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Impact of pyrolysis process conditions on the structure of biochar obtained from apple waste

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Biochar is an eco-friendly carbon material whose properties allow it to be used as a sorbent for wastewater treatment or soil remediation. The paper presents the results of research related to the pyrolysis process of apple waste after supercritical CO₂ extraction with the simultaneous use of physical activation. The research assessed the influence of the temperature of the pyrolysis process and steam activation on the structural properties of the obtained biochar, i.e. specific surface, porous structure, and presence of functional groups. The results obtained confirmed that lower temperature pyrolysis produces biochar characterised by the presence of functional groups and ordered structure. On the other hand, high temperature pyrolysis with simultaneous steam activation determines microporosity and high values of the specific surface area. Taking into consideration pollutant sorption mechanisms (physical and chemical sorption), the obtained biochar materials can be used as sorbents in water and wastewater treatment.

Contemporary EU economic development strategies impose on the member states the obligation to implement the principles of circular economy. One of them is the production of waste-derived materials, which helps ensure closed-loop recycling. An example of a waste-derived material is biochar, which is an environmentally friendly product, whose properties allow for its wider application. Biochar is gradually replacing traditionally produced active carbon for wastewater treatment and soil remediation¹⁻⁴. Plant-based biomass used for the production of biochar should come from waste, constitute no competition for human food, and require no multistage pretreatment⁵⁻⁷. Plant-based biomass can be divided into three main groups: agriculture residues, herbaceous biomass, and forest residues. Many types of biomass (e.g. fruit or vegetable pulps from the food industry, brewery spent grains, straw husk, sugarcane bagasse, switchgrass or wood chips) can be used to produce biochar⁸⁻¹⁰.

Biochar is obtained as a result of thermal conversion of plant-based biomass-a process which can employ different thermal decomposition methods (e.g. pyrolysis, gasification, torrefaction or hydrothermal carbonization)¹¹⁻¹³, in addition to bio-oil and gas. The method most suitable for biochar production is pyrolysis, i.e. the heating of biomass in the absence of oxygen. There are three types of pyrolysis: (i) conventional/slow pyrolysis; (ii) fast pyrolysis; and (iii) ultra-fast/flash pyrolysis¹⁴. Each of them varies in terms of temperature, heating rate, and residence time¹⁴⁻¹⁶. Process parameters determine biochar's structural properties. Researchers prove that high temperature, low pressure, and high heating rate of the pyrolysis process produce biochar with higher carbon content and large specific surface area^{15,17}. Additionally, structural properties of biochar also depend on the type of the surface activation method used. Physical activation with steam, most commonly at 700-900 °C, helps expand the specific surface area of biochar and its microporous structure, as well as introduce functional groups on biochar surface^{18,19}. Compared to chemical activation, physical activation reduces the number of unit steps necessary to perform, which reduces the production costs of biochar material. At the same time, it also makes it possible to reduce the use of chemical reagents in favor of more environmentally friendly components (in this case, water vapor). Adsorption of pollutants on the surface of physically activated biochar takes place mainly in multilayers using van der Waals forces. Such binding of pollutants is an easily reversible process compared to chemisorption. This aspect significantly facilitates the regeneration of the deposit, and thus has a positive impact on the economics of using such biochar materials in industry.

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Given its specific properties, biochar can be used in different applications. Studies mainly focus on the use of biochar to remove pollutants from water or soil, i.e. cationic aromatic dyes (methylene violet, methylene blue, etc.), fertilizers (pesticides, herbicides, carbofurans, atrazine, etc.), antibiotics and medicines (ibuprofen, tetracyclines, sulfamethazine, etc.), polycyclic aromatic hydrocarbons (naphthalene, phenanthrene, nitrotoluene, etc.) or volatile organic compounds (benzene, furan, butanol, etc.)^{20–23}. Apart from organic pollutants, biochar can also remove inorganic compounds like non-biodegradable heavy metals. Experiments in this regard concern the removal of the following ions: Pb^{2+} , Cu^{2+} , $Cr^{3+24-26}$. Biochar is also studied for adsorption of compounds in industrial and municipal wastewaters, i.e.: NO_3^- , NH_4^+ or $H_2S^{27,28}$.

The removal of pollutants with the use of biochar can happen during physical or chemical sorption. These mechanisms require different biochar properties. In physical sorption, a developed specific surface area of carbon materials is crucial. On the other hand, in chemical sorption, biochar should be characterised by the presence of functional groups capable of binding pollutants^{24,29}. Therefore, the selection of a suitable carbon material with proper characteristics is of key importance when managing biochar as a pollutant sorbent.

The main aims of the paper are focused on the determination of the impact of pyrolysis conditions (i.e. temperature, steam activation) on structural properties of carbon material obtained from apple waste, as well as on analysis of the development of the specific surface area, porosity, surface functional groups, biochar structure.

Materials and methods

Materials

Biochar was produced from apple biomass, a residue from the supercritical CO_2 extraction of antioxidants used in the production of cosmetics. The biomass used had low moisture content (<10%).

Pyrolysis

To produce biochar, 100 g of apple waste were pyrolysed. Pyrolysis was conducted with (BA600, BA700, BA800) or without (B300, B700) steam activation to determine the impact of pyrolysis condition on biochar properties. A Czylok FCF-V12RM muffle furnace was used. The cascade temperature control programs used are presented in Table 1.

Additionally, steam activation was carried out for samples marked BA600, BA700, and BA800. According to literature data, is carried out at high temperatures of up to 900 °C for 45–60 min^{19,29}. For this reason, the production of biochar at 300 °C with steam activation was abandoned. Whereas, the physical activation performed with steam at temperatures in the range of 600–800 °C was intended to show the changes that occur in biochar when the temperature of the pyrolysis process increases with simultaneous activation. Activation started at 60 min of pyrolysis and lasted 45 min. Steam flow rate was 8.0 dm³/min. After pyrolysis, the samples were left in the furnace for 12 h to cool to room temperature. Carbon dioxide (flow rate of 5.0 dm³/min) was used as a protective gas during heating and cooling processes.

Structural properties of biochar produced

The biochar produced was characterised in terms of its structural properties, i.e. surface morphology, porosity, presence of functional groups, and graphitization degree. To this end, ground biochar samples (fraction < 2 mm) were used.

SEM/EDS microscopy

Surface morphology and elemental composition of the biochar produced was analysed using a Hitachi SU-70 electron microscope with an X-ray microanalyzer (EDS). The analyses were conducted under the following

Temperature program of pyrolysis		B300	B700	BA600	BA700	BA800	
Step 1	Temp. [°C]		200	200	200	200	200
	Time [min]	15	15	15	15	15	
	Rate [°C/min]		12	12	12	12	12
	Hold [min]		15	15	15	15	15
	Temp. [°C]		250	650	550	650	750
Step 2	Time [min]		15	15	15	15	15
	Rate [°C/min]		3	30	23	30	37
	Hold [min]	15	15	15	15	15	
Step 3	Temp. [°C]		300	700	600	700	800
	Time [min]		15	15	15	15	15
	Rate [°C/min]		3	3	3	3	3
	Hold [min]		15	15	15	15	15
Activation H ₂ O _(g)	Temp. [°C]				600	700	800
	Time of processes [min]	Start	-	-	60	60	60
		End			105	105	105

Table. 1. Temperature program of the apple waste pyrolysis process.

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conditions: magnification: $\times\,500$ and $\times\,1000;$ accelerating voltage: 15 kV; inclination angle 30°; and vacuum 10⁻⁸ Pa.

Physical adsorption

Porosity was measured based on N_2 adsorption isotherms (77 K) determined with a Quantachrome AUTOSORB IQ analyzer. Prior to analysis, the samples were degassed under vacuum (10⁻⁷ bar) at 350 °C for 12 h. The specific surface area, volume, and average size of pores, as well as the volume and surface area of the micropores were calculated from the data obtained in the physisorption process. Specific surface area was determined using the multipoint Brunauer–Emmett–Teller (BET) method. The volume and average size of pores were calculated using the quenched solid density functional theory (QSDFT) for characterisation of mico- and mesoporous carbons. The volume and surface of micropores was calculated with the t-plot method.

FTIR spectroscopy

Functional groups on the surface of biochar were identified with an FTIR 6200 Fourier spectroscope. Infrared spectra were recorded using a Pike diamond ATR detector. For spectral measurements, a TGS detector was used in the spectral range of 4000–650 cm⁻¹ and resolution of 4 cm⁻¹.

Raman spectroscopy

The crystal structure of the biochar obtained was analysed using a Jasco NRS-5100 Raman spectroscope. Raman spectra were taken at room temperature using laser excitation at 532 nm for an exposure time of 210 s. The spectra were recorded for wavenumbers ranging from 300 to 3200 cm⁻¹ and resolution of 3.62 cm^{-1} .

Results and discussion

SEM/EDS microscopy

The biochar produced was also characterised in terms of its structural properties, i.e. surface morphology, porosity, presence of functional groups, and crystal structure. Figure 1 presents the images of biochar surface morphology taken with an electron microscope at magnification \times 500 and \times 1000.

The analysis of the microscopic images shows that the biochar produced has an irregular, fibrous, and threedimensional structure. The EDS analysis presented in Table 2 indicates that in addition to the main components (i.e. 57–80% carbon and 9–30% oxygen, depending on the pyrolysis process), biochar also contained nitrogen (1–10%), sodium (approx. 2%), potassium (approx. 5%), and chlorine (approx. 2%)—these elements were probably present in the biomass pyrolysed or could constitute residues of synthetic fertilizers used in horticulture.

Different forms of precipitates (depending on the pyrolysis process) could be observed on all biochar samples analysed. In the case of samples B700, BA600, and BA700, precipitates had the form of microspheres. As for samples B300 and BA800, they had the form of particle agglomerates and needles, respectively. The EDS analysis of these structures showed that their elemental composition differed from that of biochar. A higher content of sodium (approx. 8%), chlorine (approx. 22%), and potassium (approx. 29%) was observed, which suggests that precipitates are crystalised sodium and potassium chloride salts. Pyrolysis at 800 °C melts sodium and potassium salts (KCl and NaCl melting points: 775 and 801 °C, respectively). During cooling, these salts start to crystalise, which explains the change in the structure of crystallites in the BA800 biochar sample compared to other biochar samples analysed.

Physical adsorption

Nitrogen 77 K adsorption and desorption isotherms were determined using a Quntachrome sorption analyzer to measure porosity of the biochar produced. Based on the physisorption data, the specific surface area, volume, and average size of pores, as well as the volume and surface area of the micropores were determined with the following methods: MBET, QSDFT, and t-plot. The obtained results are presented in Table 3.

To determine the isotherm types, the IUPAC classification was applied³⁰. All biochar samples but B300 were characterised by isotherm I typical of microporous materials. Such an isotherm is characterised by an intensive deposition of adsorbent at low p/p_0 pressure ratios. Biochar obtained as a result of steam activation is characterised by adsorption isotherm type I(a) typical of materials with narrow micropores (<1 nm). Isotherm obtained for sample B700 is isotherm type I(b) characteristic of materials containing wider micropores and narrow mesopores (<2.5 nm). On the other hand, isotherm obtained for sample B300 is isotherm type III characteristic of nonporous or macroporous materials. The matching of isotherm types is consistent with the results of the average pore size obtained using the QSDFT method. Steam-activated biochar samples had pores less than 1 nm wide (BA600: 0.889 nm, BA700: 0.753 nm, and BA800 0.753 nm), and B700: 1.299 nm. This confirms that the biochar samples analysed are microporous materials³¹. The widest pores (4.323 nm) were observed in biochar pyrolysed at 300 °C. Given its small specific surface area, it can be stated that this biochar is a low-porous material.

The obtained results of the specific surface area calculated with the MBET method show big differences depending on the type of pyrolysis. Both pyrolysis temperature and steam activation of biochar play an important role. Increasing pyrolysis temperature enables increasing specific surface area of biochar by progressively degrading organic matter and removing pore-blocking substances^{32,33}. Specific surface area can also be increased through physical activation of biochar materials²⁹. Activation was conducted in parallel with pyrolysis during the last 45 min of this process. Steam temperature depended on the final temperature of pyrolysis (Table 1). The higher the steam activation temperature, the larger the specific surface area of the material, which is caused by quicker removal of carbon from the surface and evaporation of volatile substances contained in the material. This has also been confirmed by other authors³⁴⁻³⁷. For example, Rajapaksha et al.³⁷ compared properties of pure and











Figure 1. (continued)

Element	B300	B700	BA600	BA700	BA800
Carbon [% _{mas.}]	57.64	71.28	61.11	83.05	55.12
Oxygen [% _{mas.}]	30.14	12.89	22.25	9.16	22.84
Nitrogen [% _{mas.}]	5.89	4.19	7.82	1.31	3.86
Sodium [% _{mas.}]	0	3.34	2.96	1.29	6.18
Chloride [% _{mas.}]	1.36	2.48	1.35	1.38	4.52
Potassium [%mas.]	4.97	5.82	4.51	3.81	7.48

 Table 2. EDS analysis for produced biochars.

Biochar sample	Isotherm type	$S_{\rm MBET} \left[m^2/g \right]$	D _m [nm]	V _t [cm ³ /g]	V _{mic} [cm ³ /g]	$S_{mic} [m^2/g]$	S _{mic} /S _{MBET} [%]
B300	III	14.4	4.323	0.015	ND	ND	ND
B700	I(b)	314.7	1.299	0.138	0.112	287.0	91.20
BA600	I(a)	408.3	0.889	0.236	0.130	323.9	79.33
BA700	I(a)	647.0	0.753	0.250	0.202	587.9	90.87
BA800	I(a)	1 120.0	0.753	0.502	0.288	911.1	81.35

Table 3. Comparison of textural parameters of pure and steam-activated biochar. S_{MBET} specific surface area; D_m average pore width; V_t total volume of pores, V_{mic} volume of micropores; S_{mic} micropore surface; ND not detected.

steam-activated biochar obtained from tea waste at 300 and 700 °C and found that at a lower steam temperature the specific surface area grew six times compared to the initial value and 76 times at higher steam temperature.

Pure biochar (i.e. not activated with steam) is characterised by a relatively small specific surface area, which gradually grows when the pyrolysis temperature is increased (14.4 m²/g for B300, and 314.7 m²/g for B700). This relationship was also observed in the case of steam-activated biochar (408 m²/g for BA600, 647 m²/g for BA700, and 1120 m^2/g for BA800 (Table 3). As specific surface area grows, the total pore volume also increases. This relationship has a very strong linear correlation, which is confirmed by the Pearson correlation coefficients of 0.98 for the correlation between S_{MBET} and V_t and 0.99 for the correlation between S_{mic} and V_{mic} . Simultaneous increase in the specific surface area and pyrolysis temperature has also been confirmed by other researchers³⁸⁻⁴¹. Jin et al. (2020) produced biochar from grape waste (from various stages of wine production) during pyrolysis conducted at 300, 500, and 700 °C (time: 2 h, heating rate: 10 °C/min). They found that different temperature and biomass composition results in the production of biochar with different BET surface. Biochar from grapes before fermentation has specific surface area between 1.03 and 4.10 m²/g, while biochar from grapes after enzymatic hydrolysis—between 82.2 and 485 m²/g; however, in both cases, specific surface area increases in parallel with the increase in pyrolysis temperature³⁸. Elnour et al. (2019) analysed the impact of pyrolysis temperature on microstructural properties of biochar obtained from date palm waste. The analysis of specific surface area showed that biochar produced at 700 °C has $S_{BET} = 249.130 \text{ m}^2/\text{g}$, i.e. 122 times higher than in the case of biochar obtained at 300 °C ($S_{BET} = 2.04 \text{ m}^2/\text{g})^{42}$. Han et al., who produced biochar from switchgrass, hardwood, and softwood, came to similar conclusions. By increasing the temperature and introducing steam activation to the pyrolysis process, they produced biochar with a specific surface area greater by an average of 3.5 times⁴³. Bouchelta et al.⁴⁴ analysed the impact of temperature and steam activation on textural properties of biochar from date waste and found that introducing steam as an activator at peak temperatures increases microporosity of the material, which is probably due to the elimination of volatile substances produced during pyrolysis, as a result of which centres active on the biochar surface open and new pores are formed in the structure.

Based on the percentage relationship between the microporous area and specific surface area (S_{mic}/S_{MRFT}), it was also observed that the main factor affecting biochar microporosity is the temperature of the pyrolysis process (Table 3). Although B300 can be considered a nonporous material, when the pyrolysis temperature was increased, the biochar's specific surface area became dominated by micropores (91%). In the case of activated biochar, the authors observed that there were more micropores in the specific surface area of the BA700 biochar compared to BA600 (90% and 70%, respectively). Further increase in the pyrolysis temperature to 800 °C decreased the micropore area to 81%. This could be due to the breakdown of micropores and, thus, an increase in the number of mesopores at high physical activation temperatures^{45,46}. This is also confirmed by the H4 hysteresis loop observed for biochar BA800 (Fig. 2). This isotherm type indicates the micro-mesoporous nature of the sample. Such hysteresis is characterised by a rapid descend of the desorption curve at the p/p_0 value characteristic of the adsorption isotherm of 77 K nitrogen of approx. 0.4–0.5. This has been confirmed by other authors as well. Zhou et al. (2018) analysed the impact of activation time and steam activation temperature on the porosity and specific surface area of tea-based biochar. They found that the creation of mesopores at high activation temperatures (700–900 °C) was caused by the degradation of the micropore walls due to too rapid reactions between steam and carbon, involving only the outer surface of the biomass. Similar relationship was observed for the correlation between the steam activation time and temperature—the longer the activation time and the higher the activation temperature, the worse porosity of biochar⁴⁷.



Figure 2. N_2 adsorption isotherms at 77 K for pure (B300, B700) and steam-activated (BA600, BA700, and BA800) biochar.



Figure 3. FTIR spectra of: (**a**) pure biochar (B300 and B700); (**b**) steam-activated biochar (BA600, BA700, and BA800).

Comparing the $S_{\rm mic}/S_{\rm MBET}$ relationship of B700 (91.2%) and BA700 (90.9%) samples, it can be concluded that the temperature of the pyrolysis process, regardless of the activation used, is the main factor determining the number of micropores in the biochar structure.

FTIR spectroscopy

The authors analysed the presence of functional groups on biochar surface to determine the impact of pyrolysis on biochar characteristics. The obtained FTIR spectra are presented in Fig. 3.

The analysis of the spectra shows that bands are present only in the case of the B300 biochar, which suggests the presence of functional groups on its surface. Higher pyrolysis temperatures (>600 °C) cause the disappearance of functional groups containing hydrogen and oxygen as a result of deoxygenation and dehydration of the biomass³⁵. The B300 biochar has a band located at 3302 cm⁻¹ and corresponding to stretching vibrations of the -O-H bond⁴⁸⁻⁵⁰. At initial pyrolysis stages, at a temperature between 200 and 500 °C, initial degradation takes place, during which cellulose and hemicellulose are pyrolysed. At this stage, biochar is formed as a result of inter- and intramolecular reactions. Cellulose degradation reactions include decarboxylation, dehydrogenation, deoxygenation, and aromatization. At 300 °C, hydrogen bonds break down, resulting in a dehydration reaction, and as the temperature increases, hemicellulose transforms and surface functional groups (mainly hydroxyl and methoxyl groups) are reduced^{51,52}.

In the case of the B300 biochar, peaks indicative of the presence of valence vibrations of the C=O group at 1689 cm⁻¹ (a band probably originating from aromatic cyclocarbonyl) were also observed⁵³. The presence of the 1584 cm⁻¹ band indicates the presence of stretching vibrations of the C=C bonds in the aromatic ring⁵⁴, while the 1378 cm⁻¹ band corresponds to deformation vibrations of –O–H from alcohol or phenol derivatives⁵⁵. The stretching vibration at 1180 cm⁻¹ comes from the C–O bond. In the 783 cm⁻¹ region, the observed band comes from bending vibrations of the = C–H group from polycyclic aromatic hydrocarbons²⁵.

The analysis of the other spectra obtained indicates the absence of functional groups in all samples produced at temperatures above 600 °C (Fig. 3). Thus, it can be concluded that the presence of functional groups on biochar surface is strongly dependent on the temperature of the pyrolysis process. At higher temperatures, the number of oxygen, carboxyl or phenolic functional groups decreases as a result of their degradation. Banik et al. also showed that as the temperature increases, the number of oxygen heteroatoms from hydroxyl groups decreases due to the predominance of condensed polycyclic aromatic structures^{17,29}.



Figure 4. Raman spectra of: (**a**) pure biochar (B300 and B700); (**b**) steam-activated biochar (BA600, BA700, and BA800).

	Non- activation		Steam activation			
Sample of biochar	B300	B700	BA600	BA700	BA800	
$\frac{I_D}{I_G}$	0.06	0.50	0.25	0.55	0.82	

Table 4. Raman intensity ratios of biochar samples.

Raman spectroscopy

The biochar produced was characterised in terms of the order of the structure with Raman spectroscopy. The obtained spectra are presented in Fig. 4.

Two bands characteristic of carbons were detected in all types of the biochar produced: G-band at wavenumber 1592 cm⁻¹ and D-band at 1350 cm⁻¹. The G-band specifying the graphene structure originates from stretching vibrations of carbon bond pairs corresponding to stretching vibrations with symmetry E_{2g}^2 caused by the vibration of sp² carbon bond pairs (C=C). Therefore, the biochar produced was characterised by the presence of ring structures, which may be associated with the presence of ordered carbon structures. The D-band, also known as the defect band, characterises the degree of amorphousness of carbon structures. It corresponds to A_{1g} and is caused by stretching vibrations of carbon C–C bond pairs and condensed aromatic rings. The emerging peaks in the range between 3400 and 2650 cm⁻¹ are mainly attributed to aliphatic and aromatic vCH stretching vibrations resulting from combustion-induced structural modifications⁵⁶. Determination of the I_D/I_G intensity ratio is an important aspect of assessing the order of the carbon structure. The lower the I_D/I_G value, the higher the degree of carbon graphitization. The relationships calculated for the biochar produced are presented in Table 4.

The analysis of the I_D/I_G ratio found that biochar produced at the lowest temperature (i.e. 300 °C) had the most ordered structure. For this biochar, the I_D/I_G ratio was 0.06. As a result of a higher pyrolysis temperature, the biochar structure became disordered, as indicated by higher values of the I_D/I_G ratios ranging between 0.25 and 0.82. Biochar produced at 800 °C after steam activation had the most disordered structure and highest I_D/I_G ratio = 0.82. Comparing the I_D/I_G of biochar produced at 700 °C not activated with steam (I_D/I_G = 0.50) and steam activated (I_D/I_G = 0.55), it can be assumed that steam activation has no significant impact on structure order. The increase in the I_D/I_G ratio parallel to the increase in the pyrolysis temperature is attributed to the evolution of gas species, including CH₄, CO₂, CO and H₂O, formed during pyrolysis⁵⁷. The impact of higher temperature

of pyrolysis on the degree of amorphousness of biochar has also been confirmed by other authors. Chatterjee et al. also observed an increase in I_D/I_G ratios as a result of an increase in the temperature of pyrolysis of different biomass types. For biochar obtained from miscanthus pyrolysed at 500 °C, the ratio is 0.65, and at 800 °C – 0.88. In the case of corn stover, this ratio grew from 0.58 at 500 °C to 0.88 at 800 °C and for sugarcane bagasse from 0.59 to 1.01^{57} . Amdani et al. analysed beech- and pine tree-derived biochar and observed that the I_D/I_G ratio increased by 17% and 30%, respectively, when the pyrolysis temperature increased from 200 to 350 °C⁵⁸.

Conclusions

Based on the obtained results it can be concluded that pyrolysis with steam activation enables production of microporous biochar with a developed specific surface area. The authors found that (i) elevated pyrolysis temperature increases the specific surface area and microporosity of biochar and that (ii) simultaneous steam activation further develops their specific surface area. The temperature of the pyrolysis process has a major impact on the presence of functional groups and the order of the biochar structure. Low temperature (300 °C) determines the creation of functional groups and a high degree of biochar graphitization. High temperature, on the other hand, causes the disappearance of functional groups and an amorphous structure of biochar. The conclusions were formulated based on the SEM/EDS, FTIR, Raman, and physisorption analyses of the biochar structure.

Data availability

The datasets used and analysed during the current study available from the corresponding author on reasonable request. All data generated or analysed during this study are included in this published article.

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References

- 1. Ambaye, T. G. *et al.* Mechanisms and adsorption capacities of biochar for the removal of organic and inorganic pollutants from industrial wastewater. *Int. J. Environ. Sci. Technol.* **18**, 3273–3294. https://doi.org/10.1007/s13762-020-03060-w (2021).
- Parmar, S., Daki, S. & Shrivastav, A. Chapter 21- Application of biochar for soil remediation. *Biochar and its Application in Biore*mediation. https://doi.org/10.1007/978-981-16-4059-9_21 (2021).
- Acosta-Luque, M. P. et al. Remediation of Pb-contaminated soil using biochar-based slow-release P fertilizer and biomonitoring employing bioindicators. Sci. Rep. 13, 1657. https://doi.org/10.1038/s41598-022-27043-8 (2023).
- Dobrzyńska, J., Wysokińska, A. & Olchowski, R. Raspberry stalks-derived biochar, magnetic biochar and urea modified magnetic biochar - Synthesis, characterization and application for As(V) and Cr(VI) removal from river water. J. Environ. Sci. 316, 115260. https://doi.org/10.1016/j.jenvman.2022.115260 (2022).
- Turmel, M. S. et al. Crop residue management and soil health: A systems analysis. Agric. Syst. 134, 6–16. https://doi.org/10.1016/j. agsy.2014.05.009 (2015).
- Chen, N. & Pilla, S. A comprehensive review on transforming lignocellulosic materials into biocarbon and its utilization for composites applications. *Compos. Part C Open Access* 7, 100225. https://doi.org/10.1016/j.jcomc.2021.100225 (2022).
- Ding, Z. et al. Production of biochar from tropical fruit tree residues and ecofriendly applications–A review. Bioresour. Technol. 376, 128903. https://doi.org/10.1016/j.biortech.2023.128903 (2023).
- Yaashikaa, P. R., Senthil Kumar, P., Varjani, S. A. & Saravanan, A. Advances in production and application of biochar from lignocellulosic feedstocks for remediation of environmental pollutants. *Bioresour. Technol.* 292, 122030. https://doi.org/10.1016/j.biort ech.2019.122030 (2019).
- El-Gamal, E. H. *et al.* Potential bioremediation of lead and phenol by sunflower seed husk and rice straw-based biochar hybridized with bacterial consortium: A kinetic study. *Sci. Rep.* 13, 21901. https://doi.org/10.1038/s41598-023-49036-x (2023).
- 10. Anwar, T. *et al.* Synergistic effect of biochar-based compounds from vegetable wastes and gibberellic acid on wheat growth under salinity stress. *Sci. Rep.* **13**, 19024. https://doi.org/10.1038/s41598-023-46487-0 (2023).
- Al-Rumaihi, A., Shahbaz, M., Mckay, G., Mackey, H. & Al-Ansari, T. A review of pyrolysis technologies and feedstock: A blending approach for plastic and biomass towards optimum biochar yield. *Renew. Sustain. Energy Rev.* 167, 112715. https://doi.org/10. 1016/j.rser.2022.112715 (2022).
- Yadav, K. & Jagadevan, S. Chapter 3 Influence of process parameters on synthesis of biochar by pyrolysis of biomass: an alternative source of energy. *Recent Advances in Pyrolysis*. https://doi.org/10.5772/intechopen.88204 (2019).
- Meyer, S., Glaser, B. & Quicker, P. Technical, economical, and climate-related aspects of biochar production technologies: A literature review. *Environ. Sci. Technol.* 45, 9473–9483. https://doi.org/10.1021/es201792c (2011).
- Safarian, S. Performance analysis of sustainable technologies for biochar production: A comprehensive review. *Energy Rep.* 9, 4574–4593. https://doi.org/10.1016/j.egyr.2023.03.111 (2023).
- Ortiz, L. R. et al. Influence of pyrolysis temperature and bio-waste composition on biochar characteristics. Renew. Energy 155, 837–847. https://doi.org/10.1016/j.renene.2020.03.181 (2020).
- Li, S., Harris, S., Anandhi, A. & Chen, G. Predicting biochar properties and functions based on feedstock and pyrolysis temperature: A review and data syntheses. J. Clean. Prod. 215, 890–902. https://doi.org/10.1016/j.jclepro.2019.01.106 (2019).
- Banik, C., Lawrinenko, M., Bakshi, S. & Laird, D. A. Impact of pyrolysis temperature and feedstock on surface charge and functional group chemistry of biochars. J. Environ. Qual. 47, 452–461. https://doi.org/10.2134/jeq2017.11.0432 (2018).
- Feng, D. et al. Synergetic effects of biochar structure and AAEM species on reactivity of H₂O-activated biochar from cyclone air gasification. Int. J. Hydrog. Energy 42, 16045–16053. https://doi.org/10.1016/j.ijhydene.2017.05.153 (2017).
- Panwar, N. L. & Pawar, A. Influence of activation conditions on the physiochemical properties of activated biochar: A review. Biomass Conv. Bioref. 12, 925–947. https://doi.org/10.1007/s13399-020-00870-3 (2022).
- Cao, X. *et al.* Simultaneous immobilization of lead and atrazine in contaminated soils using dairy-manure biochar. *Environ. Sci. Technol.* 45, 4884–4889. https://doi.org/10.1021/es103752u (2011).
- Feng, D. et al. Adsorption characteristics of norfloxacin by biochar prepared by cassava dreg: Kinetics, isotherms and thermodynamic analysis. Bioresources 10(4), 6751–6768. https://doi.org/10.15376/biores.10.4.6751-6768 (2015).
- Gonzalez, J. M. et al. Atrazine sorption by biochar, tire chips, and steel slag as media for blind inlets: A kinetic and isotherm sorption approach. J. Water Resour. Protec. 8, 1266–1282. https://doi.org/10.4236/jwarp.2016.813097 (2016).
- Li, Y. et al. Characterization of modified biochars derived from bamboo pyrolysis and their utilization for target component (furfural) adsorption. Energy Fuels 28(8), 5119–5127. https://doi.org/10.1021/ef500725c (2014).
- Kolodynska, D. et al. Kinetic and adsorptive characterization of biochar in metal ions removal. Chem. Eng. J. 197, 295–305. https:// doi.org/10.1016/j.cej.2012.05.025 (2012).

- Kaźmierczak, B., Molenda, J. & Swat, M. The adsorption of chromium (III) ions from water solutions on biocarbons obtained from plant waste. *Environ. Technol. Innov.* 23, 101737. https://doi.org/10.1016/j.eti.2021.101737 (2021).
- 26. Wei, Y. *et al.* Enhanced lead and copper removal in wastewater by adsorption onto magnesium oxide homogeneously embedded hierarchical porous biochar. *Bioresour. Technol.* **365**, 128146. https://doi.org/10.1016/j.biortech.2022.128146 (2022).
- Zhang, K. *et al.* Linking litter production, quality and decomposition to vegetation succession following agricultural abandonment. Soil Biol. Biochem. 57, 803–813. https://doi.org/10.1016/j.soilbio.2012.08.005 (2013).
- Selenius, M. et al. Removing siloxanes and hydrogen sulfide from landfill gases with biochar and activated carbon filters. Waste Manag. 167, 31–38. https://doi.org/10.1016/j.wasman.2023.05.006 (2023).
- Sajjadi, B., Chen, W. & Egiebor, N. O. A comprehensive review on physical activation of biochar for energy and environmental applications. *Rev. Chem. Eng.* 35(6), 735–776. https://doi.org/10.1515/revce-2017-0113 (2019).
- Sing, K. S. W. *et al.* Reporting physisorption data for gas/solid system with special reference to the determination of surface area and porosity. *Pure Appl. Chem.* 57(4), 603–619. https://doi.org/10.1351/pac198557040603 (1985).
- Jaramillo, J., Álvarez, P. M. & Gómez-Serrano, V. Oxidation of activated carbon by dry and wet methods. Surface chemistry and textural modifications. *Fuel Process. Technol.* 91, 1768–1775. https://doi.org/10.1016/j.fuproc.2010.07.018 (2010).
- Savova, D. et al. Biomass conversion to carbon adsorbents and gas. Biomass Bioenergy 21, 133–142. https://doi.org/10.1016/S0961-9534(01)00027-7 (2001).
- Kim, K. H., Kim, J. Y., Cho, T. S. & Choi, J. W. Influence of pyrolysis temperature on physicochemical properties of biochar obtained from the fast pyrolysis of pitch pine (*Pinus rigida*). *Bioresour. Technol.* 118, 158–162. https://doi.org/10.1016/j.biortech.2012.04. 094 (2012).
- 34. Ahmad, M. *et al.* Speciation and phytoavailability of lead and antimony in a small arms range soil amended with mussel shell, cow bone and biochar: EXAFS spectroscopy and chemical extractions. *Chemosphere* **95**, 433–441. https://doi.org/10.1016/j.chemo sphere.2013.09.077 (2014).
- Joseph, S. D. et al. An investigation into the reactions of biochar in soil. Aust. J. Soil Res. 48, 501–515. https://doi.org/10.1071/ SR10009 (2010).
- Rajapaksha, A. U. *et al.* Enhanced sulfamethazine removal by steam-activated invasive plant-derived biochar. *J. Hazard. Mater.* 290, 43–50. https://doi.org/10.1016/j.jhazmat.2015.02.046 (2015).
- Rajapaksha, A. U. *et al.* Pyrolysis condition affected sulfamethazine sorption by tea waste biochars. *Bioresour. Technol.* 166, 303–308. https://doi.org/10.1016/j.biortech.2014.05.029 (2014).
- Jin, Q. et al. Grape pomace and its secondary waste management: Biochar production for a broad range of lead (Pb) removal from water. Environ. Res. 186, 109442. https://doi.org/10.1016/j.envres.2020.109442 (2020).
- Mohan, D. *et al.* Modeling and evaluation of chromium remediation from using low cost biochar, a green adsorbent. *J. Hazard. Mater.* 188, 319–333. https://doi.org/10.1016/j.jhazmat.2011.01.127 (2011).
- 40. Zhang, J. et al. Multiscale visualization of the structural and characteristic changes of sewage sludge biochar oriented towards potential agronomic and environmental implication. Sci. Rep. 5, 9406. https://doi.org/10.1038/srep09406 (2015).
- Hasnan, F. I. et al. Characterization of biochar derived from tapioca skin. IOP Conf. Ser. Mater. Sci. Eng. 334, 012016. https://doi. org/10.1088/1757-899X/334/1/012016 (2018).
- Elnour, A. Y. et al. Effect of pyrolysis temperature on biochar microstructural evolution, physicochemical characteristics, and its influence on biochar/polypropylene composites. Appl. Sci. 9, 1149. https://doi.org/10.3390/app9061149 (2019).
- Han, Y., Boateng, A. A., Qi, P. X., Lima, I. M. & Chang, J. Heavy metal and phenol adsorptive properties of biochars from pyrolyzed switchgrass and woody biomass in correlation with Surface properties. *J. Environ. Manag.* 118, 196–204. https://doi.org/10.1016/j. jenvman.2013.01.001 (2013).
- Bouchelta, C., Medjram, M. S., Bertrand, O. & Bellat, J. P. Preparation and characterization of activated carbon from date stones by physical activation with steam. J. Anal. Appl. Pyrolysis 82, 70–77. https://doi.org/10.1016/j.jaap.2007.12.009 (2008).
- Fu, J. et al. Effects of temperature, oxygen and steam on pore structure characteristics of coconut husk activated carbon powders prepared by one-step rapid pyrolysis activation process. *Bioresour. Technol.* **310**, 123413. https://doi.org/10.1016/j.biortech.2020. 123413 (2020).
- Fan, M., Marshal, L. W., Daugaard, D. & Brown, R. C. Steam activation of chars produced from oat hulls and corn stover. *Bioresour. Technol.* 93, 103–107. https://doi.org/10.1016/j.biortech.2003.08.016 (2004).
- Zhou, J., Luo, A. & Zhao, Y. Preparation and characterisation of activated carbon from waste tea by physical activation using steam. J. Air Waste Manag. Assoc. 68(12), 1269–1277. https://doi.org/10.1080/10962247.2018.1460282 (2018).
- Wystalska, K. & Kwarciak-Kozłowska, A. The effect of biodegradable waste pyrolysis temperatures on selected biochar properties. *Materials* 14, 1644. https://doi.org/10.3390/ma14071644 (2021).
- Varjani, S. *et al.* Treatment of wastewater from petroleum industry: Current practices and perspectives. *Environ. Sci. Poll. Res.* 27, 27172–27180. https://doi.org/10.1007/s11356-019-04725-x (2020).
- Enaime, G., Baçaoui, A., Yaacoubi, A. & Lübken, M. Biochar for wastewater treatment-conversion technologies and applications. *Appl. Sci.* 10, 3492. https://doi.org/10.3390/app10103492 (2020).
- Amalina, F. et al. Advanced techniques in the production of biochar from lignocellulosic biomass and environmental applications. Clean. Mater. 6, 100137. https://doi.org/10.1016/j.clema.2022.100137 (2022).
- 52. Wang, S., Dai, G., Yang, H. & Luo, Z. Lignocellulosic biomass pyrolysis mechanism: A state-of-the-art review. *Prog. Energy Combust. Sci.* **62**, 33–86. https://doi.org/10.1016/j.pecs.2017.05.004 (2017).
- Zhang, M. *et al.* Chitosan modification of magnetic biochar produced from *Eichhornia crassipes* for enhanced sorption of Cr(vi) from aqueous solution. *RSC Adv.* 5, 46955–46964. https://doi.org/10.1039/C5RA02388B (2015).
- Zhang, H., Chen, C., Gray, E. M. & Boyd, S. E. Effect of feedstock and pyrolysis temperature on properties of biochar governing end use efficacy. *Biomass Bioenergy* 105, 136–146. https://doi.org/10.1016/j.biombioe.2017.06.024 (2017).
- Zornoza, R. *et al.* Stability, nutrient availability and hydrophobicity of biochars derived from manure, crop residues, and municipal solid waste for their use as soil amendments. *Chemosphere* 144, 122–130. https://doi.org/10.1016/j.chemosphere.2015.08.046 (2016).
- Francioso, O. *et al.* Structural characterization of charcoal size-fractions from a burnt *Pinus pinea* forest by FT-IR, Raman and surface-enhanced Raman spectroscopies. *J. Mol. Struct.* 994, 155–162. https://doi.org/10.1016/j.molstruc.2011.03.011 (2011).
- Chatterjee, R. *et al.* Effect of pyrolysis temperature on physicochemical properties and acoustic-based amination of biochar for efficient CO₂ adsorption. *Front. Energy Res.* 8, 1–18. https://doi.org/10.3389/fenrg.2020.00085 (2020).
- Amdani, R. Z., Ossler, F. & Brackmann, C. Raman spectroscopy for characterizing porous carbon optical sensors and sensing congress. OSA Tech. Dig. https://doi.org/10.1364/LACSEA.2020.LTh4F.3 (2020).

Author contributions

W.B., M.Ł., J.M. prepared the research concept and methodology; W.B., M.Ł. performed the planned tests; W.B. wrote the original draft; W. B., M.Ł., J.M. reviewed & edited the manuscript. All authors have read and approved the final version of the manuscript.

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Competing interests

The authors declare no competing interests.

Additional information

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