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Whole Genome Phylogeny of *Bacillus* by Feature Frequency Profiles (FFP)

Aisuo Wang^{1,2} & Gavin J. Ash^{2,3}

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Fifty complete *Bacillus* genome sequences and associated plasmids were compared using the “feature frequency profile” (FFP) method. The resulting whole-genome phylogeny supports the placement of three *Bacillus* species (*B. thuringiensis*, *B. anthracis* and *B. cereus*) as a single clade. The monophyletic status of *B. anthracis* was strongly supported by the analysis. FFP proved to be more effective in inferring the phylogeny of *Bacillus* than methods based on single gene sequences [16S rRNA gene, *GryB* (gyrase subunit B) and *AroE* (shikimate-5-dehydrogenase)] analyses. The findings of FFP analysis were verified using kSNP v2 (alignment-free sequence analysis method) and Harvest suite (core genome sequence alignment method).

Members of the genus *Bacillus* comprise gram-positive, spore forming, rod-shaped, aerobic bacteria. Three species of the *Bacillus* (*Bacillus thuringiensis*, *Bacillus anthracis* and *Bacillus cereus*) have a huge impact on human activities. For example, *B. anthracis* is the cause of the acute and often lethal disease anthrax¹, which is therefore of a concern as a possible agent in biological warfare; *B. thuringiensis* is extensively used in the biological control of insects due to its ability to produce parasporal protein crystals with insecticidal activity²; *B. cereus* is an opportunistic human pathogen involved in food-poisoning incidents and contaminations in hospitals¹. Some strains of *B. cereus* have been developed as a useful biological control agent in the suppression of fungi and crop disease³.

While the phenotypes of these *Bacillus* species are different, their intra and inter phylogenetic relationships are not clear. Several approaches have been used to classify *B. thuringiensis* strains, including rRNA gene sequences², amplified fragment length polymorphisms (AFLP)², restriction fragment length polymorphisms (RFLPs) in small subunit (SSU) rRNA sequences⁴, *GryB* (gyrase subunit B) and *AroE* (shikimate-5-dehydrogenase) gene sequences⁵. The results of these approaches suggest that there is a high level of sequence homology among the strains of *B. thuringiensis*. Similarly, overall genetic studies have shown that *B. thuringiensis* and *B. cereus* are essentially identical⁶. *B. anthracis* can only be distinguished from *B. thuringiensis* and *B. cereus* through microbiological and biochemical tests⁷. Since these genetic methods are not able to easily distinguish different members of *B. thuringiensis*, *B. anthracis* and *B. cereus*, it becomes necessary to look for some more easily recognizable markers.

With the advent and development of next generation sequencing technologies, a great deal of sequencing data has been generated in recent years. The rapid accumulation of whole genome data of *Bacillus* species in Genbank makes it possible for comparisons of genomic differences over the entire genome that can't be identified in analyses of specific single gene sequences. However, the size of the whole genome data poses great challenges on alignment-based algorithms, which are effective in dealing with closely related sequences but are unable to evaluate the recombination, shuffling, and rearrangement events of the whole genomes⁸. Thus, alignment-free sequence analysis approaches, such as FFP (Feature Frequency Profile), provide attractive alternatives over alignment-based approaches.

¹NSW Department of Primary Industries, Wagga Wagga Agricultural Institute, PMB, Wagga Wagga, NSW, 2650, Australia. ²Graham Centre for Agricultural Innovation, Locked bag 588, Wagga Wagga, NSW, 2678, Australia. ³School of Agricultural and Wine Sciences, Charles Sturt University, Wagga Wagga, NSW, Australia. Correspondence and requests for materials should be addressed to G.A. (email: gash@csu.edu.au)

FFP is a new method used to study the whole genome phylogeny based on k -mers^{9–11}. In this method, the number of features of a particular length l that occur in a particular genome is counted and assembled into a FFP vector. FFPs from different species are then compared using the Jensen–Shannon (JS) Divergence¹². A neighbor-joining phylogenetic tree can thus be constructed based on the resulting distance matrix. Compared to the traditional multiple sequences alignment (MSA) based method, the alignment free FFP method can compare both genic and non-genic regions of the whole genome at higher speed; it can incorporate a wide variety of genomic features into each comparison including intron deletions, exon sequence indels, transposable element insertions, base transversions in coding sequences, and some rare genomic changes such as short interspersed element/long interspersed element (SINE/LINE) insertions¹³. Benefitting from these advantages, this method has been successfully applied to resolving relationships among *Escherichia coli* and *Shigella strains*¹⁰, prokaryotes⁹ and mammals¹³.

In this study, we reconstructed the whole-genome phylogeny of *Bacillus* (with an emphasis on *B. thuringiensis*, *B. anthracis* and *B. cereus*) using the FFP approach, with an aim to better understand the phylogenetic relationships that exist among them. To validate the usefulness of FFP method, we also processed the data with kSNP v2 (alignment-free sequence analysis method) and Harvest Suite (core genome sequence alignment method). For comparison purpose, we constructed phylogenetic trees inferred from three single genes: 16s rRNA genes, *GyrB* and *AroE*, whose DNA sequences were extracted from the corresponding genomes.

Results

The phylogenetic results based on the whole genome data. The phylogenetic tree inferred from the whole genome data of 51 taxa (Table 1) (including 23 *B. thuringiensis* strains, nine *B. anthracis* strains, 11 *B. cereus* strains, three *B. subtilis* strains, one *B. licheniformis* strain, one *B. weihenstephanensis* strain, one *B. clausii* strain, one *B. halodurans* strain and one *E. coli* strain) is presented in Fig. 1. A cluster (I) containing all the *B. thuringiensis*, *B. anthracis* and *B. cereus* strains apart from other *Bacillus* members under study can be recognized (with an exception of *B. weihenstephanensis*). This cluster could be further sub-divided into at least five sub-clusters (I-a to I-e, Fig. 1). The sub-cluster I-b contains all nine *B. anthracis* strains (*B. anthracis* str. A0248, *B. anthracis* str. A16, *B. anthracis* str. A16R, *B. anthracis* str. Ames, *B. anthracis* str. ‘Ames Ancestor’, *B. anthracis* str. CDC 684, *B. anthracis* str. H9401, *B. anthracis* str. Sterne, *B. anthracis* str. SVA11), whereas the sub-cluster of I-a and I-d contain exclusively *B. thuringiensis* strains (*B. thuringiensis* BMB171, *B. thuringiensis* Bt407, *B. thuringiensis* DAR 81934, *B. thuringiensis* HD-771, *B. thuringiensis* IBL 200, *B. thuringiensis* IBL 4222, *B. thuringiensis* serovar andalousiensis BGSC 4AW1, *B. thuringiensis* serovar berliner ATCC 10792, *B. thuringiensis* serovar chinensis CT-43, *B. thuringiensis* serovar huazhongensis BGSC 4BD1, *B. thuringiensis* serovar kurstaki str. HD73, *B. thuringiensis* serovar kurstaki str. T03a001, *B. thuringiensis* serovar kurstaki str. YBT-1520, *B. thuringiensis* serovar monterrey BGSC 4AJ1, *B. thuringiensis* serovar pakistani str. T13001, *B. thuringiensis* serovar pondicheriensis BGSC 4BA1, *B. thuringiensis* serovar pulsiensis BGSC 4CC1, *B. thuringiensis* serovar sotto str. T04001, *B. thuringiensis* serovar thuringiensis str. IS5056, *B. thuringiensis* YBT-1518). Three *B. cereus* strains (*B. cereus* ATCC 10987, *B. cereus* Q1 and *B. cereus* AH187) form a monophyletic clade in sub-cluster I-c, whilst four other *B. cereus* strains (*B. cereus* AH1271, *B. cereus* AH1273, *B. cereus* AH603, *B. cereus* AH621) are closely grouped with *Bacillus weihenstephanensis* KBAB4 in sub-cluster I-e.

In the topology of this NJ tree, three *B. subtilis* strains (*B. subtilis* BSn5, *B. subtilis* subsp. spizizenii str. W23, *B. subtilis* subsp. subtilis str. 168) form a monophyletic clade, which is further grouped with *B. licheniformis* DSM 13 = ATCC 14580. These *Bacillus* strains together with the remaining *Bacillus* members (*B. clausii* KSM-K16 DNA, *B. halodurans* C-125 DNA) are placed near the outgroup *E. coli* (*Escherichia coli* BL21(DE3)).

Validation of FFP results. The NJ tree inferred from the kSNP analyses of the whole genome is presented in Fig. 2. The monophyly of *B. anthracis* was confirmed with high bootstrap support (100). A monophyletic clade containing 16 *B. thuringiensis* isolates was recognized (clade *Bacillus thuringiensis*). All the *B. anthracis*, *B. cereus*, *B. thuringiensis* and *B. weihenstephanensis* form a monophyletic clade (*Bacillus cereus sensu lato*), which is separated from the remaining *Bacillus* species examined in this study. Outside this major clade, the monophyly of *B. subtilis* was confirmed (100 bootstrap support).

The core SNP matrix resulted from the kSNP analysis provided a direct visualization of the relationships among all the *Bacillus* species studied (Fig. 3). There was no variation between the core SNPs of *B. anthracis* and *B. thuringiensis*, whilst only single variation was found for two *B. cereus* strains (*Bacillus cereus* AH603 and *Bacillus cereus* AH621) and *B. weihenstephanensis*. The variation of core SNP increased to two among the *B. subtilis* species and the *B. licheniformis* DSM 13 = ATCC 1458034. The sharp increase of core SNP variations in *B. halodurans* C-12533 and *B. clausii* KSM-K16 (4 and 5 respectively) revealed their distant relationships to the remaining *Bacillus* species.

Our effort in using Harvest suite to analyse all the species examined in FFP was not successful. The shared core genome among all the studied taxa was too low (less than 1%) to let the Parsnp program to work. This is because Parsnp is designed for intraspecific alignments and requires $\geq 97\%$ average nucleotide identity among input genomes. The Parsnp started to work when *Bacillus* species other than the member of *Bacillus cereus sensu lato* were excluded from the analysis. The final alignment and the resulting NJ tree are presented in Figs 4–6. The NJ tree distinguished two highly supported clades (100 in

Species	Assess No.	Genome size (bp)	Plasmid (Accession No., Size in bp)
<i>Bacillus anthracis</i> str. A0248	CP001598.1	5227419	Pxo1(CP001599.1, 181677); Pxo2(CP001597.1, 94830)
<i>Bacillus anthracis</i> str. A16	CP001970.1	5227898	pXO1(CP001971.1, 181764); pXO2(CP001972.1, 94839)
<i>Bacillus anthracis</i> str. A16R	CP001974.1	5227683	pXO1(CP001975.1, 181763)
<i>Bacillus anthracis</i> str. Ames ²¹	AE016879.1	5227293	Nil
<i>Bacillus anthracis</i> str. Ames Ancestor ²⁹	AE017334.2	5227419	pXO1(AE017336.2, 181677); pXO2(AE017335.3, 94830)
<i>Bacillus anthracis</i> str. CDC 684	CP001215.1	5230115	pXO1(CP001216.1, 181773); pXO2(CP001214.1, 94875)
<i>Bacillus anthracis</i> str. H9401 ³⁰	CP002091.1	5218947	BAP1(CP002092.1, 181700); BAP2(CP002093.1, 94824)
<i>Bacillus anthracis</i> str. Sterne	AE017225.1	5228663	Nil
<i>Bacillus anthracis</i> str. SVA11 ³¹	CP006742.1	5210966	Pxo1(CP006743.1, 181793); pXO2(CP006744.1, 94758)
<i>Bacillus cereus</i> 03BB102	CP001407.1	5269628	p03BB102_179(CP001406.1, 179680)
<i>Bacillus cereus</i> AH1271	CM000739.1	5656704	Nil
<i>Bacillus cereus</i> AH1272	CM000740.1	5789540	Nil
<i>Bacillus cereus</i> AH1273	CM000741.1	5790501	Nil
<i>Bacillus cereus</i> AH187	CP001177.1	5269030	pAH187_12(CP001178.1, 12481); pAH187_270(CP001179.1, 270082); pAH187_3(CP001181.1, 3091); pAH187_45(CP001180.1, 45173)
<i>Bacillus cereus</i> AH603	CM000737.1	5799451	Nil
<i>Bacillus cereus</i> AH621	CM000719.1	5674808	Nil
<i>Bacillus cereus</i> AH820	CP001283.1	5302683	pAH820_10(CP001286.1, 10915); pAH820_272(CP001285.1, 272145); pAH820_3(CP001284.1, 3091)
<i>Bacillus cereus</i> ATCC 10987 ³²	AE017194.1	5224283	pBc10987(AE017195.1, 208369)
<i>Bacillus cereus</i> E33L	CP000001.1	5300915	pE33L466(CP000040.1, 466370); pE33L5(CP000041.1, 5108); pE33L54(CP000042.1, 53501); pE33L8(CP000043.1, 8191); pE33L9(CP000044.1, 9150)
<i>Bacillus cereus</i> Q1 ³³	CP000227.1	5214195	pBc239(CP000228.1, 239246); pBc53(CP000229.1, 52766)
<i>Bacillus clausii</i> KSM-K16 ³⁴	AP006627.1	4303871	Nil
<i>Bacillus halodurans</i> C-125 ³⁵	BA000004.3	4202352	Nil
<i>Bacillus licheniformis</i> DSM 13 = ATCC 14580 ³⁶	AE017333.1	4222645	Nil
<i>Bacillus subtilis</i> BSn5 ³⁷	CP002468.1	4093599	Nil
<i>Bacillus subtilis</i> subsp. spizizenii str. W23 ³⁸	CP002183.1	4027676	Nil
<i>Bacillus subtilis</i> subsp. subtilis str. 168 ³⁹⁻⁴¹	CM000487.1	4214547	Nil
<i>Bacillus thuringiensis</i> BMB171 ⁴²	CP001903.1	5643051	pBMB171(CP001904.1, 312963)
<i>Bacillus thuringiensis</i> Bt407 ⁴³	CM000747.1	6026843	BTB_15p(CP003892.1, 15189); BTB_2p(CP003897.1, 2062); BTB_502p(CP003890.1, 501911); BTB_5p(CP003896.1, 5518); BTB_6p(CP003895.1, 6880); BTB_78p(CP003891.1, 77895); BTB_7p(CP003894.1, 7635); BTB_9p(CP003898.1, 8513); BTB_8p(CP003893.1, 8240);
<i>Bacillus thuringiensis</i> DAR 81934 ²³	CM001804.1	5628425	Nil
<i>Bacillus thuringiensis</i> HD-771	CP003752.1	5883036	p01(CP003753.1, 171030); p02(CP003754.1, 168999); p03(CP003755.1, 69876); p04(CP003756.1, 65470); p05(CP003757.1, 45262); p06(CP003758.1, 14056); p07(CP003759.1, 9070); p08(CP003760.1, 8574)
<i>Bacillus thuringiensis</i> IBL 200	CM000758.1	6731790	Nil
<i>Bacillus thuringiensis</i> IBL 4222	CM000759.1	6616432	Nil
<i>Bacillus thuringiensis</i> serovar andalousiensis BGSC 4AW1	CM000754.1	5488844	Nil
<i>Bacillus thuringiensis</i> serovar berliner ATCC 10792	CM000753.1	6260142	Nil
<i>Bacillus thuringiensis</i> serovar chinensis CT-43 ⁴⁴	CP001907.1	5486830	pCT127(CP001908.1, 127885); pCT14(CP001909.1, 14860); pCT281(CP001910.1, 281231); pCT51(CP001911.1, 51488); pCT6880(CP001912.1, 6880); pCT172(CP001913.1, 72074); pCT8252(CP001914.1, 8252); pCT83(CP001915.1, 83590); pCT8513(CP001916.1, 8513); pCT9547(CP001917.1, 9547)
<i>Bacillus thuringiensis</i> serovar finitimus YBT-020 ⁴⁵	CP002508.1	5235490	pBMB26(CP002509.1, 187880); pBMB28(CP002510.1, 139013)
Continued			

Species	Assess No.	Genome size (bp)	Plasmid (Accession No., Size in bp)
<i>Bacillus thuringiensis</i> serovar <i>huazhongensis</i> BGSC 4BD1	CM000756.1	6231196	Nil
<i>Bacillus thuringiensis</i> serovar <i>konkukian</i> str. 97-27	AE017355.1	5237682	pBT9727(CP000047.1, 77112)
<i>Bacillus thuringiensis</i> serovar <i>kurstaki</i> str. HD73 ⁴⁶	CP004069.1	5646799	pAW63(CP004072.1, 71777); pHT11(CP004073.1, 11769); pHT7(CP004076.1, 7635); pHT73(CP004070.1, 77351); pHT77(CP004071.1, 76490); pHT8_1(CP004074.1, 8513); pHT8_2(CP004075.1, 8241)
<i>Bacillus thuringiensis</i> serovar <i>kurstaki</i> str. T03a001	CM000751.1	5527568	Nil
<i>Bacillus thuringiensis</i> serovar <i>kurstaki</i> str. YBT-1520	CP004858.1	5602265	pBMB11(CP004863.1, 11769); pBMB2062(CP004859.1, 2062); pBMB293(CP004861.1, 293574); pBMB422(CP004860.1, 422692); pBMB53(CP004862.1, 53838); pBMB67(CP004869.1, 67159); pBMB7635(CP004867.1, 7635); pBMB7921(CP004866.1, 7921); pBMB8240(CP004865.1, 8240); pBMB8513(CP004864.1, 8513); pBMB94(CP004868.1, 94568)
<i>Bacillus thuringiensis</i> serovar <i>monterrey</i> BGSC 4AJ1	CM000752.1	6489024	Nil
<i>Bacillus thuringiensis</i> serovar <i>pakistani</i> str. T13001	CM000750.1	6037513	Nil
<i>Bacillus thuringiensis</i> serovar <i>pondicheriensis</i> BGSC 4BA1	CM000755.1	6031475	Nil
<i>Bacillus thuringiensis</i> serovar <i>pulsiensis</i> BGSC 4CC1	CM000757.1	6002603	Nil
<i>Bacillus thuringiensis</i> serovar <i>sotto</i> str. T04001	CM000749.1	6107746	Nil
<i>Bacillus thuringiensis</i> serovar <i>thuringiensis</i> str. IS5056 ⁴⁷	CP004123.1	5491935	pIS56-107(CP004134.1, 107431); pIS56-11(CP004127.1, 11331); pIS56-15(CP004128.1, 15185); pIS56-16(CP004129.1, 16206); pIS56-233(CP004135.1, 233730); pIS56-285(CP004136.1, 285459); pIS56-328(CP004137.1, 328151); pIS56-39(CP004130.1, 39749); pIS56-6(CP004124.1, 6880); pIS56-63(CP004131.1, 63864); pIS56-68(CP004132.1, 68616); pIS56-8(CP004125.1, 8251); pIS56-85(CP004133.1, 85134); pIS56-9(CP004126.1, 9671)
<i>Bacillus thuringiensis</i> str. <i>Al Hakam</i> ⁴⁸	CP000485.1	5257091	pALH1(CP000486.1, 55939)
<i>Bacillus thuringiensis</i> YBT-1518	CP005935.1	6002284	pBMB0229(CP005936.1, 45206); pBMB0230(CP005937.1, 49195); pBMB0231(CP005938.1, 146276); pBMB0232(CP005939.1, 171593); pBMB0233(CP005940.1, 240661)
<i>Bacillus weihenstephanensis</i> KBAB ⁴⁹	CP000903.1	5262775	pBWB401(CP000904.1, 417054); pBWB402(CP000905.1, 75107); pBWB403(CP000906.1, 64977); pBWB404(CP000907.1, 52830)
<i>Escherichia coli</i> BL21(DE3)	AM946981.2	4558947	Nil

Table 1. Source of sequence data.

bootstrap value): one including all the *B. anthracis* strains and the other including sixteen *B. thuringiensis* strains. The whole topology of this NJ tree is highly similar to that of the NJ trees inferred from FFP and kSNP analyses (Figs 1 and 2).

The Gingr visualization of NJ tree and the genome alignment (core genome based) displayed multiple conserved regions (represented by the SNP heatmap) throughout the entire genome across 44 members of *Bacillus cereus sensu lato* (Figs 5 and 6). These conserved regions are scattered throughout the genome but showed more density in four regions (500–1500 bp; 11000–15000 bp, 36000–46000 bp and 52000–53000 bp). When being zoomed, the Gingr visualization turned the SNP heatmap into vertical lines, which revealed the phylogenetic signature of several clades [in this case within the fully-aligned *dpaA* gene (BC3801)] (Fig. 6).

The phylogenetic results based on the single gene data. Three NJ trees inferred from the data of three single genes (16s rRNA gene, *GyrB* and *AroE*), are shown in Figs 7–9 respectively. These trees did not support the monophyletic status of *B. anthracis*. The clades that contain *B. anthracis* strains also contain other *Bacillus* species (e.g. *B. cereus* AH820 in Clade II of Fig. 8, and *B. thuringiensis* serovar *monterrey* BGSC 4AJ1 in Clade D of Fig. 9). Among the total 23 *B. thuringiensis* strains studied, some strains form monophyletic sub-clades in trees inferred from *GyrB* (Clade V, Fig. 8) and *AroE* (Clade A and C, Fig. 9), but the monophyletic status of the whole *B. thuringiensis* strains cannot be confirmed by these analyses. Similarly, *B. cereus* proved to be a paraphyletic group by all NJ trees inferred from data of

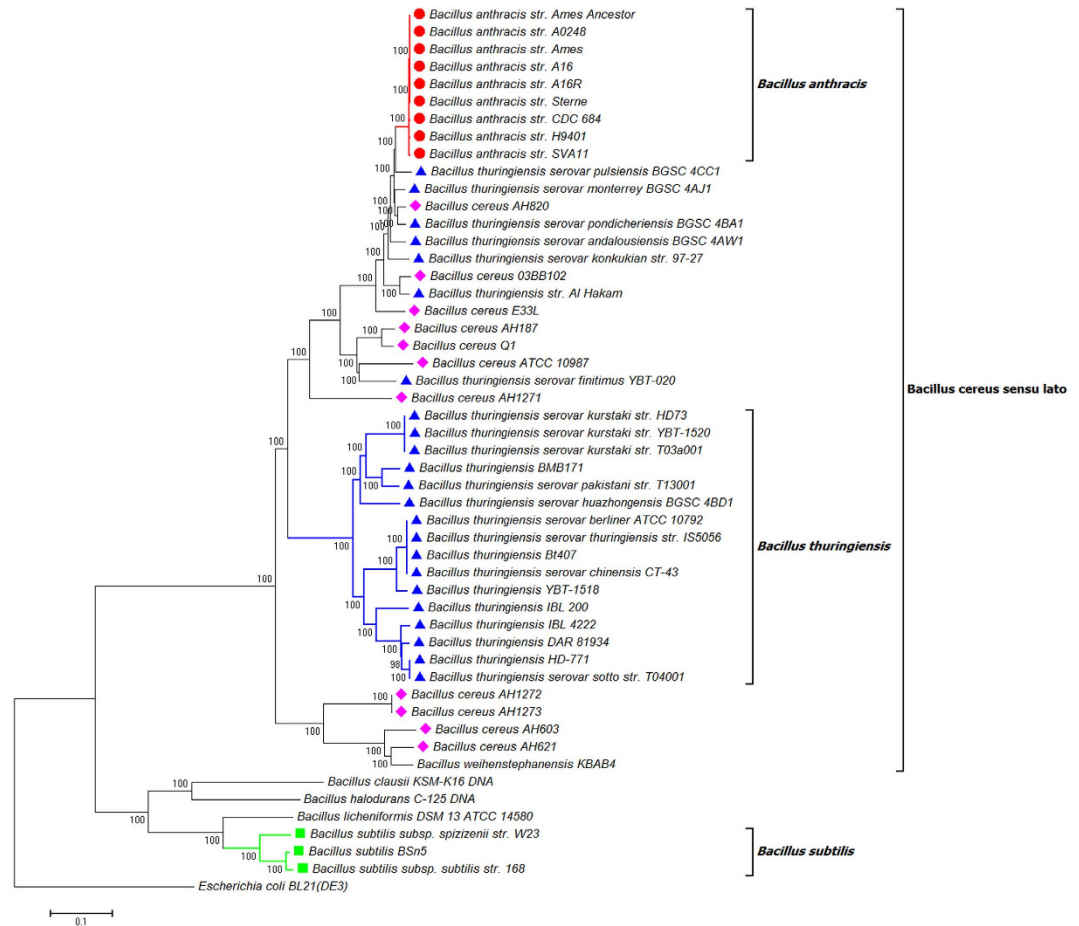


Figure 2. Phylogenetic tree of 50 *Bacillus* strains. The tree was constructed using the NJ algorithm based on all SNP matrix inferred from the kSNP V2 analysis. *Escherichia coli* BL21 (DE3) (AM946981.2) was used as an outgroup in the analysis. The bootstrap confidence values were generated using 1,000 permutations. Different symbols were allocated to represent different species: Blue triangle for *Bacillus thuringiensis*; Pink diamond for *Bacillus cereus*; Red circle for *Bacillus anthracis*; Green Square for *Bacillus subtilis*.

Discussion

Our phylogenetic analysis based on the FFP features of the whole genome and associated plasmids resulted in a major cluster containing all strains of *B. thuringiensis*, *B. anthracis* and *B. cereus* separated from other recognised *Bacillus* members. When strains of same species were grouped together and subject to pairwise distance analysis, the groups of *B. thuringiensis*, *B. anthracis* and *B. cereus* formed a monophyletic clade in the NJ tree (Fig. 10). These results clearly suggest the close relationship among *B. thuringiensis*, *B. anthracis* and *B. cereus* species, and are in agreement with earlier results from DNA-DNA hybridization analysis and Multi Locus Enzyme Electrophoresis (MEE), which showed high identity among *B. anthracis*, *B. cereus*, and *B. thuringiensis* strains¹⁴. These three species have been grouped under the name of *Bacillus cereus sensu lato*¹⁵ despite their obvious difference in phenotype and pathological effects, which are resulted from the genetic difference in plasmid rather than in chromosome¹. The results of present study appear to support the classification of *Bacillus cereus sensu lato* when using genomic sequences only (data not shown). Greater resolution of recognised species was achieved when plasmid sequences were added to the analysis.

In the present study, *B. weihenstephanensis* strain KBAB4 was found to be very closely grouped with the major cluster I-d consisting of all *B. thuringiensis* isolates and proximal to cluster I-e (*B. cereus*) and cluster I-c (a cluster containing both *B. thuringiensis* and *B. cereus* strains) (Fig. 1). *B. weihenstephanensis* is a member of the *Bacillus cereus sensu lato*, and has high similarities with *B. thuringiensis* and *B. cereus* in terms of its ecological features such as producing cereulide as *B. cereus* and being psychrotolerance as some *B. thuringiensis* isolates^{16,17}. Soufiane and Cote (2009)⁵ revealed the close relationship between *B. weihenstephanensis* and some *B. thuringiensis* serovars based on the 16S rRNA, *GyrB* and *AroE* gene sequences. Our results support their research and provide further evidence for the classification of *Bacillus cereus sensu lato*.

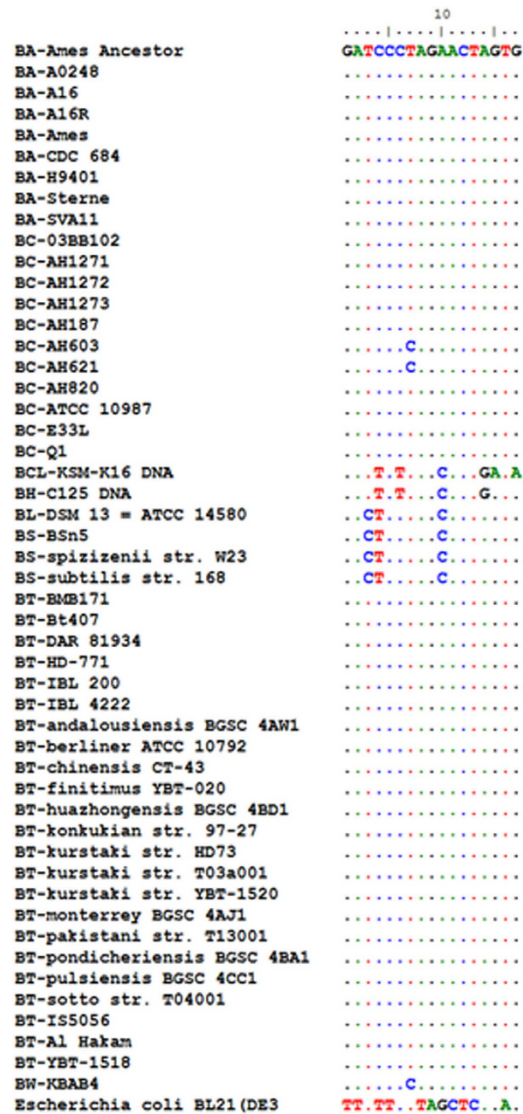


Figure 3. Core SNP matrix inferred from kSNP v2 (BA: *Bacillus anthracis*, BC: *Bacillus cereus*, BCL: *Bacillus clausii*, BH: *Bacillus halodurans*, BL: *Bacillus licheniformis*, BS: *Bacillus subtilis*, BT: *Bacillus thuringiensis*, BW: *Bacillus weihenstephanensis*).

The NJ tree inferred from the whole genome sequences of these bacteria species not only revealed the close relationship among *B. thuringiensis*, *B. anthracis* and *B. cereus*, but also confirmed the monophyly of *B. anthracis* (I-b, Fig. 1). Previous studies using other techniques have all stated that *B. anthracis* is the most monomorphic species among *B. thuringiensis*, *B. anthracis* and *B. cereus*^{18–20}. Our results confirmed the genetic homogeneity of *B. anthracis* but failed to elucidate the evolutionary relationships between *B. anthracis* and the remaining two species. *B. anthracis* has been regarded to be evolved from a *B. cereus* ancestor through acquisition of key plasmid-encoded toxin, capsule and regulatory loci²¹. Such a relationship did not appear in our phylogenetic analyses based on FFP analysis of whole genome data (Fig. 1). The *B. anthracis* clade is proximal to a number of isolates of *B. cereus* and *B. thuringiensis* which have been associated with disease or toxicity in humans.

The findings of FFP analyses were fully supported by SNP phylogenies construed by kSNP (alignment – free sequence analysis method) and Parsnp (core genome alignment method). By comparing the NJ trees inferred from FFP analysis (Fig. 1) and kSNP analysis (Fig. 2), we found a high level of similarity between two phylogenies. The clades of I - b and I - d clades in FFP tree are consistent with the *Bacillus anthracis* and *Bacillus thuringiensis* clades in kSNP tree, whilst the cluster I in FFP tree is corresponding to the clade of *Bacillus cereus sensu lato* in kSNP tree. While the core genome SNP tree constructed by Parsnp failed to cover all the species studied, the exclusion of other *Bacillus* species from the major cluster was actually a support for the monophyly of *Bacillus cereus sensu lato*. This is because Parsnp is limited in intraspecific alignment and can only tolerate genomes with high similarity (>=97%). Genomes from other species will be automatically excluded from the full alignment²².

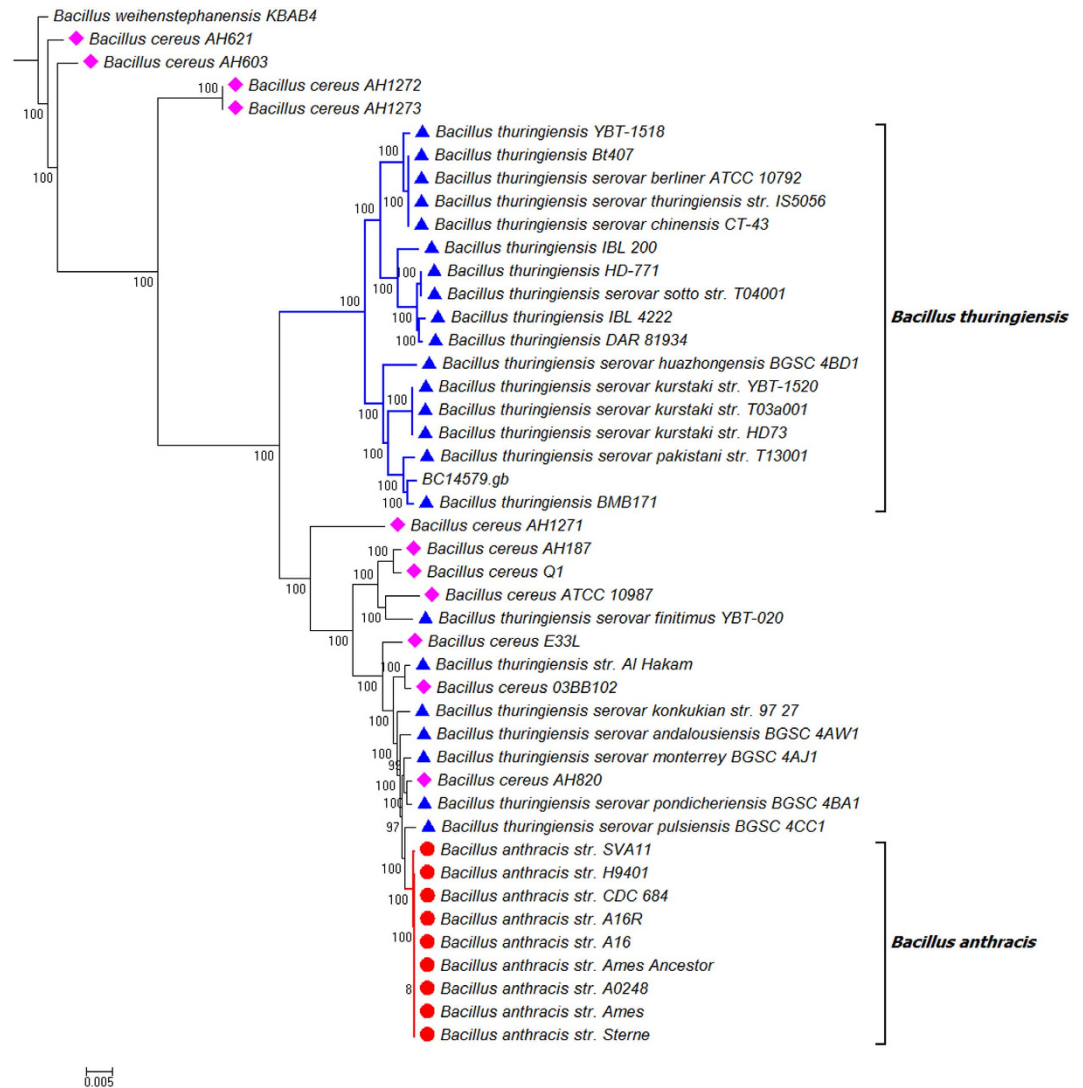


Figure 4. NJ tree of 44 *Bacillus* strains. The tree was constructed using Parsnp and annotated in MEGA 6.0. The NJ algorithm was based on the Core Genome SNP data of the 44 *Bacillus* strains. *Bacillus weihenstephanensis* KBAB4 was used as an outgroup in the analysis. The bootstrap confidence values were generated using 1,000 permutations. Different symbols were allocated to represent different species: Blue triangle for *Bacillus thuringiensis*; Pink diamond for *Bacillus cereus*; Red circle for *Bacillus anthracis*. The Genbank file of *Bacillus cereus* ATCC 14579 (represented as BC14579) was used as reference in the Parsnp analysis.

Within the core genome SNP tree constructed by Parsnp and visualized by Gingr, the monophyly of *B. anthracis* and a subclade covering 16 *B. thuringiensis* strains were confirmed, which is consistent with the results of FFP analysis. The tree also revealed the paraphyly of *B. cereus* and *B. thuringiensis*, which is similar to the findings of FFP and kSNP analyses. By zooming the alignment files from genome level to nucleotide level via the fisheye zoom feature of Gingr²², we noticed the SNP variations across different strains of *B. cereus* and *B. thuringiensis* that affects the topology of the trees. For *B. cereus*, the most variable region falls on an area between the gene of *Cytochrome d ubiquinol oxidase subunit II* and the gene of *Alanine racemase* (around 121456 bp). There are more SNP sites at this region among four *B. cereus* strains (*B. cereus* AH603, *B. cereus* AH621, *B. cereus* AH1272 and *B. cereus* AH1273), which contributed the distant placement of these four strains from the remaining *B. cereus* strains in the NJ tree. Similarly, we found a region (around 1006988 bp) with high SNP density in *B. thuringiensis* (starting from the gene of *Lysr-type transcriptional regulator* and ending at the gene of *Thiamine/molybdopterin biosynthesis protein*). The distantly placed *B. thuringiensis* strains (such as *B. thuringiensis* serovar *finitimus* YBT-020 and *B. thuringiensis* str. *Al Hakam*) generally have more SNP sites in this region than that of the remaining *B. thuringiensis* strains. It is not clear why some *B. cereus* and *B. thuringiensis* strains have more SNP variations at these particular genome regions than that of other strains, and what are the impacts of these

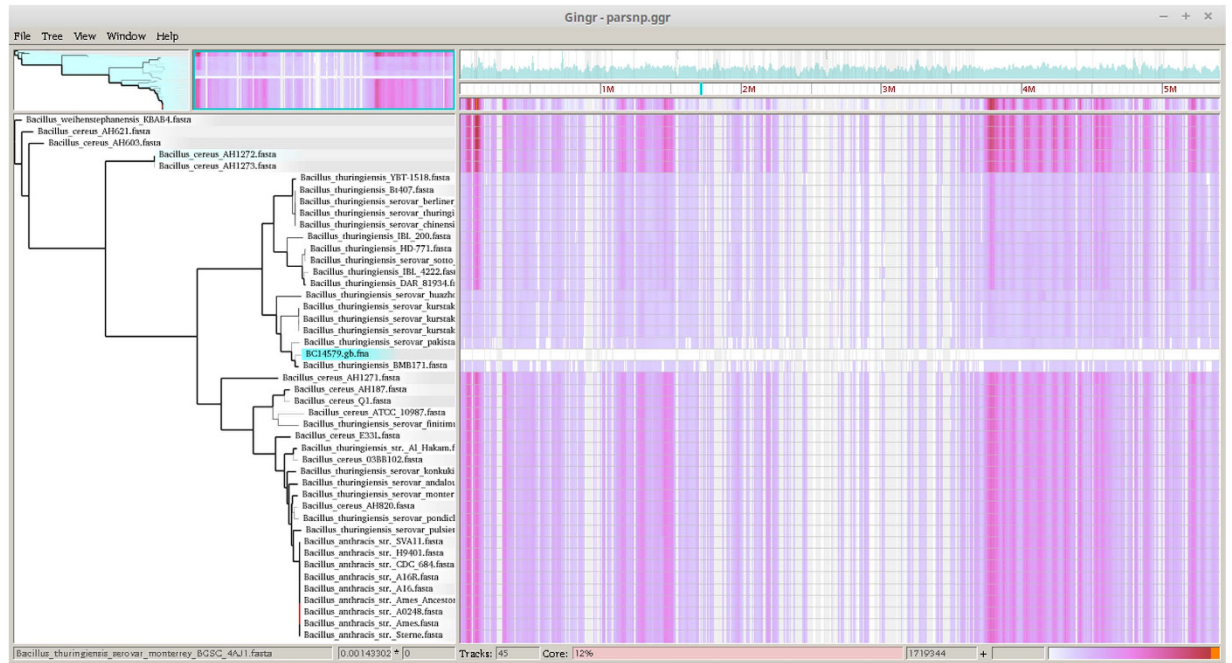


Figure 5. Gingr visualization of 44 *Bacillus* genomes aligned with Parsnp. The leaves of the reconstructed phylogenetic tree (left) are paired with their corresponding rows in the multi-alignment.

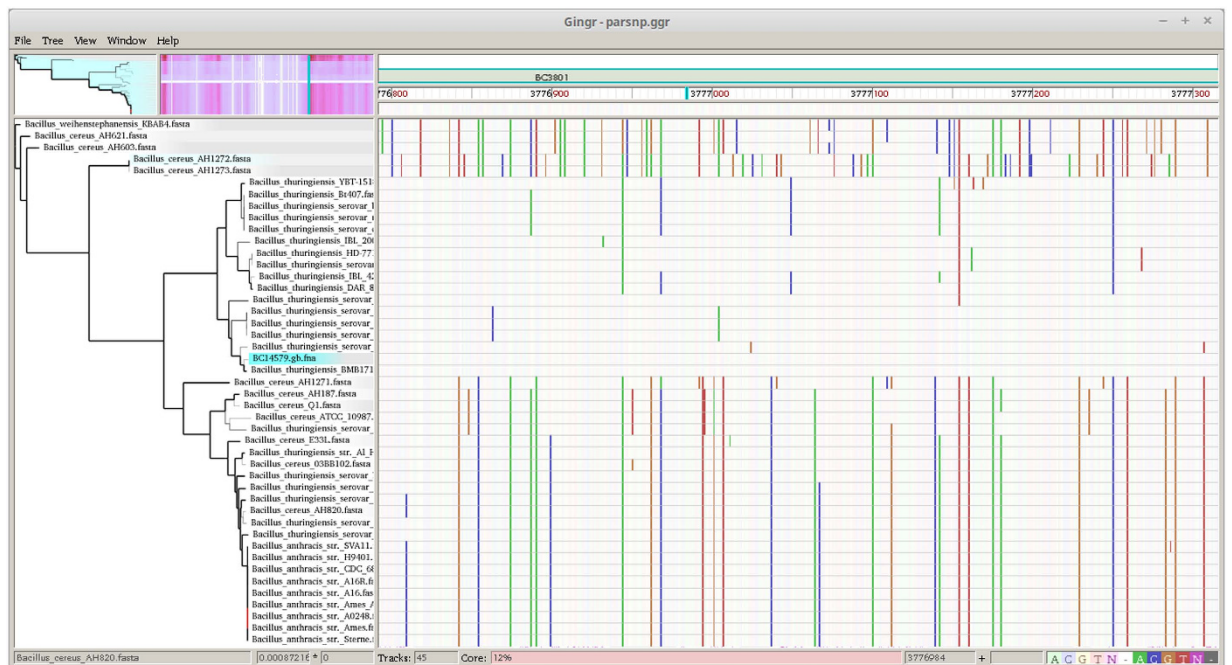


Figure 6. Gingr visualization of 44 *Bacillus* genomes aligned with Parsnp. The visual layout is the same as Fig. 5, but unlike Fig. 5, the genome alignment was zoomed to reveal the phylogenetic signature of several clades, in this case within the fully-aligned BC3801 (dipicolinate synthase subunit A).

SNP variations on the phenotype, function and pathogenicity of these *Bacillus* strains. More research is thus required to answer these questions.

In contrast to the FFP analysis based on the whole genome data, our phylogenetic analysis based on single gene data (16S rRNA gene, *GyrB* and *AroE*) were unable to clearly distinguish between *Bacillus* species examined. The 16S rRNA gene sequence analysis clustered all the sequences together and provided poor resolution for the relationships between each strain. Similarly, our analysis based on two protein coding genes, *GyrB* and *AroE*, were unable to separate *B. thuringiensis*, *B. anthracis* and *B. cereus*

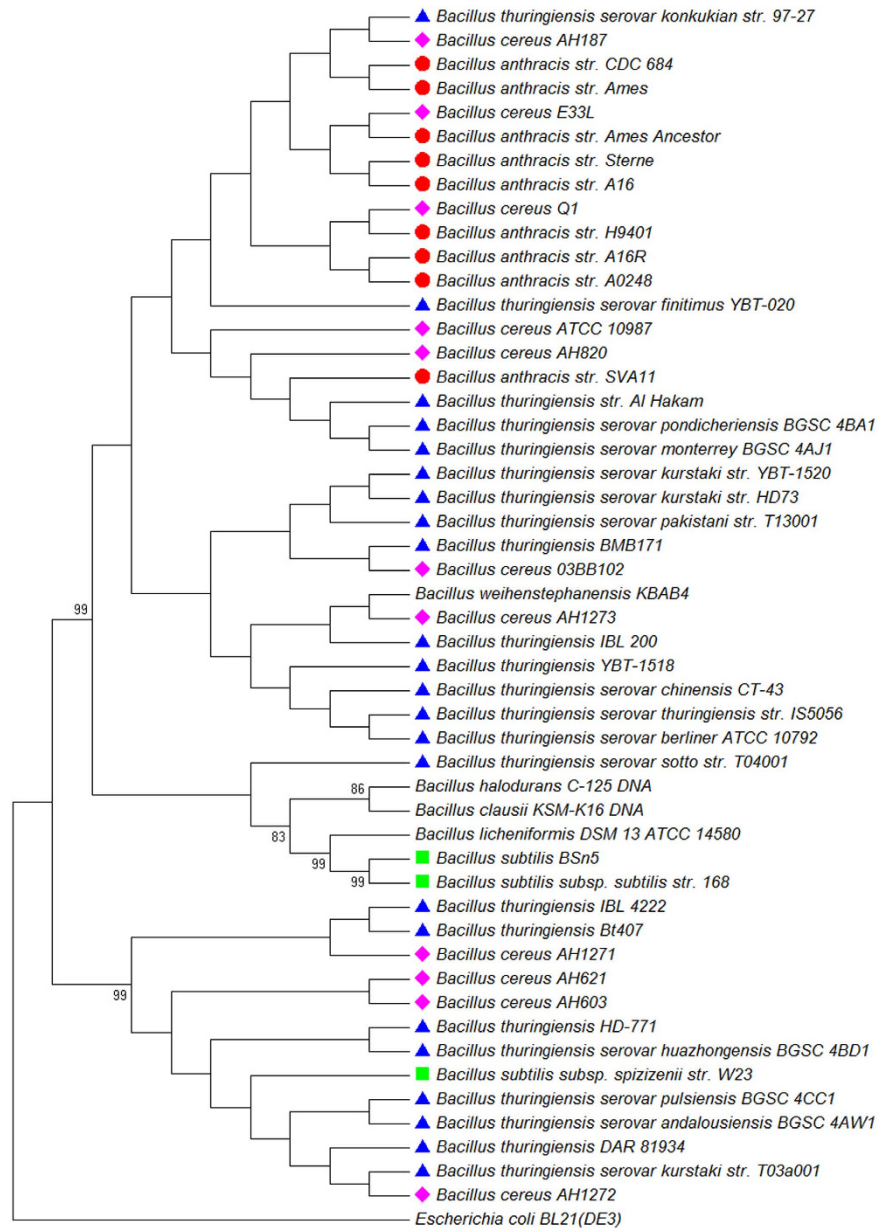


Figure 7. Neighbor-joining tree constructed based on the sequences of the 16S rRNA gene from 50 *Bacillus* strains. *Escherichia coli* BL21 (DE3) (AM946981.2) was used as an outgroup in the analysis. The bootstrap confidence values were generated using 1,000 permutations. Different symbols were allocated to represent different species: Blue triangle for *Bacillus thuringiensis*; Pink diamond for *Bacillus cereus*; Red circle for *Bacillus anthracis*; Green Square for *Bacillus subtilis*.

from other *Bacillus* members while they provided support for the monophyletic position of *B. subtilis*. A further analysis using the concatenated sequence of these genes failed to provide any better analysis (data not shown).

These results suggest that FFP analysis of the combined genomic and plasmid sequence data allows for comparisons of genomic differences that can't be identified in analyses of specific single gene sequences and provides greater resolution of species belonging to *B. cereus sensu lato* than other techniques such as MLST, AFLP or single gene sequence analysis. Furthermore, the availability and reduced cost of whole genome sequencing can be used without extensive gene annotation to provide robust phylogenetic analysis of new isolates as they become available.

Materials and Methods

Source of sequence data. The genome sequence of *Bacillus thuringiensis* strain DAR 81934 was from our previous research²³. Genome sequences of other 49 *Bacillus* and one *E. coli* [*Escherichia coli* BL21(DE3)] (used as outgroup) were retrieved from GenBank (Table 1). The retrieved genome sequences

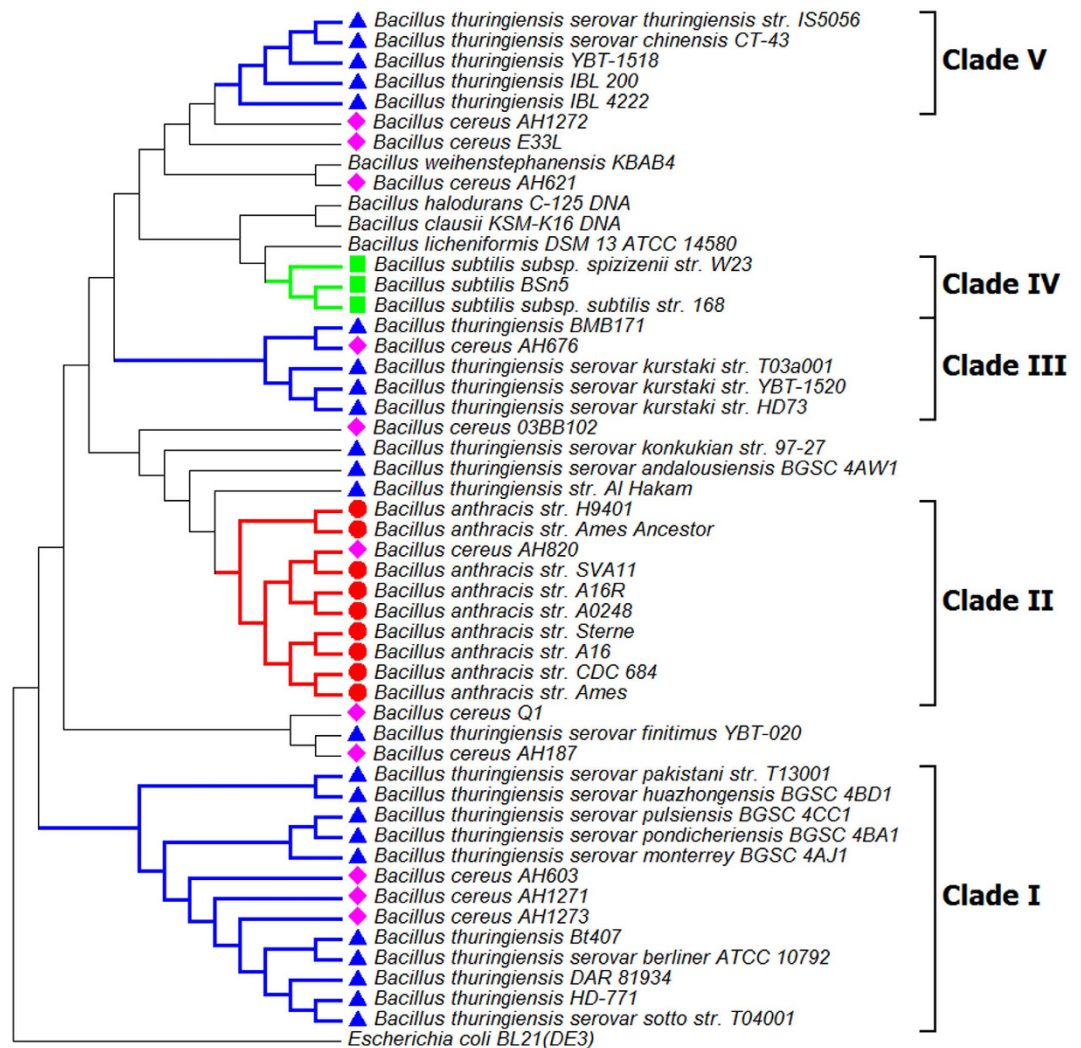


Figure 8. Neighbor-joining tree constructed based on the sequences of the *GyrB* from 50 *Bacillus* strains. *Escherichia coli* BL21 (DE3) (AM946981.2) was used as an outgroup in the analysis. The bootstrap confidence values were generated using 1,000 permutations. Different symbols were allocated to represent different species: Blue triangle for *Bacillus thuringiensis*; Pink diamond for *Bacillus cereus*; Red circle for *Bacillus anthracis*; Green Square for *Bacillus subtilis*.

cover both the main chromosome sequences and the plasmid sequences (if any) of each species. The nucleotide sequences of three single nuclear genes for these taxa: 16s rRNA, *GyrB* and *AroE*, were extracted from the corresponding whole genome sequences.

Phylogeny analysis via FFP. The whole genome sequences of the 51 taxa were converted to multi-Fasta format before being uploaded to FFP -3.19¹⁰, where the different forms of genome partitions were compared between species, and NJ trees were constructed based on the Jensen-Shannon divergences matrix from each type of genome partition. By following the recommendations of the program, we used the tools of *ffpvocab* and *ffpre* to find the right range of lengths to use ($l = 20$ was finally chosen in the analysis). We also conducted bootstrapping (1000) to assess the FFP phylogenetic analysis. The outcome of the bootstrap analysis was imported into Phylip 3.2²⁴ to create a consensus distance matrix, which was further processed in MEGA 6.0²⁵ to display the final NJ tree.

Validation of FFP results. We applied two programs to validate the outcomes of FFP analysis. The first program is *kSNP v2.1.2*²⁶, an alignment-free sequence analysis method with the capacity to build whole genome phylogeny on single nucleotide polymorphisms (SNPs) in whole genome data. We examined the same datasets of FFP by running *kchooser* to find the optimum k -mer size (19) prior to the *kSNP* analysis, and including the flag of “-j” in the command line to estimate Neighbor Joining trees based on all SNPs and core SNPs. The resulting all-SNPs-matrix was imported into MEGA 6.0²⁵ for NJ

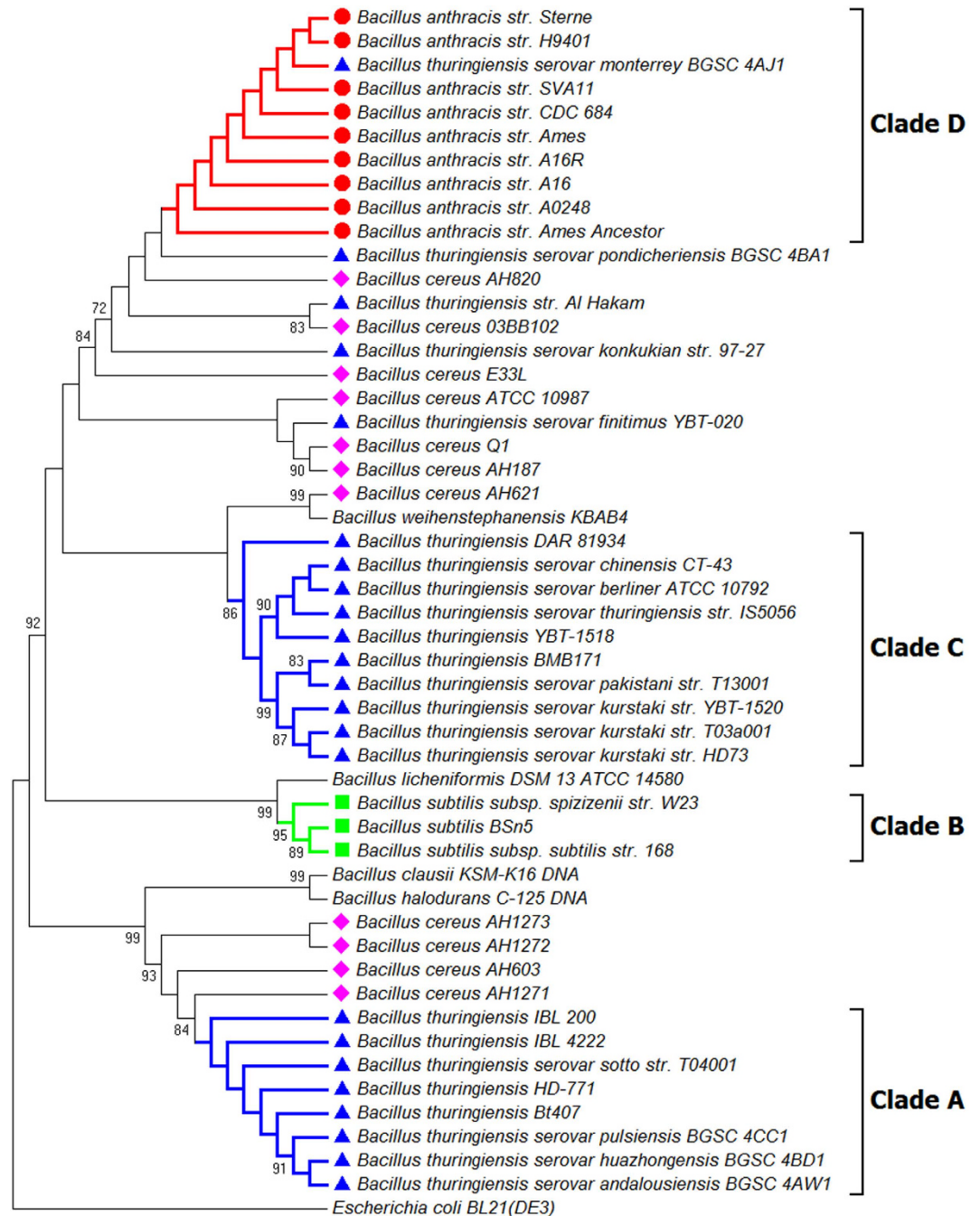


Figure 9. Neighbor-joining tree constructed based on the sequences of the *AroE* from 50 *Bacillus* strains. *Escherichia coli* BL21 (DE3) (AM946981.2) was used as an outgroup in the analyse. The bootstrap confidence values were generated using 1,000 permutations. Different symbols were allocated to represent different species: Blue triangle for *Bacillus thuringiensis*; Pink diamond for *Bacillus cereus*; Red circle for *Bacillus anthracis*; Green Square for *Bacillus subtilis*.

tree construction. The core-SNP-smatrix was applied to demonstrate the core SNP differneces across all examined genomes.

The Harvest Suite²² (including Parsnp and Gingr) was also applied to validate the FFP outcome. We aligned genomes studied in FFP and built NJ phylogentic trees through Parsnp, a fast core-genome multi-aligner, and vusualized the alignment and trees with Gingr, a dynamic visual platform. The default parameters recommanded by the program were followed during the whole analysis.

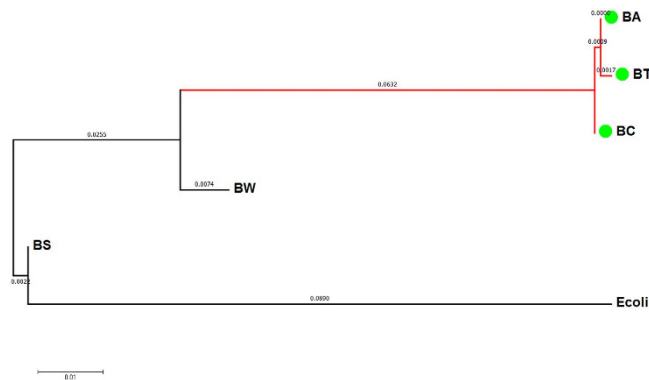


Figure 10. Pairwise Distance of the Bacillus numbers. The scale bar represents a 1% sequence difference (BA:*B. anthracis*; BT:*B. thuringiensis*; BC:*B. cereus*; BW:*B. weihenstephanensis*; BS:*B. subtilis* and *B. clausii*, *B. halodurans*, *B. licheniformis*) are placed near the outgroup *E. coli* (*Escherichia coli* BL21(DE3)).

Single gene based phylogeny. The retrieved single gene sequences of 16S rRNA, *GyrB* and *AroE* were imported into MEGA 6.0 for sequence alignment (Clustal W²⁷) and phylogenetic analysis (Neighbor-Joining²⁸). The Kimura 2-parameter model was selected by executing the function of “Find Best DNA/Protein Models” prior to the Phylogenetic analyses. Statistical confidence on the inferred tree topology was assessed with 1,000 bootstrap replications.

References

- Helgason, E. *et al.* *Bacillus anthracis*, *Bacillus cereus*, and *Bacillus thuringiensis*—One species on the basis of genetic evidence. *Applied and Environmental Microbiology* **66**, 2627–2630 (2000).
- Schnepf, E. *et al.* *Bacillus thuringiensis* and its pesticidal crystal proteins. *Microbiol Mol Biol Rev* **62**, 775–806 (1998).
- Silo-Suh, L. A. *et al.* Biological activities of two fungistatic antibiotics produced by *Bacillus cereus* UW85. *Appl Environ Microbiol* **60**, 2023–30 (1994).
- Joung, K. B. & Cote, J. C. Phylogenetic analysis of *Bacillus thuringiensis* serovars based on 16S rRNA gene restriction fragment length polymorphisms. *J Appl Microbiol* **90**, 115–22 (2001).
- Soufiane, B. & Cote, J. C. Discrimination among *Bacillus thuringiensis* H serotypes, serovars and strains based on 16S rRNA, *gyrB* and *aroE* gene sequence analyses. *Antonie Van Leeuwenhoek International Journal of General and Molecular Microbiology* **95**, 33–45 (2009).
- Helgason, E. *et al.* Genetic diversity of *Bacillus cereus* B-thuringiensis isolates from natural sources. *Current Microbiology* **37**, 80–87 (1998).
- Ticknor, L. O. *et al.* Fluorescent amplified fragment length polymorphism analysis of Norwegian *Bacillus cereus* and *Bacillus thuringiensis* soil isolates. *Applied and Environmental Microbiology* **67**, 4863–4873 (2001).
- Cheng, J. K., Cao, F. L. & Liu, Z. H. AGP: A Multimethods Web Server for Alignment-Free Genome Phylogeny. *Molecular Biology and Evolution* **30**, 1032–1037 (2013).
- Jun, S. R., Sims, G. E., Wu, G. H. A. & Kim, S. H. Whole-proteome phylogeny of prokaryotes by feature frequency profiles: An alignment-free method with optimal feature resolution. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 133–138 (2010).
- Sims, G. E., Jun, S. R., Wua, G. A. & Kim, S. H. Alignment-free genome comparison with feature frequency profiles (FFP) and optimal resolutions. *Proceedings of the National Academy of Sciences of the United States of America* **106**, 2677–2682 (2009).
- Sims, G. E. & Kim, S. H. Whole-genome phylogeny of *Escherichia coli*/Shigella group by feature frequency profiles (FFPs). *Proceedings of the National Academy of Sciences of the United States of America* **108**, 8329–8334 (2011).
- Lin, J. H. Divergence Measures Based on the Shannon Entropy. *Ieee Transactions on Information Theory* **37**, 145–151 (1991).
- Sims, G. E., Jun, S. R., Wu, G. A. & Kim, S. H. Whole-genome phylogeny of mammals: Evolutionary information in genic and nongenic regions. *Proceedings of the National Academy of Sciences of the United States of America* **106**, 17077–17082 (2009).
- Priest, F. G., Kaji, D. A., Rosato, Y. B. & Canhos, V. P. Characterization of *Bacillus*-*Thuringiensis* and Related Bacteria by Ribosomal-Rna Gene Restriction-Fragment-Length-Polymorphisms. *Microbiology-Uk* **140**, 1015–1022 (1994).
- Rasko, D. A., Altherr, M. R., Han, C. S. & Ravel, J. Genomics of the *Bacillus cereus* group of organisms. *Fems Microbiology Reviews* **29**, 303–329 (2005).
- Bartoszewicz, M., Bideshi, D. K., Kraszewska, A., Modzelewska, E. & Swiecicka, I. Natural isolates of *Bacillus thuringiensis* display genetic and psychrotrophic properties characteristic of *Bacillus weihenstephanensis*. *Journal of Applied Microbiology* **106**, 1967–1975 (2009).
- Soufiane, B. & Cote, J. C. *Bacillus thuringiensis* Serovars bolivia, vazensis and navarrensensis Meet the Description of *Bacillus weihenstephanensis*. *Current Microbiology* **60**, 343–349 (2010).
- Ash, C., Farrow, J. A., Dorsch, M., Stackebrandt, E. & Collins, M. D. Comparative analysis of *Bacillus anthracis*, *Bacillus cereus*, and related species on the basis of reverse transcriptase sequencing of 16S rRNA. *Int J Syst Bacteriol* **41**, 343–6 (1991).
- Keim, P. *et al.* Molecular evolution and diversity in *Bacillus anthracis* as detected by amplified fragment length polymorphism markers. *Journal of Bacteriology* **179**, 818–824 (1997).
- Van Ert, M. N. *et al.* Global Genetic Population Structure of *Bacillus anthracis*. *Plos One* **2** (2007).
- Read, T. D. *et al.* The genome sequence of *Bacillus anthracis* Ames and comparison to closely related bacteria. *Nature* **423**, 81–6 (2003).
- Treangen, T. J., Ondov, B. D., Koren, S. & Phillippy, A. M. The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. *Genome Biol* **15**, 524 (2014).
- Wang, A., Pattemore, J., Ash, G., Williams, A. & Hane, J. Draft genome sequence of *Bacillus thuringiensis* strain DAR 81934, which exhibits molluscicidal activity. *Genome Announc* **1**, e0017512 (2013).

24. Felsenstein, J. PHYLIP - Phylogeny Inference Package (Version 3.2). *Cladistics* **5**, 164–166 (1989).
25. Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* **30**, 2725–9 (2013).
26. Gardner, S. N. & Hall, B. G. When whole-genome alignments just won't work: kSNP v2 software for alignment-free SNP discovery and phylogenetics of hundreds of microbial genomes. *PLoS One* **8**, e81760 (2013).
27. Thompson, J. D., Higgins, D. G. & Gibson, T. J. Clustal-W—Improving the Sensitivity of Progressive Multiple Sequence Alignment through Sequence Weighting, Position-Specific Gap Penalties and Weight Matrix Choice. *Nucleic Acids Research* **22**, 4673–4680 (1994).
28. Saitou, N. & Nei, M. The Neighbor-Joining Method—a New Method for Reconstructing Phylogenetic Trees. *Molecular Biology and Evolution* **4**, 406–425 (1987).
29. Ravel, J. *et al.* The Complete Genome Sequence of *Bacillus anthracis* Ames “Ancestor”. *Journal of Bacteriology* **191**, 445–446 (2009).
30. Chun, J. H. *et al.* Complete Genome Sequence of *Bacillus anthracis* H9401, an Isolate from a Korean Patient with Anthrax. *Journal of Bacteriology* **194**, 4116–4117 (2012).
31. Aringogren, J., Finn, M., Bengtsson, B. & Segerman, B. Microevolution during an Anthrax Outbreak Leading to Clonal Heterogeneity and Penicillin Resistance. *Plos One* **9**(2), e89112, doi: 10.1371/journal.pone.0089112 (2014).
32. Rasko, D. A. *et al.* The genome sequence of *Bacillus cereus* ATCC 10987 reveals metabolic adaptations and a large plasmid related to *Bacillus anthracis* pXO1. *Nucleic Acids Research* **32**, 977–988 (2004).
33. Xiong, Z. H. *et al.* Complete Genome Sequence of the Extremophilic *Bacillus cereus* Strain Q1 with Industrial Applications. *Journal of Bacteriology* **191**, 1120–1121 (2009).
34. Kobayashi, T. *et al.* Purification and Properties of an Alkaline Protease from Alkalophilic *Bacillus* Sp Ksm-K16. *Applied Microbiology and Biotechnology* **43**, 473–481 (1995).
35. Takami, H. *et al.* An improved physical and genetic map of the genome of alkaliphilic *Bacillus* sp. C-125. *Extremophiles* **3**, 21–28 (1999).
36. Veith, B. *et al.* The complete genome sequence of *Bacillus licheniformis* DSM13, an organism with great industrial potential. *Journal of Molecular Microbiology and Biotechnology* **7**, 204–211 (2004).
37. Deng, Y. *et al.* Complete Genome Sequence of *Bacillus subtilis* BSn5, an Endophytic Bacterium of *Amorphophallus konjac* with Antimicrobial Activity for the Plant Pathogen *Erwinia carotovora* subsp. *carotovora*. *Journal of Bacteriology* **193**, 2070–2071 (2011).
38. Zeigler, D. R. The genome sequence of *Bacillus subtilis* subsp. *spizizenii* W23: insights into speciation within the *B. subtilis* complex and into the history of *B. subtilis* genetics. *Microbiology-Sgm* **157**, 2033–2041 (2011).
39. Barbe, V. *et al.* From a consortium sequence to a unified sequence: the *Bacillus subtilis* 168 reference genome a decade later. *Microbiology-Sgm* **155**, 1758–1775 (2009).
40. Belda, E. *et al.* An updated metabolic view of the *Bacillus subtilis* 168 genome. *Microbiology-Sgm* **159**, 757–770 (2013).
41. Kunst, F. *et al.* The complete genome sequence of the Gram-positive bacterium *Bacillus subtilis*. *Nature* **390**, 249–256 (1997).
42. He, J. *et al.* Complete Genome Sequence of *Bacillus thuringiensis* Mutant Strain BMB171. *Journal of Bacteriology* **192**, 4074–4075 (2010).
43. Sheppard, A. E., Poehlein, A., Rosenstiel, P., Liesegang, H. & Schulenburg, H. Complete Genome Sequence of *Bacillus thuringiensis* Strain 407 Cry. *Genome Announc* **1**(1), e00158–12, doi: 10.1128/genomeA.00158-12 (2013).
44. He, J. *et al.* Complete Genome Sequence of *Bacillus thuringiensis* subsp. *chinensis* Strain CT-43. *Journal of Bacteriology* **193**, 3407–3408 (2011).
45. Zhu, Y. G. *et al.* Complete Genome Sequence of *Bacillus thuringiensis* Serovar *finitimus* Strain YBT-020. *Journal of Bacteriology* **193**, 2379–2380 (2011).
46. Liu, G. *et al.* Complete genome sequence of *Bacillus thuringiensis* subsp. *kurstaki* strain HD73. *Genome Announc* **1**, e0008013 (2013).
47. Murawska, E., Fiedoruk, K., Bideshi, D. K. & Swiecicka, I. Complete genome sequence of *Bacillus thuringiensis* subsp. *thuringiensis* strain IS5056, an isolate highly toxic to *Trichoplusia ni*. *Genome Announc* **1**, e0010813 (2013).
48. Challacombe, J. F. *et al.* The complete genome sequence of *Bacillus thuringiensis* A1 Hakam. *J Bacteriol* **189**, 3680–1 (2007).
49. Lapidus, A. *et al.* Extending the *Bacillus cereus* group genomics to putative food-borne pathogens of different toxicity. *Chem Biol Interact* **171**, 236–49 (2008).

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Author Contributions

A.W. analysed the data and produced the results. A.W. and G.A. contributed equally in to writing the manuscript.

Additional Information

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