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J.Z. (jwzou21@njau.
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(siemann@rice.edu)Decomposition of *Phragmites australis*
litter retarded by invasive *Solidago*
canadensis in mixtures: an antagonistic
non-additive effectLing Zhang^{1,2}, Yaojun Zhang¹, Jianwen Zou^{1,3} & Evan Siemann^{1,2}¹College of Resources & Environmental Sciences, Nanjing Agricultural University, Nanjing 210095, China, ²Department of Ecology & Evolutionary Biology, Rice University, Houston, TX 77005, USA, ³Department of Horticulture, Cornell University, Ithaca, NY 14853, USA.

Solidago canadensis is an aggressive invader in China. *Solidago* invasion success is partially attributed to allelopathic compounds release and more benefits from AM fungi, which potentially makes the properties of *Solidago* litter different from co-occurring natives. These properties may comprehensively affect litter decomposition of co-occurring natives. We conducted a field experiment to examine litter mixing effects in a *Phragmites australis* dominated community invaded by *Solidago* in southeast China. *Solidago* had more rapid mass and N loss rate than *Phragmites* when they decomposed separately. Litter mixing decreased N loss rate in *Phragmites* litter and increased that of *Solidago*. Large decreases in *Phragmites* mass loss and smaller increases in *Solidago* mass loss caused negative non-additive effect. *Solidago* litter extracts reduced soil C decomposition and N processes, suggested an inhibitory effect of *Solidago* secondary compounds. These results are consistent with the idea that nutrient transfer and secondary compounds both affected litter mixtures decomposition.

Biological invasions alter ecosystem services across the globe. Invasive plants are frequently fast-growing, use high levels of soil nutrients¹, and produce higher quality litter (e.g. higher N content, lower C:N ratio)^{2–4}. Higher quality litter following invasions can be important in altering ecosystem C and N cycling through litter decomposition^{2,5}. Litter from invasive plants with higher N concentration and lower C:N ratio usually decomposes faster and releases more N than native species, in line with a number of cases that an N-fixing species invades communities where N fixers are absent^{2,4,6,7}. On the other hand, secondary metabolites released by decomposing litter might reduce nutrient releasing rate of co-occurring plant litter via litter mixing^{8,9}. While many studies examining the impacts of invasive species focus on the decomposition of litter from a given species decomposing in isolation, native ecosystems are generally invaded over a prolonged time period and invasions are spatially patchy so different plant litters often co-occur and decompose in mixtures in natural conditions. Therefore, inferences from the studies of individual invasive species are limited given that there may be chemical and/or biological interactions among different co-occurring species^{2,10}.

Solidago canadensis L. (Canada goldenrod, hereafter “*Solidago*”) is an invasive perennial forb introduced from North America into China as an ornamental plant in 1913 and subsequently escaped to surrounding areas¹¹ and is aggressively invading *Phragmites australis* (Cav.) Trin. ex Steud. (common reed, hereafter “*Phragmites*”) stands in southeast China¹². *Phragmites* is a perennial grass that maintains living rhizomes year round. *Phragmites* has thick aboveground stalks that create micro-environments in which little sunlight reaches the ground and few other native species can survive. Several explanations have been given for *Solidago*’s successful invasion. Consistent with Enhanced Mutualism Hypothesis^{11,13}, invasive *Solidago* plants benefited more from AM fungi than native plants, potentially leading to higher soil nutrient up-taking ability and hence higher litter quality^{13,14}. Other studies showed that release of allelopathic compounds (e.g. phenolics, flavones, saponins) by invasive *Solidago* plants exhibited inhibitory effects on native plant neighbors and soil pathogens in the introduced range, providing the evidence in support of Novel Weapons Hypothesis^{15,16}. Given that litter of invasive *Solidago* has higher nutrients concentration and allelopathic compounds, these chemical traits may have mixing effect on litter decomposition in the invasion stand of *Phragmites* communities by *Solidago*^{15–18}.

Non-additive effects of litter mixing on litter mass loss and nutrient release relative to the litters of component species decomposed in isolation have been well documented^{10,17–19}. Overall decomposition of litter mixtures may be accelerated or retarded through non-additive effects^{17–20}. The first review on litter mixture decomposition studies found that positive non-additive effects as a result of synergistic interactions among component litter types are the most common result of litter mixture studies¹⁹. A key factor driving synergistic interactions could be transfer of nutrients among component litter types. Through nutrient transfers mediated by physical or biological means (e.g. leaching, fungi) in mixtures, nutrients released from rapidly decaying, higher quality litter can facilitate decay in adjacent, more recalcitrant litters, resulting in an overall positive non-additive mixing effect. Since *Solidago* litter generally decomposes in mixtures with *Phragmites* when *Solidago* invades *Phragmites* stands, synergistic interactions between higher *Solidago* quality litter and lower *Phragmites* ones could reduce the quality differences between the two species litter types in mixtures^{17,21,22}. Thereby, we may expect that nutrient transfer from *Solidago* to *Phragmites* would cause synergistic decomposition effects.

While synergistic interactions among different species litters are relatively common in mixture, indeed, other studies have reported antagonistic non-additive effects, in which decay rates of mixed litters were slower than expected from the component litters of mixtures decomposed separately^{8–10,23,24}. One possible explanation for antagonistic effects is given by the evidence that secondary compounds produced by one component species have inhibitory effects on the decomposition of other component litters in mixtures^{8,9}. Movement of secondary compounds (e.g. phenolic compounds, tannins) among litter types can be caused simply by leaching, incurring an antagonistic interactions among litters in mixtures^{17,19}. Based on results of allelopathic chemicals release and its inhibition effect on soil organic C decomposition and N process reported in *Solidago*^{15,16,25}, we may expect that allelopathic compounds released by *Solidago* would result in an antagonistic non-additive effect on litter mixture decomposition²⁶. Therefore, it would be of interest to know the effects of mixing litter from *Solidago* with both higher litter quality and secondary compounds content with that from native species *Phragmites* with lower litter quality and secondary compounds content on litter mixture decomposition.

Non-additive effects may vary over the course of decomposition due to changes in litter physical and chemical properties^{18,19,27,28}. The literature on mixing effects and their variation over time emphasizes the need for more experiments investigating the effects of mixing litter of native and invasive species on litter mass and N dynamics. As litter decomposition proceeds in mixture over time, changes in the relative amount of component litters would alter the way litter decays in the mixed pool, especially when the invasive plant *Solidago* is potentially allelopathic to the co-occurring *Phragmites* species¹⁵. For instance, secondary chemicals released by decomposing *Mikania micrantha* in mixtures with *Pinus massoniana* litter causes a shift from a positive effect to an inhibitory effect on decomposition of mixtures after three months²⁸.

Here, we investigated litter mixing effects in a natural *Phragmites* dominated community invaded by *Solidago* in southeast China. We conducted a pair of field experiments to examine potential effects of *Solidago* invasion on soil C and N processes from litter mixing, nutrient transfer, and secondary compounds. Specifically, we proposed that: (1) *Solidago* litter with lower C:N ratio would decompose faster than *Phragmites* litter (H_1); (2) positive non-additive mixing effects on litter mixtures would initially be observed by possible transfer of N from *Solidago* litter to *Phragmites* litter (H_2); (3) secondary chemicals released by decomposing *Solidago* litter would inhibit soil C and N process and would subsequently have an antagonistic non-additive mixing effect on *Phragmites* litter decomposition in mixtures as decomposition proceeds^{28,29} (H_3).

Results

Litter C, N concentration and C:N ratio. Initial litter C and N concentration both differed between *Phragmites* and *Solidago* species. While *Phragmites* litter was higher in C (441.9 ± 5.0 g kg⁻¹ in *Solidago* vs. 537.7 ± 9.7 g kg⁻¹ in *Phragmites*, $F_{1,4} = 76.68$, $P = 0.0009$), initial N concentration of *Solidago* litter (7.9 ± 0.03 g kg⁻¹) was higher than that of *Phragmites* litter (7.3 ± 0.1 g kg⁻¹; $F_{1,4} = 45.86$, $P = 0.0025$). *Solidago* litter was characterized by a significantly lower initial C:N ratio (56.0 ± 0.5) relative to that of *Phragmites* (73.8 ± 0.6 ; $F_{1,4} = 564.15$, $P < 0.0001$). Compared with litters decaying alone, *Solidago* N concentration was significantly lower in mixtures at the second and third measurements ($P = 0.0021$; Fig. 1b) but *Phragmites* N concentration was significantly higher in mixtures than in isolation ($P = 0.0004$; Fig. 1c). Except for differences in initial N concentration, *Solidago* N concentration did not significantly differ with that of *Phragmites* in mixtures ($P = 0.67$; Fig. 1d). Whether decaying alone or in mixture, the C:N ratio of litter decreased over time for both species. Over 12 months, C:N ratios of *Solidago* and *Phragmites* litters in single species bags dropped down to 42.0 ± 1.3 and 64.5 ± 5.1 , respectively. In mixtures, C:N ratio of *Solidago* and *Phragmites* litters reached similar values over 12 months (48.3 ± 4.7 vs. 40.9 ± 1.2).

Litter mass loss. *Solidago* litter decomposed more rapidly than *Phragmites* litter when they decomposed in single species bags ($P < 0.0001$; Fig. 2a). The first order decay constant (k) of *Solidago* litter was twice that of *Phragmites* (Table 1). Litter mixing accelerated litter mass loss rate of *Solidago* ($P = 0.0003$; Fig. 2b) but decreased that of *Phragmites* ($P = 0.0003$; Fig. 2c). For mass loss, litter mixing increased $k_{Solidago}$ and decreased $k_{Phragmites}$ by 53% and 38%, respectively, when calculated by the single negative exponential model (Table 1). During the first 6 months, non-additive mixing effect on litter mass was, on average, slightly positive but not statistically significant (Table 2). Over 12 months, however, significant negative effects on litter mass loss were observed. Compared with predicted remaining mass, up to 15% more mass remained over 12 months of litter decomposition in mixtures.

Litter N release. When decomposed alone, *Solidago* litter had faster N release than *Phragmites* litter ($P = 0.0023$; Fig. 3a). K for N release of *Solidago* litter was approximately two times that of *Phragmites* litter (Table 1). While *Phragmites* litter N decreased 17% decomposing alone, it was increased by 17% in litter mixtures over 12 months ($P = 0.0004$; Fig. 1c). An initial non-significant positive effect became a slightly significant negative effect over 12 months (Table 2). Nitrogen release over 12 months was 7% lower in mixtures than predicted from decomposition in isolation.

Litter extract effects on soil C and N processes. *Solidago* litter extracts had significant effects on soil organic carbon (SOC) concentration ($F_{3,12} = 14.91$, $P = 0.0002$), N mineralization ($F_{3,8} = 41.51$, $P < 0.0001$) and nitrification ($F_{3,8} = 39.52$, $P < 0.0001$) processes. After 6 months of extract treatments, SOC concentration was significantly greater for plots receiving the medium or high concentrations of extract treatments (Fig. 4a). Meanwhile, both nitrification and N mineralization rates with high level extracts were lower than those that received control, low or mid treatments (Fig. 4bc).

Discussion

In the present study, mixing of *Solidago* and *Phragmites* litter resulted in non-additive effects on decomposition compared to litter from each species decomposing in isolation. As predicted (H_1), *Solidago* litter with higher initial N concentration and lower C:N ratio had faster mass loss and N release rate than that of native *Phragmites* when each was decomposed alone. Litter mixing decreased litter N concentration of *Solidago* but increased that of

Figure 1 | Litter N concentration dynamics (mean $\pm$ se) of *Phragmites* and *Solidago* when each was decomposed alone or in mixtures. Results of MANOVA are shown in Table 3, and P -values referring to results of follow-up contrasts are shown.

Phragmites, partially in support of the second hypothesis (H_2), suggesting possible N transfer from *Solidago* litter to *Phragmites* litter, but further studies using isotope labeling method are necessary before solid conclusions could be drawn²¹. However, possible N transfer in mixtures was not associated with significant positive effect on litter mixture decomposition. In accordance with the third hypothesis (H_3), instead, there was a negative non-additive mixing effect over 12 months of decay in mixtures, possibly due to release of inhibitory compounds by *Solidago*. For litter mass loss, there was a shift in mixing effect from a slightly but not significantly positive interaction to a significantly negative effect. A negative effect of secondary chemicals on C and N processes was supported by the litter extracts experiment, which is also consistent with the prediction of the third hypothesis (H_3).

Our study demonstrated that litter mass loss and N release from *Solidago* were faster than that of *Phragmites* when each was decomposed separately (Table 1; Figs. 2a and 3a). Invasive species are frequently characterized by higher assimilation of soil nutrients relative to native species, and consequently produce high quality litter^{30,31},

leading to faster mass loss and N releasing rate. In this study, when each was decomposed alone, up to 64% and 71% of initial N was released by *Phragmites* and *Solidago* litter over 12 months decomposition, respectively (Fig. 3a), which is in keeping with some previous studies on nutrient release rates by invasive plants^{2,3,7}. For example, a recent study on invasive shrub *Lonicera maackii* reported 96% of the initial N was released within the first year of decomposition³. Another study found that 50% of initial N was released from native plant litter during the period when the litter from the invasive shrub *Berberis thunbergii* lost 83% of initial N⁷. Increased N pool in standing biomass was reported in invasive *Solidago gigantea* relative to that of the adjacent natives¹⁴. Indeed, higher N concentration and lower C:N ratio of invasive species relative to native plant neighbors are usually associated with rapid growth of invasive plants. Thereby, decomposition of invasive plant litter may often be expected to proceed more rapidly than decomposition of native litter.

Mixing *Solidago* litter with that of *Phragmites* caused significant negative effects over 12 months of decomposition. Non-additive mixing effects were observed in most reviews of litter mixtures, with

Figure 2 | Litter mass loss dynamics (mean $\pm$ se) of *Phragmites* and *Solidago* when each was decomposed alone or in mixtures. Results of MANOVA are shown in Table 3, and P -values referring to results of follow-up contrasts are shown. Statistical significance at the level of $\alpha = 0.025$.

both positive and negative effects occurring^{17,19,28,32}. Transfer of nutrients among litters in mixtures has been demonstrated leading to positive non-additive effects³³, while the release of secondary compounds has been linked to negative effects²⁶. For instance, during a succession in Alaskan taiga, secondary chemicals released by low quality balsam poplar litter were found to inhibit decomposition of alder litter and N mineralization in alder soils²³. Since secondary compounds (e.g. polyacetynes, diterpenoids, triterpenoid saponins, and phenolics) have been reported in *Solidago*^{16,34}, chemicals from the litter of invasive *Solidago* plants were also possibly released to the

surrounding *Phragmites* litter when both were mixed together, retarding decomposition of *Phragmites* litter in this study. Unfortunately, chemical concentrations and transfer dynamics were not directly measured, which is a limitation of this study^{35–37}.

Soil processes could be involved in inhibitory effects of *Solidago* secondary compounds from leaf litter on litter decomposition (e.g. enhancement of microbial substrate, reduced soil N mineralization)^{17,19,23}. The results of litter extracts experiment showed that soil SOC decomposition and N mineralization were reduced by extract treatments, particularly in the high-level treatment (Fig. 4). In a

Table 1 | Decay constants (k , month⁻¹) of litter mass loss and N release during litter decomposition in both single- (k_{alone}) and mixed-species litterbags (k_{mixed}) over 12-month field incubation. Statistics were calculated by the single negative exponential model $M_t = M_0 e^{-kt}$ as described in the text. Four data points are used to estimate k value in fitting the model. Coefficients of determination R^2 indicate goodness of fit of the data to the single negative exponential model

Variables	Species	K_{alone}					K_{mixed}				
		Estimate	S.E.	t ratio	P value	R^2	Estimate	S.E.	t ratio	P value	R^2
Mass loss	<i>Solidago</i>	0.17	0.01	12.52	0.006	0.99	0.26	0.04	6.95	0.020	0.98
	<i>Phragmites</i>	0.08	0.01	6.98	0.020	0.97	0.05	0.01	6.10	0.026	0.96
N release	<i>Solidago</i>	0.16	0.02	8.93	0.012	0.99	0.31	0.07	4.17	0.053	0.95
	<i>Phragmites</i>	0.09	0.03	2.88	0.102	0.84	0.04	0.01	3.76	0.064	0.88

Table 2 | Predicted and observed litter mass or N remaining (% initial) (mean $\pm$ se) in mixed-species litters after 3, 6 and 12 months of decomposition. Predicted remaining mass was calculated by litter mass measured in single-species litterbags. Differences between predicted and observed values were examined by ANOVA. Statistical significance at the level of $\alpha = 0.025$

Time	Mass remaining (%)				Nitrogen remaining (%)			
	Predicted	Observed	F value	P value	Predicted	Observed	F value	P value
3 months	68.73 $\pm$ 1.33	65.65 $\pm$ 2.31	1.37	0.27	59.37 $\pm$ 2.39	53.23 $\pm$ 4.14	1.64	0.23
6 months	45.89 $\pm$ 0.68	44.60 $\pm$ 1.17	0.91	0.36	48.09 $\pm$ 1.45	47.79 $\pm$ 2.52	0.01	0.92
12 months	29.78 $\pm$ 0.71	34.38 $\pm$ 1.24	9.75	0.011	33.30 $\pm$ 0.88	38.02 $\pm$ 1.52	7.20	0.023

previous study, soil SOC and ammonium contents in areas invaded by *Solidago* were 1.2 and 3.3 times those of the native uninvaded areas, respectively²⁵. However, soil nitrate in the invaded area was only less than a half of that in the native areas²⁵, indicating *Solidago* invasion might have inhibited nitrification process, which is consistent with our findings that both soil SOC decomposition and nitrification process were inhibited in the *Solidago* invaded area than in an area dominated only by *Phragmites* where *Solidago* was not present (Fig. 4). Such inhibitory effect on soil process of secondary compounds released from *Solidago* is consistent with the negative effects in litter mixture decomposition^{8,26}.

The litter mixing effects varied over the time course of decay have been frequently observed in litter mixture studies^{18,19,27,28,38,39}. As we predicted, litter mixing effects were shifted from a slight positive interaction within the earlier stages to a significant negative non-additive effect over 12 months in this study. The way in which this non-additive effect varied over the course of decomposition is possibly determined by a combined outcome of changes in concentrations of nutrient availability and carbohydrates during the different stages of decay. A shift from a positive effect to an inhibitory effect from mixing litters was also found after three months in the decomposition of litter of invasive *Mikania micrantha* mixing with that of

Figure 3 | Litter N release dynamics (mean $\pm$ se) of *Phragmites* and *Solidago* when each was decomposed alone or in mixtures. Results of MANOVA were shown in Table 3, and P-values referring to results of follow-up contrasts were shown. Statistical significance at the level of $\alpha = 0.025$.

Figure 4 | SOC concentration (a), N mineralization (b) and nitrification (c) rates (mean $\pm$ se) as affected by *Solidago* litter extracts treatment. Treatments marked by different letters indicate significant treatment differences for contrasts conducted following a significant one-way ANOVA.

native *Pinus massoniana* plants²⁸. Despite of the limitation of this study that allelopathic chemical concentration and its transfer dynamics were not directly measured, our results suggest that simultaneously examining nutrient and secondary chemical dynamics of component litters in mixtures may give an insight into why the litter mixing effect was not accurately predicted by initial litter nutrient chemistry^{40,41}, especially in invasion cases by potentially allelopathic species.

Solidago is a widespread, problematic invader in China, Australia and Europe^{42–44}. Currently, *Solidago* is aggressively invading farmlands, roadsides and open areas in cities in southeast China¹². To

control invasive *Solidago* species or restore invaded ecosystems, it is necessary to understand the mechanisms of *Solidago* invasion and its impacts on ecosystem processes. Several hypotheses have been proposed and approved in invasive *Solidago* species in previous studies, such as Resource Availability Hypothesis, Enhanced Mutualism Hypothesis, and Novel Weapons Hypothesis^{15,25,37,45}, indicating that invasive *Solidago* has higher nutrients concentration and allelopathic compounds. The results of this study suggest that mixing effect of these properties by invasive *Solidago* on litter decomposition might have played an important role in its invasion success. Consistent with our hypothesis (H₃), allelopathic compounds released by *Solidago* might have retarded litter decomposition of co-occurring natives, and also restrained soil mineralization and nitrification processes at the high extract level, which may increase the soil C and N sink capacity retained in *Solidago* invaded ecosystems. On the other hand, faster nutrient cycling and more efficient use of nutrient through mutualistic soil-microbe interaction would give *Solidago* a competitive advantage over the co-occurring natives in the invasive range.

By examining litter mixing effects of phylogenetically paired native and invasive species, Hickman *et al.* posited the time course scheme of changes in nutrient cycling following invasion¹⁰. As proposed by Hickman *et al.*¹⁰, the invasive species impacts on nutrient cycling may be small during the earlier stage of an invasion, when the invasive plant is less abundant than native species. As invasion proceeds, increasingly abundant invasive plants will progressively remove native plants, and thereby litter mixing effects of invasive plants with native plants would generate more profound impacts on nutrient cycling in the invaded area¹⁰. In this study, N mineralization and nitrification inhibited by *Solidago* litter extracts will effectively reduce N leaching loss in wetland ecosystem, which is supported by a field investigation, showing that *Solidago* invasion significantly increased soil organic carbon and ammonium contents and soil microbial parameters (biomass, activity, and functional diversity) relative to the non-invaded area in the introduced Chinese range²⁵. The slowing down SOC decomposition and N cycling might be advantageous for *Solidago* invasion by providing C source for soil microbial communities and N nutrients for plant growth. In particular, *Solidago* litter retarded decomposition of native *Phragmites* litter, also build up SOC and N pools available for the later recruitment of invasive *Solidago*. As *Solidago* aggressively displaces native *Phragmites* plants and ultimately forms monospecific stands, *Solidago* decomposing alone will increase decomposition rates. This study has implications for element cycling changes following native *Phragmites* communities invaded by *Solidago* species, particularly as the scale *Solidago* invasion increases with time¹⁰. *Solidago* invasion would result in a shift from native *Phragmites* dominated communities with slow litter decomposition rate to invasive *Solidago* communities with fast litter decomposition rate. As invasion proceeds, more C would be decomposed and input belowground, enhancing the SOC pools and C sequestration ability in the invaded areas. However, this implication could not be generalized before more species invading multi-site with multi-year time courses are considered in future studies.

Methods

Study site. Experiments were conducted in an invaded seasonal wetland ecosystem located at the Nanjing Agricultural University experimental station (6 m elevation, 32°01'N 118°37'E), Jiangsu province, China. Mean annual temperature is 25°C, with monthly means from -10°C (January) to 32.5°C (July). Annual precipitation is 980 mm. Soils have high clay content and are classified as Gleysols or hydromorphic soils. This location was flooded by the Yangtze River until the early 20th century. Some areas are dominated by native *Phragmites australis*, but *Phragmites* stands are currently threatened by *Solidago* invasions. Other plant species co-exist with *Phragmites* and *Solidago* in this location including *Alternanthera philoxeroides*, *Eragrostis pilosa*, *Erigeron annuus*, *Coryza sumatrensis*, *Polygonum perfoliatum*, *Acalypha australis*, *Eleusine indica*, *Bidens pilosa*, *Digitaria sanguinalis*, *Abutilon theophrasti*, *Amaranthus mangostanus* and *Xanthium sibiricum*. However, the most abundant species in this experimental station was *Phragmites*, and it remains the most abundant species after *Solidago* invaded 5~ years ago. Currently, *Phragmites* coverage of this area is approximately 50% while *Solidago* takes around 30%.

Table 3 | Repeated-measures analysis of variance on the difference in litter N concentration, litter mass and N remaining of *Solidago* and *Phragmites* litter decomposed alone or in mixture. Results of follow-up contrasts were shown in Figs. 1–3. Statistical significance at the level of $\alpha = 0.025$

Factor	df	N concentration(g kg ⁻¹)		Mass remaining (%)		N remaining (%)	
		F value	P value	F value	P value	F value	P value
Between subjects							
Species	1,8	44.93	0.0002	858.94	<0.0001	341.04	<0.0001
Litter mixing	1,8	1.05	0.34	0.00	0.95	6.69	0.03
Species×Litter mixing	1,8	53.75	<0.0001	74.21	<0.0001	140.86	<0.0001
Within subjects							
Time	2,7	19.66	0.001	141.76	<0.0001	91.22	<0.0001
Time×Species	2,7	1.61	0.27	0.26	0.78	2.00	0.21
Time×Litter mixing	2,7	1.96	0.21	2.90	0.12	10.54	0.008
Time×Species×Litter mixing	2,7	1.01	0.41	1.18	0.36	1.33	0.33

Litterbag experiment. Litter decomposition rates of both focal species were examined via an *in situ* litterbag experiment¹⁹. In October 2009, we hand collected completely developed, mature leaf litter samples from different *Solidago* and *Phragmites* individuals located in the invaded area. Sun leaves and shade leaves were evenly collected by individuals, cut into a size of litter length at 3 ~ 6 cm and pooled together to produce a combined sample by species. Air dried litter samples (about 10 g) of a single species were placed in nylon litterbags (1 mm² mesh, 10 × 15 cm). Another set of bags contained mixtures of *Solidago* and *Phragmites* litter (about 5 g of each species - thoroughly mixed). Litterbags were sealed and randomly deployed in March 2010 in a single block where both species co-occur. All litterbags were approximately 2 m from each other and at least 5 m from the edge of the plant standings. Litterbags were fixed to the soil surface by anchored flags after the original litter layer of the specific location was removed. Three replicates of each treatment were retrieved after decomposition of 3, 6 or 12 months and others were analyzed (0 months). When bags were retrieved, litter samples were processed by carefully removing dirt or other contaminants, separated by species, dried, and weighed for mass, and C and N measurement by an elemental analyzer. The morphological traits difference in *Solidago* and *Phragmites* leaf litter samples decomposed less than 12 months enabled us to identify component litter by species from litter mixtures. Leaf litter that decomposed 18 months was not used for analysis as litter fragments could not be readily identified. Litter samples at 0 months were directly dried for water, C and N estimation without deployment. Litter subsamples at each time points were oven dried at 60°C to constant weight and the litter data presented were converted to that oven dry weight (Supplementary Material).

***Solidago* litter extracts experiment.** An additional experiment was performed to examine the effects of *Solidago* compounds on SOC and N processes. Within a patch dominated by *Solidago* at the center of invaded stand, twelve 0.5 × 0.5 m plots were established in March prior to the 2010 growing season and assigned to one of four concentration treatments with three replicates. The distance between plots was 2 m. Establishing plots within *Solidago* site dominated patch enabled us to exclude any possible effects of other plant litter extract on soil C and N process. When plots were established, *Solidago* litter, stems and roots were removed from plots and plots were isolated from the adjacent soils by PVC quadrat frame (0.5 × 0.5 m) buried flush with the soil surface. *Solidago* leaf litter collected during the 2009 growing season was air dried then soaked in distilled water (1 g per 10 ml) for 48 hours to extract chemical compounds^{46,47}. The solution was filtered and diluted by distilled water to 3 concentrations (g ml⁻¹): 0.002 (low), 0.050 (mid) and 0.100 (high). From April to October 2010, each plot received 800 ml of one solution biweekly (total = 9600 ml). Control plots received distilled water. Concentrations of the extract applied were decided according to an investigation of the mean annual litter input of *Solidago* in this area. Specifically, the low level extract treatment was related to one tenth of, with the mid and high level related to 2.5 and 5.5 times the mean annual litter input, respectively. After 6 months of treatment, top 20 cm soil samples were collected from each plot for SOC estimation by an elemental analyzer and N process study by lab incubation approach (Supplementary Material).

Data analysis. In the litter decomposition experiment, one-way ANOVAs were used to examine differences in initial litter C and N concentrations. For both litter types (i.e. alone or in mixture), we calculated litter decay constants (k , month⁻¹) to examine litter mixing effects on litter decomposition rates. The k values were estimated using Maximum Likelihood from a negative exponential model using four time points over 12 months as follows^{48,49}:

$$M_t = M_0 e^{-kt}$$

where M_t is litter mass (or N_t for N) remaining at time t ($t = 3, 6, 12$ months), M_0 is initial litter biomass at time $t = 0$ (or N_0 for N), k is a first-order litter decay constant (Table 1).

To examine effects of litter mixing on decomposition rates, we compared observed mass remaining in mixed-species bags to predicted mass remaining of litter mixtures

based on the observed mass of single plant litter samples. We generated predicted values using a permutation approach with the set of all combinations of the two single-species results. The predicted litter mass remaining in mixed-species bags was calculated by an equation as follows^{50,51}:

$$\text{Predicted mass remaining} = \frac{M_S}{M_S + M_P} \times R_S + \frac{M_P}{M_S + M_P} \times R_P$$

where R refers to the remaining litter mass of a species ($S = \text{Solidago}$, $P = \text{Phragmites}$) when they decomposed in isolation and M refers to the initial litter mass of a species in litter mixture.

Similar to other litter mixture analysis^{52,53}, we used one-way ANOVAs for each time period to test for differences between predicted and observed percent mass remaining and N remaining of the litter in mixtures (Table 2). As pointed out by Gartner and Cardon¹⁹, nevertheless, an ANOVA of observed vs. predicted values could violate the assumption of homogeneity of variance in ANOVA. It should also be noted that design with blocking might help in accounting for a source of variation unrelated to our treatments. Repeated-measure ANOVAs (MANOVA) were used to test the dependence of N concentration, mass and N remaining on plant species and litter mixing treatments⁵⁴, followed by a set of contrasts to examine differences as affected by species within litter type or by litter mixing within species (Table 3; Figs. 1–3). In the repeated-measure ANOVAs, the between subject effects refer to litters, and the within subject effects refer to the three census times. Since litter mass and N remaining were analyzed twice, statistical significance was at the level of $\alpha = 0.05/2$. Otherwise, $\alpha = 0.05$ was used to test the statistical significance.

One-way ANOVA followed by a set of contrasts was used to examine extracts effect on SOC and N processes in the extract experiment (Fig. 4).

No transformation was conducted if data meet the assumption of ANOVA, otherwise data was square-root transformed as necessary in this study. Statistical analyses were conducted in SAS 9.0 and JMP 9.0 (SAS Inc., Cary, NC, USA).

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Author contributions

L.Z., J.W.Z. and E.S. designed the research; L.Z. and Y.J.Z. performed the experiment; L.Z., J.W.Z. and E.S. analyzed data; L.Z., J.W.Z. and E.S. wrote the main manuscript text; L.Z., Y.J.Z., J.W.Z. and E.S. reviewed the manuscript.

Additional information

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