **25**, 402–408 (2001).
44. Wei, X. N. *et al.* The wheat calcium-dependent protein kinase *TaCPK7-D* positively regulates host resistance to sharp eyespot disease. *Mol. Plant Pathol.*, doi:10.1111/mpp.12360 (2015).

## Acknowledgements

The authors are very grateful to Profs Jizeng Jia (Institute of Crop Science, CAAS), Daowen Wang (Chinese Academy of Sciences, China), Huaigu Chen (Jiangsu Academy of Agricultural Sciences, China), and Jinfeng Yu

(Shandong Agricultural University, China) for their kind providence of the RILs materials, the original vector p35S:GFP, and *R. cerealis* strains R0301 and WK207. This study was supported by the programs of a National Natural Science Foundation of China (Grant no. 31471494) and a National “Key Sci-Tech” Project (2016ZX08002001-4).

### Author Contributions

Z.Z. designed the research and wrote the paper. T.S., W.R. and X.L. performed the cloning, sequencing, subcellular localization, EMSA, VIGS and functional assays. H.X. and L.D. conducted transformation.

### Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

**Competing financial interests:** The authors declare no competing financial interests.

**How to cite this article:** Shan, T. *et al.* The wheat R2R3-MYB transcription factor TaRIM1 participates in resistance response against the pathogen *Rhizoctonia cerealis* infection through regulating defense genes. *Sci. Rep.* **6**, 28777; doi: 10.1038/srep28777 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>