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## “Cation-Stitching Cascade”: exquisite control of terpene cyclization in cyclooctatin biosynthesis

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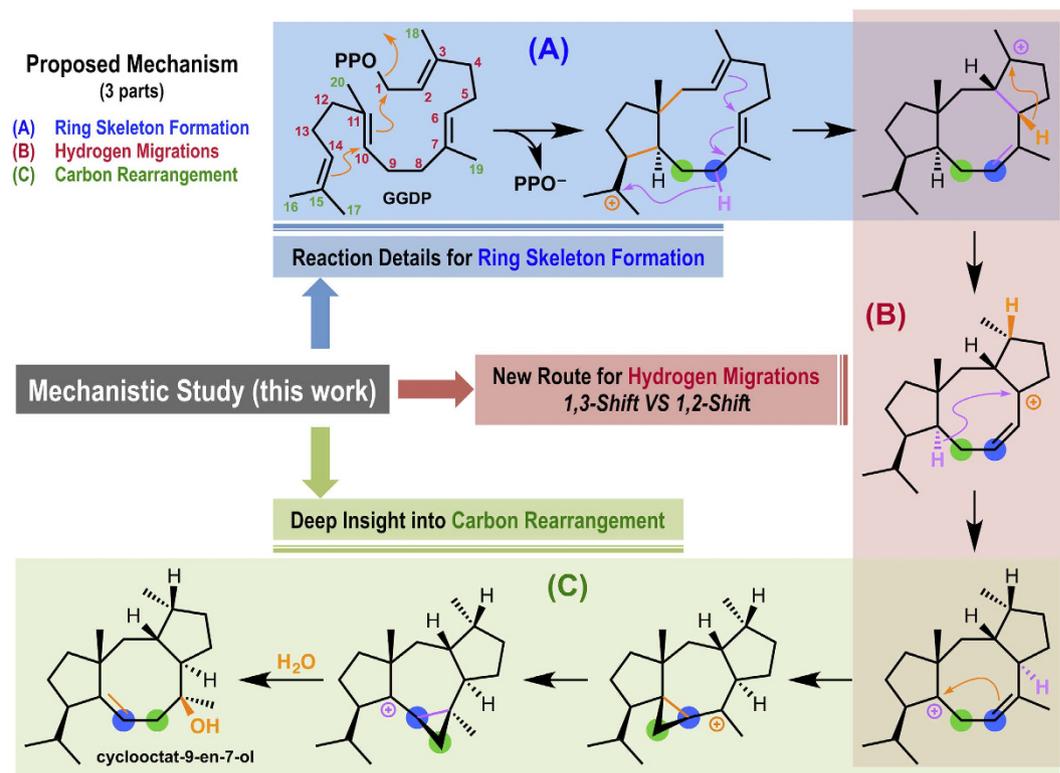
Terpene cyclization is orchestrated by terpene cyclases, which are involved in the biosynthesis of various cyclic natural products, but understanding the origin and mechanism of the selectivity of terpene cyclization is challenging. In this work, we describe an in-depth mechanistic study on cyclooctatin biosynthesis by means of theoretical calculations combined with experimental methods. We show that the main framework of cyclooctatin is formed through domino-type carbocation transportation along the terpene chain, which we call a “cation-stitching cascade”, including multiple hydrogen-shifts and a ring rearrangement that elegantly determine the stereoselectivity.

Terpenoids or isoprenoids are a large and highly diverse group of natural products; over 55,000 terpenoids with a wide range of chemical structures (including monocyclic or multi-cyclic fused ring structures) and various biological activities have been reported to date<sup>1</sup>. This structural diversity is generated in the course of the cyclization reactions catalyzed by terpene cyclases, which provide remarkable examples of exquisite enzymatic control of the course of a chemical reaction cascade<sup>2–4</sup>.

We have established that CotB2, a diterpene cyclase<sup>5</sup> identified in a soil-living filamentous bacterium *Streptomyces melanosprofaciens*, provides an especially elegant example of synthetic virtuosity and stereochemical control; CotB2 synthesizes the characteristic 5-8-5 fused ring system of cyclooctat-9-en-7-ol (PD), containing six chiral carbons, by cyclizing the achiral and acyclic C<sub>20</sub> substrate (SM), geranylgeranyldiphosphate (GGDP)<sup>5</sup>. Among various diterpene cyclases, CotB2 specifically produces PD as a single product with the correct C–C bond connectivity and stereochemistry. Thus, CotB2 appears to precisely pre-organize the conformation of SM, control the conformations of all reactive intermediates, and properly regulate the reactivities of the cation intermediates in its active site pocket.

We previously proposed the mechanism of the CotB2-catalyzed reaction cascade (Fig. 1), based on a powerful combination of *in vivo* tracer experiments using uniformly <sup>13</sup>C-labeled glucose and *in vitro* reactions of regiospecifically <sup>2</sup>H-substituted SM<sup>6</sup>. The reaction cascade is divided into three parts, ring skeleton construction (Part A), hydrogen migrations (Part B), and carbon–carbon backbone rearrangement (Part C). Part A begins with CotB2-catalyzed dissociation of the pyrophosphate group followed by initial cyclization to produce a cation intermediate with a bicyclic 5–11 fused-ring skeleton. Subsequent 1,5-hydrogen-transfer from C8 to C15 provides an allylic cation, which undergoes intramolecular cyclization reaction with the C2–C3 olefin to form a cationic intermediate bearing the tricyclic 5-8-5 fused-ring system. The second part (Part B, Fig. 1) involves long-range cation transfer via multiple hydrogen shifts. The cationic intermediate likely undergoes a 1,3-H shift from C6 to C3 and a successive 1,5-H shift from C10 to C6 to generate the homoallylic cation. In Part C, formation of the cyclopropylcarbanyl cation, which features a C8–C9–C10 cyclopropyl ring, is followed by interconversion of

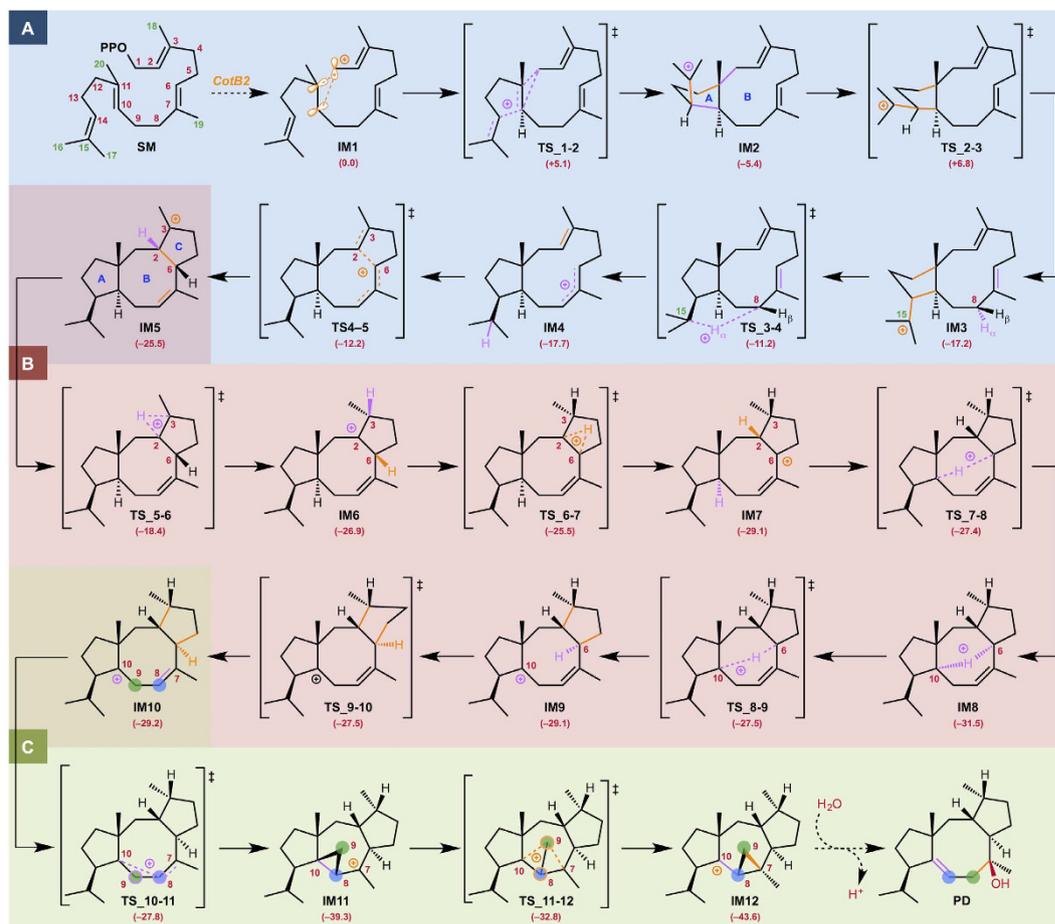
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**Figure 1.** Proposed biosynthetic route to cyclooctat-9-en-7-ol. Putative carbocations involved in the conversion of geranylgeranyl diphosphate (GGDP) to cyclooctat-9-en-7-ol.

cyclopropylcarbinylium cations. Finally, C7–C8 ring opening with capture of water by the homoallylic cation results in the formation of **PD**. Although our previous isotope tracing experiments and NMR spectroscopic investigations demonstrated the presence of the unusual carbon–carbon bond rearrangement, less is known about other intermediates and the mechanisms of other reactions on the pathway to the final product. For instance, in Part B, an alternative cascade involving two successive non-concerted 1,2-H shifts, firstly C2 to C3 yielding a cation at C2, then C6 to C2, could not be excluded in the previous work. Recent advances in theoretical methods, coupled with the enormous increase in available computational power, have enabled chemists to calculate transition structures and reaction pathways for complex molecules<sup>7–10</sup>. Density functional theory (DFT) combined with the AFIR (artificial force induced reaction) method and Reaction Plus method was employed herein to corroborate the experimental results and to comprehensively unveil this biosynthetic pathway.

All calculations were carried out at the M062X/6-31G\*\* level<sup>11</sup> using Gaussian 09<sup>12</sup>, GRRM11<sup>13</sup>, and Reaction Plus<sup>14</sup> programs. The full reaction pathway for the conversion of geranylgeranyl diphosphate (GGDP: **SM**) to cyclooctat-9-en-7-ol (**PD**) and the energy diagram are presented in Figs 2 and 3, respectively. We will first discuss the CotB2-mediated carbocation formation and successive tricyclic skeleton construction part (Part A). Dissociation of the pyrophosphate of **SM** yields an allylic carbocation (**IM1**) partially stabilized by a cation- $\pi$  interaction with a distal C10–C11 double bond<sup>15</sup>. Cation-mediated annulation proceeds smoothly to form a bicyclic intermediate (**IM2**), which undergoes sequential conformational change<sup>16</sup> to give a more stable intermediate (**IM3**) with an activation energy of 12.2 kcal/mol. **IM3** contains two non-equivalent hydrogens at the C8 carbon atom. The  $\alpha$ -hydrogen (downward) atom of the two hydrogens can migrate smoothly<sup>17</sup> along the intrinsic reaction coordinate to the C15 carbocation with 6.6 kcal/mol stabilization energy for formation of the more stable allylic cation (**IM4**). **TS\_3-4** is conformationally more favorable than **TS\_3-4B** (1,5- $\beta$ -H shift) by ca. 40 kcal/mol. The second annulation then occurs smoothly with an activation barrier of 5.5 kcal/mol to form the 5-8-5 fused-ring skeleton (**IM5**) with a large stabilization energy ( $\Delta G = 13.3$  kcal/mol). The overall gain to construct the tricyclic structure **IM5** is rather large ( $-25.5$  kcal/mol from **IM1**). In Part B, multiple hydrogen shifts occur successively (**IM5**→**IM6**→**IM7**→**IM8**→**IM9**), leading to long-range cation transfer<sup>18</sup>. After the tandem 1,2-H shifts (**IM5**→**IM7**) on the C-ring to yield a new allylic cation (the reaction pathway is discussed in detail below), a 1,5-H shift across the B-ring gives a homoallylic cation intermediate (**IM9**). As for Part C, DFT calculations of the C8–C9 bond rearrangement process were essentially consistent with our previously proposed reaction mechanism based on isotope tracing experiments and NMR spectroscopic investigations. After the conformational change of **IM9** into **IM10** with a very small activation barrier, the C7–C8  $\pi$  bond of the homoallylic cation in **IM10** approaches the cation centre to form a cyclopropylcarbinylium cation intermediate (**IM11**) with an unexpectedly large stabilization energy ( $\Delta G = -11.4$  kcal/mol). **IM11** has a highly distorted cyclopropane ring in which participation of the elongated C8–C10  $\sigma$  orbital partially contributes to stabilization of the neighboring C7 carbocation, as judged from the elongation of the C8–C10 bond (1.67 Å) as well as the short C7–C8 bond (1.38 Å)<sup>19</sup>. An unusual C–C bond



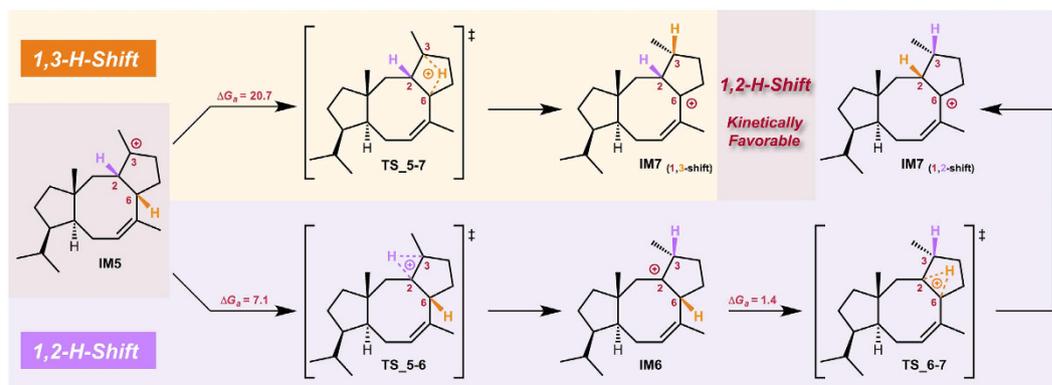
**Figure 2.** Computed reaction pathways and potential energy changes for the “cation-stitching cascade” from SM to PD. Potential energies (kcal/mol, Gibbs free energies calculated at the M062X/6-31G\*\* level) relative to IM1 are shown in parentheses.



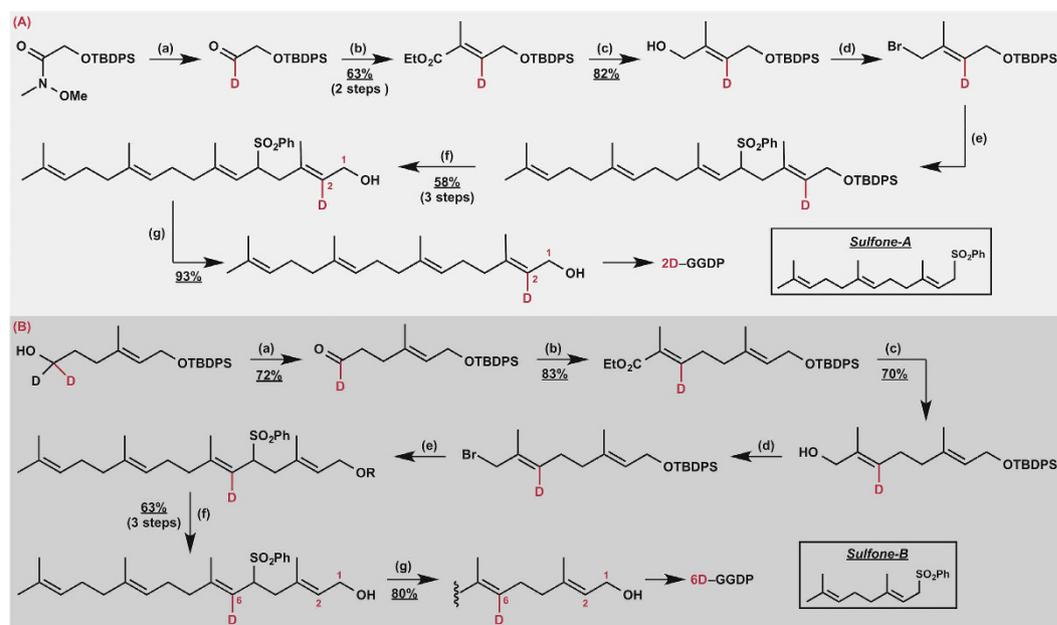
**Figure 3.** Computed potential energy profiles for the “cation-stitching cascade” reaction. Potential energies (kcal/mol, Gibbs free energies calculated at the M062X/6-31G\*\* level) relative to IM1 are shown in red.

rearrangement then takes place *via* a unique bicyclobutonium cation<sup>20</sup> transition state (TS<sub>11-12</sub>) with very low activation energy (6.5 kcal/mol) to give another cyclopropylcarbinyl cation (IM12). IM12 is energetically more stable than IM11 by 4.3 kcal/mol owing to the strain magnitudes of their cyclopropane rings. Finally, a hydration reaction yields the final product (PD). Although we should keep in mind the possibility that the CotB2 may be involved in the final step, our preliminary calculations indicate that non-enzymatic hydration of IM12 with water molecules would also be likely to proceed smoothly to provide PD.

The energy diagram (Fig. 3) immediately suggests that this is a thermodynamically and kinetically favorable biosynthetic reaction cascade: 1) activation barriers are all low enough for the reactions to proceed smoothly at ambient temperature, 2) the entire energy profile descends as the reactions proceed, and 3) the overall exothermicity



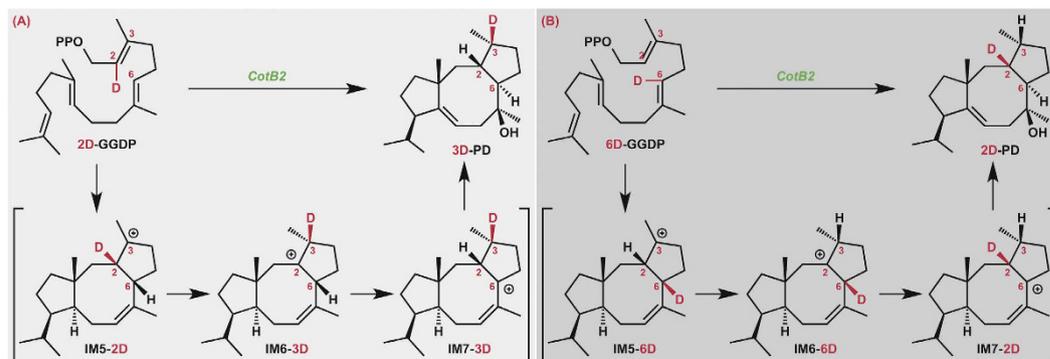
**Figure 4.** Comparison of the two possible hydrogen shift pathways explored theoretically. Energy changes (kcal/mol, Gibbs free energies calculated at the M062X/6-31G\*\* level) are shown together with arrows.



**Figure 5.** Regioselective synthesis of the deuterated GGDPs. (A) Reaction conditions: (a)  $\text{LiAlD}_4$  (1.0 equiv.),  $\text{Et}_2\text{O}$ ,  $-60^\circ\text{C}$ , 0.5 h; (b)  $\text{EtO}_2\text{C}-\text{C}(\text{CH}_3)=\text{PPh}_3$  (1.2 equiv.),  $\text{CH}_2\text{Cl}_2$ , rt, 1 h; (c)  $\text{LiAlH}_4$  (1.5 equiv.),  $\text{Et}_2\text{O}$ ,  $-40$  to  $-20^\circ\text{C}$ , 1 h; (d)  $\text{CBr}_4$  (1.3 equiv.),  $\text{PPh}_3$  (1.3 equiv.),  $\text{CH}_2\text{Cl}_2$ , rt, 0.5 h; (e) *Sulfone-A* (1.5 equiv.), *t*-BuOK (4.0 equiv.), THF,  $-20^\circ\text{C}$ , 1 h; (f) TBAF (4.0 equiv.), THF, rt, 3 h; (g)  $\text{LiBHET}_3$  (5.0 equiv.),  $\text{Pd}(\text{dppp})\text{Cl}_2$  (5.0 mol%), THF,  $0^\circ\text{C}$ , 1 h. (B) Reaction conditions: (a) PCC (2.0 equiv.),  $\text{CH}_2\text{Cl}_2$ , rt, 3 h; (b)  $\text{EtO}_2\text{C}-\text{C}(\text{CH}_3)=\text{PPh}_3$  (1.2 equiv.),  $\text{CH}_2\text{Cl}_2$ , rt, 2 h; (c)  $\text{LiAlH}_4$  (1.5 equiv.),  $\text{Et}_2\text{O}$ ,  $-40$  to  $-20^\circ\text{C}$ , 1 h; (d)  $\text{CBr}_4$  (1.3 equiv.),  $\text{PPh}_3$  (1.3 equiv.),  $\text{CH}_2\text{Cl}_2$ , rt, 0.5 h; (e) *Sulfone-B* (1.5 equiv.), *t*-BuOK (4.0 equiv.), THF,  $-20^\circ\text{C}$ , 1 h; (f) TBAF (4.0 equiv.), THF, rt, 3 h; (g)  $\text{LiBHET}_3$  (5.0 equiv.),  $\text{Pd}(\text{dppp})\text{Cl}_2$  (5.0 mol%), THF,  $0^\circ\text{C}$ , 2 h.

is very large<sup>21</sup>. The first annulation and conformational change ( $\text{IM2} \rightarrow \text{TS}_2\text{-3}$ ) is the rate-determining step. Remarkably, the cyclopropane ring formation and carbon-carbon bond rearrangement each require much lower activation energy than this step.

We next studied in detail the 1,3-cation shift reaction mechanism ( $\text{IM5} \rightarrow \text{IM7}$ ), which is the last puzzle in the biosynthesis. Two pathways are possible, i.e., direct 1,3-hydrogen shift and sequential double 1,2-hydrogen shift (Fig. 4). We would expect *a priori* that the concerted 1,3-H shift is a reasonable route, because of the stereoselectivity and step-economy, and so it was found. The 1,3-H shift from the C6 to the C3 position takes place as a single event with an overall energy loss of 20.7 kcal/mol. We could also identify the TSs for the two successive non-concerted 1,2-H shifts: the first 1,2-H shift from the C2 to C3 via  $\text{TS}_5\text{-6}$  yielding  $\text{IM6}$  followed by the second one from C6 to C2 via  $\text{TS}_6\text{-7}$  generating an allyl cation  $\text{IM7}$ . The activation energies for the first and second 1,2-H shifts are 7.1 kcal/mol and only 1.5 kcal/mol, respectively. The more than 10 kcal/mol difference of energy between the 1,3- and the tandem 1,2-transfer translates into an over  $10^{10}$ -fold difference of reaction rate at room temperature; hence the sequential double 1,2-hydrogen shift should predominate over direct 1,3-hydrogen shift in the microorganism.



**Figure 6.** Isotope tracing experiments for the biosynthesis of cyclooctatin.

To validate the successive 1,2-hydrogen migrations suggested by our calculation, we next incubated recombinant CotB<sup>6</sup> individually with two regioselectively deuterated GGDPs, **2D-** and **6D-GGDP** (which were successfully prepared with >99% D incorporation ratio *via* the multistep procedure shown in Fig. 5 and characterized by means of high-resolution mass analysis and NMR spectroscopy as shown in the SI). Comparison of the deuterated product generated in each reaction by means of <sup>1</sup>H NMR spectroscopy with natural abundance **2** (Fig. S1–S2) allowed assignment of the positions of deuteration in each product, thereby establishing that **2D-** and **6D-GGDP** were converted to **3D-** and **2D-PD**, respectively (Fig. 6). The formation of **3D-** and **2D-PD** from **2D-** and **6D-GGDP**, respectively, is fully consistent with the occurrence of successive 1,2-hydrogen migrations from C2 to C3 and C3 to C6 during the cyclization reaction cascade.

In summary, our computational and experimental studies have clarified the whole biosynthetic route to cyclooctatin. DFT calculations reveal that the core framework of cyclooctatin is constructed via tandem carbocation transfer (“cation-stitching cascade”), involving several hydrogen-shifts as well as cyclopropylcarbanyl cation rearrangement. Such a reaction cascade provides high efficiency and selectivity, and is well supported by the results of the deuteration experiments. Thus, the combination of theoretical calculations and experimental studies has provided detailed insight into this biosynthetic mechanism. This approach may also throw light on the evolution of particular biosynthetic pathways, and provide clues for designing new synthetic routes for complex functional molecules.

## Methods

All calculations were performed with the Gaussian 09 program<sup>12</sup>. Geometry optimizations were performed in the gas phase at the M062X/6-31G\*\* level<sup>11</sup>, without any symmetry restrictions, and the nature of the minima was verified by frequency calculations within the harmonic approximation. IRC for all TS was performed with GRRM11<sup>13</sup> and/or Reaction Plus<sup>14</sup> programs based on Gaussian 09. Gibbs free energy was used as the basis for discussion in this study. The Cartesian coordinates of all species are collected in Supporting Information. Details for the synthesis of deuterated GGDPs and the corresponding deuterium tracing experiments are described in Supporting Information as well.

## References

- Christianson, D. W. Structural Biology and Chemistry of the Terpenoid Cyclases. *Chem. Rev.* **106**, 3412–1442 (2006).
- Zhang, Q. & Tiefenbacher, K. Terpene cyclization catalysed inside a self-assembled cavity, *Nat. Chem.* **7**, 197–202 (2015).
- Pronin, S. V. & Shenvi, R. A. Synthesis of highly strained terpenes by non-stop tail-to-head polycyclization, *Nat. Chem.* **4**, 915–920 (2012).
- Smanski, M. J., Peterson, R. M., Huang, S. X. & Shen, B. Bacterial Diterpene Synthases: New Opportunities for Mechanistic Enzymology and Engineered Biosynthesis. *Curr Opin Chem Biol.* **16**, 132–141 (2012).
- Kim, S. Y. *et al.* Cloning and heterologous expression of the cyclooctatin biosynthetic gene cluster afford a diterpene cyclase and two P450 hydroxylases. *Chem. Biol.* **16**, 736–743 (2009).
- Meguro, A. *et al.* An Unusual Terpene Cyclization Mechanism Involving a Carbon–Carbon Bond Rearrangement. *Angew. Chem. Int. Ed.* **54**, 4353–4356 (2015).
- Tantillo, D. J. Walking in the woods with quantum chemistry—application of quantum chemical calculations in natural products research. *Nat. Prod. Rep.* **30**, 1079–1086 (2013).
- Tantillo, D. J. Biosynthesis via carbocations; Theoretical studies on terpene formation. *Nat. Prod. Rep.* **28**, 1035–1053 (2011).
- Hong, Y. J. & Tantillo, D. J. A potential energy surface bifurcation in terpene biosynthesis, *Nat. Chem.* **1**, 384–389 (2009).
- Hong, Y. J. & Tantillo, D. J. Biosynthetic consequence of multiple sequential post-transition-state bifurcations, *Nat. Chem.* **6**, 104–111 (2014).
- Zhao, Y. & Truhlar, D. G. The M06 suite of density functionals for main group thermochemistry, thermochemical kinetics, noncovalent interactions, excited states, and transition elements: two new functional and systematic testing of four M06-class functionals and 12 other functionals. *Theor. Chem. Acc.* **120**, 215–241 (2008).
- Frisch, M. J. *et al.* Gaussian 09 Revision D.01, Gaussian, Inc., Wallingford CT. Full citations are given in supporting information.
- Maeda, S., Osada, Y., Morokuma, K. & Ohno, K. *GRRM11, Version 11.03*. Full citations are given in supporting information.
- <http://www.hpc.co.jp/chem/react1.html>.
- Cossi, M. & Barone, V. Solvent effect on vertical electronic transitions by the polarizable continuum model. *J. Chem. Phys.* **112**, 2427–2435 (2000).
- Concomitant C–Ring Expansion and D–Ring Formation in Lanosterol Biosynthesis from Squalene without Violation of Markovnikov’s Rule. *J. Am. Chem. Soc.* **124**, 10286–10287 (2002).

17. Chiba, R., Minami, A., Gomi, K. & Oikawa, H. Identification of Ophiobolin F Synthase by a Genome Mining Approach: A Sesterterpene Synthase from *Aspergillus clavatus*. *Org. Lett.* **15**, 594–597 (2013).
18. Hong, Y. J. & Tantillo, D. J. Feasibility of Intramolecular Proton Transfers in Terpene Biosynthesis – Guiding Principles. *J. Am. Chem. Soc.* **137**, 4134–4140 (2015).
19. Olah, G. A., Kelly, D. P., Jeuell, C. L. & Porter, R. D. Stable Carbonium Ions. XCVIII. The Nonclassical Cyclopropylcarbinyl Cation. *J. Am. Chem. Soc.* **92**, 2544–2546 (1970).
20. Hong, Y. J., Giner, J. L. & Tantillo, D. J. Bicyclobutonium Ions in Biosynthesis – Interconversion of Cyclopropyl-Containing Sterols from Orchids. *J. Am. Chem. Soc.* **137**, 2085–2088 (2015).
21. Wendt, K. U., Poralla, K. & Schulz, G. E. Structure and Function of a Squalene Cyclase. *Science* **277**, 1811–1815 (1997).
22. Hong, Y. J. & Tantillo, D. J. The energetic viability of an unexpected skeletal rearrangement in cyclooctatin biosynthesis. *Org. Biomol. Chem.*, **13**, 10273–12738 (2015).

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## Author Contributions

H.S. performed the calculations and analysed the results with the assistance of Y.M. and N.T. K.T., M.N. and T.K. performed and analysed the deuteration experiments. K.S., S.U., Y.T. and T.S. synthesized the deuterated GGDP precursors. H.S., C.W., T.K. and M.U. discussed the results and wrote the manuscript with feedback from all authors. M.U. supervised the overall research.

## Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

**Competing financial interests:** The authors declare no competing financial interests.

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