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Seed dormancy, climate changes, desertification and soil use transformation threaten the Mediterranean endemic monospecific plant Petagnaea gussonei

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This study investigated the germination capacity (endogenous factor) of Petagnaea qussonei (Spreng.) Rauschert, an endemic monospecific plant considered as a relict species of the ancient Mediterranean Tertiary flora. This investigation focused also on the temporal trends of soil-use, climate and desertification (exogenous factors) across the natural range of P. gussonei. The final germination percentage showed low values between 14 and 32%, the latter obtained with GA₃ and agar at 10 °C. The rising temperatures in the study area will further increase the dormancy of P. gussonei, whose germination capacity was lower and slower at temperatures higher than 10 °C. A further limiting factor of P. qussonei is its dormancy, which seems to be morpho-physiological. Regarding climate trends, in the period 1931–2020, the average temperature increased by 0.5 °C, from 15.4 to 15.9 °C, in line with the projected climate changes throughout the twenty-first century across the Mediterranean region. The average annual rainfall showed a relatively constant value of c. 900 mm, but extreme events grew considerably in the period 1991–2020. Similarly, the land affected by desertification expanded in an alarming way, by increasing from 21.2% in 2000 to 47.3% in 2020. Soil-use changes created also a complex impacting mosaic where c. 40% are agricultural areas. The effective conservation of P. gussonei should be multilateral by relying on germplasm banks, improving landscape connectivity and vegetation cover, and promoting climate policies.

Keywords *Petagnaea gussonei*, Mediterranean endemism, Seed biology, Climate trends, Environmentally sensitive areas, CORINE Land Cover

Human activities are drastically altering the planet's biosphere, leading to an unprecedented biodiversity crisis^{1,2}. With nearly three-quarters of Earth's surface altered by human activities³, the degradation of ecosystems and natural habitats is continuing at an alarming pace, despite decades of effort to achieve all targets aimed at halting biodiversity loss⁴. The Mediterranean region, in particular, is a world hotspot of plant biodiversity that hosts 10% of all known vascular plants, amounting to c. 25,000 species of which 13,000 are endemic taxa^{5–7}. Endemic plants are of great interest for a variety of reasons that encompass inherent geographic limitation, usually threatened conservation status, and important ecological roles such as specialist pollinators and/or dispersers⁸. The Mediterranean region is thus a key area for the conservation of global plant biodiversity. However, effective measures of species conservation require a multivariate approach that considers factors acting both intrinsically (endogenously) and extrinsically (exogenously) on biodiversity. Seed dormancy, for example, is a complicated feature of plants that reflects how a species adapted to its habitat conditions^{9,10}, consequently, germination behavior is very important for the conservation of plant species^{11,12}. Climate change is accelerating biodiversity loss at an alarming rate³, and is likely to intensify over the next few decades unless substantive mitigation efforts are implemented¹³. The Mediterranean region has been especially identified as highly vulnerable to desertification¹⁴, triggered by

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deep soil-use changes¹⁵, and high sensitivity to climate changes^{13,16}. The resulting habitat fragmentation reduces landscape connectivity, and increases extinction risk¹⁷.

This study investigated some important endogenous and exogenous factors that can influence the conservation strategy of any plant species: seed dormancy, climate trends, desertification and soil-use changes. This study focused in particular on Petagnaea gussonei (Spreng.) Rauschert, a Mediterranean endemic monospecific plant (fam. Apiaceae, subfam. Saniculoideae, syn. Petagnia saniculifolia Guss., Sison gussonii Spreng., Sison gussonianum Balb. ex DC., Petagnaea saniculaefolia (Spreng.) Caruel). Specifically, this species is considered as a palaeoendemic relict of the Sicilian Tertiary flora¹⁸, narrowly distributed across the north-eastern mountains of Sicily (Italy). Petagnaea gussonei is categorized as Endangered (EN) according to the IUCN Red List Criteria^{19,20}. It is listed in Appendix I of the Bern Convention²¹, and in Annexes II and IV of the Habitats Directive 92/43/EEC²². Petagnaea gussonei has been also included among the "Top 50 Mediterranean Island Plants"23. Although P. gussonei is protected by local and national laws and conventions, measures for the conservation of this endangered plant species are only slowly being introduced. Moreover, P. gussonei has been poorly investigated with respect to its germination behavior²⁴. Knowledge of seed dormancy and optimal propagation techniques are indeed crucial for implementing conservation programs to counteract the risk of species extinction, and for successful restoration and reintroduction projects. Similarly, despite the endangered conservation status of this plant species, no study examined the climate and soil-use temporal changes occurred within the distributional area of P. gussonei. This endemic species should be a priority for conservation biology in Sicily, and effective protection strategies and management actions should ensure the persistence of P. gussonei under current and future climate and environmental changes. This study specifically aimed to shed light both on the seed dormancy of P. gussonei, and on the multi-temporal trends of climate, desertification and soil-use that affect the natural range size of P. gussonei.

Materials and methods

Study area

This study was conducted in north-eastern Sicily (Italy), within the Nebrodi Mountains (province of Messina, Fig. 1). The Nebrodi are geomorphologically complex, and occupy the central sector of the orographic chain along northern Sicily, which is the natural extension of the Italian Apennine Mountains. The Nebrodi stretch for c. 70 km, and their highest peak is Mount Soro (1847 m asl), surrounded by a vast beechwood. Distinctive features of the Nebrodi landscape are the so called *fiumare*, namely short, wide gravel-bed streams characterized by steep slopes, a large mouth, a seasonally variable flow and a torrential regime with catastrophic transport of solid material following heavy rainfall. In 1993, the Sicilian regional government created the Nebrodi Park, the largest protected area in Sicily (c. 86,000 ha). From a geological point of view, the area is part of a collisional system, developed since the Late Cretaceous, as the result of the convergence between the European and African-Adriatic plates^{25,26}. The climate is Mediterranean, with a long and dry summer. The annual mean temperature is c. 15 °C (with a minimum of -5 °C in winter and a maximum of 35 °C in summer); the annual rainfall ranges from 600 to 1300 mm. From a bioclimatic point of view²⁷, the territory investigated lies in the oceanic pluviseasonal Mediterranean belt, with meso-supramediterranean thermotype and subhumid-humid ombrotype.

Biology and ecology of Petagnaea gussonei

The plant Petagnaea gussonei (Spreng.) Rauschert is an isolated taxon that has affinities with the genera Sanicula L. and Eryngium L. However, the exact relationships of P. gussonei remain unclear²⁸⁻³¹. Known in Italy as "falsa sanicola" or "Petagna di Gussone" (Fig. 2), it is a perennial herbaceous plant (hemicryptophyte), 20-40 cm high, with peltate basal leaves having five equal lobes and small teeth at the margins; cauline leaves are tripartite and sessile. Inflorescences are in the axil of two opposite tripartite bract leaves. The small, numerous, white flowers are grouped in dichotomy cymes: a central hermaphrodite flower to which three male flowers are attached. According to³¹, each group, with one hermaphrodite flower and three male flowers, should be considered as a highly reduced umbellet, and the inflorescence as a composite umbel. Flowering occurs from early February to May, fruiting from May to July. The fruits are unilocular achenes. Although numerous studies investigated the inflorescence and fruits of *P. gussonei*^{28,32,33}, its reproductive biology remains still poorly understood, e.g., with regard to kind of fertilization, pollinators, seeds production and seed viability. Petagnaea gussonei can spread rapidly by propagating asexually through rhizome cleavages³⁴. This species is restricted to few localities in the Nebrodi Mountains, mostly in inaccessible deep valleys and gorges, within an altitude range of 300-1400 m³⁵. The species occurs on hydromorphic soil along small streams and on the edge of rivulets where water flows slowly³⁵. Petagnaea gussonei lives mainly in mixed woodland areas of deciduous trees (e.g., Quercus pubescens Willd., Quercus cerris L., rarely Fagus sylvatica L.), but it can be also found in hazel groves (Corylus aveilana L.)¹⁹.

Occurrence data of Petagnaea gussonei

Localities with the presence of *P. gussonei* were obtained from different sources^{19,36,37}, and direct exploration across the Nebrodi Mountains (Table 1). Only records with global positioning system coordinates and detailed localities were used. This study included the occurrences covering the known geographic distribution of the species. Records with uncertain locations, or localities where the species became extinct, were removed. *Petagnaea gussonei* occurs partly within the Nebrodi Park and other smaller nature reserves, partly in various sites not included in protected areas (Fig. 1). During field exploration, an occurrence site never recorded before was discovered in a remote place (site n. 20, see Fig. 1 and Table 1).

Germination tests

Little information is available about the germination patterns of *P. gussonei* in response to various regimes of temperature and light²⁴. Numerous studies investigated instead other taxa of the *Saniculoideae* subfamily such



Figure 1. Study area that shows the natural range of Petagnaea gussonei (red circle).

as Sanicula L.^{38,39}, Astrantia L.⁴⁰, Alepidea F. Delaroche⁴¹, Eryngium L.^{42,43}. The germination tests of this study aimed to identify the dormancy-breaking requirements of *P. gussonei* in terms of temperature, light, gibberellic acid (GA₃) and germination medium (agar, blotting paper). All diaspores (hereafter achenes or fruits) of *P. gussonei* were collected in July 2022 from one of the largest native populations along the "Torrente Fiumetto" site (n. 17, Fig. 1, Table 1), which is about 2.5 km long, with an altitudinal range of 400–750 m asl. Given the generally low fruit production of *P. gussonei* and its endangered conservation status, the "Torrente Fiumetto" population was selected because of its highest density and capacity of reproductive output. Specifically, in agreement with national and international protocols^{44,45}, fruits were randomly collected from at least 20 different individuals of *P. gussonei*, each of them far away from the others at a variable distance of 10–50 m, along an altitudinal gradient (400–750 m asl), in order to obtain an adequate degree of genetic diversity. The mean fruit mass was calculated by weighing 5 replicates of 20 fruits. The average weight (for 100 fruits) was measured using a balance with an accuracy of 0.001 g. The dimensions (length, width) of 10 randomly selected fruits were measured using a stereoscopic microscope. Fruit details were 100-fruit mass: 0.27 g ± 0.03; fruit length: 3.6 mm ± 0.32; fruit width: 2.5 mm ± 0.21. According to a revised version of Martin's⁴⁶ key for types of seeds⁴⁷, *P. gussonei* may have a linear underdeveloped embryo.

All fruits were preliminarily stored in paper bags at controlled conditions (20 °C and 40% of relative humidity), for approximately 3 weeks⁴⁴. Fruits that were not used in the tests were stored at -18/-20 °C for long-term ex situ conservation. Germination tests were performed in the laboratories of the Germplasm Bank of the Department of Biological, Geological and Environmental Sciences (Catania University, Italy). Fruits used in the experiments were washed with sterile distilled water, were not scarified and accompanying structures were



Figure 2. (A) Habitat of *Petagnaea gussonei*; (B) habit of *P. gussonei*; (C) inflorescences of *P. gussonei*; (D) germinated seeds of *P. gussonei*.

Site number	Site name	Municipality districts	Coordinates	Altitude (m asl)	Protection status
1	Vallone Mangalaviti	Longi	37° 58' N-14° 46' E	1230	Nebrodi Park
2	Stagno Mangalaviti	Longi	37° 57′ N-14° 46′ E	1300	Nebrodi Park
3	Torrente Contrada Contrasto	Longi	37° 57′ N-14° 46′ E	1330	Nebrodi Park
4	Affluente Vallone Linari (Contrada Cufò)	Galati Mamertino	37° 59′ N-14° 46′ E	1150	Nebrodi Park
5	Affluente Vallone S. Pietro (Contrada Cufò)	Galati Mamertino	37° 59′ N-14° 46′ E	1180	Nebrodi Park
6	Vallone Arcangelo (Contrada Pulici)	Galati Mamertino	37° 59′ N-14° 47′ E	1150	Nebrodi Park
7	Vallone Solazzo Salmieri (Contrada Acquasanta)	Tortorici	37° 56′ N-14° 51′ E	1400	Nebrodi Park
8	Vallone Runcillo (Contrada Acquasanta)	Tortorici	37° 56′ N-14° 51′ E	1400	Nebrodi Park
9	Torrente Galati (Gole del Catafurco)	Galati Mamertino	37° 59′ N-14° 48′ E	900	Nebrodi Park
10	Torrente Galati (path Madonna del Catafurco)	Galati Mamertino	37° 59′ N-14° 48′ E	890	Nebrodi Park
11	Torrente Suta (Contrada Monacelle)	Galati Mamertino	38° 00' N-14° 48' E	920	Nebrodi Park
12	Vallone Crocetta	Longi	38° 00' N-14° 45' E	760	No protected area
13	Sorgente Patirà	Tortorici	38° 01' N-14° 48' E	780	Natura 2000 site
14	Vallone Calagna	Tortorici	38° 01′ N-14° 48′ E	600	Nature Reserve
15	Vallone Calagna (reserve entrance)	Tortorici	38° 01′ N-14° 49′ E	520	Nature Reserve
16	Vallone Calagna (west periphery of Tortorici)	Tortorici	38° 01′ N-14° 49′ E	490	Nature Reserve
17	Torrente Fiumetto	Galati Mamertino	38° 01′ N-14° 47′ E	600	Natura 2000 site
18	Torrente Fiumetto (near Pizzo Bufana)	Galati Mamertino	38° 02′ N-14° 47′ E	500	Natura 2000 site
19	Torrente Potame	Tortorici	38° 02' N-14° 50' E	700	No protected area
20	Vallone Sant'Andrea	Tortorici	38° 01′ N-14° 51′ E	840	No protected area
21	Torrente Monte Cuculone	Ucria	38° 01′ N-14° 53′ E	850	No protected area
22	Vallone Ruggeri	S. Salvatore di Fitalia	38° 04' N-14° 47' E	500	No protected area
23	Vallone Vina	S. Salvatore di Fitalia	38° 04' N-14° 48' E	370	No protected area
24	Vallone S. Adriano Vecchio	S. Salvatore di Fitalia	38° 03′ N-14° 48′ E	450	No protected area
25	Vallone S. Lucia	S. Salvatore di Fitalia	38° 03′ N-14° 48′ E	400	No protected area
26	Sorgente Castell'Umberto	Castell'Umberto	38° 05′ N-14° 48′ E	600	No protected area

 Table 1. Occurrence sites of Petagnaea gussonei.

not removed. All tests were carried out in temperature and light controlled growth chambers. The light in each chamber was provided by white fluorescent tubes (Osram FL 40 SS W/37), with photosynthetic photon flux density of 40 μ mol m⁻² s⁻¹. To assess the seed dormancy breaking of *P. gussonei*, both 1% agar and blotting paper, combined with 0 (control), 250 mg L⁻¹ gibberellic acid (GA₃) solution, were used at different constant thermoperiods (10 and 15 °C), and at the photoperiod of 12/12 h (light/dark). Each treatment was carried out with four replicates of 20 achenes put in 9-cm Petri dishes. A total of four different experimental treatments were carried out: 1) blotting paper (three layers) moistened with GA₃ at 10 °C constant, 12/12 h; 2) blotting paper (three layers) moistened with GA₃ at 10 °C constant, 12/12 h; 4) agar added with GA₃ at 15 °C constant, 12/12 h.

Optimal germination at lower temperature is a characteristic of Mediterranean plants⁴⁸. Low incubation temperatures were chosen because they seem the most suitable for the germination of *P. gussonei* under natural conditions. Cold temperatures are specifically required to break the dormancy of *P. gussonei*²⁴. Low-temperature requirement for dormancy break has been frequently observed in *Saniculoideae* species³⁹, resulting in late winter or in early spring germination and emergence of seedlings. Seeds were considered to have germinated when radicle was 1-mm long⁴⁹. All experimental treatments lasted 60 days and were checked every 24 h. At the end of the incubation period, the viability of each remaining fruit was estimated by cut test. Each fruit was classified as viable/fresh, empty or died⁴⁵. Achenes with white and firm embryos were considered viable. The cut test showed that the $80.1 \pm 7.1\%$ seeds are empty (embryoless seeds are quite common in the *Apiaceae*); $15 \pm 1.7\%$ are dead; $4.9 \pm 0.53\%$ viable (not germinated). Viable seeds that did not germinate at the end of each test were not further investigated.

Germination parameters

The germination process of *P. gussonei* seeds was analysed through several parameters such as capacity, rate, time, speed and homogeneity, which were calculated with the following formulas:

(a) Final germination percentage (FGP)⁵⁰,

$$\text{FGP} = \left(\frac{\sum_{i=1}^{k} n_i}{N}\right) \times 100$$

where n_i = number of seeds germinated on the *ith* day; k = 60, number of days of experiment duration; N = 20, total number of seeds put in a Petri dish;

(b) Mean germination time $(MGT)^{51}$,

$$MGT = \frac{\sum_{i=1}^{k} n_i t_i}{\sum_{i=1}^{K} n_i}$$

where t_i = number of days between the start of the experiment and the *ith* day; (c) *Mean germination rate* (MGR), calculated as the reciprocal of MGR⁵²,

$$MGR = 1/MGT$$

- (d) *First day of germination* (FDG), expressed as the day on which the first germination event occurs⁵³;
- (e) Last day of germination (LDG), expressed as the day on which the last germination event occurs⁵³;
- (f) Median germination time $(T_{50})^{54}$,

$$T_{50} = t_i + \frac{\left(\frac{N+1}{2} - n_i\right) \times \left(t_j - t_i\right)}{n_i - n_i}$$

where N = final number of germinated seeds; n_i and n_j are the total number of seeds germinated by adjacent counts at time t_i and t_j , with $n_i < N/2 < n_i$;

(g) Coefficient of variation of the mean germination time $(CV_t)^{52}$,

$$CV_t = (s_t/MGT) \times 100$$

where the variance of the mean germination time is expressed as:

$$s_t^2 = \frac{\sum_{i=1}^k n_i (t_i - \text{MGT})^2}{\sum_{i=1}^k n_i - 1}$$

(h) Coefficient of velocity of germination (CVG)⁵⁵,

$$CVG = \frac{\sum_{i=1}^{k} n_i}{\sum_{i=1}^{k} n_i t_i} \times 100$$

(i) *Germination Rate Index* (GR)⁵⁶,

$$GRI = G_1/1 + G_2/2 + \dots + G_i/i$$

where G_1 is the final germination percentage (*FGP*₁) on day 1, G_2 is the final germination percentage (*FGP*₂) on day 2, and so on;

(j) *Germination Index* (GI)⁵⁷,

 $GI = (60 \times n_1) + (59 \times n_2) + \dots + (1 \times n_{60})$

where $n_1, n_2, ..., n_{60}$, are respectively the number of seeds germinated on the first, second,...*ith* day.

Climate trends: multi-temporal processing

This study aimed to identify the trends of annual temperature and rainfall in the distributional area of *P. gussonei* (Figs. 3 and 4). Specifically, the variations of temperature and rainfall were processed over a 90-year period, distributed in four temporal intervals: 1931–1960, 1961–1990, 1991–2020, 1931–2020. The raw data were recorded by the weather stations within the natural range of *P. gussonei*, and were acquired from the website of the Sicilian Regional Government⁵⁸. The annual mean values of temperature and rainfall, and their resulting trends, were processed and arranged in an excel spreadsheet.

Areas sensitive to desertification: multi-temporal processing

Desertification trend in the distributional area of *P. gussonei* was processed through the MEDALUS approach (Mediterranean Desertification and Land Use), funded by the European Commission⁵⁹. The desertification sensitive areas are identified on the basis of an index (Environmental Sensitive Areas to Desertification index, ESAI) that integrates bio-physical data on environmental quality (climate, soil and vegetation) with management (anthropogenic) factors⁶⁰. Three types of environmental sensitive areas (ESAs) to desertification can be distinguished in the MEDALUS approach (Fig. 5): (a) critical ESAs for areas already highly degraded, (b) fragile ESAs for areas in which any change in the delicate natural and human activity balance can lead to desertification, and (c) potential ESAs for areas threatened under significant climate change or if a particular combination of land use practices is implemented. The software ArcGIS version 10.2 was used to process ESAs.

CORINE Land Cover: multi-temporal processing

The European CORINE Land Cover (CLC) project was used to investigate the soil-use changes occurred in the natural range of *P. gussonei*. In particular, CLC III level was used to identify soil-use classes in 1990, 2000, 2006, 2012 and 2018 (Fig. 6). The CLC GIS data were collected from SINANET⁶¹. Regarding soil uses in 1958, this study obtained land cover data from the "map of Italy's soil-use"⁶². The equivalence between CNR-TCI classes and CLC classes was reported in Fig. 6. All soil-use data were turned into shapefiles, which were then processed through the software ArcGIS version 10.2.

Statistical analysis

The Wilcoxon signed ranks test was used when two related samples were compared. In case of unrelated pairs of samples, the Mann–Whitney *U*-test was carried out. The Friedman test was instead used when the comparison of multiple related samples was needed. The Kruskal–Wallis *H*-test was carried out when multiple unrelated samples were compared. To detect significant differences between pairs, contrasts were carried out with the Wilcoxon signed ranks test (related pairs) and the Mann–Whitney *U*-test (unrelated pairs). Conducting multiple contrasts may increase the Type I error rate. The initial level of risk ($\alpha = 0.05$) was therefore adapted according to the Bonferroni formula $\alpha_B = \alpha/k$, where α_B is the adapted level of risk, and k is the number of comparisons. The Kendall rank correlation coefficient was applied to check for significant monotonic correlations. The degree of significance was set at 0.05. Statistical processing was performed with the statistical package IBM SPSS Version 27.0.

Plant materials statement

The plant collection and use were in accordance with all the relevant guidelines. Collection permission was granted by the Italian Ministry of the Environment. Specimens were collected and identified by the authors of this article. Voucher specimens were stored at the Seed Bank of the Department of Biological, Geological and Environmental Sciences (Catania University, Italy).

Results and discussion

Germination behavior of Petagnaea gussonei

The final germination percentage (FGP) of the four experimental treatments showed significantly different values, which ranged from 14% (GA₃, 15 °C, no agar) to 32% (GA₃, 10 °C, with agar) (Table 2). The results suggest that the germination behavior of *P. gussonei* is not only sensitive to modest temperature oscillations (10–15 °C), but also to the presence of agar as germination medium. Indeed, at 10 °C, FGP varied from 18% (no agar) to 32% (with agar); similarly, at 15 °C, FGP was 14% without agar, and 23% with agar. In particular, germination capacity benefited from lower temperatures by shifting from 14% (15 °C) to 18% (10 °C), and the use of agar as growth medium acted as a stronger germination booster thanks to which FGP increased by 14% at 10 °C, and by 9.0% at 15 °C. No achene germinated under the control condition during the whole experimental period. The best result of FGP was 32%, which was obtained at 10 °C and with agar, thus implying that low temperatures (10 °C) and grow medium (agar) are the best experimental germination conditions.

The values of the mean germination time (MGT) showed that the variation of temperature played instead a primary role in influencing germination velocity (Table 2). Specifically, at 10 °C, MGT values were 30.3 days (no agar) and 32.5 days (with agar), whereas at 15 °C, the values were 36 days (no agar) and 34.4 (with agar). The seeds of *P. gussonei* reported thus a faster germination at lower temperatures (10 °C), in agreement with the first day



Figure 3. Temperature trends in the distributional area of *Petagnaea gussonei* over 1931–1960, 1961–1990, 1991–2020, 1931–2020.



Figure 4. Rainfall trends in the distributional area of *Petagnaea gussonei* over 1931–1960, 1961–1990, 1991–2020, 1931–2020.

of germination (FDG) that was 26 without agar, and 25 with agar. The lower temperatures seem also to prompt the germination of more seeds on the same day, as reported by the coefficient of velocity of germination (CVG), whose values at 10 °C were 3.30 (no agar) and 3.08 (with agar), whereas 2.78 (no agar) and 2.91 (with agar) at 15 °C (Table 2). Regarding the germination rate index (GRI) and the germination index (GI), the highest values were found at the lowest temperature (10 °C), specifically (without agar) GRI = 0.50%/day, and GI = 92, which respectively increased to GRI = 0.94%/day, and GI = 171, thanks to the addition of agar. Similarly, the median



Figure 5. Areas sensitive to desertification in 1960, 2000, 2020, within the natural range of *Petagnaea gussonei* (different letters show significant differences – p < 0.05 – among 1960, 2000 and 2020 within the same class).

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germination time (T₅₀) showed that the fastest results of 50% of the germinated seeds were reached at lower temperature (10 °C) and with agar, in particular T₅₀ = 28.3 days (10 °C, no agar), and T₅₀ = 31 days (10 °C, with agar). Overall, the germination parameters corroborated the role of low temperatures and agar, whose combined use makes the germination of *P. gussonei* seeds faster and higher.

Studies on dormancy and germination of species of the *Saniculoideae* subfamily are scarce⁹. Only a few attempts have been made to resolve dormancy-breaking requirements in *Saniculoideae* species. However, most studies agree with the general high dormancy in *Saniculoideae* species, including *P. gussonei*²⁴ showed that only high concentrations of GA₃ had some effect on *P. gussonei* seed germination (GA₃ = 1000 mg l⁻¹), which occurred between 4 and 5 weeks in $12 \pm 1.2\%$ of the seeds incubated in Petri dishes, in a growth chamber set at 18-23 °C. In particular²⁴, pointed out that high temperatures may result in a low germination of *P. gussonei* seeds, whereas low temperatures are an important factor in breaking seed dormancy in *P. gussonei*, as found by these authors for the pots placed in natural conditions, which were exposed to colder temperatures. In another study³⁸, investigated the factors affecting dormancy break of *Sanicula europaea* L., which is a species very close to *P. gussonei*, and the only representative of *Sanicula* occurring in western Europe. These authors found that the incubated seeds of *S. europaea*, previously pre-treated with cold stratification, may reach a germination rate of c. 70% in a faster way and at low temperatures of 5–10 °C. Other studies with similar species were in line with the above findings, as found for instance in *Sanicula canadensis* L., *Eryngium maritimum* L. and *Eryngium vuccifolium* Michx., which all require cold temperatures for dormancy break^{63–65}. In agreement with dormancy patterns reported for similar



Figure 6. Soil-use changes according to CNR-TCI in 1958, and CORINE Land Cover (CLC) III level in 1990, 2000, 2006, 2012, 2018, within the natural range of *Petagnaea gussonei*.

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species, the seed germination in *P. gussonei* seems significantly favored under low temperatures (5–10 °C). However, *P. gussonei* shows a lower germination rate when compared to other *Saniculoideae* species^{38,39,42}.

Most *Apiaceae* species generally have seeds containing an underdeveloped embryo at the moment of dispersal⁴⁶. Prior to germination, the underdeveloped embryo has to grow to a critical length inside the seed. These seeds are defined as morphologically dormant (MD). In seeds of numerous temperate species with MD, an additional physiological obstacle preventing embryo growth and/or germination occurs. Ref⁶⁶ defined this

Treatment	FGP (%)	MGT (days)	MGR	FDG (days)	LDG (days)	CVt	CVG	GRI (%/day)	GI	T ₅₀ (days)
GA ₃ , blotting paper, 10 °C	17 ± 2.4^{a}	30.3 ± 2.4^a	0.033 ± 0.004^{a}	26 ± 3.1^a	39 ± 5.2^a	14.9 ± 2.3^{a}	3.3 ± 0.41^a	0.50 ± 0.07^a	92 ± 12.3^{a}	28.3 ± 2.9^a
GA ₃ , blotting paper, 15 °C	14 ± 1.6^{b}	36 ± 4.1^{b}	$0.028 \pm 0.003^{\rm b}$	32 ± 2.7^{b}	40 ± 3.5^b	15.7 ± 1.9^{b}	2.8 ± 0.31^b	$0.28\pm0.03^{\rm b}$	50 ± 4.3^{b}	32 ± 2.6^{b}
GA ₃ , agar, 10 °C	$32 \pm 2.5^{\circ}$	$32.5 \pm 3.5^{\circ}$	$0.031 \pm 0.02^{\circ}$	$25 \pm 2.1^{\circ}$	$37 \pm 3.0^{\circ}$	$15.8 \pm 1.4^{\circ}$	$3.1\pm0.23^{\circ}$	$0.94 \pm 0.07^{\circ}$	$171 \pm 13.2^{\circ}$	32 ± 2.4^{c}
GA ₃ , agar, 15 °C	23 ± 1.9^{d}	34.4 ± 2.9^{d}	0.029 ± 0.002^{d}	29 ± 2.2^{d}	42 ± 3.3^d	10.4 ± 1.2^d	2.9 ± 0.23^d	0.73 ± 0.06^d	133 ± 10.6^d	33.5 ± 2.8^d

Table 2. Germination parameters of *Petagnaea gussonei*. 1) FGP, final germination percentage; MGT, mean germination time; MGR, mean germination rate; FDG, first day of germination; LDG, last day of germination; CV_v, coefficient of variation of germination time; CVG, coefficient of velocity of germination; GRI, germination rate index; GI, germination index; T₅₀, median germination time; 2) different letters show significantly different values, among different treatments, within the same parameter (p < 0.05).

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combined dormancy as morpho-physiological dormancy (MPD). Physiological dormancy in MPD seeds is usually broken before or during elongation of the embryo⁹. Therefore, in order to germinate, MPD seeds require a dormancy-breaking pretreatment. The significant role of GA₃ on breaking *P. gussonei* dormancy may suggest that the seeds of *P. gussonei* seem to have mainly MPD. Moreover, this study showed that *P. gussonei* seeds start germinating after 4–5 weeks, and in MPD seeds, embryo growth/radicle emergence requires a considerably longer period of time than in MD seeds⁶⁷. Similarly, various studies showed that several species of the *Sanicula* genus, closely related to *P. gussonei*, have MPD seeds^{38,63,68}. High temperatures and dormancy are undoubtedly two important factors that limit *P. gussonei* germination, which is further affected by the ongoing climate changes.

Climate trends in the distributional area of Petagnaea gussonei

The climate patterns showed fluctuating trends, especially over 30-year intervals (Figs. 3 and 4). In the period 1931–1960, the average values of temperature decreased moderately by 0.2 °C, specifically from 15.4 to 15.2 °C; in the following period of 1961–1990, the average temperature increased very significantly by 1.1 °C, from 15.1 to 16.2 °C; in turn, the last thirty years, 1991–2020, showed a decrease by 1.0 °C, from 16.2 to 15.2 °C. Overall, in the period of 1931–2020, the average temperature increased by 0.5 °C, in particular from 15.4 to 15.9 °C. Similarly, the average values of rainfall showed oscillations over periods of thirty years (Fig. 4): during 1931–1960, the average rainfall declined by 200 mm, from 1050 to 850 mm; in the time interval of 1961–1990, the decrease in rainfall from 950 to 800 mm, thus by 150 mm, was significant but slightly lower; instead, in the final period of 1931–2020, although the general trend of the average rainfall showed a relatively constant value of 920 mm, the significant decline in rainfall during the first 60 years (1931–1960 and 1961–1990) was overall compensated by the increase in rainfall in the last 30 years (1991–2020), during which the rainfall regime of the study area showed considerable extreme events.

The Mediterranean region is warming and becoming increasing arid⁶⁹. Rainfall has begun to either decrease in the long term, mainly in the dry season, or, as found in this study, not to change significantly⁷⁰. In either case, however, a steady increase in temperatures has led to greater aridity, which generates frequent extreme events such as floods, severe heat, longer droughts and wildfires⁷¹⁻⁷³. Moreover, this study showed that the rise by 0.5 °C during 1931-2020 is overall in line with future climate change projections, which expect an increase in air temperatures by 0.3-4.0 °C throughout the twenty-first century over the Mediterranean region, under an intermediate emission climate change scenario⁶⁹. Given the high evidence of large climate shifts, the Mediterranean region has been identified as a hotspot of future climate changes⁷⁴⁻⁷⁶. This ongoing climate crisis is inevitably worsening the conditions of natural ecosystems and wildlife in the Mediterranean region, which is considered as a global biodiversity hotspot^{6,77,78}. The Mediterranean biodiversity, indeed, accounts for c. 20% of world's vascular plants⁷⁹. In particular, the combination of a marked seasonality, long-term fluctuations, and a diversified geological mosaic, favored an exceptional accumulation of endemic plants⁸⁰, among which 36% are narrow endemics, i.e., they grow only in a single area or have a narrow geographic range⁸¹. Many of these endemic species persisted over geological times^{82,83}, in areas where the local (or regional) climate was buffered in comparison with the global climate changes. These areas are named refugia⁸⁴, and many of them are located in the mountains of southern Europe⁸⁵, where heterogenous topography results in a variety of microclimates providing suitable habitats during both warm and cold periods⁸⁶⁻⁸⁸. In the long term, these refugia not only provided suitable habitats for many taxa from the Northern Europe, but also acted as the locations of post-glacial recolonization when temperatures rose again at the end of the Ice Age⁸⁹⁻⁹¹.

The species of this study, namely *P. gussonei*, shows the above mentioned biological, geological and geographical peculiarities of Mediterranean plant biodiversity. *Petagnaea gussonei* is indeed a typical endemic plant distributed within a very narrow area (<20-km diameter) acting as refugia across a southern Mediterranean mountain range (Nebrodi Mountains), which is also geologically old, complex and heterogenous^{25,26}. However, current predictions of climate changes show serious threats to this unique biological heritage. Even minor atmospheric shifts can lead to substantial changes in climate, making Mediterranean mountains potentially more vulnerable than the other Mediterranean zones⁹²⁻⁹⁴. Some predictions report that a change in the biogeographic characteristics of these mountains is already occurring⁹⁵⁻⁹⁷. With increasing temperatures, many species across the world have already shifted their ranges to more suitable habitats, moving upwards in elevation or towards the poles^{98,99}, especially in mountains and at high elevation¹⁰⁰. Moreover, although many plant endemics are protected in reserves, some species may locally disappear because their niche lower limit passes over the local highest elevation¹⁰¹. In particular, current reserve networks, in the absence of functional corridors, may be inadequate on any scale to ensure long-term persistence of rare and endangered species¹⁰². It is generally believed that the risk of extinction under climate changes strongly reflects the inability of species to shift to suitable habitats¹⁰³. However, the velocity of the predicted climate changes is very likely to exceed the migration capability of many range-restricted species¹⁰⁴, especially because these species are often habitat specialists and weak dispersers. This is the case of Mediterranean narrow mountain endemics, like *P. gussonei*, which are expected to be particularly sensitive to environmental shifts, as several of these plants are extremely specialized and have evolved low dispersal ability^{105–107}. Climate changes alter also the main stages of plant phenology¹⁰⁸ and sexual reproduction¹⁰⁹, thus making these species more vulnerable to meteorological extreme events, and affecting plant–pollinator interaction¹¹⁰. However, still few studies have examined the effects of future climate changes on Mediterranean plant species, mainly on woody species, whereas information on endemic or rare herbaceous plants is even poorer. It is therefore of the utmost importance to assess the impact of climate changes on Mediterranean plant biodiversity, with a special focus on range-restricted mountain endemics like *P. gussonei*.

Desertification trends in the distributional area of Petagnaea gussonei

The areas sensitive to desertification showed overall different trends across the tree study periods (Fig. 5). Firstly, it is considerable how the "not affected" areas (class 1) increased from 14.3% in 1960, to 70.0% in 2000. During this 40-year period (1960–2000), the significant expansion of areas not sensitive to desertification is likely to reflect the positive events that occurred within the distributional range of *P. gussonei*. These events include the abandoning of cultivated fields, the consequent recovery and spread of natural vegetation, as well as the creation of wide protected areas such as the Nebrodi Mountains Park, which is the largest nature reserve in Sicily covering a surface of c. 86,000 ha. In the period 1960-2000 (Fig. 5), the results of this study showed also a moderate-low increase in the areas classified as potential, fragile 1 and fragile 2; in turn, the categories fragile 3, critical 1, critical 2, critical 3 reported a significant decline, the most considerable of which was the category "critical 3" that decreased from 25 to 0%. However, the general positive trend of desertification during 1960–2000 suffered a significant reversal of the tendency in the period 2000-2020 (Fig. 5). During this 20-year interval, all classes showed significantly worse values in the distributional area of P. gussonei: the increase in areas sensitive to desertification was on average two and tree fold compared to the previous 40-year period. In particular, the most dramatic result was the collapse of the "not affected" areas, which declined by half from 70% (2000) to 35% (2020). The factors, which contributed to this serious worsening of the areas sensitive to desertification, likely include high human impact with consequent natural habitat alteration, and massive erosive phenomena associated with extreme events, such as violent rains resulting from the ongoing climate changes.

Soil and landscape degradation, driven by human activities and climate variation, are considered key factors of desertification¹¹¹. Degraded soils, in particular, often show significant erosion processes that can be due to, e.g., unsustainable land management¹¹². Inappropriate changes in soil use include, on the one hand, removal or disturbance of surface cover, e.g., by fire, cultivation, intensive tillage and ploughing^{113,114}; on the other hand, loss of vegetation cover and lack of adequate soil conservation practices¹¹². Climate changes resulting in more intense storms may also be an important driver of soil erosion¹¹⁵. However, desertification occurs at different temporal and spatial scales, and is influenced by several factors that make the assessment of the process extremely complex¹¹⁶. Moreover, robust information is often insufficient to provide indicators that allow precise and efficient measurement of desertification effects¹¹⁷. The distributional range of P. gussonei falls within the heart of a hotspot of desertification: the Mediterranean basin. The Mediterranean region has been identified as particularly vulnerable to soil degradation¹⁴. In 2017, 25% of European land (411,000 km²), especially in southern Europe, was identified as being at high or very high risk of desertification, a 14% increase since 2008¹¹⁸. The Mediterranean region has the overall highest erosion rates within the EU¹¹⁹, the lowest levels of soil organic matter¹²⁰, and severe salinization problems¹²¹. It also has high abundance of shallow soils¹²², strong and increasing human pressures¹²³, and high climate change vulnerability^{13,16}. The main causes of soil degradation and increasing land sensitivity to desertification in the Mediterranean basin are primarily human-induced¹²⁴, and are generally more pronounced in areas with semi-arid or dry climate conditions, with water being the main factor limiting ecosystem performance, resilience and recovery^{125,126}. In rural areas, soil degradation occurs mainly through deforestation or unsustainable cropping, irrigation or grazing practices, which, in turn, stem from the socioeconomic conditions of the local people¹²⁷.

Across the Mediterranean region, increasing abandonment of extensive livestock systems raises wildfire risk and affects the current land footprint through changes in the mosaic landscape¹²⁸. Changes to agricultural systems, together with other land-use changes, are leading to critical levels of habitat loss¹⁶. This is of particular concern because the Mediterranean region is characterized by extraordinary biodiversity, with large numbers of endemic species¹²⁹. Unfortunately, the endemic plant *P. gussonei* is distributed in an area prone to land degradation and desertification due to both unfavorable bio-physical conditions and adverse human activities. Such factors include erratic precipitation (occurring mainly in winter season), high summer temperature and frequent seasonal droughts, poor and erodible soils, diversified landscapes, extensive human-induced deforestation and forest losses due to frequent wildfires, land abandonment and deterioration. Managing land in a sustainable manner means using land without damaging ecological processes or reducing biological diversity. It requires the maintenance of key components of the environment, such as biodiversity, ecological integrity and natural capital, as well as the conservation of a rich set of relationships among the constitutive elements of the system. Sustainable land management is a straightforward tool not only for the recovery of arid land, but also for the conservation of local biodiversity hotspots such as the Nebrodi Mountains where *P. gussonei* grows, and where landscape resilience could be improved by creating further protected areas and by increasing vegetation cover.

Soil-use changes in the distributional area of Petagnaea gussonei

CORINE Land Cover (CLC) classes showed numerous and complex changes from 1958 to 2018 (Table 3). Continuous and discontinuous urban fabric occupies just c. 1.5% of the *P. gussonei* range, and was constant across the 60-year study period. Agricultural areas (CLC class 2) declined considerably from 83.6% in 1958 to 40.3% in 2018. Forests and semi-natural areas (CLC class 3), instead, increased significantly from 16% in 1958 to 58.1% in 2018. Water bodies (CLC class 5) showed very low values, which were c. 0.3% in 1958 and less than 0.1% in 2018. Regarding the agricultural areas, the dominant class in 2018 was land mainly occupied by agriculture, with significant areas of natural vegetation (2.4.3.); fruit trees and berry plantations (class 2.2.2.) were dominant from 1990 to 2012 (20.4–22.2%), but drastically declined to 0.4% in 2018. Among forests and semi-natural areas, the dominant class throughout the study period was broad-leaved forest (3.1.1.), which reached a peak of 28.1% in 2012, and moderately declined to 22% in 2018. Other significant classes were sparsely vegetated areas (3.3.3.) and mixed forest (3.1.3.), which in 2018 were respectively 13.6 and 9.8% of the distributional area of *P. gussonei*. Overall, the occurrence sites of *P. gussonei* benefited from a drastic decrease in agricultural areas, and from a significant increase in forested and semi-natural areas, especially in terms of broad-leaved and mixed forests. However, farmland still occupies 40.3% of *P. gussonei* natural range, and 13.6% of the study area is characterized by sparse vegetation covering only 10–50% of surface, which is therefore highly vulnerable to erosion processes.

Landscape transformation is a key threat to ecosystem integrity, and the negative impacts extend far beyond the boundaries of the transformed land^{130,131}. Land-use changes caused by human activities have contributed directly to global biodiversity loss¹³². Land-use changes, in particular, are considered the major cause of natural habitat fragmentation and alteration due to the sprawl of rural and urban areas¹³³. Habitat fragmentation may decrease species diversity directly or indirectly by reducing the core area or causing habitat isolation^{134,135}. Fragmentation reduces also habitat connectivity and may increase the risk of extinction by altering the spatial configuration and community composition¹⁷. Halting biodiversity loss attributed to anthropogenic drivers, such as land-use changes, is thus one of the most urgent challenges for human society. For terrestrial ecosystems, the conversion of natural habitats to agricultural land is the primary threat¹³⁶. An estimated 40% of the Earth's ice-free land has already been converted to cropland and pastures^{137,138}. This global estimate is in line with the study area whose agricultural component decreased by half over a 60-year period. Despite this seemingly positive trend, the natural range of *P. gussonei* still falls within an area significantly impacted by farmland. However, the dominant CORINE agricultural class of the study area is defined as "*land mainly occupied by agriculture*" but also "*with significant areas of natural vegetation*" (2.4.3.), thus suggesting that landscape connectivity can be a feasible and effective guiding principle of conservation. To prevent local species extinctions, and to ensure the

	1958*		1990		2000		2006		2012		2018	
CORINE Land Cover classes	ha	%	ha	%	ha	%	ha	%	ha	%	ha	%
1.1.1. Continuous urban fabric	n.a	n.a	230 ^a	0.5	239 ^a	0.6	194 ^b	0.5	203 ^c	0.5	205 ^c	0.5
1.1.2. Discontinuous urban fabric		n.a	424 ^a	1.0	448 ^a	1.1	448 ^a	1.1	436 ^a	1.0	436 ^a	1.0
2.1.1. Non-irrigated arable land		32.1	658 ^b	1.6	658 ^b	1.6	954°	2.3	1153 ^d	2.7	1111 ^d	2.6
2.1.2. Irrigated arable land		0.4	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2.2.1. Vineyards		0.6	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2.2.2. Fruit trees and berry plantations		21.9	8658 ^b	20.4	8646 ^b	20.4	9387 ^c	22.2	9484 ^c	22.4	150 ^d	0.4
2.2.3. Olive groves	2962ª	7.0	3991 ^b	9.4	3970 ^b	9.4	3970 ^b	9.4	3870 ^c	9.1	2809 ^d	6.6
2.3.1. Pastures and uncultivated productive land		16.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2.4.1. Annual crops associated with permanent crops		5.6	192 ^b	0.5	192 ^b	0.5	192 ^b	0.5	80 ^c	0.2	119 ^d	0.3
2.4.2. Complex cultivation patterns		n.a	134 ^a	0.3	134 ^a	0.3	134 ^a	0.3	110 ^b	0.3	68 ^c	0.2
2.4.3. Land mainly occupied by agriculture, with significant areas of natural vegetation	n.a	n.a	1623ª	3.8	1623ª	3.8	3720 ^b	8.8	1932 ^c	4.6	12,792 ^d	30.2
3.1.1. Broad-leaved forest	5948 ^a	14.1	9125 ^b	21.5	9125 ^b	21.5	7124 ^c	16.8	11,912 ^d	28.1	9316 ^e	22.0
3.1.2. Coniferous forest	n.a	n.a	74 ^a	0.2	74 ^a	0.2	130 ^b	0.3	72 ^a	0.2	72 ^a	0.2
3.1.3. Mixed forest	798 ^a	1.9	784 ^a	1.9	784 ^a	1.9	1263 ^b	3.0	1506 ^c	3.6	4149 ^d	9.8
3.2.1. Natural grassland	n.a	n.a	4264 ^a	10.1	4264 ^a	10.1	4907 ^b	11.6	6440 ^c	15.2	2575 ^d	6.1
3.2.2. Moors and heathland	n.a	n.a	3118 ^a	7.4	3118 ^a	7.4	n.a	n.a	n.a	n.a	2,642 ^b	6.2
3.2.3. Sclerophylous vegetation	n.a	n.a	1545 ^a	3.6	1545 ^a	3.6	4501 ^b	10.6	5018 ^c	11.8	51 ^d	0.1
3.2.4. Transitional woodland/shrub	n.a	n.a	6630 ^a	15.6	6630ª	15.6	5,352 ^b	12.6	75°	0.2	61 ^d	0.1
3.3.2. Bare rock	n.a	n.a	86 ^a	0.2	86 ^a	0.2	n.a	n.a	n.a	n.a	n.a	n.a
3.3.3. Sparsely vegetated areas	n.a	n.a	710 ^a	1.7	710 ^a	1.7	43 ^b	0.1	n.a	n.a	5766 ^c	13.6
5.1.1. Water courses	n.a	n.a	112 ^a	0.3	112 ^a	0.3	38 ^b	0.1	41 ^c	0.1	n.a	n.a
5.1.2. Water bodies	n.a	n.a	9 ^a	0.02	9 ^a	0.02	9 ^a	0.02	36 ^b	0.09	36 ^b	0.09

Table 3. Areas (ha) of soil-use classes in 1958, 1990, 2000, 2006, 2012, 2018, within the natural range of *Petagnaea gussonei*. "n.a." is data not available; *soil-use classes in 1958 are provided by CNR-TCI, whose equivalence with CORINE Land Cover classes is reported in Fig. 6; different letters show significantly different values, among different years, within the same CORINE Land Cover class.

Scientific Reports | (2024) 14:8235 |

long-term maintenance of biodiversity, it is necessary to connect patches of remaining natural habitat¹³⁹. In this context, the implementation of conservation networks of natural or restored vegetation have become a favored mitigation approach in regions where arable cropping occurs within naturally forested landscapes^{140,141}. These ecological networks comprise mostly linear patches of remnant land (often connecting protected areas outside ecological networks), connected together through corridors and stepping-stone habitat^{142,143}.

Ecological networks are an effective approach that integrate environmental management strategies and landscape planning, and can be understood by different actors¹⁴⁴⁻¹⁴⁶. However, landscape connectivity for plants is mainly linked to their ability to disperse between habitat patches via propagules. Their dispersal is only successful if habitat patches are sufficiently connected^{147,148}, or if it is facilitated by suitable landscape features¹⁴⁹. Indeed, the ability of plants to disperse in fragmented landscapes also depends on their dispersal strategy^{150,151}. Overall, biodiversity loss largely depends on the temporal patterns of environmental changes, e.g., when biological response cannot match the rate of soil-use changes¹⁵². Whatever their alteration, restoring landscapes to their primitive ecological configuration is conservation utopia. Reference environmental conditions cannot exist because Earth's systems change in a natural way. Moreover, landscapes with a mosaic of human-affected soil uses may be biodiversity-friendly only to a certain point, but not completely. Improving landscape connectivity should be considered as a flexible and proactive solution for preserving biodiversity in most areas with different patterns of land-use changes, including the distributional area of P. gussonei, which can greatly benefit from dispersal corridors, especially because of its high capacity of asexual reproduction. Dispersal corridors provide also habitats for pollinators, which may contribute to the relatively low sexual reproduction of P. gussonei. Besides global and regional drivers of biodiversity decline, the consequences of local land-use decisions are major factors of this global decline, but they can be also the starting point for a biodiversity recovery.

Conclusions

The endemic plant P. gussonei showed critical conditions due to exogenous and endogenous factors. Dormancy affects endogenously P. gussonei, whose poor germination capacity may be further lower and slower as a result of the ongoing climate changes. Species like P. gussonei, with peculiar requirements for breaking dormancy, make inevitably the role of germplasm banks fundamental in ex situ conservation programs. Species respond also to exogenous factors that may act on different space-time scales. The natural range of P. gussonei falls within a narrow area where the exogenous impact of climate changes and desertification showed alarming trends, which especially worsened in the last few decades. Similarly, P. gussonei is distributed across a complex mosaic where the exogenous action of soil-use changes created a fragmented landscape. This conservation issue seems a dead end, but a feasible approach is possible to tackle it. The first step is to consider that these exogenous and endogenous factors interact, therefore coping with one factor helps dealing with the other three. Improving ecological connectivity can be a starting point to reverse the trends of the three exogenous factors, and to reduce the endogenous factor by favoring the dormancy-breaking of P. gussonei. Indeed, creating ecological networks is a realistic approach to conservation because c. 30% of P. gussonei range is characterized by "land mainly occupied by agriculture" but "with significant areas of natural vegetation", which can be interconnected through dispersal corridors. The genic flux of the P. gussonei metapopulation will benefit from widespread landscape connectivity, thus compensating the limiting consequences of a high dormancy. However, other biodiversity-friendly interventions should be considered, such as increasing plant cover in those areas with scarse vegetation in order to mitigate the erosion processes associated with desertification. All these actions will contribute to make P. gussonei more resilient to the ongoing climate changes. The second step is political, and consists in involving different stakeholders according to the kind of factor. Scientists have to continue their job to shed further light on the ecophysiology of *P. gussonei* seeds. Local communities play an invaluable role in influencing the soil uses that shape the distributional area of P. gussonei. Policy makers act on a wider scale, and can take concrete actions against climate changes by setting their agenda. A better biodiversity-friendly world is possible only if each stakeholder does its part and is open to dialogue with the other stakeholders.

Data availability

The data that support the findings of this study are not openly available due to reasons of sensitivity, and are available from the corresponding author upon reasonable request. Data are located in controlled access data storage at the Department of Biological, Geological and Environmental Sciences (Catania University, Italy).

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References

- 1. Pimm, S. L., Russell, G. J., Gittleman, J. L. & Brooks, T. M. The future of biodiversity. Science 269, 347-350 (1995).
- 2. Ceballos, G. et al. Accelerated modern human-induced species losses: entering the sixth mass extinction. Sci. Adv. 1, e1400253 (2015).
- IPBES. Global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services (eds Brondizio, E. S., Settele, J., Díaz, S. & Ngo, H. T.) 1148 (IPBES secretariat, Bonn, Germany) (2019). https://zenodo.org/records/6417333.
- SCBD (Secretariat of the Convention on Biological Diversity). Global Biodiversity Outlook, 5 (Montreal); https://www.cbd.int/ gbo/gbo5/publication/gbo-5-en.pdf (2020).
- Greuter, W. Botanical diversity, endemism, rarity and extinction in the Mediterranean area: An analysis based on the published volumes of Med-checklist. *Botanika Chronika* 10, 63–79 (1991).
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A. B. & Kent, J. Biodiversity hotspots for conservation priorities. Nature 403, 853–858 (2000).

- Cuttelod, A., García, N., Abdul Malak, D., Temple, H. & Katariya, V. The Mediterranean: A biodiversity hotspot under threat. In *The 2008 Review of The IUCN Red List of Threatened Species* (eds Vié, J. C. *et al.*) (IUCN, Gland, 2008).
- Meynecke, J. O. Effects of global climate change on geographic distributions of vertebrates in North Queensland. *Ecol. Model.* 174, 347–357 (2004).
- 9. Baskin, C. C. & Baskin, J. M. Seeds Ecology, Biogeography, and Evolution of Dormancy and Germination (Academic Press, San Diego, California, 1998).
- 10. Van Assche, J., Van Nerum, D. & Darius, P. The comparative germination ecology of nine *Rumex* species. *Plant Ecol.* **159**, 131–142 (2002).
- 11. Donohue, K., de Casas, R. R., Burghardt, L., Kovach, K. & Willis, C. G. Germination, post germination adaptation and species ecological ranges. *Annu. Rev. Ecol. Evolut. Syst.* **41**, 293–319 (2010).
- 12. Pedrini, S. & Dixon, K. W. International principles and standards for native seeds in ecological restoration. *Restor. Ecol.* 28, S286–S303 (2020).
- IPCC. Climate Change and Land: An IPCC Special Report on Climate Change, Desertification, Land Degradation, Sustainable Land Management, Food Security, and Greenhouse Gas Fluxes in *Terrestrial Ecosystems* (eds Shukla, P. R., Skea, J. & Calvo Buendia, E.) (2019).
- 14. Lahmar, R. & Ruellan, A. Soil degradation in the Mediterranean region and cooperative strategies. *Cahiers Agric*. **16**(4), 318–323 (2007).
- 15. Pachauri, R. K., Allen, M. R., Barros, V. R., Broome, J., Cramer, W. et al. Climate change 2014: Synthesis report. Contribution of Working Groups I, II and III to the fifth assessment report of the Intergovernmental Panel on Climate Change (2014).
- Bourlion, N. & Ferrer, R. The Mediterranean region's development and trends: framework aspects. FAO and Plan Bleu. 2018. State of Mediterranean Forests 2018. Food and Agriculture Organization of the United Nations, Rome and Plan Bleu, Marseille. Chapter 1, 2–15 (2018).
- 17. May, F. *et al.* The geometry of habitat fragmentation: Effects of species distribution patterns on extinction risk due to habitat conversion. *Ecol. Evol.* **9**(5), 2775–2790 (2019).
- Wolff, H. Umbelleferae–Saniculoideae in Das Pflanzenreich (ed. Engler, A.), IV, 228 (Hf. 61) Leipzig, Germany, 1–305 [Petagnaea in pp. 23, 273–274] (Wilhelm Engelmann, 1913).
- Gianguzzi, L., La Mantia, A. & Lo Presti, R. M. Distribuzione, ecologia e status conservativo delle stazioni di *Petagnaea gussonei* (Sprengel) Rauschert (Apiaceae) nell'area dei Monti Nebrodi (Sicilia nord-orientale). *Naturalista Siciliano* 28(1), 265–326 (2004).
- 20. De Montmollin, B. & Strahm, W. The Top 50 Mediterranean Island Plants. Wild Plants at the Brink of Extinction, and What is Needed to Save Them (IUCN, Gland, 2005).
- 21. Bern Convention on the Conservation of European Wildlife and Natural Habitats. https://www.coe.int/en/web/bern-convention (1979).
- 22. Habitats Directive. Council Directive 92/43/EEC on the Conservation of natural habitats and of wild fauna and flora. https://ec.europa.eu/environment/nature/legislation/habitatsdirective/index_en.htm (1992).
- 23. Top 50 Mediterranean Island Plants. https://top50.iucn-mpsg.org/species/38 (2017).
- 24. De Castro, O. *et al.* Old sleeping Sicilian beauty: Seed germination in the palaeoendemic Petagnaea gussonei (Spreng.) Rauschert (Saniculoideae, Apiaceae). *Plant Biol.* **17**, 1095–1098 (2015).
- Lentini, F., Carbone, S. & Grasso, M. Carta geologica della Provincia di Messina. Scale 1:50.000, 3 sheets. Nota illustrativa a cura di Lentini F., Catalano S. & Carbone S; Firenze (S.EL.CA, 2000).
- Corrado, S., Aldega, L., Balestrieri, M. L., Maniscalco, R. & Grasso, M. Structural evolution of the sedimentary accretionary wedge of the alpine system in Eastern Sicily: Thermal and thermochronological constraints. *Geol. Soc. Am. Bull.* 121(11–12), 1475–1490 (2009).
- 27. Rivas-Martínez, S., Sáenz, S. R. & Penas, A. Worldwide bioclimatic classification system. Glob. Geobot. 1, 1-634 (2011).
- Liu, M., van Wyk, B. E. & Tilney, P. M. The taxonomic value of fruit structure in the subfamily Saniculoideae and related African genera (Apiaceae). *Taxon* 52, 261–270 (2003).
- Calviño, C. I. & Downie, S. R. Circumscription and phylogeny of Apiaceae subfamily Saniculoideae based on chloroplast DNA sequences. Mol. Phylogenet. Evol. 44, 175–191 (2007).
- Calviño, C. I., Martínez, S. G. & Downie, S. R. Morphology and biogeography of Apiaceae subfamily Saniculoideae as inferred by phylogentic analysis of molecular data. Am. J. Bot. 95(2), 196–214 (2008).
- Kadereit, J. W., Repplinger, M., Schmalz, N., Uhink, C. H. & Wörz, A. The phylogeny and biogeography of Apiaceae subf. Saniculoideae tribe Saniculeae: From south to north and south again. *Taxon* 57, 365–382 (2008).
- 32. Scharhag, C. & Claßen-Bockhoff, R. Architecture and flowering sequence in the Apiaceae-Saniculoideae. In *Systematics* (eds Gradstein, S. R. *et al.*) (Universitätsverlag Göttingen, Göttingen, 2008).
- De Castro, O., Colombo, P., Gianguzzi, L. & Perrone, R. Flower and fruit structure of the endangered species *Petagnaea gussonei* (Sprengel) Rauschert (Saniculoideae, Apiaceae) and implications for its reproductive biology. *Plant Biosyst.* 149(6), 1042–1051 (2015).
- De Castro, O., Cennamo, P. & De Luca, P. Analysis of the genus Petagnaea Caruel (Apiaceae), using new molecular and literature data. Plant Syst. Evolut. 278(3), 239–249 (2009).
- Gianguzzi, L. Petagnaea gussonei (Sprengel) Rauschert. Schede per una Lista Rossa della Flora vascolare e crittogamica Italiana. Informatore Botanico Italiano 43(2), 412–416 (2011).
- De Castro, O., Marino, G., Gianguzzi, L. & Guida, M. A survey on conservation status of *Petagnaea gussonei* (Apiaceae), an endemic species to Nebrodi Mountains (Sicily, Italy). *Delpinoa* 48, 11–19 (2006).
- De Castro, O. *et al.* Genetic structure in the paleoendemic and endangered Petagnaea gussonei (Spreng.) Rauschert (Saniculoideae, Apiaceae) and implications for its conservation. *Plant Syst. Evolut.* 299, 209–223 (2013).
- Vandelook, F. & Van Assche, J. A. Deep complex morphophysiological dormancy in *Sanicula europaea* (Apiaceae) fits a recurring pattern of dormancy types in genera with an Arcto-Tertiary distribution. *Botany* 86, 1370–1377 (2008).
- Hawkins, T. S., Baskin, C. C. & Baskin, J. M. Morphophysiological dormancy in seeds of three eastern North American Sanicula species (Apiaceae subf. Saniculoideae): Evolutionary implications for dormancy break. *Plant Species Biol.* 25, 103–113 (2010).
- Torchyk, S. P. & Titok, V. V. Biological peculiarities of Astrantia major L. under ex situ conditions. *Rev. Bot.* 1(8), 125–129 (2014).
 Mulaudzi, R. B., Kulkarni, M. G., Finnie, J. F. & Van Staden, J. Optimizing seed germination and seedling vigour of *Alepidea amatymbica* and *Alepidea natalensis*. *Seed Sci. Technol.* 37(2), 527–533 (2009).
- 42. Necajeva, J. & Ievinsh, G. Seed dormancy and germination of an endangered coastal plant *Eryngium maritimum* (Apiaceae). *Estonian J. Ecol.* **62**, 150–161 (2013).
- Wolkis, D., Blackwell, S. & Villanueva, S. K. Conservation seed physiology of the ciénega endemic, *Eryngium sparganophyllum* (Apiaceae). Conserv. Physiol. 8, 1–9 (2020).
- 44. Bacchetta, G., Fenu, G., Mattana, E., Piotto, B. & Virevaire, M. Manuale per la raccolta, studio, conservazione e gestione ex situ del germoplasma. Manuali e Linee Guida APAT 37/2006, Roma (APAT, 2006).
- 45. ISTA. International rules for seed testing 2017. The International Seed Testing Association (ISTA). Bassersdorf, Switzerland (2017).
- 46. Martin, A. C. The comparative internal morphology of seeds. Am. Midl. Nat. 36, 513-660 (1946).

- 47. Baskin, C. C. & Baskin, J. M. Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination 2nd edn. (Elsevier Academic Press, 2014).
- Thanos, C. A., Georghiu, K. & Skarou, F. Glaucium flavum seed germination an ecophysiological approach. Anna. Bot. 63, 121–130 (1989).
- 49. ENSCONET. ENSCONET Germination Recommendations (2009).
- 50. Scott, S., Jones, R. & Williams, W. Review of data analysis methods for seed germination. Crop Sci. 24, 1192–1199 (1984).
- 51. Orchard, T. Estimating the parameters of plant seedling emergence. Seed Sci. Tech. 5, 61-69 (1977).
- Khan, S. et al. Quantifying temperature and osmotic stress impact on seed germination rate and seedling growth of Eruca sativa Mill. via hydrothermal time model. Life 12(3), 400 (2022).
- 53. Al-Mudaris, M. A. Notes on various parameters recording the speed of seed germination. J. Agric. Trop. Subtrop. 99, 147–154 (1998).
- 54. Coolbear, P., Francis, A. & Grieson, D. The effect of low temperature pre-sowing treatment on the germination performance and membrane integrity of artificially aged tomato seeds. *J. Exp. Bot.* **35**(160), 1609–1617 (1984).
- Jones, K. & Sanders, D. The influence of soaking pepper seed in water or potassium salt solutions on germination at three temperatures. J. Seed Tech. 11, 97–102 (1987).
- 56. Esechie, H. Interaction of salinity and temperature on the germination of Sorghum. J. Agron. Crop Sci. 172, 194–199 (1994).
- Benech Arnold, R. L., Fenner, M. & Edwards, P. J. Changes in germinability, ABA content and ABA embryonic sensitivity in developing seeds of Sorghum bicolor (L.) Moench. induced by water stress during grain filling. *New Phytol.* 118, 339–347 (1991).
- Sicilian Regional Government. Meteorological records. https://www.regione.sicilia.it/isituzioni/regione/strutture-regionali/ presidenza-regione/autorita-bacino-distretto-idrografico-sicilia/annali-idrologici (2022).
- Kosmas, C., Ferrara, A., Briassouli, H. & Imeson, I. Methodology for mapping ESAs to desertification in *The MEDALUS Project Mediterranean Desertification and Land Use. Manual on Key Indicators of Desertification and Mapping Environmentally Sensitive Areas to desertification* (eds. Kosmas, C., Kirkby, M. & Geeson, N.) 31–47 (1999).
- Coscarelli, R., Minervino, I. & Sorriso-Valvo, M. Methods for the characterization of areas sensitive to desertification: an application to the Calabrian territory (Italy), in *Geomorphological Process and Human Impacts in River Basins*. Proceedings of the International Conference held at Solsona, Catalonia, Spain, May 2004. *IAHS Publ.* 299, (2005).
- 61. SINANET. CORINE Land Cover. https://groupware.sinanet.isprambiente.it/uso-copertura-e-consumo-di-suolo/library/copertura-del-suolo/corine-land-cover (2018).
- 62. CNR-TCI. Carta dell'utilizzazione del suolo. Foglio n. 22. Touring Club Italiano (1958).
- Baskin, C. C. & Baskin, J. M. Germination ecophysiology of herbaceous plant species in a temperate region. Am. J. Bot. 75, 286-305 (1988).
- 64. Walmsley, C. A. & Davy, A. J. Germination characteristics of shingle beach species, effects of seed ageing and their implications for vegetation restoration. J. Appl. Ecol. 34, 131–142 (1997).
- Schutte, B. & Knee, M. The effects of rudimentary embryos and elevated oxygen on seed dormancy of Eryngium yuccifolium Michx. (Apiaceae). Seed Sci. Technol. 33, 53–62 (2005).
- 66. Nikolaeva, M. G. Factors controlling the seed dormancy pattern. In *The Physiology and Biochemistry of Seed Dormancy and Germination* (ed. Khan, A. A.) 51–74 (North-Holland, Amsterdam, Netherlands, 1977).
- 67. Baskin, J. M. & Baskin, C. C. A classification system for seed dormancy. Seed Sci. Res. 14, 1-16 (2004).
- Hawkins, T. S., Baskin, J. M. & Baskin, C. C. Seed morphology, germination phenology, and capacity to form a seed bank in six herbaceous layer Apiaceae species of the Eastern Deciduous Forest. *Castanea* 72, 8–14 (2007).
- 69. IPCC. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (Cambridge University Press, Cambridge, 2013).
- Christensen, J. H. et al. Regional climate projections. In Climate Change, 2007: The Physical Science Basis Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change 848–940 (Cambridge University Press, Cambridge, 2007).
- Peñuelas, J. et al. Evidence of current impact of climate change on life: a walk from genes to the biosphere. Glob. Change Biol. 19, 2303–2338 (2013).
- Mariotti, A., Pan, Y. T., Zeng, N. & Alessandri, A. Long-term climate change in the Mediterranean region in the midst of decadal variability. *Clim. Dyn.* 44(5–6), 1437–1456 (2015).
- 73. Scoccimarro, E., Gualdi, S., Bellucci, A., Zampieri, M. & Navarra, A. Heavy precipitation events over the Euro-Mediterranean region in a warmer climate: results from CMIP5 models. *Reg. Environ. Chang.* **16**(3), 595–602 (2016).
- 74. Giorgi, F. Climate change hot-spots. Geophys. Res. Lett. 33(8), L08707 (2006).
- 75. Diffenbaugh, N. S. & Giorgi, F. Climate change hotspots in the CMIP5 global climate model ensemble. *Clim. Chang.* **114**, 813–822 (2012).
- Cramer, W. *et al.* Climate change and interconnected risks to sustainable development in the Mediterranean. *Nat. Clim. Chang.* 8, 972–980 (2018).
- Médail, F. & Quézel, P. Hot-spots analysis for conservation of plant biodiversity in the Mediterranean basin. Ann. Mi. Bot. Gard. 84, 112–127 (1997).
- Médail, F. & Quézel, P. Biodiversity hotspots in the Mediterranean Basin: setting global conservation priorities. *Conserv. Biol.* 13, 1510–1513 (1999).
- Cowling, R. M., Rundel, P. W., Lamont, B. B., Arroyo, M. K. & Arianoutsou, M. Plant diversity in Mediterranean-climate regions. *Trends Ecol. Evol.* 11, 362–366 (1996).
- 80. Médail, F. et al. What is a tree in the Mediterranean basin hotspot? A critical analysis. For. Ecosyst. 6, 1–19 (2019).
- Thompson, J. D., Lavergne, S., Affre, L., Gaudeul, M. & Debussche, M. Ecological differentiation of Mediterranean endemic plants. *Taxon* 54, 967–976 (2005).
- Van der Wiel, A. M. & Wijmstra, T. A. Palynology of the lower part (78–120m) of the core Tenaghi Philippon, Middle Pleistocene of Macedonia. Greece. Rev. Palaeobot. Palynol. 52, 73–88 (1987).
- Biltekin, D. et al. Anatolia: A long-time plant refuge area documented by pollen records over the last 23million years. Rev. Palaeobot. Palynol. 215, 1–22 (2015).
- 84. Bennett, K. D. & Provan, J. What do we mean by "refugia"?. Quat. Sci. Rev. 27(27-28), 2449-2455 (2008).
- Väre, H., Lampinen, C., Humphries, C. & Williams, P. Taxonomic diversity of vascular plants in the European alpine areas. In Alpine Biodiversity in Europe. Ecological Studies (eds Nagy, L. et al.) 133–148 (Springer, Berlin, 2003).
- 86. Bennett, K. D., Tzedakis, P. C. & Willis, K. J. Quaternary refugia of North European trees. J. Biogeogr. 18, 103–115 (1991).
- 87. Taberlet, P. & Cheddady, F. R. Quaternary refugia and persistence of biodiversity. *Science* **297**, 2009–2010 (2002).
- 88. Magri, D. Persistence of tree taxa in Europe and quaternary climate changes. *Quat. Int.* **219**, 145–151 (2010).
- Tzedakis, P. C., Lawson, I. T., Frogley, M. R., Hewitt, G. M. & Preece, R. C. Buffered tree population changes in a Quaternary refugium: Evolutionary implications. *Science* 297, 2044 (2002).
- Bhagwat, S. A. & Willis, K. J. Species persistence in northerly glacial refugia of Europe: A matter of chance or biogeographical traits?. J. Biogeogr. 35, 464–482 (2008).
- 91. Médail, F. & Diadema, K. Glacial refugia influence plant diversity patterns in the Mediterranean Basin. J. Biogeogr. 36, 1333–1345 (2009).

- 92. Lionello, P. *et al.* The Mediterranean climate: An overview of the main characteristics and issues. In *Mediterranean Climate Variability* (eds Lionello, P. *et al.*) 1–26 (Elsevier, Amsterdam, 2006).
- Nogués-Bravo, D., Araújo, M. B., Errea, M. P. & Martínez-Rica, J. P. Exposure of global mountain systems to climate warming during the 21st century. *Glob. Environ. Change* 17, 420–428 (2007).
- 94. Nogués-Bravo, D., Araújo, M. B., Lasanta, T. & López, J. I. Climate change in Mediterranean Mountains during the 21st century. Ambio 37, 280–285 (2008).
- Sanz-Elorza, M., Dana, E. D., González, A. & Sobrino, E. Changes in the high-mountain vegetation of the Central Iberian Peninsula as a probable sign of global warming. *Anna. Bot.* 92, 273–280 (2003).
- 96. Wilson, R. J. *et al.* Changes to the elevational limits and extent of species ranges associated with climate change. *Ecol. Lett.* **8**, 1138–1146 (2005).
- 97. Ruiz-Labourdette, D., Nogués-Bravo, D., Saínz Ollero, L., Schmitz, M. F. & Pineda, F. Forest composition in Mediterranean mountains is projected to shift along the entire elevational gradient under climate change. *J. Biogeogr.* **39**, 162–176 (2012).
- Hughes, L. Biological consequences of global warming: Is the signal already apparent?. *Trends Ecol. Evol.* 2(15), 56–61 (2000).
 Lenoir, J., Gegout, J. C., Marquet, P. A., De Ruffray, P. & Brisse, H. A significant upward shift in plant species optimum elevation
- during the 20th century. Science 5884(320), 1768–1771 (2008).
- 100. Walther, G. R., Beissner, S. & Burga, C. A. Trends in the upward shift of alpine plants. J. Veg. Sci. 5(16), 541-548 (2005).
- Trivedi, M. R., Morecroft, M. D., Berry, P. M. & Dawson, T. P. Potential effects of climate change on plant communities in three montane nature reserves in Scotland, UK. *Biol. Conserv.* 6(141), 1665–1675 (2008).
- Araujo, M. B., Cabeza, M., Thuiller, W., Hannah, L. & Williams, P. H. Would climate change drive species out of reserves? An assessment of existing reserve-selection methods. *Glob. Change Biol.* 9(10), 1618–1626 (2004).
- Engler, R. *et al.* Predicting future distributions of mountain plants under climate change: does dispersal capability matter?. *Ecography* 32, 34–45 (2009).
- 104. Pearson, R. G. Climate change and the migration capacity of species. Trends Ecol. Evol. 21, 111-113 (2006).
- Lavergne, S., Thuiller, W., Molina, J. & Debussche, M. Environmental and human factors influencing rare plant local occurrence, extinction and persistence: A 115-year study in the Mediterranean region. J. Biogeogr. 32, 799–811 (2005).
- Thuiller, W., Lavorel, S. & Araújo, M. B. Niche properties and geographical extent as predictors of species sensitivity to climate change. *Glob. Ecol. Biogeogr.* 14, 347–357 (2005).
- 107. Isik, K. Rare and endemic species: Why are they prone to extinction?. Turk. J. Bot. 35, 411-417 (2011).
- 108. Menzel, A. & Fabian, P. Growing season extended in Europe. Nature 6721(397), 659-1659 (1999).
- Hedhly, A., Hormaza, J. I. & Herrero, M. Global warming and sexual plant reproduction. *Trends Plant Sci.* 1(14), 30–36 (2009).
 Morin, X. *et al.* Variation in cold hardiness and carbohydrate concentration from dormancy induction to bud burst among
- provenances of three European oak species. *Tree Physiol.* **6**(27), 817–825 (2007). 111. Herrmann, S. M. & Hutchinson, C. F. The changing contexts of the desertification debate. *J. Arid. Environ.* **63**, 538–555 (2005).
- Herrinann, S. M. & Hutchinson, C. F. The changing contexts of the desertification debate. J. Arta. Environ. 65, 556–555 (2005).
 Borrelli, P. et al. An assessment of the global impact of 21st century land use change on soil erosion. Nat. Commun. 8, 1–13
- (2017). 113. Shakesby, R. A. *et al.* Impacts of prescribed fire on soil loss and soil quality: an assessment based on an experimentally-burned
- Shakesby, R. A. *et al.* Impacts of prescribed fire on soil loss and soil quality: an assessment based on an experimentally-burned catchment in Central Portugal. *Catena* 128, 278–293 (2015).
- Panagos, P., Borrelli, P. & Poesen, J. Soil loss due to crop harvesting in the European Union: A first estimation of an underrated geomorphic process. Sci. Total Environ. 664, 487–498 (2019).
- Borrelli, P. et al. Land use and climate change impacts on global soil erosion by water (2015–2070). Proc. Natl. Acad. Sci. 117(36), 21994–22001 (2020).
- Vieira, R. M. D. S. P. et al. Desertification risk assessment in Northeast Brazil: Current trends and future scenarios. Land Degrad. Dev. 32, 224–240 (2021).
- 117. Vendruscolo, J. *et al.* Monitoring desertification in semiarid Brazil: Using the desertification degree index (DDI). *Land Degrad. Dev.* **32**, 684–698 (2021).
- 118. Pravalie, R., Patriche, C. & Bandoca, G. Quantification of land degradation sensitivity areas in southern and central southeastern Europe. New results based on improving DISMED methodology with new climate data. *Catena* **158**, 309–320 (2017).
- Panagos, P. et al. A soil erosion indicator for supporting agricultural, environmental and climate policies in the European Union. Remote Sens. 12, 1365 (2020).
- Aguilera, E., Lassaletta, L., Saz-Cobena, A., Garnier, J. & Vallejo, A. The potential of organic fertilizers and water management to reduce N₂O emissions in Mediterranean climate cropping systems A review. *Agric. Ecosyst. Environ.* 164, 32–52 (2013).
- Stolte, J., Tesfai, M., Oygarden, L., Kvaemo, S., Keizer, J. et al. Soil Threats in Europe. Status, Methods, Drivers and Effects on Ecosystem Services. A Review Report, Deliverable 2.1 of the RECARE Project. European Union. https://doi.org/10.2788/828742 (2016).
- 122. Lagacherie, P. *et al.* Managing Mediterranean soil resources under global change: Expected trends and mitigation strategies. *Reg. Environ. Chang.* **18**, 663–675 (2018).
- 123. Guittonny-Philippe, A. *et al.* Constructed wetlands to reduce metal pollution from industrial catchments in aquatic Mediterranean ecosystems: A review to overcome obstacles and suggest potential solutions. *Environ. Int.* **64**, 1–16 (2014).
- 124. Feoli, E., Giacomich, P., Mignozzi, K., Oztürk, M. & Scimone, M. Monitoring desertification risk with an index integrating climatic and remotely-sensed data: an example from the coastal area of Turkey. *Manag. Environ. Qual.* **14**, 10–21 (2003).
- 125. Lavado Contador, J. F., Schnabel, S., Gomez Gutierrez, A. & Pulido Fernandez, M. Mapping sensitivity to land degradation in Extremadura, SW Spain. Land Degrad. Dev. 20(2), 129–144 (2009).
- Ferrara, C., Salvati, L. & Tombolini, I. An integrated evaluation of soil resource depletion from diachronic settlement maps and soil cartography. *Geoderma* 232–234, 394–405 (2014).
- 127. Imeson, A. Desertification, Land Degradation and Sustainability (Wiley, Chichester, 2012).
- 128. Shakesby, R. Post-wildfire soil erosion in the Mediterranean: review and future research directions. *Earth Sci. Rev.* **105**(3-4), 71–100 (2011).
- EC. The Mediterranean Region. European Commission (accessed 26 February 2024); https://ec.europa.eu/environment/nature/ natura2000/biogeog_regions/mediterranean/index_en.htm (2020).
- Samways, M. J., Bazelet, C. S. & Pryke, J. S. Provision of ecosystem services by large scale corridors and ecological networks. Biodivers. Conserv. 19(10), 2949–2962 (2010).
- 131. Sage, R. F. Global change biology: A primer. Glob. Chang. Biol. 26(1), 3–30 (2020).
- 132. Newbold, T. et al. Global effects of land use on local terrestrial biodiversity. Nature 520(7545), 45-50 (2015).
- Foltête, J. C., Girardet, X. & Clauzel, C. A methodological framework for the use of landscape graphs in land-use planning. Landsc. Urban Plann. 124, 140–150 (2014).
- Piekielek, N. B. & Hansen, A. J. Extent of fragmentation of coarse-scale habitats in and around US National Parks. *Biol. Conserv.* 155, 13–22 (2012).
- 135. Aguilar, R. et al. Habitat fragmentation reduces plant progeny quality: A global synthesis. Ecol. Lett. 22(7), 1163–1173 (2019).
- Phillips, H. R. P., Newbold, T. & Purvis, A. Land-use effects on local biodiversity in tropical forests vary between continents. Biodiv. Conserv. 26, 2251–2270 (2017).
- 137. Foley, J. A. et al. Global consequences of land use. Science 309, 570-574 (2005).

- Ramankutty, N., Evan, A. T., Monfreda, C. & Foley, J. A. Farming the planet: 1. Geographic distribution of global agricultural lands in the year 2000. *Glob. Biogeochem. Cycl.* 22(1), GB1003 (2008).
- 139. Fahrig, L. & Merriam, G. Conservation of fragmented populations. Conserv. Biol. 8, 50-59 (1994).
- Morandin, L. A. & Kremen, C. Hedgerow restoration promotes pollinator populations and exports native bees to adjacent fields. Ecol. Appl. 23, 829–839 (2013).
- 141. Wilson, S. *et al.* Influence of crop type, heterogeneity and woody structure on avian biodiversity in agricultural landscapes. *Ecol. Indic.* 83, 218–226 (2017).
- 142. Bouwma, I. M., Jongman, R. H. & Butovsky, R. O. Indicative Map of the Pan-European Ecological Network for Central and Eastern Europe. Technical Background Document (ECNC, 2002).
- 143. Jones-Walters, L. Pan-European ecological networks. J. Nat. Conserv. 15(4), 262-264 (2007).
- 144. De Montis, A. et al. Urban-rural ecological networks for landscape planning. Land Use Pol. 50, 312-327 (2016).
- 145. Keeley, A. T. H. *et al.* New concepts, models, and assessments of climate-wise connectivity. *Environ. Res. Lett.* **13**(7), 073002 (2018).
- 146. Sahraoui, Y. *et al.* Integrating ecological networks modelling in a participatory approach for assessing impacts of planning scenarios on landscape connectivity. *Landsc. Urban Plann.* **209**, 104039 (2021).
- 147. Fahrig, L. & Merriam, G. Habitat patch connectivity and population survival. Ecology 66, 1762-1768 (1985).
- 148. Bowne, D. R. & Bowers, M. A. Interpatch movements in spatially structured populations: A literature review. *Landsc. Ecol.* **19**, 1–20 (2004).
- Taylor, P. D., Fahrig, L., Henein, K. & Merriam, G. Connectivity is a vital element of landscape structure. Oikos 68, 571–573 (1993).
- Vittoz, P. & Engler, R. Seed dispersal distances: a typology based on dispersal modes and plant traits. Bot. Helv. 117, 109–124 (2007).
- 151. Boscutti, F., Sigura, M., De Simone, S. & Marini, L. Exotic plant invasion in agricultural landscapes: A matter of dispersal mode and disturbance intensity. *Appl. Veg. Sci.* 21(2), 250–257 (2018).
- 152. Ordonez, A., Martinuzzi, S. & Radelo, V. C. Combined speeds of climate and land-use change of the conterminous US until 2050. *Nat. Clim. Chang.* **4**, 1–6 (2014).

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Author contributions

The authors declare that they contributed equally to each part of the manuscript. G. B.: conceptualization, data curation, formal analysis, funding acquisition, investigation, methodology, resources, supervision, validation, visualization, writing original draft, writing review & editing. V. V.: conceptualization, data curation, formal analysis, funding acquisition, investigation, methodology, resources, supervision, validation, writing original draft, writing review & editing.

Competing interests

The authors declare no competing interests.

Additional information

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