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Task-specific odorant receptor expression in worker antennae indicates that sensory filters regulate division of labor in ants

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Division of labor (DOL) is a characteristic trait of insect societies, where tasks are generally performed by specialized individuals. Inside workers focus on brood or nest care, while others take risks by foraging outside. Theory proposes that workers have different thresholds to perform certain tasks when confronted with task-related stimuli, leading to specialization and consequently DOL. Workers are presumed to vary in their response to task-related cues rather than in how they perceive such information. Here, we test the hypothesis that DOL instead stems from workers varying in their efficiency to detect stimuli of specific tasks. We use transcriptomics to measure mRNA expression levels in the antennae and brain of nurses and foragers of the ant *Temnothorax longispinosus*. We find seven times as many genes to be differentially expressed between behavioral phenotypes in the antennae compared to the brain. Moreover, half of all odorant receptors are differentially expressed, with an overrepresentation of the 9-exon gene family upregulated in the antennae of nurses. Nurses and foragers thus apparently differ in the perception of their olfactory environment and task-related signals. Our study supports the hypothesis that antennal sensory filters predispose workers to specialize in specific tasks.

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Division of labor (DOL) is an important organizing principle of complex biological systems that arose independently during three of the major evolutionary transitions¹. DOL was originally formulated in the context of the production process in human societies², but specialization to specific tasks is also found within cells and across many organisms¹. Examples range from bacteria, where clonal populations are divided into subpopulations focusing on different activities^{3–6} to multicellular organisms with differentiation of cells into different tissues and organs, and individuals performing specific roles in animal societies^{7,8}. To understand the evolution of complex life, it is therefore essential to investigate the mechanisms that underlie DOL.

DOL in insect societies results from individuals specializing in the performance of specific tasks. In addition to the reproductive DOL between fertile queens and functionally sterile workers, there is a behavioral DOL among workers that specialize in tasks such as brood care, foraging, nest building, and defense^{9–11}. Several factors can affect task specialization, including age^{12,13}, nutrition^{14,15}, morphology¹⁶, genotype^{17–19}, experience²⁰, and colony size^{21–23}. In leafcutter ant species, among others, behavioral specialization of morphologically distinct groups of workers contributes to the DOL²⁴. In most social insect species, younger workers tend to perform intranidal tasks, while older individuals perform risky activities such as nest defense and foraging outside the nest^{25,26}. Yet, such specialization among workers remains flexible, as foragers can revert to perform brood care when needed^{27,28}.

Several molecular mechanisms have been implicated in the regulation of DOL. Task specialization is associated with transcriptional changes in the worker brain^{29–34}. Molecular pathways such as the insulin/insulin-like signaling (IIS), vitellogenin (Vg), and juvenile hormone (JH) pathways are involved in the regulation of worker behavior^{35–40}. Functional manipulations have confirmed that the expression of key genes in the worker brain controls task specialization^{41–43}. Behavioral variation among workers is also associated with signaling of biogenic amines (e.g., dopamine, octopamine, tyramine, serotonin), which act as neurotransmitters or neuromodulators involved in the modulation of the responsiveness to task-associated stimuli^{44–47}.

Self-organization and collective behavior in insect societies are maintained via the exchange of chemical information⁴⁸. Social insects communicate primarily through glandular pheromones and complex mixtures of long-chain hydrocarbons on their cuticle. These cuticular hydrocarbons (CHC) facilitate recognition of nestmates, developmental stages, castes, sexes, and species^{49,50}. Social insects perceive chemical information via different types of sensilla in the antennae, small receptor organs embedded in the integument that are connected to sensory neurons⁵¹⁻⁵³. Decoding the identities of chemical compounds relies on odorant receptors (ORs) located within each sensillum^{54,55}. ORs are transmembrane proteins expressed in the dendrites of olfactory receptor neurons (ORN). The largely conserved OR coreceptor (Orco) of insects⁵⁶ is required for odorant recognition in the dendritic membrane: it forms an ion channel with specific OR, which determines the sensitivity and specificity of the ORN⁵⁷. Odorant molecules penetrate through the antenna cuticular pores and are transported by odorantbinding proteins to the ORN membrane, where they interact with receptors, leading to the generation of action potentials^{58,59}. ORN axons relay signals from the sensilla to the glomeruli of the antennal lobes in the insect brain, which are the first processing unit for olfactory information. Then, ORN make synaptic contact with the projection neurons and local neurons, which transfer information to the central brain $^{60-62}$.

Several lines of evidence indicate that OR genes play central roles in the regulation of social life of insects. First, social insect species typically harbor large numbers of OR genes^{63–67}. Second, some OR gene families have specifically expanded during social evolution, such as the 9-exon subfamily in ants, which appears to serve an important function in the perception of CHC^{63,66,68–71}. Third, species that evolved socially parasitic strategies resulting in reduced behavioral repertoires show a strong and convergent reduction in the number of OR genes⁷². Finally, experimentally produced mutant ants that lack the *Orco* gene (coding for the coreceptor necessary for OR to properly function) show impaired social behavior^{73,74}.

Current DOL models bring together the chemical nature of social insect communication and variation among workers in their response to chemical cues. They posit that flexible response thresholds to task-related chemical stimuli serve as regulators of worker specialization⁷⁵. Workers take on a particular task when the stimulus intensity exceeds their individual threshold for this task. Therefore, individuals with lower thresholds for a given task are more likely to perform it than those with higher thresholds⁷⁶. Individual response decisions and task performance are modulated via numerous parameters on different time scales^{7,77–79}, and despite extensive research on DOL and task allocation, these mechanisms are not fully understood.

As the name suggests, response threshold models are based on an individual's response to certain stimuli and the variation in their responsiveness over time. However, these studies do not include in their models how these cues are processed^{65,79–85}. The fact that the processing of signal information has been primarily described in the insect brain (see ref. 86) may suggest that thresholds and associated responses are set in the central nervous system, possibly regulated via molecular pathways in the brain that correlate with behavioral variation. We propose that DOL models would benefit from considering odor sensitivity as a potential upstream sensory filter that may affect task specialization. Along these lines, we hypothesize that behavioral variation among workers may also stem from their ability and/or efficiency in detecting different signals. For example, we propose that individuals that specialize in brood care do so because they are more sensitive to brood cues, rather than (or in addition to) being more likely to respond to similar levels of brood cues. To test the hypothesis that a sensory filter regulates inter-individual behavioral variation, and thus the DOL in social insects, we investigated transcriptional signals in the brain and antennae of workers that specialize in either brood care or foraging behavior in the ant Temnothorax longispinosus. We found that (i) behavioral variation was associated with more extensive transcriptomic differences in the antennae than in the brain, (ii) these differences included a large proportion of the OR gene repertoire, and (iii) individuals specializing in brood care overexpressed an OR gene family putatively involved in detecting social cues. These findings support our hypothesis that the peripheral nervous system, acting as a sensory filter, plays an important role in regulating behavioral differences between workers and thus in the DOL of social insects.

Results

Identification of *Temnothorax longispinosus* behavioral phenotypes. Task specialization in the ant *T. longispinosus* is neither genetically fixed nor rigid, but can change with age and in response to colony needs³⁹. We conducted behavioral observations of seven *T. longispinosus* laboratory colonies to identify individuals that specialize in brood care behavior (hereafter referred to as nurses), and others that specialize in foraging (hereafter referred to as foragers) (Fig. 1A; see "Methods" section for more details). The grouping of workers into the two behavioral categories was based on the frequency of their location inside or outside the colony and on their behavior, especially



Fig. 1 Variation in behavior, brain and antenna gene expression between nurses and foragers. A Boxplot showing behavioral differences between ants selected for transcriptomic analysis. Each black dots represent individual ants. For differential expression analysis, each sample contains the pooled RNA of seven ants of the respective behavioral phenotype. B Venn diagrams showing the number of DEGs that were upregulated in nurses (red) and foragers (turquoise) for both antennae and brain tissues. Principal component analysis (PCA) plots based on all expressed genes for C antenna and D brain samples with 95% confidence level presented as ellipses. The color of each sample represents task (red = nurse, turquoise = forager), and the shape the colony of origin.

whether they showed brood care or foraging behavior (Dataset S1). Individuals identified as nurses interacted with the brood in $54\% \pm 22\%$ (mean \pm sd) of the observations, and were recorded outside the nest in $1\% \pm 3\%$ of the observations. On the contrary, foragers were found outside the nest in $42\% \pm 26\%$ of the observations but interacted with the brood in only $2\% \pm 4\%$ of the observations.

Larger task-associated transcriptomic changes in the antennae than in the brain. To investigate transcriptomic variation between nurses and foragers, we used RNA-seq to generate seven nurse and seven forager brain and antenna samples, each consisting of pooled tissue from seven workers of a single colony. Of the 14,837 genes annotated in the T. longispinosus genome, we found 91% (13,494) and 92% (13,683) to be expressed (FPKM > 0) in the brain and the antennae, respectively. To investigate gene expression differences between nurses and foragers, we compared full models that included task as an explanatory variable to reduced models that did not using the likelihood ratio test (LRT) method implemented in DEseq2. The influence of colony identity was controlled for by including it as an explanatory variable in both full and reduced models. A Benjamini–Hochberg adjusted p value of 0.05 was set as a threshold to obtain genes whose variation was significantly explained by behavioral specialization in each tissue. We detected 339 differentially expressed genes (DEGs) in the brain (223 upregulated in nurses, 116 in foragers), and 2267 in the antennae (1241 upregulated in nurses, 1026 in foragers; Dataset S2 and Fig. 1B). We found an overlap of 162 DEGs between the two tissues, including 143 DEGs that showed differences in the same direction across tissues. Principal component analyses (PCA) for both brain and antenna data reveal that most samples clustered by behavioral phenotype rather than colony of origin (Fig. 1C, D).

OR gene expression differs between antennae of nurses and foragers. To investigate whether odorant perception differs between nurses and foragers, we focused our attention on the expression of OR genes in the antennae. We found that all 419 previously annotated OR genes in the genome of T. longispinosus⁷² were expressed in the antennae, and that 50% (209/419) of them were differentially expressed between nurses and foragers. Specifically, 64 OR genes were upregulated in nurses (15% of all OR genes), and 145 in foragers (35% of all ORs) (Fig. 2). Then, we studied which OR subfamilies were preferentially expressed in nurses and foragers. The 64 OR genes overexpressed in nurses belonged to three OR subfamilies, while the 145 OR genes upregulated in foragers were distributed among 19 OR subfamilies. Foragers overexpressed 27, 19, 8, and 8 OR genes from the L, V, P, and H subfamilies, respectively, while no OR genes from these subfamilies were upregulated in nurses (Dataset S3). We found that 63% (27/43) of the genes from the L subfamily and 73% (8/11) from the P subfamily were overexpressed in foragers, which represents a significant overrepresentations (L subfamily: Fisher's test, odds ratio = 3.67, p value < 0.001; P subfamily: Fisher's test, odds ratio = 5.25, p value < 0.02). For the V and H subfamilies we did not find such an overrepresentation, likely due to lower gene numbers (V subfamily: Fisher's test, odds ratio = 1.64, p value = 0.17; H subfamily: Fisher's test, odds ratio = 1.71, *p* value = 0.30). On the other hand, we found that 80% (51/64) of the OR genes overexpressed in nurses belong to the 9-exon subfamily. This results in an overrepresentation of the 9-exon subfamily in genes that were overexpressed in nurses (Fisher's test, odds ratio = 22.18,



Fig. 2 OR expression in the antennae differs between nurses and foragers. A Among the DEGs overexpressed in nurses and foragers, the most represented OR subfamilies are indicated by different colors. The red dotted line represents the significance threshold of our differential expression analysis. **B**, **C** Boxplots representing the expression of **B** Orco and OR257, the gene with the highest expression difference in terms of log₂ FoldChange in the 9-exon subfamily, and **C** OR041, OR045 and OR318, the genes with the highest expression difference in terms of log₂ FoldChange in the H, L, and P subfamilies, respectively.

p value < 0.001), with 45% (51/114) of this subfamily being overexpressed in nurses. In contrast, only 6% (8/137) of the 9-exon subfamily was overexpressed in foragers, which is less than expected by chance (Fisher's test, odds ratio = 0.09, *p* value < 0.001). We also found that nurses overexpressed *Orco* compared to foragers (FDR *p* value = 0.001).

Genes in biogenic amine pathways vary in expression between nurses and foragers. Biogenic amines have been implicated in the regulation of behavior, and they may affect sensory perception^{46,87–90}. We therefore screened our lists of brain and antennal DEGs for biogenic amine pathway genes. We detected five differentially expressed genes in the antennae that are associated with biogenic amine signaling. Genes in the serotonin (5-hydroxytryptamine, DBV15_11483), tyramine (*tyramine beta-hydroxylase*, DBV15_00422) and octopamine (octopamine receptor, LOC112465659) pathways were upregulated in the antennae of foragers, while nurses showed a higher expression of genes in dopamine (dopamine 1-like receptor 2, DBV15_07611) and octopamine (octopamine beta2 receptor, DBV15_10418) pathways (Fig. 3A-E). These five genes were also expressed in the brain, but not differentially expressed between nurses and foragers. Also, no other biogenic amine pathway genes were found to be differentially expressed in the brain.

Association between behavioral variation and molecular pathways in brain and antennae. Our analysis revealed expression differences of genes expressed in the brains of nurses and foragers involved in the regulation of task specialization in social insects. Previous studies have found that Vg genes and the associated JH and IIS/TOR pathways are important endocrine networks that play central roles in the regulation of lifespan, fertility and behavior in bees and ants^{36,91-96}. Therefore, we searched and found evidence in our RNA-seq data for task-associated expression of genes involved in the metabolism, biosynthesis, and regulation of these pathways in the brain and antennae (Table S1). These genes included conventional Vg (LOC112466671), Vg-like A per;43 (named/classified as DBV15_03138), venom carboxylesterase-6-like (DBV15_11528), and protein takeout (DBV15_08771) overexpressed in the brain of nurses. Meanwhile, allatostatin A-like (LOC112454443) and insulin-like growth factor I (IGF1) (LOC112454447) were found overexpressed in the brain of foragers. Remarkably, venom carboxylesterase-6-like and IGF1 were overexpressed in the antennae of nurses and foragers respectively. Additionally, high expression levels of the zinc-finger transcription factor *Krüppel homolog-1* (*Kr-h1*) were detected in the antennae of foragers (DBV15_06330) (Fig. 3F). The expression of this gene has been correlated with caste and behavioral differences in the brain of social insects^{93,97-100}, but little is known about its function and gene targets in the antennae.

We performed a GO enrichment analysis to gain a deeper understanding of the biological processes represented in the lists of DEGs. This analysis detected many enrichments based on single one or few genes, and we mention below a few interesting processes with the number of genes driving the enrichment in parentheses (see Table S2 for complete list). Genes that were upregulated in the brain of nurses were enriched for biological processes such as *translation* (19 genes), *cellular iron ion*



Fig. 3 Expression patterns of candidate DEGs of nurse and forager antennae known to regulate division of labor in social insects^{44-47,93,97-100}. Boxplots showing the expression of **A**-**E** some genes from different biogenic amines pathways, while **F** one behavioral candidate gene (Table S1). Gene identity is shown in parentheses.

homeostasis (2 genes), and catabolic process (6 genes); while translation (22 genes), endocytosis (6 genes), glucose metabolic process (4 genes), and regulation of cell cycle (3 genes) were enriched in the antennae. In the list of genes that were overexpressed in the brain of foragers, we found that enriched processes included peptide metabolic process (4 genes), methionyltRNA aminoacylation (1 gene), positive regulation of type I interferon production (1 gene), and cell redox homeostasis (1 gene); while phosphatidylinositol phosphorylation (4 genes), inositol phosphate dephosphorylation (3 genes), carbohydrate metabolic process (17 genes), innate immune response (2 genes), and nucleotide catabolic process (3 genes) were enriched in the antennae.

Discussion

In this study, we found evidence supporting the hypothesis that variation among social insect workers in their ability to perceive different chemical signals could contribute to the regulation of task allocation, and thus to DOL in insect societies. To do so, we analyzed brain and antenna transcriptomes of nurses and foragers of the ant T. longispinosus. We report several lines of evidence that support our hypothesis. First, we found almost seven times as many genes to be differentially expressed between nurses and foragers in the antennae as in the brain, indicating that peripheral sensory organs may have an important function in the task specialization process of social insect workers. Second, we found that half of all OR genes of the T. longispinosus genomes are differentially expressed between the antennae of nurses and foragers, suggesting that behavioral specialization is associated with different sensory filters that result in specific perceptions of the chemical environment. Third, our analyses revealed that nurses and foragers upregulated distinct families of OR genes, indicating that their sensory filters may target different types of chemical cues, possibly adjusted to the tasks they perform. Finally, we detected several genes in multiple biogenic pathways to be differentially expressed between the antennae of nurses and foragers, potentially involved in fine-tuning the sensitivity of the odorant filters.

Many organisms have evolved sensory filters to focus on only a subset of all environmental cues¹⁰¹⁻¹⁰⁴. In insects, the peripheral olfactory filtering system enables individuals to detect and discriminate odors that convey ecologically relevant information used to mediate important behaviors such as courtship, locomotion and navigation to avoid predators and locate food or nesting sites¹⁰⁵. For example, the mosquito Anopheles gambiae strongly responds to odorant components of its vertebrate hosts that provide meals¹⁰⁶. Sensory filters have been selected for because they limit the amount of information perceived by the organism, and thus the energy and time required by the brain to process it¹⁰⁵. Within insect colonies, individuals typically exhibit many morphological and physiological traits associated with increased efficiency in task specialization. These can be understood as adaptations that allow individuals to become more proficient at their task due to learning, training, or the perception of valuable task-related information, allowing the colony to avoid the cost of task switching¹⁰⁷. In this context, it is interesting to find that T. longispinosus workers may have different sensory filters that would serve as a basis for their task specialization, as workers would mostly perceive chemical cues that pertain to their tasks. Our findings indicate that the sensory filter of ant workers is dynamic, and its changes may underlie their behavioral maturation.

In this study, we report a higher number of DEGs between nurses and foragers in the antennae than in the brain. The brain is a heterogeneous tissue in which different cell types perform very specific functions, which differ greatly in their gene expression¹⁰⁸⁻¹¹⁰. Therefore, using the whole brain for transcriptome analyses could make it more difficult to identify genes that are differentially expressed only in specialized parts of the brains of nurses or foragers. In comparison, the antennae might have a more uniform cell composition, which could facilitate the identification of DEGs. In contrast to this prediction, a transcriptome comparison of two different behavioral phenotypes of worker honey bees revealed that transcriptional differences are much more pronounced in the antenna than in the separately studied brain parts including the mushroom body, antennal lobe and central brain¹¹¹. Furthermore, Chandra et al.¹¹². used wholebrain RNA-seq in several ant species to detect differential expression of the gene Ilp2 between castes, a gene that was later

found to be expressed in only about 15 cells of the pars intercerebralis. This led us to conclude that it may be slightly more difficult to identify DEGs in the brain than in the antennae, but this unlikely explains the nearly seven-fold difference in the number of DEGs.

Age, genetic background, social environment, individual experiences and hormones influence behavioral differences between workers leading to DOL^{13,20,23,113–115}. In our study, we expect that age differed between individuals identified as nurses and those identified as foragers, although we did not measure it directly. However, we know from previous studies on T. longispinosus that workers live between one to three years and switch from brood carer to forager about one year after eclosion, when new generation of workers emerged and taken over the care of the brood^{43,116}. Previous experiments designed to disentangle gene expression associated with behavioral specialization, age and fertility showed that behavioral specialization is much more strongly associated with gene expression than age and fertility in T. $longispinosus^{39}$. We propose that the molecular and physiological regulators such as JH, Vg, biogenic amines, and nutritional status known to regulate task specialization could drive different OR expression patterns, which in turn would produce behavioral variation via contrasting abilities to detect different sets of odors. This hypothesis is supported by several studies (reviewed in ref. ¹¹⁷) showing that the physiological condition of an animal can influence the level of receptor expression, including mating status, oviposition, feeding, circadian rhythm, experience, and aging. Alternatively, we cannot exclude that the exposure to different odors that is associated with performing different tasks may at least in part have affected OR gene expression in the antennae. However, such an effect is unlikely to explain the largescale variation in gene expression, as there is very limited evidence that the mere exposure to odors influences the expression of the gene coding for the OR that binds this odor^{118,119}.

According to our sensory filter hypothesis, ant workers would be expected to primarily detect chemical cues that correspond to their tasks. We found that the 9-exon subfamily of OR was overrepresented in genes that were upregulated in the antennae of nurses. Interestingly, recent studies have shown a rapid expansion of the 9-exon subfamily in ants, and several lines of evidence indicate that OR genes from this subfamily mediate complex social interactions in ant colonies⁶⁷⁻⁷⁰. First, comparisons of antennal transcriptomes revealed that 9-exon OR genes are expressed more frequently in workers than in males, suggesting a role in social communication among workers^{63,70}. Second, representative OR from the 9-exon subfamily can detect CHC extracts from several castes⁷¹. According to McKenzie et al.⁷⁰, 9-exon OR genes were first expressed in solitary ancestors of aculeate wasps and facilitated CHC discrimination, likely for prey or mate recognition, with a lineage giving rise to the ancestors of ants. Moreover, OR genes of the 9-exon subfamily were convergently lost in socially parasitic ant species that lost the ability to perform brood care or foraging⁷², suggesting that they are essential for the performance of these worker tasks. Our finding of an overexpression of 9-exon OR genes in the antennae of nurses is in line with these previous studies, and suggests that many of these receptors have important functions within the nest, such as sensing chemical cues from the larvae, queen, or other workers, and/or that they are less important for sensing task-related stimuli or other signals outside the nest. Among the OR genes overexpressed in nurses was also the co-receptor Orco, which is widely expressed in olfactory sensory neurons and nearly unchanged in sequence in distant insect taxa¹²⁰⁻¹²². ORs form a unique class of heteromeric cation channels composed of two related heptahelical subunits: a divergent OR subunit that confers odor specificity, and the co-receptor Orco subunit^{123,124}. Since those functional receptors would increase the sensitivity of the workers to

odors, we propose that the overexpression of *Orco* may indicate higher olfactory sensitivity to odors in the antennae of nurses compared to foragers. This would be supported by previous studies that showed that changes in *Orco* expression can be indicative of physiological conditions and sensory receptivity^{125,126}.

The behavioral transition from nursing to foraging may be triggered by a lower efficiency in detecting brood cues via the downregulation of specific OR genes (e.g., from the 9-exon subfamily). This would result in ants moving farther away from the brood, and this change in spatial location may trigger the behavioral transition to outside tasks^{127,128}. In addition to being less efficient at detecting brood cues, the sensory filter of foragers may also become fine-tuned to detect a more diverse set of odors. Foragers overexpressed a greater number of OR gene subfamilies compared to nurses (19 and 3 for foragers and nurses, respectively), which may indicate that the olfactory system of foragers could perceive the more diverse chemical environment outside the nest. Similar to the 9-exon subfamily, the L subfamily has also been expanded in social insects^{64,66,67}, and along with the P and H subfamilies, it has been lost in socially parasitic ants^{72,129}. Interestingly, the OR genes from the L, P, V, and H subfamilies have been upregulated in foragers, and thus may have a taskspecific function, such as recognition of chemical cues related to environmental perception or recruitment cues outside the nest. OR genes belonging to subfamilies L, H and V have been shown to be highly responsive to long-chain hydrocarbons and are overexpressed in the antennae of males and workers of the ant Harpegnathos saltator¹³⁰. In addition, several ORs of the H subfamily have been proposed to act as putative floral odorant detectors in the antennae of honey bees¹³¹.

Finding task-specific variation in OR gene expression raises the question as to which molecular mechanisms regulate those changes. Variation in biogenic amines levels have been identified as one of the leading causes of behavioral plasticity and specialization of social insects to different tasks (reviewed in ref. ¹³²). Functional manipulation of biogenic amines has led to changes in behavior, dominance status and reproductive activity, as well as shifts in worker task performance¹³³⁻¹³⁶. Previous studies on insects revealed that olfaction-guided behavior is mediated by biogenic amine receptors in the antenna, and their expression is involved in fine-tuning the sensitivity of the olfactory system^{89,117}. Signal transduction of biogenic amine receptors is mediated by G protein-coupled receptors (GPCRs) located on the cell membrane, which trigger different signaling cascades that lead to increased or decreased cAMP level and Ca2+ release¹³⁷⁻¹⁴⁰. For example, modified concentration cAMP and intracellular Ca²⁺ levels due to octopamine-induced signal transduction in the moth Manduca sexta¹⁴¹ activate Orco, leading to changes in ORN sensitivity^{142,143}. Our results suggest that the biogenic amine signaling pathway may modulate the sensory filtering function of insect antennae and alter sensitivity to various signals. We found that genes encoding tyramine and its precursor, octopamine, are upregulated in forager antennae, similar to genes involved in serotonin signaling. Tyramine and dopamine (which was upregulated in nurses) have been implicated in modulating taste and olfactory receptor neurons, while serotonin may serve as a neurotransmitter and neurohormone in antennal vessels and mechanosensory organs⁸⁹. Serotonin influences foraging activity⁸⁸ and regulates food intake in many animals^{144–146}. Dopamine signaling also plays an important role in controlling the insect circadian clock and mediating clockcontrolled behavioral phenotypes such as locomotion^{147,148}. In our focal species T. longispinosus, inside workers were found to exhibit a stronger circadian rhythmicity than foragers, which may be regulated via differences in the acetylation of histone proteins¹⁴⁹.

Changes in behavior and olfactory sensitivity in insects could be related to the expression of genes involved in IIS, target of rapamycin (TOR), JH and Vg pathways, according to age, circadian rhythm, mating and feeding status¹¹⁷. For example, appetite state in D. melanogaster is signaled by insulin, which upregulates a peptide receptor on the olfactory receptor cells that innervate the DM1 glomerulus. Activation of the DM1 glomerulus is enough to drive the fly to reach for food¹⁵⁰. Recent studies have shown that pheromone release and odor sensitivity appear to be under JH control in Schistocerca gregaria and Locusta migratoria, which could lead to behavioral changes^{151–155}. Finally, an experimental downregulation of Vg-like A in T. longispinosus workers resulted in decreased brood care behavior and a lower sensitivity to broodrelated chemical cues, suggesting changes in odor perception and olfactory-driven decision making⁴³. Our results reveal that genes associated with all these pathways were differentially expressed in the brain and antennae between nurses and foragers, predicting a link between their role in task-associated behavioral changes and the regulation of odor perception. Given the central role of IIS, Vg, JH, and TOR pathways in regulating division of labor in social insects, and our finding of task-associated patterns of the antennal expression of genes from multiple biogenic amines, we hypothesize that these modulators and hormones could be involved in the regulation of the olfactory filter. How the detailed molecular mechanisms of these pathways in the brain are causally linked to the complex changes in olfactory perception in the antennae should therefore be investigated next.

Conclusion

Our transcriptomic analyses of the brain and antennae of T. longispinosus nurses and foragers provide support to our hypothesis that behavioral variation and task specialization in ant workers are regulated via differences in olfactory perception. We predict that antennal physiology acts as sensory filters that limit the type and amount of chemical information passed to the brain. This would allow workers to target relevant chemical information from the environment and discriminate signal from noise without using energetically costly processing by the central nervous system. We argue that this sensory filter is flexible and regulated through changes in physiological conditions such as age, nutrition, and hormones. Variation among workers in their efficiency to detect specific chemical cues would result in task specialization and division of labor. The information perceived by the peripheral ORs is transmitted to the primary brain center of the olfactory pathway, the glomeruli of the antennal lobes. The question now arises whether there are differences between nurses and foragers in the morphology or physiology of the antennal lobes. A limited subset of active ORs and glomeruli might be easier to process and less energy consuming. Our study opens novel avenues of research to better understand the role of sensory filters in controlling DOL in insect societies.

Methods

Sample collection and behavioral determination. A total of seven colonies of the ant *T. longispinosus* were selected with an average colony size of 110 ± 31.5 workers (mean \pm sd, Dataset S4) at the moment when the workers were sampled. The ants were collected in the forests of the Edmund Niles Huyck Preserve, Renssellearville, NY, USA (42°31′41.0″N 74°09′38.8″W), in June of 2018 with permission. Upon collection, we housed each colony in a plaster-floored nesting box (43 cm \times 28 cm \times 10 cm) divided into three chambers containing a single slide nest, in which the colony relocated. A slide nest is an artificial nesting site comprised of a small Plexiglas cavity sandwiched between two glass microscope slides. Colonies were established at the Johannes

Gutenberg University in Mainz, Germany, under a 14 h:10 h light:dark photoperiod at 18 °C to a 22 °C temperature. We provided honey and water ad libitum and fed crickets to the colony twice a week. To allow for visible behavioral division of labor between workers of the two behavioral phenotypes, we marked, observed and recaptured ants from inside and outside the nest. We defined foragers as workers that perform outsidenest tasks, including gathering and searching for food and water and exploring the environment surrounding the nest, while nurses remained inside the dark nest and cared for the ant brood. A total of 69 workers inside (from the brood pile) and 76 workers outside the nest were marked with fine colored metal wires (0.02 mm Elektrisola, Eckenhagen, Germany). To immobilize the workers, they were placed with their heads and part of the thorax in a notch of a soft sponge without prior anesthesia. Then we marked the ants with a very thin loop around the petiole. It was then checked that the wires did not interfere with the ants' movement. We performed behavioral observations every 2 h, four times a day for 5 days (total = 20 scans), in which we noted down how many times an individual performed brood care and foraging behavior and the position in the nest (Table S3). Based on these behavioral observations, the marked individuals found outside the nest, exploring the surroundings for food or water, were identified as foragers. These workers usually do not care for the brood and do not frequently reside on brood piles, as ant colonies organize themselves spatially in a way that reduces contact between foragers and brood¹⁵⁶. We identified nurses as workers that remain inside the nest in direct contact with brood and were unlikely to leave the nest. Foragers were found in $42\% \pm 26\%$ of the observations outside of the nest, whereas nurses spend only $1\% \pm 3\%$ outside. In contrast, nurses were interacting with the brood in $54\% \pm 22\%$ of the observations, whereas we found that foragers only did this only in $2\% \pm 4\%$ of the observations. We scanned the behavior of workers over 20 observations, albeit earlier studies have shown that a single observation makes it possible to group T. longispinosus and other ants reliably into nurses and foragers that differ in behavior^{116,156}, gene expression³⁹ and CHC composition⁴³. Furthermore, spatial location can alone can predict behavior in Temnothorax workers¹⁵⁷. We focused in this study on individuals highly specialized on either foraging or brood care. Workers that performed both tasks regularly were not included in this study. After all observations were completed, the marked nurses and foragers were collected, directly frozen in liquid nitrogen and stored at -80 °C until further processing for dissection and pooling according to behavioral state and colony.

RNA extraction and sequencing. For RNA extraction, we removed both antennae and stored them in a 1.5 ml Eppendorf tube containing 50 µl TRIzol (Invitrogen), cut the head off and fixed it on a slide with melted dental wax. We then made an incision around the head with a surgical scalpel and removed the head capsule with forceps to expose the intact brain. Finally, we carefully pulled the brain out of the head capsule and removed the remains of other tissues that were connected to it. The dissected brain was transferred to a 1.5 ml Eppendorf tube containing 20 µl PBS. Each dissection was completed in less than 5 min to prevent RNA degradation. We dissected brain and antennae tissues from 48 nurses and 49 foragers. We pooled the brains and antennae from seven workers from each behavioral state and colony. The only exception was the "GO" colony (NY18 E110), for which we pooled only 6 brains and 12 antennae from 6 nurses (Dataset S4) due to the loss of one sample during the dissection process. Immediately after dissection of each brain and antennae, the Eppendorf tubes were kept on dry ice while we dissected the remaining individuals. Brain and antennae tissues were homogenized with a pestle. Sample brains were transferred separately to a 1.5 ml Eppendorf tube containing 50 μ l of TRIzol. We added 50 μ l chloroform to each brain and antenna samples, gently inverted for 30 s and then centrifuged samples at 12,000 × g for 15 min at 4 °C. We collected the resulting supernatant and precipitated RNA with 25 μ l 70% ethanol. We conducted the subsequent RNA extraction with the RNeasy Mini Kit (Qiagen), following the manufacturer's instruction. The resulting 28 samples (14 brains and 14 antennae) were stored at -80 °C until library preparation.

RNA-seq libraries were prepared by Novogene Company Limited, Cambridge, UK, using the NEBNext Ultra RNA Library Prep Kit for Illumina according to the manufacturer's protocol. After amplification and purification, 28 libraries were sequenced on an Illumina NovaSeq 6000 S4 flow cell platform using a paired-end 150 bp. Approximately 43 million raw reads were generated from each library (Dataset S4).

Gene expression analyses. Raw data obtained from Novogene were checked using FastQC v.0.11.9¹⁵⁸, and Illumina adapters were removed using Trimmomatic v.0.36159. The protein-coding genes of T. longispinosus together with the manual OR annotations (GCA_004794745.1)¹⁶⁰; and the congener T. curvispinosus (GCA_003070985.1) were retrieved from the NCBI database and we used Liftoff v.1.6.1 tool¹⁶¹ to assign these annotations to the recently published T. longispinosus genome⁷². In total, 10,029 of 13,061 (~77%) annotated protein-coding genes were assigned from the original T. longispinosus assembly (genes identified as "DBV15") and 4808 were assigned from T. curvispinosus (genes identified as "LOC"), for a total of 14,837. For gene expression analysis, reads were mapped to our T. longispinosus genome assembly, and the read counts table was generated using STAR 2.7.0¹⁶² with default settings. Detailed mapping statistics for each sample is available in Dataset S4. We used the deseq2 v1.16.1 package for R to identify differentially expressed genes¹⁶³. To avoid biased results due to low read counts, we removed from the counting matrix those genes for which less than 10 of the reads mapped to at least 6 of our 14 samples (n - 1 of the smallest sample)size). Then, we conducted a differential gene expression analysis with DESeq2¹⁶⁴. We began with comparisons between nurses and foragers using the ~Colony+Task model, followed by a likelihood ratio test (LRT) approach, with colony ID as a fixed factor. Genes were considered differentially expressed if the false discovery rate (FDR), using Benjamini-Hochberg procedure, had an adjusted *p* value of ≤ 0.05 . The resulting lists of DEGs refer to genes that are overexpressed and underexpressed in foragers compared to nurses. We used the online tool Venny v.2.1 (https://bioinfogp.cnb.csic.es/ tools/venny) to generate a Venn diagram containing the DEGs associated with task and tissues. Separation of differentially expressed genes by task was visualized by performing principal component analysis (PCA) with a 95% confidence ellipse using the ggplot2 v3.4.2 package for R¹⁶⁵. For PCA, we used the transformed reads of filtered transcriptomes from all contigs using the plotPCA function provided by DESeq2. Samples "GO", "BG" and "YY" showed a divergent expression pattern. To ensure that our results were not influenced by these deviating samples, we re-run the DEseq2 analyses repeatedly removing sample after sample. This resulted show slight shifts in the number of DEGs in the antennae (2267 with all samples vs. 1960 without "GO"; 1861 without "BG"; and 1862 without "YY") and in the brain (339 with all samples vs. 227 without "GO"; 272 without "BG"; and 346 without "YY"). However, the main findings remained similar and a large number of differentially expressed ORs were always found in all analyses

(209 with all samples vs. 195 without "GO"; 209 without "BG"; and 179 without "YY"). Finally, OR genes that were upregulated in each behavioral phenotype were visualized in a volcano plot using ggplot2. All statistical tests and graphical visualizations were performed in RStudio v.1.4.1106¹⁶⁶.

Identification of behavior candidate genes and ORs. We used gene annotations based on a BlastX search of the T. longispinosus transcriptome compared to a list of different invertebrate proteomes (i.e., Acromyrmex echinatior, Apis mellifera, Camponotus floridanus, Drosophila melanogaster, Harpegnathos saltator, Odontomachus brunneus, Temnothorax curvispinosus) downloaded from the NCBI database with an E-value of 1e-5 and below. Clusters containing more than one sequence match per species were reduced to a single specimen based on the highest blast score. We constructed orthogroups across all of the above species using OrthoFinder¹⁶⁷, including amino acid sequences from the T. longispinosus proteome¹⁶⁰, and retained orthogroups containing caste DEGs (Dataset S5) to again compare potential behavioral candidate genes previously identified as involved in regulating the division of labor in social insects^{39,42,94,100,155,168-171}. GO enrichment analysis was performed with TopGO v.2.44.0 for R using a Fisher's exact test for the different gene sets compared to the whole genome with the weight01 algorithm¹⁷². Only annotated GO terms with a *p* value of ≤ 0.05 were considered significantly enriched.

OR protein sets were clustered across multiple ant species using OrthoFinder to derive orthologous groups and identify subfamilies for each OR in *T. longispinosus*. To associate orthogroups with previously identified OR subfamilies, we used OR annotation in *Atta cephalotes, Acromyrmex echinatior* from Engsontia et al.⁶⁷, and *Camponotus floridanus, Harpegnathos saltator*, and *Solenopsis invicta* from Zhou et al.^{63,66}. Missing subfamily information was labeled as "unassigned" (Dataset S3).

Statistics and reproducibility. The experiments were performed in seven replicates. Each sample contains the pooled RNA from either the antennae or brain of seven ant workers of the respective behavioral phenotype belonging to the same colony, with the colony representing the level of replicates. Bar graphs show next to median and quartiles, the individual data points. Statistical analyses were performed in RStudio v.1.4.1106, bioinformatics analyses in Bash, and scripts for both are available on Mendeley. Gene expression analyses were performed using DEseq2, and to exclude the influence of outliers, these were removed individually, and results presented without them.

Reporting summary. Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

Numerical source data found in Supplementary Data S1 was used to create Fig. 1A and Data S2 for Figs. 2B, C and 3. Raw sequencing reads generated for this study have been deposited in NCBI under BioProject PRJNA926589. Any remaining information can be obtained from the corresponding author upon request.

Code availability

Data analysis and visualization for this study was done using code written in R and Bash, which can be found on Mendeley Data (https://doi.org/10.17632/yyg46xmph6.3).

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M.A.C., R.L., and S.F. conceived the study; M.A.C. conducted the research; M.A.C., M.M., R.L., and D.V.H. contributed to the development of the data analytical protocol; M.A.C. analyzed the data; and M.A.C., R.L., M.M., P.B., and S.F. wrote the manuscript.

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