

SCIENTIFIC REPORTS

OPEN

Titanium Dioxide Photocatalytic Polymerization of Acrylamide for Gel Electrophoresis (TIPAGE) of Proteins and Structural Identification by Mass Spectrometry

Wenyang Zhang, Zhiwei Yuan, Lulu Huang, Jie Kang, Ruowei Jiang & Hongying Zhong

Polyacrylamide gel electrophoresis (PAGE) coupled with mass spectrometry has been well established for separating, identifying and quantifying protein mixtures from cell lines, tissues or other biological samples. The copolymerization process of acrylamide and bis-acrylamide is the key to mastering this powerful technique. In general, this is a vinyl addition reaction initiated by free radical-generating reagents such as ammonium persulfate (APS) and tetramethylethylenediamine (TEMED) under basic pH and degassing experimental condition. We report herein a photocatalytic polymerization approach that is based on photo-generated hydroxyl radicals with nanoparticles of titanium dioxide. It was shown that the polymerization process is greatly accelerated in acidic condition when ultraviolet light shots on the gel solution containing TiO₂ nanoparticles without degassing. This feature makes it very useful in preparing Triton X-100 acid urea (TAU) gel that has been developed for separating basic proteins such as histones and variants in acidic experimental condition. Additionally, the presence of titanium dioxide in the gel not only improves mechanistic property of gels but also changes the migration pattern of different proteins that have different affinities to titanium dioxide.

Since the invention in last century, polyacrylamide gel electrophoresis remains the most powerful separation technique for proteomics^{1–3} so far. By using this technique, thousands of cellulosic proteins can be simultaneously fractionated and displayed with unparalleled high resolution in a gel. Coupled to mass spectrometry, it becomes an indispensable tool for protein structural identification^{4–6}. Polyacrylamide gels are usually formed by copolymerization of acrylamide and bis-acrylamide with ammonium persulfate (APS) and tetramethylethylenediamine (TEMED) as the initiator and catalyst. There are mainly three steps for such gel preparation. (A) Initiation of free radicals. Free radicals are generated from ammonium persulfate and TEMED accelerates the rate of formation of free radicals from ammonium persulfate. (B) Elongation of polymer chains. Resultant free radicals further convert acrylamide monomers to free radicals that can react with unactivated acrylamide monomers to start elongation of polymer chains. In the meanwhile, the elongating polymer chains are crosslinked with bis-acrylamide, resulting in the formation of characteristic pores. (C) Termination of polymerization cascade. As the polymerization reaction proceeds, acrylamide monomers are gradually consumed and polymer chain elongation stops. For all these reactions, the pH value of the gel solution should be kept between 8 and 9 in order for ammonium persulfate and TEMED to act efficiently. Additionally, because TEMED is prone to oxidative reaction which causes the loss of catalytic activity, the gel solution should also be degassed under a vacuum of 125 torr or better for at

Mass Spectrometry Center for Structural Identification of Biological Molecules and Precision Medicine, Key Laboratory of Pesticides and Chemical Biology, Ministry of Education, College of Chemistry, Central China Normal University, Wuhan, Hubei 430079, P. R. China. Correspondence and requests for materials should be addressed to H.Z. (email: hyzhong@mail.ccnu.edu.cn)

least 15 minutes at room temperature. The major challenge of this protocol is the analysis of basic proteins such as histones and variants in acidic condition under which it takes very long time to complete polymerization process.

Histones and variants represent a very important class of proteins that contribute to the encoding of epigenetic information and regulation of gene expression^{7–9}. Because of their similar molecular masses and extensive post-translational modifications, the resolution of current sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS PAGE) is not enough for fine separation of these proteins. Considering on their insolubility in neutral and basic solution, acidic polyacrylamide gels with urea and Triton X-100 as additives have been developed to separate acid extracted histones and histone variants on the basis of differences in charges^{10–12}. In this technique, proteins are denatured by Triton X-100 or urea without affecting the charge of proteins. Under acidic running condition, the charge of proteins is solely determined by the number of protonated groups. Different modification levels of acetylation and phosphorylation of histone isoforms are mostly favored by this approach because they greatly affect the migration of histones through reduction of the positive charge of a Lys or introduction of an additional negative charge respectively. Despite the great success, preparation of acidic polyacrylamide gels is still very difficult by using ammonium persulfate and TEMED as the initiator and catalyst for polymerization reaction in acidic condition. It usually takes more than 1 hour to complete gelation process. In particular, the mechanistic property of resultant gels is not strong due to incomplete chain elongation and crosslinking.

Photocatalytic production of electron-hole pairs in the aqueous suspension of titanium dioxide under ultraviolet irradiation has been reported^{13–15} for many years. It is well-established that superoxide (O_2^-) and hydroxyl radical ($\cdot OH$) can be generated by O_2 capture of photoexcited electrons in conduction band and hole oxidation of H_2O respectively. More than that, semiconductor nanoparticles have been increasingly recognized to initiate polymerization process with light excitation^{16–22}. The photopolymerization technique provides a new way to prepare polymer/inorganic nanocomposites. This work is aimed to investigate the polymerization reaction of acrylamide and bis-acrylamide initiated by hydroxyl radicals in acidic condition. Compared with Azo-based initiator such as VA-086 which is also used for introducing hydroxyl group into terminal of polymer^{23–25}, semiconductor nanoparticles not only speed up the polymerization process under light irradiation but also improve the mechanical properties because of the formation of cross-linked nanocomposites. The so called TiO_2 -PAGE approach (titanium dioxide photocatalytic polymerization of acrylamide for gel electrophoresis) should be able to provide an alternative approach for proteomics.

Experimental Section

Preparation of Triton-Acid-Urea (TAU) Polyacrylamide Gel by Titanium Dioxide Photocatalytic Polymerization Reaction.

All chemicals and apparatus are listed in Supplementary Notes 1. The components of separating gel and stacking gel were listed in Supplementary Table 1. The stacking gel was prepared by using ammonium persulfate and TEMED as the initiator and catalyst. As for the separating gel, nanoparticles of titanium dioxide were mixed with other chemicals after they were completely dissolved in water. The mixture was gently stirred until a uniform suspension of titanium dioxide was formed and polymerization process started. Air bubbles were carefully removed with a pipette. Then the mixture was gently poured to the gel casting model to 3 cm below top of plate and pure water was overlaid on the top. The separating gel was irradiated with ultraviolet light for 20 minutes to allow adequate polymerization. At room temperature (23–25 °C), gelation can complete in about 10 minutes. Additional 10 minutes is necessary to ensure full reaction. The stacking gel was casted on the top of the separating gel. After poured off water from the top of the polymerized separating gel, the stacking gel solution containing APS and TEMED was poured to 1.5 cm below the edge of the glass plate and a comb was inserted. Then pure water was overlaid on the top and it was allowed to polymerize for 1 hour. The gel should be made freshly right before downstream electrophoresis.

Triton-Acid-Urea (TAU) Gel Electrophoresis with Titanium Dioxide Photocatalytically Polymerized Acrylamide Gel.

In order to remove un-reacted residues, it is recommended to pre-run the freshly prepared gel at 100 volts for at least 1 hour. Histones were extracted with H_2SO_4 as previously reported²⁶. Lyophilized histone powder (3 mg) or standard casein/de-phosphorylated casein was dissolved in 600 μl acidic sample buffer containing 0.36 g urea, 50 μl glacial acetic acid and 30 μl 40 mg/ml $CuSO_4$ as well as 60 μl 0.2% Pyronin Y. $CuSO_4$ was added to the sample buffer so that to chelate with histones and improve separation resolution²⁶. Pyronin Y was used a marker to visualize running front line. Freshly prepared separating gel and stacking gel are installed in a mini gel system for electrophoresis experiments. About 50 μg histone proteins (10 μl sample buffer) or 10 μg standard casein/de-phosphorylated casein (2 μl sample buffer) were loaded into each lane. The running buffer contains 5% acetic acid solution. In order to concentrate the sample, first set the work voltage as 30 Volts and allow the gel to run for about 45 minutes. When the red dye pyronin Y was condensed into a straight line at the bottom of the stacking gel, increase the work voltage to 100 Volts and allow it to run for about 200 minutes. Finally, the TAU gel was stained as described in Supplementary Notes 2. Mass spectrometric identification of gel separated proteins was performed with a Waters MALDI Synapt G2 HDMS system (MA, USA) in positive ion and sensitivity mode as described in Supplementary Notes 3.

Results and Discussion

Chemical Polymerization Versus Photochemical Polymerization. As shown in Supplementary Fig. 1(A–C), ammonium persulfate and TEMED start the free radical reaction cascade. All three steps including initiation, elongation and termination are required to react in basic pH gel forming solution at which initiators are effective. The free base form of TEMED is essential for initiation reaction. At low pH, TEMED may become protonated, resulting in very slow polymerization. Careful examination of these reactions also reveals that the presence of oxygen may cause oxidation of TEMED and inhibition of polymerization reaction. Instead, the mechanism of photochemical polymerization was shown in Fig. 1. With ultraviolet irradiation, hydroxyl radicals

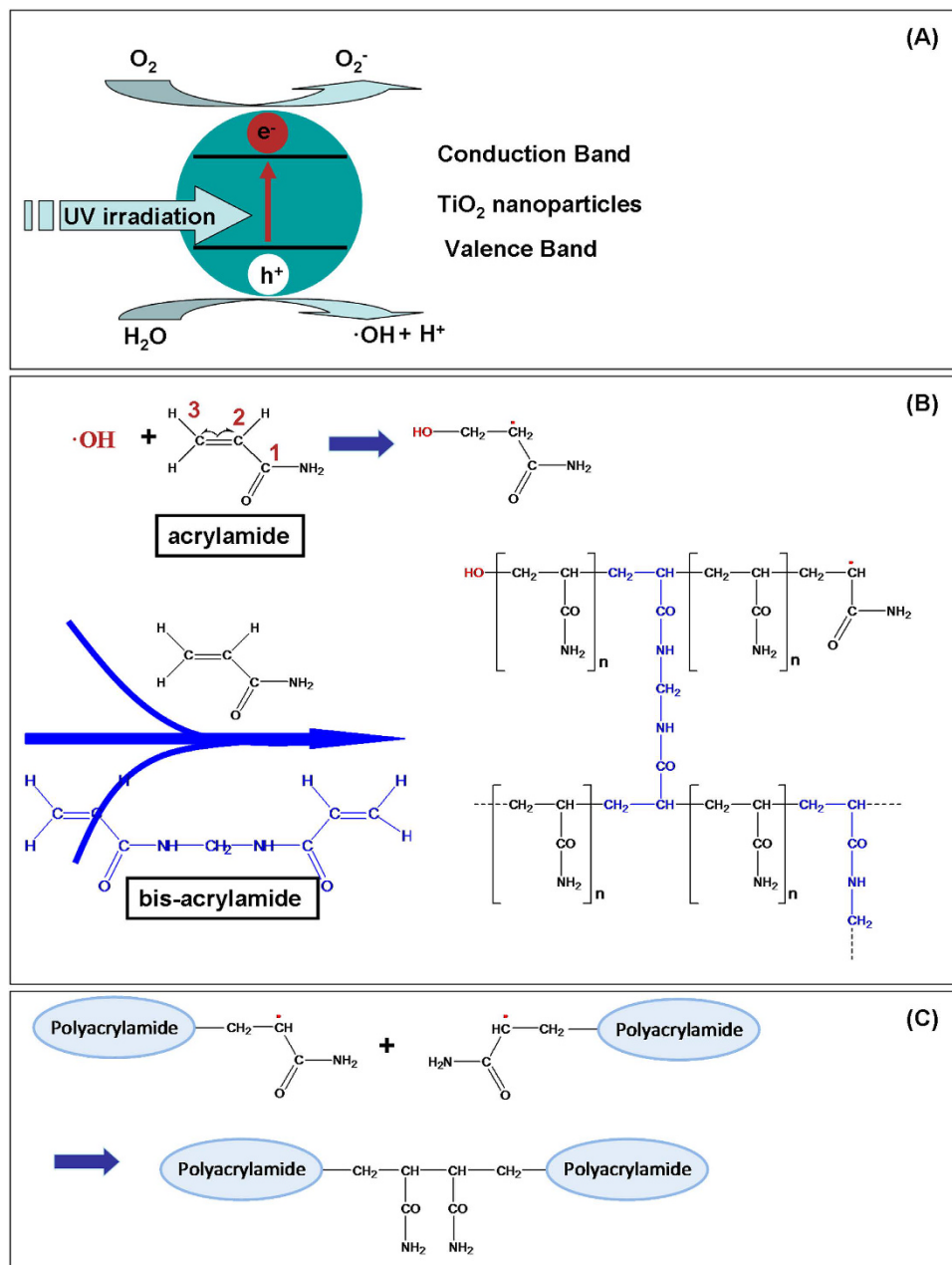


Figure 1. Mechanism of photochemical polymerization of acrylamide. (A) Photocatalytic generation of hydroxyl radicals on titanium dioxide nanoparticles. (B) Elongation of polymer chains and crosslinking with bis-acrylamide monomers. (C) Termination of the polymerization reaction.

are readily generated as shown in Fig. 1(A). Hydroxyl radical is a highly chemical reactive species which forms an important part of radical chemistry. Its role in pollutant degradation has been extensively investigated^{26–28}. This work demonstrates the polymerization reaction cascade initiated by hydroxyl radicals. As shown in Fig. 1(B), there are three carbon atoms present in acrylamide monomer molecules. Charge distribution in this molecule is calculated in Supplementary Notes 4 and shown in Supplementary Fig. 2. Clearly, the 3rd carbon atom has relatively higher electron density and less steric effect than other atoms. It becomes the most favorable site for photo-generated hydroxyl radicals to attack. Similar as that described in chemical polymerization, acrylamide monomers are converted into free radicals that can proceed to react with unactivated acrylamide monomers and the photochemical polymerization reaction cascade is started. In the meanwhile, double bonds of bis-acrylamide can also react with free radicals. The competition between hydroxyl radicals and resultant acrylamide radicals depends on the concentration of acrylamide monomers and bis monomers. Because bis acrylamide is present in much smaller quantities than acrylamide monomer, hydroxyl radicals mainly react with acrylamide monomers. Characteristic pores are formed when the elongating polymer chains are crosslinked with bis-acrylamide. Termination of the polymerization cascade was shown in Fig. 1(C). This photochemical polymerization approach

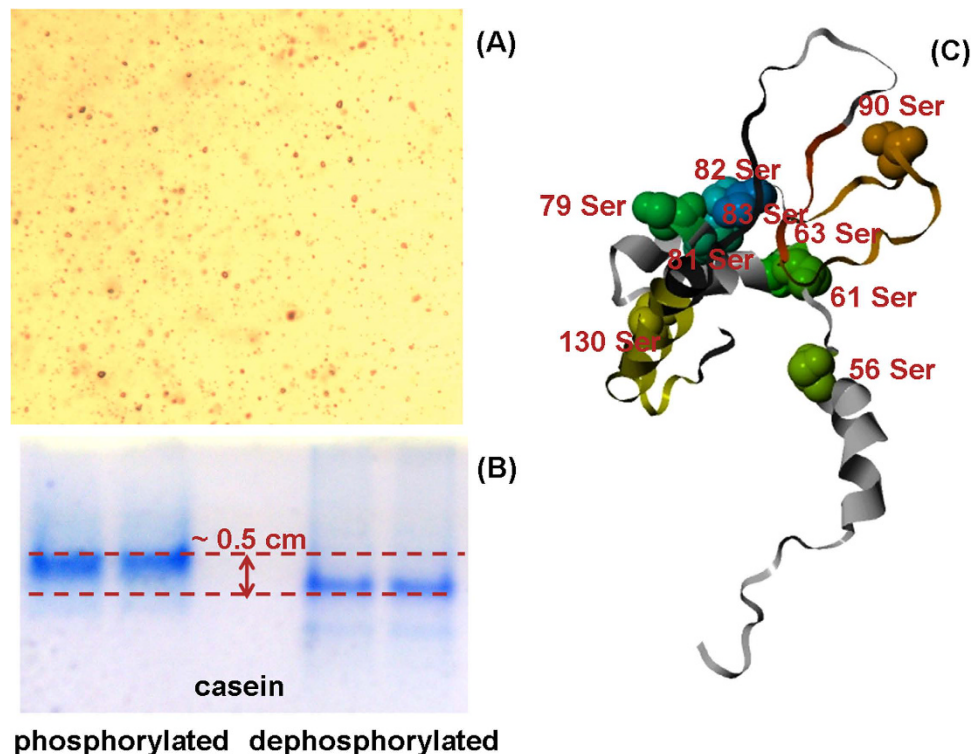


Figure 2. Electrophoretic migration of phosphorylated casein S1 and de-phosphorylated casein S1 proteins on titanium dioxide contained TAU gel. (A) Optical image of titanium dioxide contained TAU gel with 100 X magnification. (B) Gel image of phosphorylated casein and de-phosphorylated casein proteins on titanium dioxide contained TAU gel. (C) 3D structural modeling of phosphorylated casein S1 protein.

avoids the use of ammonium persulfate and TEMED. In addition, photo-generated electrons are captured by oxygen in air, which prevents oxygen to trap free radicals and thus prevent it to inhibit acrylamide polymerization. Therefore this titanium dioxide photocatalytic polymerization of acrylamide for gel electrophoresis (TIP-PAGE) approach can be used in acidic condition without degassing. It should be mentioned that acidic condition herein favors the photopolymerization reaction. In basic condition used for SDS PAGE experiments, titanium dioxide photocatalytic polymerization of acrylamide can not be observed. Additionally, although riboflavin (riboflavin-5'-phosphate) has ever been developed as an initiator for photochemical polymerization reaction, it still needs to work with ammonium persulfate or TEMED²⁹. The TIP-PAGE approach makes use of the photocatalytic properties of semiconductors that are highly active in acidic condition.

Electrophoretic Migration of Different Proteins on Titanium Dioxide Containing TAU Gel. Due to protonation and oxidization of TEMED, the polymerization process of acrylamide in acidic condition is not only very slow but also generates mechanically weak gels. The presence of titanium dioxide nanoparticles provides a substrate to differentially adsorb proteins with different affinities. In addition to photocatalytic activities, it has been found that TiO_2 can specifically bind with phosphate groups^{30–32} and glycosylation groups^{33–35} of proteins. It thus should strongly affect electrophoretic migration of phosphorylated and non-phosphorylated proteins. Figure 2(A,B) are optical images of TAU gel with TiO_2 nanoparticles under 100 X magnification and electrophoresis of phosphorylated and dephosphorylated casein proteins respectively. If the effects of TiO_2 , viscosity of the medium, differences in protein conformation and other experimental conditions are ignored, the migration rate differences between these two proteins can be simplified as the equation (1). Equation development was shown in Supplementary Notes 5.

$$\frac{v_1}{v_2} = \sqrt{\frac{m_2 z_1}{m_1 z_2}} \quad (1)$$

In the case of casein proteins whose molecular weights are more than 20 KDa, mass differences of phosphorylated or non-phosphorylated proteins do not have too much impact on their migration velocities. Essentially, charge Z determines the migration velocity v . For α -S1 casein, there are 21 lysine or arginine basic residues in total, 9 phosphorylation sites and 720 Da mass differences. The mass ratio of phosphorylated over dephosphorylated casein is about 1.03. But the charge ration of these two proteins is about 0.57 because the presence of negatively charged phosphate groups neutralizes some of the positive charges carrying on protonated lysine and arginine residues. The expected velocity ratio of phosphorylated casein over dephosphorylated casein is about 0.75. Increased electrophoretic time should result in increased differences in migration distances in the gel. When

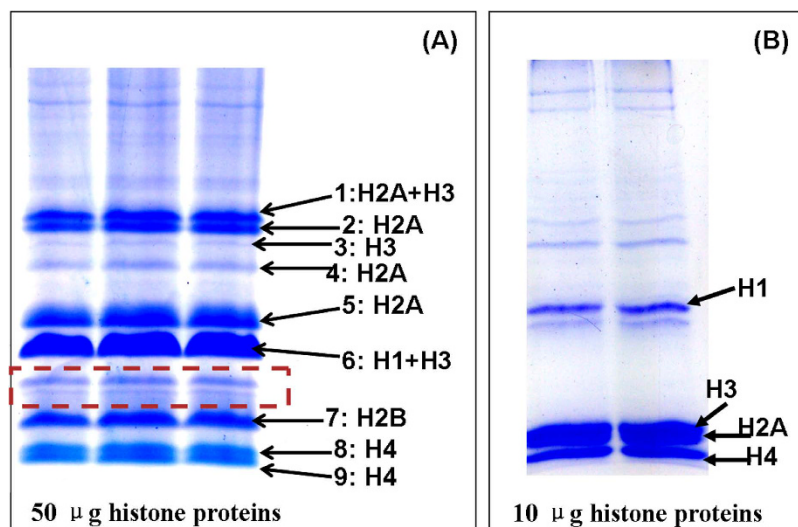


Figure 3. Analysis of acid extracted histones of male rat liver. (A) Gel image of histones with TIP -PAGE approach. (B) Gel image of the same histone proteins with regular SDS PAGE approach.

TiO_2 nanoparticles are present in the gel, electrophoretic migration velocity of phosphorylated proteins should be even slower than that of dephosphorylated proteins because of the binding of TiO_2 with exposed negatively charged phosphate groups. As shown in Fig. 2, after 200 minutes of electrophoresis with 100 volts of electrophoretic voltage, there are only about 0.5 cm differences (about 25% of the total, similar to that expected by eq. 1) in the migration distances of phosphorylated and dephosphorylated casein proteins. It seems that the effect of TiO_2 binding contributes little to the migration in this case. Looking at the 3D structural model as shown in Fig. 2(C), because no detergents have been used to denature protein structures, some of the phosphate groups are hidden in the folded protein structures, which prevent the interaction of TiO_2 with phosphate groups. It was demonstrated that protein conformation actually plays important roles in acidic TAU gel electrophoresis.

The presence of titanium dioxide nanoparticles also strengthens the gels. As described in Supplementary Notes 6, formation of crosslinked organic/inorganic polymer nanocomposites greatly improves the elastic properties of gels. In the case of APS/TEMED catalyzed polymerization reaction, gels are always very soft and weak due to incomplete polymerization and cross-linking. It is difficult to transfer those gels further for the second dimensional separation. In contrast, TiO_2 catalyzed polymerization reaction produces gels that can not only be easily transferred but also can be stretched almost twice of the original length. Additionally, the amount of titanium dioxide used for the gel does not show detectable effect on the sieving ability of the gel as shown in Supplementary Notes 7. The pore sizes of polyacrylamide gels usually range from several hundreds of nanometers (nm) to 70 nm. Sizes of nanoparticles of titanium dioxide used in this work are less than 25 nm as indicated by the manufacturers. As for nanoparticles which are much bigger or smaller than gel pores, they are washed away and do not affect the sieving ability of gels; As for nanoparticles whose sizes are equivalent to that of gel pores, they are trapped in the pores. So long as the amount of nanoparticles is in excess, it should be able to maintain the separation pattern of proteins when different amount of nanoparticles are used.

Titanium Dioxide Photocatalytic Polymerization of Acrylamide for Gel Electrophoresis (TIP -PAGE) of Acid Extracted Rat Liver Histone Proteins.

Histone proteins from rat livers have been separated by using TIP -PAGE approach. Fig. 3(A) is the gel image of these proteins. Compared with that of routine SDS PAGE shown in Fig. 3(B), it is not surprising that much more bands have been observed and several isoforms of histone proteins have been identified. All MS spectra of band 1–9 were shown in Supplementary 3–11 respectively. Compared with the same TAU gel electrophoresis without TiO_2 that we reported previously²⁶, the separation resolution of TIP -PAGE has been greatly improved with finely dissected protein bands. For example, in TAU gel electrophoresis, H4 and H2B were co-eluted as one band. But with TIP -PAGE approach, H2B and H4 have been significantly separated into at least 3 major bands (labeled as band 7, 8 and 9) with two H4 isoforms. The similar electrophoretic migration of H4 and H2B in TAU gel electrophoresis without TiO_2 nanoparticles means these proteins have very similar mass-to-charge ratios. Changes in H2B mobility on TiO_2 containing TAU gel may be ascribed to its posttranslational modifications. Examination of amino acid sequences of H2B and H4 shown in Supplementary Fig. 12 reveals that H2B have more than two times of possible phosphorylation sites than that of H4, which have strong affinities to TiO_2 nanoparticles. Although only 3–4 phosphorylation sites among these have been reported so far, this work provides interesting experimental evidences for necessities of further investigation. Similar mass spectra of gel band 8 and 9 indicate that two isoforms of H4 have exactly the same tryptic peptides extracted from in-gel digestion. It means that different isoforms of H4 histone cannot be distinguished by this bottom-up proteomics approach. It can be proposed that the differential separation of H4 proteins by TiO_2 TAU gel electrophoresis results from their differences in modifications. Attention should also be paid to the series bands just above the identified H2B band which was labeled with a dotted red square. These bands were not identified due to low abundance. But their absence in non- TiO_2 TAU gel electrophoresis also indicates

their possibilities for post-translational modifications. The separation of H2A and H3 isoforms has also been dramatically improved. Only one band for each H2A and H3 was observed in SDS PAGE. However, by using gels made with TiO₂ photocatalytic polymerization reaction, H2A histones were detected in four different bands and H3 histones were detected in 3 different bands. In contrast, the separation of H1 was not significantly changed probably because of its less modified structures. All identified peptides were listed in Supplementary Table 2. MS/MS spectra as well as database searching results were shown in Supplementary Figs 13–22. Because of enhanced mechanical properties, T_{IP}PAGE is highly reproducible for separation of histone proteins in acidic condition as shown in Supplementary Notes 8. Although the distribution of nanoparticles is not very uniform with 100 X magnification as shown in Fig. 2(A), it does not show obvious effects on protein separation.

Conclusion

With UV irradiation, hydroxyl radicals generated on TiO₂ nanoparticles can rapidly initiate polymerization reaction cascade of acrylamide without ammonium persulfate and TEMED. Hydroxyl radicals are highly reactive due to the presence of an unpaired electron. This photochemical polymerization starts as the attacking of hydroxyl radicals on the carbon atoms of acrylamide with high electron density but low steric effects. The polymer chain elongates through the reaction of resultant new free radicals with unactivated acrylamide monomers. Bis-acrylamide monomers are crosslinked with acrylamide polymer chains to form pores that can trap molecules with different sizes. TiO₂ nanoparticles are not only involved in the photochemical reaction but also change the electrophoretic mobility of those proteins with phosphoryl modifications. The T_{IP}PAGE approach is useful for separation of basic proteins such as histones in acidic condition.

References

- Vesterberg, O. History of electrophoretic methods. *J. Chromatogr. A* **480**, 3–19 (1989).
- Chen, B. & Chrambach, A. Estimation of polymerization efficiency in the formation of polyacrylamide gel, using continuous optical scanning during polymerization. *J. Biochem. Biophys. Methods* **1**, 105–116 (1979).
- Herr, A. E. & Singh, A. K. Photopolymerized cross-linked polyacrylamide gels for on-chip protein sizing. *Anal. Chem.* **76**, 4727–4733 (2004).
- Timperman, A. T. & Aebersold, R. Peptide electroextraction for direct coupling of in-gel digests with capillary LC-MS/MS for protein identification and sequencing. *Anal. Chem.* **72**, 4115–4121 (2000).
- Lu, J. J., Zhu, Z., Wang, W. & Liu, S. Coupling sodium dodecyl sulfate–capillary polyacrylamide gel electrophoresis with matrix-assisted laser desorption ionization time-of-flight mass spectrometry via a poly(tetrafluoroethylene) membrane. *Anal. Chem.* **83**, 1784–1790 (2011).
- Saveliev, S. V. *et al.* Mass spectrometry compatible surfactant for optimized in-gel protein digestion. *Anal. Chem.* **85**, 907–914 (2013).
- Strahl, B. D. & Allis, C. D. The language of covalent histone modifications. *Nature* **403**, 41–45 (2000).
- Peterson, C. L. & Laniel, M. A. Histones and histone modifications. *Curr. Biol.* **14**, R546–R551 (2004).
- Muller, M. M. & Muir, T. W. Histones: at the crossroads of peptide and protein chemistry. *Chem. Rev.* **115**, 2296–2349 (2015).
- Johms, E. W. The electrophoresis of histones in polyacrylamide gel and their quantitative determination. *Biochem. J.* **104**, 78–82 (1967).
- Panyim, S. & Chalkley, R. High resolution acrylamide gel electrophoresis of histones. *Arch. Biochem. Biophys.* **130**, 337–346 (1969).
- Shechter, D., Dormann, H. L., Allis, C. D. & Hake, S. B. Extraction, purification and analysis of histones. *Nat. Protoc.* **2**, 1445–1457 (2007).
- Shiraishi, Y., Saito, N. & Hiral, T. Adsorption-driven photocatalytic activity of mesoporous titanium dioxide. *J. Am. Chem. Soc.* **127**, 12820–12822 (2005).
- Leshuk, T. *et al.* Photocatalytic activity of hydrogenated TiO₂. *Acs Appl. Mater. Interfaces* **5**, 1892–1895 (2013).
- Krylova, G. & Na, C. Photo-induced crystallization and activation of amorphous titanium dioxide. *J. Phys. Chem. C* **119**, 12400–12407 (2015).
- Popovic, I. G., Katsikas, L. & Weller, H. The photopolymerisation of methacrylic acid by colloidal semiconductors. *Polym. Bull.* **32**, 597–603 (1994).
- Stroyuk, A. L., Granchak, V. M., Korzhak, A. V. & Kuchmii, S. Ya. Photoinitiation of butylmethacrylate polymerization by colloidal semiconductor nanoparticles. *J. Photochem. Photobiol. A: Chem.* **162**, 339–351 (2004).
- Liang, S. S., Makamba, H., Huang, S. Y. & Chen, S. H. Nano-titanium dioxide composites for the enrichment of phosphopeptides. *J. Chromatogr. A* **1116**, 38–45 (2006).
- Ni, X., Ye, J. & Dong, C. Kinetics studies of methyl methacrylate photopolymerization initiated by titanium dioxide semiconductor nanoparticles. *J. Photochem. Photobiol. A: Chem.* **181**, 19–27 (2006).
- Li, G. & Ni, X. A novel photoconductive ZnO/PVK nanocomposite prepared through photopolymerization induced by semiconductor nanoparticles. *Mater. Lett.* **62**, 3066–3069 (2008).
- Zhang, D., Yang, J., Bao, S., Wu, Q. & Wang, Q. Semiconductor nanoparticle-based hydrogels prepared via self-initiated polymerization under sunlight, even visible light. *Sci. Rep.* **3**, 1399 (2013).
- Dadashi-Silab, S., Tasdelen, M. A., Asiri, A. M., Khan, S. B. & Yagci, Y. Photoinduced atom transfer radical polymerization using semiconductor nanoparticles. *Macromol. Rapid Commun.* **35**, 454–459 (2014).
- Norton, K., Graffe, A. & Maric, M. Synthesis of hydroxyl-terminated poly(vinylcarbazole-ran-styrene) copolymers by nitroxide-mediated polymerization. *Macromol. Chem. Phys.* **208**, 496–503 (2007).
- Yang, Y. *et al.* A facile, green, and tunable method to functionalize carbon nanotubes with water-soluble azo initiators by one-step free radical addition. *Appl. Surf. Sci.* **256**, 3286–3292 (2010).
- Fujii, S. *et al.* Liquid marbles prepared from pH-responsive sterically stabilized latex particles. *Langmuir* **27**, 8067–8074 (2011).
- Zhang, W., Tang, X., Ding, M. & Zhong, H. Cu²⁺-assisted two dimensional charge-mass double focusing gel electrophoresis and mass spectrometric analysis of histone variants. *Anal. Chim. Acta* **852**, 121–128 (2014).
- Bunce, N. J., Liu, L., Zhu, J. & Lane, D. A. Reaction of naphthalene and its derivatives with hydroxyl radicals in the gas phase. *Environ. Sci. Technol.* **31**, 2252–2259 (1997).
- Gehling, W., Khachatryan, L. & Dellinger, B. Hydroxyl radical generation from environmentally persistent free radicals (EPFRs) in PM_{2.5}. *Environ. Sci. Technol.* **48**, 4266–4272 (2014).
- Riva, M., Robinson, E. S., Perraudin, E., Donahue, N. M. & Villenzve, E. Photochemical aging of secondary organic aerosols generated from the photooxidation of polycyclic aromatic hydrocarbons in the gas-phase. *Environ. Sci. Technol.* **49**, 5407–5416 (2015).
- Wang, T., Bruin, G. J., Kraak, J. C. & Poppe, H. Preparation of polyacrylamide gel-filled fused-silica capillaries by photopolymerization with riboflavin as the initiator. *Anal. Chem.* **63**, 2207–2208 (1991).

31. Torta, F., Fusi, M., Casari, C. S., Bottani, C. E. & Bachi, A. Titanium dioxide coated MALDI plate for on target analysis of phosphopeptides. *J. Proteome Res.* **8**, 1932–1942 (2009).
32. Nie, S. *et al.* Comprehensive profiling of phosphopeptides based on anion exchange followed by flow-through enrichment with titanium dioxide (AFET). *J. Proteome Res.* **9**, 4585–4594 (2010).
33. Fukuda, I. *et al.* Optimization of enrichment conditions on TiO₂ chromatography using glycerol as an additive reagent for effective phosphoproteomic analysis. *J. Proteome Res.* **12**, 5587–5597 (2013).
34. Larsen, M. R., Jensen, S. S., Jakobsen, L. A. & Heegaard, N. H. H. Exploring the sialome using titanium dioxide chromatography and mass spectrometry. *Mol. Cell Proteomics* **6**, 1778–1787 (2007).
35. Palmisano, G. *et al.* Selective enrichment of sialic acid-containing glycopeptides using titanium dioxide chromatography with analysis by HILIC and mass spectrometry. *Nat. Protoc.* **5**, 1974–1982 (2010).
36. Huang, B. Y., Yang, C. K., Liu, C. P. & Liu, C. Y. Stationary phases for the enrichment of glycoproteins and glycopeptides. *Electrophoresis* **35**, 2091–2107 (2014).

Acknowledgements

We greatly appreciate the support from National Natural Science Foundation of China (NSFC, 21175054, 31270876, 31370815, 21575046), Ministry of Science and Technology of the People's Republic of China (MSTC, 2014DFB30020), Program for Changjiang Scholars and Innovative Research Team in University (PCSIRT, no. IRT0953), Hubei Natural Science Foundation Council (HBNSFC, 2009CDA001), Research Funds of Central China Normal University from the Ministry of Education (CCNU11G01007, CCNU11C01002, 20205001536) and the Research Platform of Hubei Province for Monitoring of Pesticide Residues and Agricultural Products Safety.

Author Contributions

W.Z. performed the experiments on gel preparation, histone protein extraction and identification by mass spectrometry. Z.Y. repeated parts of experiments including gel preparation, histone extraction and mass spectrometric analysis. L.H., J.K. and R.J. have been involved in the experiments. H.Z. developed the concept, designed experiments, analyzed data and wrote the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Zhang, W. *et al.* Titanium Dioxide Photocatalytic Polymerization of Acrylamide for Gel Electrophoresis (TIP-PAGE) of Proteins and Structural Identification by Mass Spectrometry. *Sci. Rep.* **6**, 20981; doi: 10.1038/srep20981 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>