scientific reports

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OPEN Association between triglyceride glucose index and total bone mineral density: a cross-sectional study from NHANES 2011–2018

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The Homeostatic Model Assessment for Triglyceride Glucose Index (TyG) and its related indices, including triglyceride glucose-waist circumference (TyG-WC), triglyceride glucose-waist-to-height ratio (TyG-WHtR) and triglyceride glucose-body mass index (TyG-BMI), has emerged as a practical tool for assessing insulin resistance in metabolic disorders. However, limited studies have explored the connection between TyG, TyG-related indices and osteoporosis. This population-based study, utilizing data from the National Health and Nutrition Examination Survey 2011–2018, involved 5456 participants. Through weighted multivariate linear regression and smoothed curve fitting, a significant positive correlation was found between TyG, TyG-related indices and total bone mineral density (BMD) after adjusting for covariates [$\beta = 0.0124$, 95% CI (0.0006, 0.0242), P = 0.0390; $\beta = 0.0004$, 95% CI (0.0003, 0.0004), P < 0.0001; β = 0.0116, 95% CI (0.0076, 0.0156), P < 0.0001; β = 0.0001, 95% CI (0.0001, 0.0001), P < 0.0001]. In subgroup analysis, race stratification significantly affected the relationship between TyG and total BMD. Additionally, gender and race were both significant for TyG-related indices. Non-linear relationships and threshold effects with inflection points at 9.106, 193.9265, 4.065, and 667.5304 (TyG, TyG-BMI, TyG-WHtR, TyG-WC) were identified. Saturation phenomena were observed between TyG-BMI, TyG-WC and total BMD with saturation thresholds at 314.177 and 1022.0428. These findings contributed to understanding the association between TyG, TyG-related indices and total BMD, offering insights for osteoporosis prevention and treatment.

Keywords Osteoporosis, BMD, TyG, Metabolism, NHANES

Abbreviations

ALP	Alkaline phosphatase
BMD	Bone mineral density
BMI	Body mass index
BUN	Blood urea nitrogen
CPK	Creatine phosphokinase
DXA	Dual-energy X-ray absorption
Family PIR	Ratio of family income to poverty
HDL-C	High-density lipoprotein cholesterol
HECT	Hyperinsulinemic-euglycemic clamp
HOMA-IR	Homeostasis model assessment of insulin resistance
IR	Insulin resistance
LDL-C	Low-density lipoprotein cholesterol
MetS	Metabolic syndrome
NHANES	National Health and Nutrition Examination study database
OP	Osteoporosis
T2DM	Type 2 diabetes mellitus
TyG	Triglyceride glucose index
TvG-BMI	Triglyceride glucose-body mass index

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Triglyceride glucose-waist circumference
Triglyceride glucose-waist-to-height ratio
Waist circumference
Waist-to-height ratio

Osteoporosis (OP) is a systemic degenerative skeletal disorder characterized by diminished bone mineral density (BMD) and impaired skeletal microarchitecture, leading to increased skeletal fragility and fractures^{1–3}. An epidemiological investigation estimated that approximately 200 million individuals worldwide were affected by OP, which is increasing yearly⁴. As the global population continues to age, the implications of OP are expected to escalate. Projections indicate that by 2025, the direct costs associated with this condition will reach a staggering \$25.3 billion annually⁵. This will inevitably place a considerable strain on both the healthcare system and the broader social economy⁶. Undoubtedly, OP has emerged as an imperative global public health concern.

As a major determinant of bone health, the measurement of BMD is a commonly used operational tool for the diagnosis of OP⁷. Relevant studies have shown that metabolic syndrome (MetS), characterized by abdominal obesity and metabolic dysfunction, has emerged as a new risk factor for BMD reduction⁸. Insulin resistance (IR) refers to the inability of peripheral tissues to respond to insulin and is the main feature of metabolic syndrome^{9,10}. IR is a potential factor affecting bone density. However, current studies investigating the association between IR and OP have produced inconsistent results. Earlier study reported that the relationship between lumbar spine BMD and IR was negative¹¹. In contrast, a recent research showed that IR was a protective factor for OP in a specific range¹². There's also a evidence that reveals the connection between IR and OP risk in type 2 diabetes mellitus (T2DM) patients is significantly affected by gender¹³. A more precise comprehension of the correlation between IR and BMD could offer novel insights into the prevention and treatment of OP.

Hyperinsulinemic-euglycemic clamp (HECT) is widely considered the gold standard for evaluating IR¹⁴. Another prevalent method for assessing IR is the homeostasis model assessment of insulin resistance (HOMA-IR)¹⁵. However, both of the above two methods are limited in application scenarios, particularly because fasting insulin levels are infrequently measured in routine clinical settings and large-scale epidemiological studies. In contrast, the triglyceride glucose index (TyG) offers a simpler and more cost-effective way to assess IR. Numerous studies have shown that the accuracy of TyG is on par with that of HECT and HOMA-IR for estimating IR^{16,17}. Moreover, it is imperative to note that patients with high IR often experience obesity¹⁸, which is also a significant factor affecting BMD¹⁹. In recent years, many researchers have reported that combining TyG and obesity-related indices [body mass index (BMI), waist-to-height ratio (WHtR) and waist circumference (WC)] into TyG-related indices (TyG-BMI, TyG-WHtR and TyG-WC) can enhance the ability to identify IR^{20,21}. Additionally, the latest study discovered that TyG-related indices were generally more effective at predicting the progression of coronary artery calcification and identifying fatty liver than TyG^{22,23}. Despite this, the evidence regarding the superiority of TyG and TyG-related indices in assessing BMD is inconclusive.

Therefore, the present study aimed to analyze the correlation between TyG, TyG-related indices and total BMD through a large-scale cross-sectional study based on the National Health and Nutrition Examination study database (NHANES).

Methods

Data source and study population

The NHANES is a large, ongoing cross-sectional study conducted in the U.S. designed to provide objective health statistics and address emerging public health issues among the general population (http://www.cdc.gov/nchs/nhanes/)²⁴. All participants consented to the use of their data in the research, and the National Center for Health Statistics Institutional Review Board approved the survey techniques.

This study employed NHANES data from 2011 to 2018. In total, 39,156 participants completed the demographic survey, laboratory examination, and health condition questionnaire. The exclusion criteria were as follows: (1) Missing total BMD data (N = 21,282); (2) Missing data on fasting triglycerides (N = 11,259) and fasting glucose (N = 7); (3) Missing data on BMI (N = 22) and waist circumference (N = 43); (4) Patients with thyroid disease (N = 249), cancer or malignancy (N = 156) and nephropathy (N = 84); (5) Postmenopausal women (N = 462); (6) Missing data on total calcium (N = 38), low-density lipoproteins (N = 69), creatine phosphokinase (N = 15), total bilirubin (N = 5) and glycohemoglobin (N = 9). Ultimately, 5456 participants were enrolled in the research (Fig. 1). The accompanying NHANES variable codes and their utilization details are outlined in Supplementary Table S1.

BMD examination

Total BMD was measured using dual-energy X-ray absorption (DXA) on a Hologic QDR 4500A fan-beam densitometer²⁵. The screening tool is commonly used internationally to assess fragility-fracture risk. Radiologic technologists were certified and trained to administer DXA examinations. In the survey, DXA scans were available to all survey participants aged 8 and older, but they were not suitable for individuals weighing over 300 pounds, pregnant women, or those who had taken radiographic contrast material in the past 7 days.

Definitions of TyG and TyG-related indices

The indices in the study were defined as follows: WHtR is defined as WC divided by body height²⁶; TyG = ln [fast-ing triglyceride (mg/dL) × fasting glucose (mg/dL)/2]²⁷; TyG-BMI = TyG × BMI²⁸; TyG-WHtR = TyG × WHtR²³; TyG-WC = TyG × WC²⁸.





Covariates

Data on demographics, examination, and laboratory are covariates. Demographics data covered age (years), gender (male and female), race/ethnicity (Mexican American, non-Hispanic white, non-Hispanic black, other race/ethnicity), level of education (less than high school, high school or equivalent, college or above) and family income-poverty ratio (family PIR). Examination data included weight (kg), height (cm), body mass index (BMI, kg/m²) and waist circumference (WC, cm). Finally, laboratory data consisted of fasting glucose (mg/dL), fasting triglyceride (mg/dL), alkaline phosphatase (ALP, IU/L), blood urea nitrogen (BUN, mg/dL), creatine phosphokinase (CPK, IU/L), creatinine (mg/dL), phosphorus (mg/dL), total calcium (mg/dL), uric acid (mg/

dL), total bilirubin (mg/dL), glycohemoglobin (%), total cholesterol (mg/dL), high-density lipoprotein cholesterol (HDL-C, mg/dL), low-density lipoprotein cholesterol (LDL-C, mg/dL) and 250HD2+250HD3 (nmol/L).

Statistical analysis

Statistical analysis for this study was conducted using R (http://www.r-project.org) and EmpowerStats (http:// www.empowerstats.com), setting a significance level at P < 0.05. In accordance with NCHS analytical guidelines, we employed sample weights in all estimates to better represent the non-institutionalized civilian population in the USA. To investigate correlations between research variables and total BMD, participants were categorized into quartiles based on their total BMD values, which ranged from 0.670 to 1.728 g/cm². Univariate analyses and chi-square tests were then conducted to examine these correlations. Weighted multiple linear regression analysis analyses were utilized to explore the linear associations between TyG, TyG-related indices and total BMD. The multiple linear regression analysis were structured into three distinct models: Model 1 served as the baseline and involved no adjustments for any variables; Model 2 included adjustments for age, gender and race/ethnicity; Model 3 adjusted for a variety of factors, including age, gender, race/ethnicity, level of education, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 250HD2 + 250HD3. Subgroup analyses were also conducted, focusing specifically on variations across gender and race. In the final stage of our statistical analysis, smoothing curve fitting techniques and threshold effects evaluations were employed to assess any non-linear relationships between TyG, TyG-related indices and total BMD respectively.

Ethics approval and consent to participate

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. All analyses were based on data of the National Health and Nutrition Examination Survey (NHANES). The study was approved by the ethics review board of the National Center for Health Statistics. The detailed information located on the NHANES website. Written informed consent was obtained from each participant before their inclusion on the NHANES database. Detailed information on the ethics application and written informed consent are provided on the NHANES website.

Results

Baseline characteristics

After applying inclusion and exclusion criteria, 5456 participants were included in the study with a mean age of 30.33 ± 13.55 years. The sample comprised 55.65% males and 44.35% females, along with 17.19% Mexican–American people, 32.04% non-Hispanic white people, 21.13% non-Hispanic black people and 29.64% individuals from other racial backgrounds.

As shown in Table 1, the clinical characteristics of the participants are listed in stratified groups based on the four categories of total BMD. Compared to the bottom quartile, those in the top total BMD quartile were more likely to be male and older, had a higher proportion of non-Hispanic white and non-Hispanic black, exhibited a higher PIR and education level, and showed increased levels of TyG, TyG-BMI, TyG-WHtR, TyG-WC, weight, height, BMI, WC, fasting glucose, fasting triglyceride, BUN, CPK, creatinine, uric acid, total bilirubin, glycohemoglobin, total cholesterol, LDL-C and 25OHD2 + 25OHD3. Conversely, they demonstrated lower levels of ALP, phosphorus, total calcium and HDL-C (P < 0.05).

Association between TyG, TyG-related indices and total BMD

To investigate the relationships between TyG, TyG-related indices and total BMD, multiple linear regression models ware used. Table 2 presents the outcomes for three distinct models. All models exhibited a positive correlation between TyG, TyG-related indices and total BMD, TyG-BMI, TyG-WHtR and TyG-WC demonstrated a statistically correlated with total BMD (P < 0.0001). After adjusting for all confounding variables, one-unit increments in TyG, TyG-BMI, TyG-WHtR and TyG-WC were linked with 0.0124 g/cm², 0.0004 g/cm², 0.0116 g/cm², and 0.0001 g/cm² increase in total BMD, respectively. When TyG-BMI, TyG-WHtR and TyG-WC were grouped into quartiles, this association persisted statistically significant (P < 0.0001).

Subgroup analysis

A subgroup analysis was conducted to evaluate the stability of the relationship between TyG, TyG-related indices and total BMD across various demographic contexts. As illustrated in Tables 3, 4, 5 and 6, positive correlations between TyG-related indices and total BMD were significantly affected by gender stratification (P < 0.05 for interactions). Racial differences in absolute effect sizes were statistically significant in racially stratified subgroup analyses of TyG and TyG-related indices (P < 0.05 for interactions). Furthermore, positive correlations between TyG-BMI, TyG-WC and total BMD were significant not only unadjusted but also fully adjusted (P < 0.0001).

Non-linearity and saturation effect analysis between TyG, TyG-related indices and total BMD

As a result of smooth curve fitting, the non-linear association and saturation phenomenon between TyG, TyG-related indices, and total BMD have been delineated (Fig. 2). For TyG, TyG-BMI, TyG-WHtR and TyG-WC, the thresholds were 9.106, 193.9265, 4.065 and 667.5304, respectively (Tables 7, 8, 9, 10). The regression coefficient increases when TyG exceeds the threshold. By contrast, when TyG-BMI, TyG-WHtR and TyG-WC exceed their thresholds, the regression coefficient decreases accordingly. Furthermore, TyG-BMI and TyG-WC displayed saturation effects on total BMD, with saturation thresholds of 314.177 and 1022.0428, respectively.

Total BMD (g/cm ²)	Quartile 1 (0.670-1.018)	Quartile 2 (1.019–1.094)	Quartile 3 (1.095–1.167)	Quartile 4 (1.168-1.728)	P value
Age (years)	26.81±14.63	32.21±12.96	34.08±12.22	35.92±11.82	< 0.0001
Gender (%)					< 0.0001
Male	44.19	47.20	57.24	76.36	
Female	55.81	52.80	42.76	23.64	
Race/ethnicity (%)					< 0.0001
Mexican American	15.40	13.09	12.02	8.94	
Non-Hispanic White	56.08	59.23	61.18	57.98	
Non-Hispanic Black	6.87	7.11	10.27	19.82	
Other race/ethnicity	21.64	20.57	16.53	13.26	
Education level (%)					< 0.0001
Less than high school	50.56	28.58	21.52	17.38	
High school or equivalent	12.72	17.78	22.94	21.59	
College or above	36.72	53.64	55.54	61.03	
Family PIR	2.56 ± 1.64	2.55 ± 1.64	2.86±1.69	2.97 ± 1.64	< 0.0001
Weight (kg)	65.93 ± 18.85	77.93±21.39	81.68±19.86	88.45 ± 20.00	< 0.0001
Height (cm)	163.44 ± 9.15	167.61 ± 8.53	170.36 ± 8.94	174.12 ± 7.95	< 0.0001
BMI (kg/cm ²)	24.52 ± 6.13	27.69 ± 7.13	28.16±6.65	29.15±6.22	< 0.0001
WC (cm)	86.07±16.63	93.84 ± 17.40	95.40±16.58	98.19 ± 15.77	< 0.0001
Fasting glucose (mg/dL)	97.98 ± 14.60	100.44 ± 22.06	100.69 ± 22.75	104.99 ± 34.19	< 0.0001
Fasting triglyceride (mg/dL)	92.56±57.73	103.02 ± 62.96	101.87±63.33	105.12 ± 65.01	< 0.0001
ALP (IU/L)	117.46±91.24	74.95 ± 43.07	68.87±28.77	65.57±23.92	< 0.0001
BUN (mg/dL)	11.30±3.39	11.80 ± 3.56	12.36±3.70	13.28±4.18	< 0.0001
CPK (IU/L)	121.48±120.75	135.81±128.26	161.84±311.36	208.20±327.07	< 0.0001
Creatinine (mg/dL)	0.71 ± 0.16	0.80 ± 0.17	0.84 ± 0.17	0.92 ± 0.18	< 0.0001
Phosphorus (mg/dL)	4.05 ± 0.74	3.76 ± 0.59	3.67±0.55	3.63 ± 0.55	< 0.0001
Total calcium (mg/dL)	9.44 ± 0.42	9.38 ± 0.35	9.35±0.32	9.35 ± 0.33	< 0.0001
Uric acid (mg/dL)	4.96 ± 1.21	5.20 ± 1.25	5.44 ± 1.35	5.72 ± 1.35	< 0.0001
Total bilirubin (mg/dL)	0.62 ± 0.30	0.63 ± 0.32	0.67 ± 0.35	0.67 ± 0.34	< 0.0001
Glycohemoglobin (%)	5.33 ± 0.53	5.37 ± 0.70	5.39 ± 0.74	5.51 ± 1.03	< 0.0001
Total cholesterol (mg/dL)	173.32 ± 37.93	180.02 ± 39.14	181.93 ± 38.24	181.41 ± 36.04	< 0.0001
HDL-C (mg/dL)	54.65 ± 13.56	52.11 ± 13.42	52.82 ± 15.42	51.09 ± 13.21	< 0.0001
LDL-C (mg/dL)	100.18 ± 33.35	107.29 ± 34.14	108.73 ± 32.99	109.30 ± 32.27	< 0.0001
25OHD2+25OHD3 (nmol/L)	62.61 ± 22.47	63.49 ± 23.24	64.63 ± 24.00	65.60 ± 24.85	0.0075
TyG	8.24 ± 0.62	8.37 ± 0.63	8.36±0.63	8.42 ± 0.65	< 0.0001
TyG-BMI	203.71 ± 59.66	233.28 ± 68.14	237.00±65.09	246.60 ± 60.64	< 0.0001
TyG-WHtR	4.37 ± 1.01	4.72 ± 1.05	4.72 ± 1.04	4.77 ± 0.96	< 0.0001
TyG-WC	714.47 ± 172.34	790.26 ± 180.75	802.56 ± 176.23	830.79 ± 167.95	< 0.0001

Table 1. Basic characteristics of the research population based on total BMD quartiles. Mean ± SD for continuous variables: the *P* value was calculated by the weighted linear regression model. (percentage) for categorical variables: the *P* value was calculated by the weighted chi-square test. *Family PIR* the ratio of family income to poverty, *BMI* body mass index, *WC* waist circumference, *ALP* alkaline phosphatase, *BUN* blood urea nitrogen, *CPK* creatine phosphokinase, *HDL-C* high-density lipoprotein cholesterol, *LDL-C* low-density lipoprotein cholesterol, *TyG* triglyceride glucose index, *TyG-BMI* triglyceride glucose-body mass index, *TyG-WC* triglyceride glucose-waist circumference.

Supplementary analysis

To ensure the stability of the TyG and TyG-related indices data in NHANES, we performed supplementary correlation analyses between TyG and TyG-related indices and HOMA-IR as well as fasting insulin. The results revealed a positive correlation between TyG and TyG-related indices with both HOMA-IR and fasting insulin. This observation aligns with the findings of previous studies, affirming the consistency of our results with established research in the field (Supplementary Table S2–S3).

To ensure the relative stability of experimental results, missing data for Family PIR were retained, introducing potential bias in the outcomes of Model 3. To address this potential bias, additional subgroup analyses were conducted using all covariates except Family PIR (Supplementary Table S4–S7). In this subgroup analysis, no data were missing in the study population, revealing only a minor deviation compared to Model 3, and indicating no substantive impact on the experimental conclusions.

Exposure	Model 1 β (95% CI),	Model 2 β (95% CI),	Model 3 β (95% CI),			
	<i>P</i> value	<i>P</i> value	<i>P</i> value			
TyG (continuous)	0.0176 (0.0130, 0.0222)	0.0016 (- 0.0031, 0.0063)	0.0124 (0.0006, 0.0242)			
	<0.0001	0.4987	0.0390			
TyG (quartile)						
Q1	Reference	Reference	Reference			
Q2	0.0159 (0.0072, 0.0245)	0.0112 (0.0031, 0.0192)	0.0017 (- 0.0062, 0.0095)			
	0.0003	0.0065	0.6766			
Q3	0.0282 (0.0197, 0.0366)	0.0153 (0.0073, 0.0233)	0.0045 (- 0.0044, 0.0134)			
	< 0.0001	0.0002	0.3243			
Q4	0.0255 (0.0170, 0.0339)	- 0.0019 (- 0.0103, 0.0065)	- 0.0130 (- 0.0268, 0.0008)			
	<0.0001	0.6587	0.0648			
TyG-BMI (continuous)	0.0004 (0.0004, 0.0005)	0.0003 (0.0003, 0.0003)	0.0004 (0.0003, 0.0004)			
	<0.0001	< 0.0001	< 0.0001			
TyG-BMI (quartile)						
Q1	Reference	Reference	Reference			
Q2	0.0665 (0.0582, 0.0748)	0.0554 (0.0475, 0.0634)	0.0376 (0.0298, 0.0455)			
	< 0.0001	< 0.0001	<0.0001			
Q3	0.0848 (0.0766, 0.0930)	0.0633 (0.0551, 0.0715)	0.0539 (0.0452, 0.0627)			
	<0.0001	< 0.0001	< 0.0001			
Q4	0.0883 (0.0801, 0.0965)	0.0678 (0.0596, 0.0761)	0.0702 (0.0602, 0.0802)			
	<0.0001	< 0.0001	< 0.0001			
TyG-WHTR (continuous)	0.0155 (0.0126, 0.0183)	0.0080 (0.0051, 0.0109)	0.0116 (0.0076, 0.0156)			
	<0.0001	<0.0001	< 0.0001			
TyG-WHTR (quartile)						
Q1	Reference	Reference	Reference			
Q2	0.0466 (0.0380, 0.0552)	0.0402 (0.0320, 0.0483)	0.0274 (0.0195, 0.0354)			
	< 0.0001	< 0.0001	<0.0001			
Q3	0.0525 (0.0441, 0.0608)	0.0327 (0.0243, 0.0410)	0.0272 (0.0182, 0.0363)			
	<0.0001	< 0.0001	< 0.0001			
Q4	0.0539 (0.0454, 0.0623)	0.0332 (0.0246, 0.0418)	0.0347 (0.0239, 0.0455)			
	<0.0001	< 0.0001	< 0.0001			
TyG-WC (continuous)	0.0001 (0.0001, 0.0002)	0.0001 (0.0001, 0.0001)	0.0001 (0.0001, 0.0001)			
	<0.0001	< 0.0001	<0.0001			
TyG-WC (quartile)						
Q1	Reference	Reference	Reference			
Q2	0.0632 (0.0547, 0.0718)	0.0523 (0.0441, 0.0606)	0.0346 (0.0264, 0.0427)			
	<0.0001	< 0.0001	< 0.0001			
Q3	0.0756 (0.0673, 0.0839)	0.0537 (0.0452, 0.0622)	0.0470 (0.0378, 0.0562)			
	<0.0001	< 0.0001	< 0.0001			
Q4	0.0843 (0.0760, 0.0925)	0.0550 (0.0463, 0.0638)	0.0567 (0.0457, 0.0677)			
	< 0.0001	< 0.0001	<0.0001			

Table 2. Association between TyG, TyG-BMI, TyG-WHtR, TYG-WC and total BMD. Model 1: no covariates were adjusted. Model 2: age, gender, and race were adjusted. Model 3: age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted.

Discussion

To our knowledge, this is the first cross-sectional study in the United States to examine correlations between TyG, TyG-related indices and BMD, revealing the close relationship between IR and bone metabolism. It was found that IR evaluated by TyG and TyG-related indices was significantly positively correlated with BMD among Americans. This is consistent with previous studies, such as an ongoing prospective study that found TyG-BMI was positively related to BMD and geometry in non-diabetic middle-aged and elderly Chinese individuals²⁹. Moreover, a single-center retrospective study discovered that IR was positively correlated with BMD in post-menopausal women, as well as a protective factor for OP¹². As per another study, higher IR values were associated with higher BMD values in non-diabetic older adults³⁰.

Currently, the underlying mechanisms that explain the observed positive correlation between TyG, TyGrelated indices and total BMD remain incompletely understood. Several interconnected pathways could contribute to this association. Firstly, IR stimulates the secretion of insulin, and hyperinsulinemia leads to increased BMD. Insulin acts as a promoter of osteoblast proliferation and an inhibitor of osteoclast activity, thus playing an anabolic role in bone. In the state of IR, the body compensates for resistance in skeletal muscle, adipose tissue, and liver by increasing insulin secretion, leading to hyperinsulinemia. This elevated insulin level further boosts bone mass³¹. Secondly, IR may also influence bone metabolism through its impact on inflammatory responses and estrogen levels. Inflammatory mediators such as TNF- α , IL-1, and IL-9 have been shown to promote osteoclasts differentiation or inhibit the osteogenic differentiation of bone marrow mesenchymal stem cells via the activation

	Model 1	Model 2	Model 3
	β (95% CI), <i>P</i> value	β (95% CI), <i>P</i> value	β (95% CI), <i>P</i> value
Stratified by gender			
Male	0.0140 (0.0077, 0.0203)	0.0047 (- 0.0019, 0.0113)	0.0101 (- 0.0055, 0.0256)
	<0.0001	0.1622	0.2049
Female	- 0.0005 (- 0.0073, 0.0062)	- 0.0030 (- 0.0095, 0.0035)	0.0182 (0.0006, 0.0357)
	0.8730	0.3710	0.0430
P for interaction	0.0001	0.0504	0.3478
Stratified by race		~	
Mexican American	0.0301 (0.0199, 0.0404)	0.0075 (- 0.0032, 0.0181)	0.0366 (0.0090, 0.0642)
	< 0.0001	0.1688	0.0096
Non-Hispanic White	0.0225 (0.0145, 0.0305)	0.0012 (- 0.0071, 0.0094)	0.0123 (- 0.0080, 0.0326)
	< 0.0001	0.7834	0.2368
Non-Hispanic Black	0.0337 (0.0228, 0.0445)	0.0161 (0.0046, 0.0277)	0.0298 (0.0054, 0.0542)
	< 0.0001	0.0064	0.0168
Other race/ethnicity	0.0139 (0.0056, 0.0223)	- 0.0111 (- 0.0196, - 0.0027)	- 0.0056 (- 0.0265, 0.0154)
	0.0011	0.0095	0.6031
P for interaction	0.0054	0.0244	0.0198

Table 3. Subgroup analysis between TyG and total BMD. Model 1: no covariates were adjusted. Model 2: age, gender and race were adjusted. Model 3: age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted.

EDE-C and 25011D2 + 25011D5 were adjusted

	Model 1	Model 2	Model 3		
	6 (95% CI) P value	B (95% CI) P value	B (95% CI) P value		
Stratified by gender					
Male	0.0004 (0.0004, 0.0005)	0.0004 (0.0003, 0.0004)	0.0003 (0.0002, 0.0004)		
	<0.0001	< 0.0001	<0.0001		
Female	0.0003 (0.0003, 0.0004)	0.0002 (0.0002, 0.0003)	0.0004 (0.0003, 0.0005)		
	<0.0001	< 0.0001	< 0.0001		
<i>P</i> for interaction	0.0433	0.0139	0.0741		
Stratified by race					
Mexican American	0.0005 (0.0004, 0.0006)	0.0004 (0.0003, 0.0005)	0.0004 (0.0003, 0.0006)		
	< 0.0001	<0.0001	< 0.0001		
Non-Hispanic White	0.0004 (0.0003, 0.0005)	0.0003 (0.0002, 0.0003)	0.0003 (0.0002, 0.0004)		
	<0.0001	<0.0001	< 0.0001		
Non-Hispanic Black	0.0004 (0.0003, 0.0005)	0.0004 (0.0002, 0.0005)	0.0003 (0.0002, 0.0004)		
	<0.0001	<0.0001	< 0.0001		
Other race/ethnicity	0.0005 (0.0004, 0.0006)	0.0003 (0.0002, 0.0004)	0.0005 (0.0004, 0.0006)		
	<0.0001	< 0.0001	< 0.0001		
<i>P</i> for interaction	0.1024	0.0249	0.0002		

Table 4. Subgroup analysis between TyG-BMI and total BMD. Model 1: no covariates were adjusted. Model 2: age, gender and race were adjusted. Model 3: age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2+25OHD3 were adjusted.

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of the RANKL/RANK/OPG and Wnt signaling pathways³². Estrogen exerts a direct influence on osteocytes, osteoblasts, and osteoclasts, while also inhibiting osteoclast activation either directly or through osteoblasts and T-cells. The ultimate effect of estrogen on the skeletal system is a reduction in bone remodeling and resorption, while simultaneously preserving bone formation³³. Finally, the relationship between IR and bone metabolism is bidirectional, suggesting the potential for a cycle: IR impacts bone metabolism, and in turn, the state of bone health influences glucose metabolism³⁴⁻³⁸.

Furthermore, our study revealed thresholds between TyG, TyG-related indices and total BMD. As TyG exceeds the threshold, the regression coefficient increases. Conversely, when TyG-related indices exceed their thresholds, the regression coefficient decreases. The discrepancy may be caused by two factors. On the one hand, as IR progresses, insulin sensitivity significantly decreases, resulting in more insulin release³⁹. Insulin is a critical anabolic hormone in osteoblasts⁴⁰. Insulin action in osteoblasts stimulates mitosis, inhibits apoptosis, and prevents hyper-glycemia's deleterious effects on bone formation^{40,41}. Research have demonstrated that the stimulation of insulin receptors promotes proliferation and differentiation of osteoblasts in immature mice⁴². Mature osteoblasts in culture also express insulin receptors⁴³. Osteoblasts are affected by insulin through multiple signaling pathways, thus de-repressing proliferation⁴⁴, blocking the pro-apoptotic effect^{45,46}, and promoting anabolism⁴⁷. On the other hand, the involvement of obesity in IR prevents IGF-1 from binding to insulin receptors on osteoblasts, leading

	Model 1	Model 2	Model 3			
	β (95% CI), <i>P</i> value	β (95% CI), <i>P</i> value	β (95% CI), <i>P</i> value			
Stratified by gender	Stratified by gender					
Male	0.0149 (0.0108, 0.0190)	0.0076 (0.0032, 0.0120)	0.0036 (- 0.0019, 0.0091)			
	< 0.0001	0.0008	0.1962			
Female	0.0147 (0.0111, 0.0182)	0.0083 (0.0047, 0.0119)	0.0191 (0.0141, 0.0241)			
	< 0.0001	<0.0001	<0.0001			
P for interaction	0.9866	0.7375	0.0002			
Stratified by race						
Mexican American	0.0216 (0.0151, 0.0281)	0.0124 (0.0056, 0.0193)	0.0192 (0.0103, 0.0282)			
	< 0.0001	<0.0001	<0.0001			
Non-Hispanic White	0.0157 (0.0109, 0.0206)	0.0060 (0.0010, 0.0110)	0.0048 (- 0.0018, 0.0114)			
	< 0.0001	0.0184	0.1557			
Non-Hispanic Black	0.0174 (0.0109, 0.0240)	0.0128 (0.0058, 0.0198)	0.0117 (0.0032, 0.0202)			
	< 0.0001	0.0004	0.0070			
Other race/ethnicity	0.0183 (0.0128, 0.0238)	0.0077 (0.0022, 0.0133)	0.0232 (0.0160, 0.0304)			
	< 0.0001	0.0066	<0.0001			
P for interaction	0.4474	0.4230	0.0036			

Table 5. Subgroup analysis between TyG-WHtR and total BMD. Model 1: no covariates were adjusted. Model 2: age, gender and race were adjusted. Model 3: age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted.

	Model 1	Model 2	Model 3		
	β (95% CI), <i>P</i> value	β (95% CI), <i>P</i> value	β (95% CI), <i>P</i> value		
Stratified by gender					
Male	0.0001 (0.0001, 0.0002)	0.0001 (0.0001, 0.0001)	0.0001 (0.0001, 0.0001)		
	< 0.0001	< 0.0001	< 0.0001		
Female	0.0001 (0.0001, 0.0001)	0.0001 (0.0001, 0.0001)	0.0002 (0.0001, 0.0002)		
	< 0.0001	<0.0001	< 0.0001		
<i>P</i> for interaction	0.5747	0.2585	0.0027		
Stratified by race					
Mexican American	0.0002 (0.0002, 0.0002)	0.0001 (0.0001, 0.0002)	0.0002 (0.0001, 0.0002)		
	< 0.0001	<0.0001	<0.0001		
Non-Hispanic White	0.0001 (0.0001, 0.0002)	0.0001 (0.0000, 0.0001)	0.0001 (0.0000, 0.0001)		
	< 0.0001	<0.0001	< 0.0001		
Non-Hispanic Black	0.0001 (0.0001, 0.0002)	0.0001 (0.0001, 0.0001)	0.0001 (0.0001, 0.0001)		
	<0.0001	<0.0001	<0.0001		
Other race/ethnicity	0.0002 (0.0001, 0.0002)	0.0001 (0.0001, 0.0001)	0.0002 (0.0001, 0.0002)		
	<0.0001	< 0.0001	< 0.0001		
P for interaction	0.995	0.0260	0.0002		

Table 6. Subgroup analysis between TyG-WC and total BMD. Model 1: no covariates were adjusted. Model 2: age, gender and race were adjusted. Model 3: age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted.

to dysfunctional insulin signaling, which negatively affects bone remodeling^{48–50}. Obesity is also associated with the loss of Dock 7 protein and silence of Thy-1 expression, which results in higher bone resorption and higher levels of adipogenesis, leading to impaired bone formation^{48,51–53}.

In addition to the threshold effect, TyG-BMI and TyG-WC had a saturation effect on total BMD with saturation thresholds of 314.177 and 1022.0428. Saturation might be caused by genetic determinism and co-leadership interactions. Early in life, bone growth trajectories and peak bone mass were determined^{54,55}, which might account for the plateau in BMD values observed in adults after a limited increase. Pocock et al. explored the influence of genetics on BMD in 38 pairs of monozygotic twins and 27 pairs of dizygotic twins, revealing that genetics played a pivotal role in adult bone mass⁵⁶. It was determined that genetic factors accounted for 75% of the variance in BMD, regardless of gender^{57,58}. Additionally, acquired environmental factors such as calcium intake, estrogen intake, and physical exercise also had limitations in terms of increasing adult BMD⁵⁹⁻⁶¹. Furthermore, bone mass was also dominated by a complex interplay of multiple factors. Research has demonstrated a connection between adipose and bone tissues within the body, with numerous bioactive molecules maintaining bone homeostasis⁶². This bone-adipose axis may offer an explanation for the observed saturation effects. It has been found that adipose and bone tissues share a common stem cell precursor and are in a competitive relationship, with excessive fat accumulation resulting in decreased bone mass^{63,64}. Meanwhile, excessive fat accumulation has



Figure 2. Association between TyG, TyG-related indices and total BMD. Solid red line represents the smooth curve fit between variables. Blue bands represent the 95% confidence interval from the fit. Gender, race, age, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2+25OHD3 were adjusted.

Total BMD	β (95% CI)	P value
TyG		
Model I	0.0124 (0.0006, 0.0242)	0.0390
Model II		
Inflection point (K)	9.106	
<k 1<="" effect="" point="" td=""><td>0.0209 (0.0084, 0.0334)</td><td>0.0010</td></k>	0.0209 (0.0084, 0.0334)	0.0010
>K point effect 2	0.0752 (0.0422, 0.1083)	< 0.0001
Effect 2 minus effect1	0.0543 (0.0276, 0.0811)	< 0.0001
Predicted value of the equation at the folding point	1.1177 (1.1116, 1.1238)	
Log-likelihood ratio test		< 0.0001

Table 7. Saturation effect analysis of TyG on total BMD. Age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted. Model I, univariate linear regression; Model II, two-piecewise regression model.

Total BMD	β (95% CI)	P value
TyG-BMI		
Model I	0.0004 (0.0003, 0.0004)	< 0.0001
Model II		
Inflection point (K 1)	193.9265	
<k 1<="" effect="" point="" td=""><td>0.0013 (0.0011, 0.0015)</td><td>< 0.0001</td></k>	0.0013 (0.0011, 0.0015)	< 0.0001
>K point effect 2	0.0002 (0.0002, 0.0003)	< 0.0001
Effect 2 minus effect1	- 0.0011 (- 0.0013, - 0.0009)	< 0.0001
Predicted value of the equation at the folding point	1.1222 (1.1174, 1.1270)	
Log-likelihood ratio test		< 0.0001
Inflection point (K 2)	314.177	
<k 1<="" effect="" point="" td=""><td>0.0004 (0.0003, 0.0006)</td><td>< 0.0001</td></k>	0.0004 (0.0003, 0.0006)	< 0.0001
>K point effect 2	0.0000 (- 0.0001, 0.0002)	0.6523
Effect 2 minus effect1	- 0.0004 (- 0.0007, - 0.0001)	0.0025
Predicted value of the equation at the folding point	1.1386 (1.1294, 1.1479)	
Log-likelihood ratio test		0.0020

Table 8. Saturation effect analysis of TyG-BMI on total BMD. Age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted. Model I, univariate linear regression; Model II, two-piecewise regression model.

Total BMD	β (95% CI)	P value
TyG-WHtR		
Model I	0.0116 (0.0076, 0.0156)	< 0.0001
Model II		
Inflection point (K)	4.065	
<k 1<="" effect="" point="" td=""><td>0.0408 (0.0291, 0.0524)</td><td>< 0.0001</td></k>	0.0408 (0.0291, 0.0524)	< 0.0001
>K point effect 2	0.0062 (0.0017, 0.0106)	0.0065
Effect 2 minus effect1	- 0.0346 (- 0.0476, - 0.0216)	< 0.0001
Predicted value of the equation at the folding point	1.1169 (1.1117, 1.1221)	
Log-likelihood ratio test		< 0.0001

Table 9. Saturation effect analysis of TyG-WHtR on total BMD. Age, gender, race, education level, family PIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, total cholesterol, HDL-C, LDL-C and 25OHD2 + 25OHD3 were adjusted. Model I, univariate linear regression; Model II, two-piecewise regression model.

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been linked to decreased bone mass, as confirmed by animal models on high-fat diets^{65–67}. According to other studies, adipose tissue also secretes inflammatory mediators such as interleukin-1 and tumor necrosis factor- α^{68} , which inhibit BMD via the RANK/RANKL/OPG pathway⁶⁹. In summary, the saturation effects observed in TyG-BMI and TyG-WC could be attributable to a variety of factors. Despite these findings, the existing body of direct experimental evidence remains limited, necessitating further in-depth research.

Indeed, excessive IR and obesity were detrimental to the entire population. Firstly, excessive IR and obesity were associated with a number of bone-related disorders, such as increased bone fragility^{70,71} and increased risk of fractures^{19,30}. In addition, IR and obesity can lead to a variety of chronic diseases and complications, such as cardiovascular disease^{72,73}, T2DM^{74,75}, gallbladder disease^{76,77} and fatty liver^{78,79}. More seriously, high IR and excessive obesity might increase cancer risk^{80,81} and cancer-related mortality^{82,83}. Therefore, we believe that maintaining TyG and TyG-related indices at reasonable values might contribute to maintaining optimal BMD and reducing the risk of related diseases and complications.

In subgroup analysis, the racial subgroup showed significant differences in the effects of TyG and TyG-related indices on BMD, which might indicate racial differences in bone and mineral metabolism. This may be due to differences in insulin sensitivity and the relationship to triglyceride concentration among the different race/ ethnic groups. According to one research⁸⁴, African Americans and non-Hispanic Whites have greater insulin sensitivity. Moreover, African Americans have lower triglyceride levels than non-Hispanic Whites at a given level of IR. Intriguingly, a divergent study with a similar focus on adults, highlighting a negative correlation between TyG and BMD⁸⁵, introduces a notable contrast to our findings. It's crucial to acknowledge that our study included children but excluded postmenopausal females, potentially creating unique subsets within age and gender subgroups. In addition, gender had a significant effect on BMD related to TyG-related indices. This may

Total BMD	β (95% CI)	P value
TyG-WC		
Model I	0.0001 (0.0001, 0.0001)	< 0.0001
Model II		
Inflection point (K 1)	667.5304	
<k 1<="" effect="" point="" td=""><td>0.0004 (0.0003, 0.0005)</td><td>< 0.0001</td></k>	0.0004 (0.0003, 0.0005)	< 0.0001
>K point effect 2	0.0001 (0.0001, 0.0001)	< 0.0001
Effect 2 minus effect1	- 0.0003 (- 0.0004, - 0.0002)	< 0.0001
Predicted value of the equation at the folding point	1.1140 (1.1090, 1.1191)	
Log-likelihood ratio test		< 0.0001
Inflection point (K 2)	1022.0428	
<k 1<="" effect="" point="" td=""><td>0.0001 (0.0001, 0.0002)</td><td>< 0.0001</td></k>	0.0001 (0.0001, 0.0002)	< 0.0001
>K point effect 2	0.0000 (- 0.0001, 0.0001)	0.8095
Effect 2 minus effect1	- 0.0001 (- 0.0002, - 0.0001)	< 0.0001
Predicted value of the equation at the folding point	1.1562 (1.1500, 1.1625)	
Log-likelihood ratio test		< 0.0001

Table 10. Saturation effect analysis of TyG-WC on total BMD. Age, gender, race, education level, familyPIR, ALP, BUN, CPK, creatinine, phosphorus, total calcium, uric acid, total bilirubin, glycohemoglobin, totalcholesterol, HDL-C, LDL-C and 250HD2 + 250HD3 were adjusted. Model I, univariate linear regression;Model II, two-piecewise regression model.

be related to estrogen, which is an important regulator of body weight, body fat distribution and IRn^{86–89}. Given these intriguing disparities observed across racial and gender subgroups, further investigation is warranted. A thorough exploration of the underlying mechanisms driving these differences could offer valuable insights into the complex interplay between insulin resistance and bone health across diverse populations.

In summary, this was a successful large cross-sectional study that revealed a positive association between IR and BMD through TyG and TyG-related indices. Primarily, the utilization of a nationally representative cohort ensured that the results summarized the heterogeneity of the overwhelming majority of demographics in the USA. Furthermore, the large sample size enabled us to conduct subgroup analyses, classifying participants by race, thereby enhancing the generalizability of our findings. However, several limitations of this study merit cautious interpretation of the conclusions. Firstly, the cross-sectional design of the study precludes the establishment of a definitive causal link between TyG, TyG-related indices and BMD. Moreover, database constraints meant that comprehensive data on participants' lifestyle, dietary habits and bone metabolism indices, which might have impacted BMD. The inability to gather BMD data from various skeletons prevented the investigation of the consistency of the outcomes across skeletons. Finally, the absence of fracture data in our database meant we could not determine whether participants with lower TyG or TyG-related indices had a higher incidence of fractures compared to the general population.

Conclusion

This study revealed a positive association between TyG, TyG-related indices and total BMD. There was also a saturation phenomenon between TyG-BMI, TyG-WC and total BMD. TyG and TyG-related indices can serve as effective indicators for the prevention and treatment of OP. Maintaining optimal levels of TyG and TyG-related indices is critical for managing bone metabolism effectively.

Data availability

The survey data are publicly available on the Internet for data users and researchers throughout the world http://www.cdc.gov/nchs/nhanes/.

Received: 25 September 2023; Accepted: 9 February 2024 Published online: 20 February 2024

References

- Kanis, J. A., Cooper, C., Rizzoli, R. & Reginster, J. Y. European guidance for the diagnosis and management of osteoporosis in postmenopausal women. Osteoporos. Int. 30, 3–44. https://doi.org/10.1007/s00198-018-4704-5 (2019).
- Kanis, J. A., Delmas, P., Burckhardt, P., Cooper, C. & Torgerson, D. Guidelines for diagnosis and management of osteoporosis. The European Foundation for Osteoporosis and Bone Disease. Osteoporos. Int. 7, 390–406. https://doi.org/10.1007/bf01623782 (1997).
 - . Qaseem, A. *et al.* Treatment of low bone density or osteoporosis to prevent fractures in men and women: A clinical practice guideline update from the American College of Physicians. *Ann. Intern. Med.* **166**, 818–839. https://doi.org/10.7326/m15-1361 (2017).
- Kushchayeva, Y., Pestun, I., Kushchayev, S., Radzikhovska, N. & Lewiecki, E. M. Advancement in the treatment of osteoporosis and the effects on bone healing. J. Clin. Med. https://doi.org/10.3390/jcm11247477 (2022).
- 5. Dempster, D. W. Osteoporosis and the burden of osteoporosis-related fractures. Am. J. Manag. Care 17(Suppl 6), S164-169 (2011).
- Khosla, S. & Hofbauer, L. C. Osteoporosis treatment: Recent developments and ongoing challenges. *Lancet Diabetes Endocrinol.* 5, 898–907. https://doi.org/10.1016/S2213-8587(17)30188-2 (2017).

- Xue, S. et al. Age at attainment of peak bone mineral density and its associated factors: The National Health and Nutrition Examination Survey 2005–2014. Bone 131, 115163. https://doi.org/10.1016/j.bone.2019.115163 (2020).
- von Muhlen, D., Safii, S., Jassal, S. K., Svartberg, J. & Barrett-Connor, E. Associations between the metabolic syndrome and bone health in older men and women: The Rancho Bernardo Study. Osteopor. Int. 18, 1337–1344. https://doi.org/10.1007/s00198-007-0385-1 (2007).
- Haffner, S. M. et al. Prospective analysis of the insulin-resistance syndrome (syndrome X). Diabetes 41, 715–722. https://doi.org/ 10.2337/diab.41.6.715 (1992).
- 10. Grundy, S. M. Metabolic syndrome update. Trends Cardiovasc. Med. 26, 364-373. https://doi.org/10.1016/j.tcm.2015.10.004 (2016).
- 11. Kalimeri, M. *et al.* Association of insulin resistance with bone strength and bone turnover in menopausal Chinese–Singaporean women without diabetes. *Int. J. Environ. Res. Public Health* https://doi.org/10.3390/ijerph15050889 (2018).
- 12. Gu, P. *et al.* The metabolic score of insulin resistance is positively correlated with bone mineral density in postmenopausal patients with type 2 diabetes mellitus. *Sci. Rep.* **13**, 8796. https://doi.org/10.1038/s41598-023-32931-8 (2023).
- Wang, X., Jiang, L. & Shao, X. Association analysis of insulin resistance and osteoporosis risk in Chinese patients with T2DM. Ther. Clin. Risk Manag. 17, 909–916. https://doi.org/10.2147/tcrm.S328510 (2021).
- Pacini, G. & Bergman, R. N. MINMOD: A computer program to calculate insulin sensitivity and pancreatic responsivity from the frequently sampled intravenous glucose tolerance test. *Comput. Methods Prog. Biomed.* 23, 113–122. https://doi.org/10.1016/ 0169-2607(86)90106-9 (1986).
- 15. Bloomgarden, Z. T. Measures of Insulin Sensitivity. Clin. Lab. Med. 26, 611-633. https://doi.org/10.1016/j.cll.2006.06.007 (2006).
- Simental-Mendía, L. E., Rodríguez-Morán, M. & Guerrero-Romero, F. The product of fasting glucose and triglycerides as surrogate for identifying insulin resistance in apparently healthy subjects. *Metab. Syndr. Relat. Disord.* 6, 299–304. https://doi.org/10.1089/ met.2008.0034 (2008).
- 17. Guerrero-Romero, F. *et al.* The product of triglycerides and glucose, a simple measure of insulin sensitivity. Comparison with the euglycemic–hyperinsulinemic clamp. *J. Clin. Endocrinol. Metab.* **95**, 3347–3351. https://doi.org/10.1210/jc.2010-0288 (2010).
- Suren Garg, S., Kushwaha, K., Dubey, R. & Gupta, J. Association between obesity, inflammation and insulin resistance: Insights into signaling pathways and therapeutic interventions. *Diabetes Res Clin Pract* 200, 110691. https://doi.org/10.1016/j.diabres.2023. 110691 (2023).
- 19. Palermo, A. et al. BMI and BMD: The potential interplay between obesity and bone fragility. Int. J. Environ. Res. Public Health https://doi.org/10.3390/ijerph13060544 (2016).
- Lim, J., Kim, J., Koo, S. H. & Kwon, G. C. Comparison of triglyceride glucose index, and related parameters to predict insulin resistance in Korean adults: An analysis of the 2007–2010 Korean National Health and Nutrition Examination Survey. *PLoS One* 14, e0212963. https://doi.org/10.1371/journal.pone.0212963 (2019).
- 21. Er, L. K. et al. Triglyceride glucose-body mass index is a simple and clinically useful surrogate marker for insulin resistance in nondiabetic individuals. *PLoS One* 11, e0149731. https://doi.org/10.1371/journal.pone.0149731 (2016).
- Cho, Y. K. *et al.* Triglyceride glucose-waist circumference better predicts coronary calcium progression compared with other indices of insulin resistance: A longitudinal observational study. *J. Clin. Med.* https://doi.org/10.3390/jcm10010092 (2020).
- Malek, M., Khamseh, M. E., Chehrehgosha, H., Nobarani, S. & Alaei-Shahmiri, F. Triglyceride glucose-waist to height ratio: A novel and effective marker for identifying hepatic steatosis in individuals with type 2 diabetes mellitus. *Endocrine* 74, 538–545. https://doi.org/10.1007/s12020-021-02815-w (2021).
- 24. Fain, J. A. NHANES. Diabetes Educ. 43, 151. https://doi.org/10.1177/0145721717698651 (2017).
- Ouyang, Y. et al. Saturation effect of body mass index on bone mineral density in adolescents of different ages: A population-based study. Front. Endocrinol. (Lausanne) 13, 922903. https://doi.org/10.3389/fendo.2022.922903 (2022).
- Nevill, A. M., Stewart, A. D., Olds, T. & Duncan, M. J. A new waist-to-height ratio predicts abdominal adiposity in adults. *Res. Sports Med.* 28, 15–26. https://doi.org/10.1080/15438627.2018.1502183 (2020).
- Kim, J. A. et al. Association of the triglyceride and glucose index with low muscle mass: KNHANES 2008–2011. Sci. Rep. 11, 450. https://doi.org/10.1038/s41598-020-80305-1 (2021).
- Khamseh, M. E. *et al.* Triglyceride glucose index and related parameters (triglyceride glucose-body mass index and triglyceride glucose-waist circumference) identify nonalcoholic fatty liver and liver fibrosis in individuals with overweight/obesity. *Metab. Syndr. Relat. Disord.* 19, 167–173. https://doi.org/10.1089/met.2020.0109 (2021).
- Wen, Z. et al. Triglyceride glucose-body mass index is a reliable indicator of bone mineral density and risk of osteoporotic fracture in middle-aged and elderly nondiabetic Chinese individuals. J. Clin. Med. https://doi.org/10.3390/jcm11195694 (2022).
- Napoli, N. et al. Effect of insulin resistance on BMD and fracture risk in older adults. J. Clin. Endocrinol. Metab. 104, 3303–3310. https://doi.org/10.1210/jc.2018-02539 (2019).
- Conte, C., Epstein, S. & Napoli, N. Insulin resistance and bone: A biological partnership. Acta Diabetol. 55, 305–314. https://doi. org/10.1007/s00592-018-1101-7 (2018).
- Ponzetti, M. & Rucci, N. Updates on osteoimmunology: What's new on the cross-talk between bone and immune system. Front. Endocrinol. (Lausanne) 10, 236. https://doi.org/10.3389/fendo.2019.00236 (2019).
- Cauley, J. A. Estrogen and bone health in men and women. Steroids 99, 11–15. https://doi.org/10.1016/j.steroids.2014.12.010 (2015).
- Wei, J. et al. Bone-specific insulin resistance disrupts whole-body glucose homeostasis via decreased osteocalcin activation. J. Clin. Investig. 124, 1–13. https://doi.org/10.1172/jci72323 (2014).
- Lee, N. K. et al. Endocrine regulation of energy metabolism by the skeleton. Cell 130, 456–469. https://doi.org/10.1016/j.cell.2007. 05.047 (2007).
- Li, Z. et al. Glucose transporter-4 facilitates insulin-stimulated glucose uptake in osteoblasts. Endocrinology 157, 4094–4103. https://doi.org/10.1210/en.2016-1583 (2016).
- Zoch, M. L., Abou, D. S., Clemens, T. L., Thorek, D. L. & Riddle, R. C. In vivo radiometric analysis of glucose uptake and distribution in mouse bone. *Bone Res.* 4, 16004. https://doi.org/10.1038/boneres.2016.4 (2016).
- Huang, S. et al. Decreased osteoclastogenesis and high bone mass in mice with impaired insulin clearance due to liver-specific inactivation to CEACAM1. Bone 46, 1138–1145. https://doi.org/10.1016/j.bone.2009.12.020 (2010).
- Shanik, M. H. *et al.* Insulin resistance and hyperinsulinemia: Is hyperinsulinemia the cart or the horse?. *Diabetes Care* 31, S262–S268. https://doi.org/10.2337/dc08-s264 (2008).
- Thrailkill, K. M., Lumpkin, C. K. Jr., Bunn, R. C., Kemp, S. F. & Fowlkes, J. L. Is insulin an anabolic agent in bone? Dissecting the diabetic bone for clues. Am. J. Physiol. Endocrinol. Metab. 289, E735-745. https://doi.org/10.1152/ajpendo.00159.2005 (2005).
- Mieczkowska, A. *et al.* Alteration of the bone tissue material properties in type 1 diabetes mellitus: A Fourier transform infrared microspectroscopy study. *Bone* 76, 31–39. https://doi.org/10.1016/j.bone.2015.03.010 (2015).
- Thrailkill, K. M. et al. Bone formation is impaired in a model of type 1 diabetes. Diabetes 54, 2875–2881. https://doi.org/10.2337/ diabetes.54.10.2875 (2005).
- Thomas, D. M., Hards, D. K., Rogers, S. D., Ng, K. W. & Best, J. D. Insulin receptor expression in bone. *J. Bone Miner. Res.* 11, 1312–1320. https://doi.org/10.1002/jbmr.5650110916 (1996).
- Uchida, T. *et al.* Deletion of Cdkn1b ameliorates hyperglycemia by maintaining compensatory hyperinsulinemia in diabetic mice. *Nat. Med.* 11, 175–182. https://doi.org/10.1038/nm1187 (2005).

- Fang, X. *et al.* Regulation of BAD phosphorylation at serine 112 by the Ras-mitogen-activated protein kinase pathway. *Oncogene* 18, 6635–6640. https://doi.org/10.1038/sj.onc.1203076 (1999).
- 46. White, M. F. Insulin signaling in health and disease. Science 302, 1710–1711. https://doi.org/10.1126/science.1092952 (2003).
- Campos Pastor, M. M., López-Ibarra, P. J., Escobar-Jiménez, F., Serrano Pardo, M. D. & García-Cervigón, A. G. Intensive insulin therapy and bone mineral density in type 1 diabetes mellitus: A prospective study. Osteoporos. Int. 11, 455–459. https://doi.org/ 10.1007/s001980070114 (2000).
- Imerb, N., Thonusin, C., Chattipakorn, N. & Chattipakorn, S. C. Aging, obese-insulin resistance, and bone remodeling. *Mech. Ageing Dev.* 191, 111335. https://doi.org/10.1016/j.mad.2020.111335 (2020).
- Rosen, C. J. & Motyl, K. J. No bones about it: Insulin modulates skeletal remodeling. Cell 142, 198–200. https://doi.org/10.1016/j. cell.2010.07.001 (2010).
- 50. Cao, J. J. Effects of obesity on bone metabolism. J. Orthop. Surg. Res. 6, 30. https://doi.org/10.1186/1749-799X-6-30 (2011).
- Barker, T. H. & Hagood, J. S. Getting a grip on Thy-1 signaling. Biochim. Biophys. Acta (BBA) Mol. Cell Res. 1793, 921–923. https:// doi.org/10.1016/j.bbamcr.2008.10.004 (2009).
- Motyl, K. J. et al. Altered thermogenesis and impaired bone remodeling in Misty mice. J. Bone Miner. Res. 28, 1885–1897. https:// doi.org/10.1002/jbmr.1943 (2013).
- Paine, A. et al. Thy1 is a positive regulator of osteoblast differentiation and modulates bone homeostasis in obese mice. FASEB J. 32, 3174–3183. https://doi.org/10.1096/fj.201701379R (2018).
- Jones, G. & Dwyer, T. Birth weight, birth length, and bone density in prepubertal children: Evidence for an association that may be mediated by genetic factors. *Calcif. Tissue Int.* 67, 304–308. https://doi.org/10.1007/s002230001148 (2000).
- Weiler, H. A., Yuen, C. K. & Seshia, M. M. Growth and bone mineralization of young adults weighing less than 1500 g at birth. Early Hum. Dev. 67, 101–112. https://doi.org/10.1016/s0378-3782(02)00003-8 (2002).
- Pocock, N. A. et al. Genetic determinants of bone mass in adults. A twin study. J. Clin. Investig. 80, 706–710. https://doi.org/10. 1172/jci113125 (1987).
- Flicker, L. et al. Bone density determinants in elderly women: A twin study. J. Bone Miner. Res. 10, 1607–1613. https://doi.org/10. 1002/jbmr.5650101102 (1995).
- Videman, T. et al. Heritability of BMD of femoral neck and lumbar spine: A multivariate twin study of Finnish men. J. Bone Miner. Res. 22, 1455–1462. https://doi.org/10.1359/jbmr.070606 (2007).
- Tønnesen, R., Schwarz, P., Hovind, P. H. & Jensen, L. T. Physical exercise associated with improved BMD independently of sex and vitamin D levels in young adults. *Eur. J. Appl. Physiol.* 116, 1297–1304. https://doi.org/10.1007/s00421-016-3383-1 (2016).
- Watts, N. B., Nolan, J. C., Brennan, J. J. & Yang, H.-M., Group, E. S. O. S. Esterified estrogen therapy in postmenopausal women. Relationships of bone marker changes and plasma estradiol to BMD changes: A two-year study. *Menopause* 7(6), 375–382. https:// doi.org/10.1097/00042192-200011000-00002 (2000).
- Hirsch, C. ACP Journal Club Review: Dietary or supplemental calcium increase BMD by ≤ 1.8% in persons > 50 years of age. Ann. Intern. Med. 164, Jc5. https://doi.org/10.7326/acpjc-2016-164-2-005 (2016).
- 62. Gómez-Ambrosi, J., Rodríguez, A., Catalán, V. & Frühbeck, G. The bone-adipose axis in obesity and weight loss. *Obes. Surg.* 18, 1134–1143. https://doi.org/10.1007/s11695-008-9548-1 (2008).
- Gregoire, F. M., Smas, C. M. & Sul, H. S. Understanding adipocyte differentiation. *Physiol. Rev.* 78, 783–809. https://doi.org/10. 1152/physrev.1998.78.3.783 (1998).
- Akune, T. et al. PPARgamma insufficiency enhances osteogenesis through osteoblast formation from bone marrow progenitors. J. Clin. Investig. 113, 846–855. https://doi.org/10.1172/jci19900 (2004).
- Halade, G. V., El Jamali, A., Williams, P. J., Fajardo, R. J. & Fernandes, G. Obesity-mediated inflammatory microenvironment stimulates osteoclastogenesis and bone loss in mice. *Exp. Gerontol.* 46, 43–52. https://doi.org/10.1016/j.exger.2010.09.014 (2011).
- Iwaniec, U. T. et al. Body mass influences cortical bone mass independent of leptin signaling. Bone 44, 404–412. https://doi.org/ 10.1016/j.bone.2008.10.058 (2009).
- Halade, G. V., Rahman, M. M., Williams, P. J. & Fernandes, G. High fat diet-induced animal model of age-associated obesity and osteoporosis. J. Nutr. Biochem. 21, 1162–1169. https://doi.org/10.1016/j.jnutbio.2009.10.002 (2010).
- Abrahamsen, B., Bonnevie-Nielsen, V., Ebbesen, E. N., Gram, J. & Beck-Nielsen, H. Cytokines and bone loss in a 5-year longitudinal study–hormone replacement therapy suppresses serum soluble interleukin-6 receptor and increases interleukin-1-receptor antagonist: The Danish Osteoporosis Prevention Study. J. Bone Miner. Res. 15, 1545–1554. https://doi.org/10.1359/jbmr.2000.15.8. 1545 (2000).
- Khosla, S. Minireview: The OPG/RANKL/RANK system. Endocrinology 142, 5050–5055. https://doi.org/10.1210/endo.142.12. 8536 (2001).
- Dolan, E., Swinton, P. A., Sale, C., Healy, A. & O'Reilly, J. Influence of adipose tissue mass on bone mass in an overweight or obese population: Systematic review and meta-analysis. *Nutr. Rev.* 75, 858–870. https://doi.org/10.1093/nutrit/nux046 (2017).
- Cipriani, C. et al. The interplay between bone and glucose metabolism. Front. Endocrinol. (Lausanne) 11, 122. https://doi.org/10. 3389/fendo.2020.00122 (2020).
- Kim, M. S. et al. Association between adiposity and cardiovascular outcomes: An umbrella review and meta-analysis of observational and Mendelian randomization studies. Eur. Heart J. 42, 3388–3403. https://doi.org/10.1093/eurheartj/ehab454 (2021).
- Hill, M. A. et al. Insulin resistance, cardiovascular stiffening and cardiovascular disease. Metabolism 119, 154766. https://doi.org/ 10.1016/j.metabol.2021.154766 (2021).
- Scheen, A. J. & Van Gaal, L. F. Combating the dual burden: Therapeutic targeting of common pathways in obesity and type 2 diabetes. *Lancet Diabetes Endocrinol.* 2, 911–922. https://doi.org/10.1016/s2213-8587(14)70004-x (2014).
- 75. Taylor, R. Type 2 diabetes: Etiology and reversibility. Diabetes Care 36, 1047-1055. https://doi.org/10.2337/dc12-1805 (2013).
- Cortés, V. A., Barrera, F. & Nervi, F. Pathophysiological connections between gallstone disease, insulin resistance, and obesity. Obes. Rev. 21, e12983. https://doi.org/10.1111/obr.12983 (2020).
- Nervi, F. et al. Gallbladder disease is associated with insulin resistance in a high risk Hispanic population. J. Hepatol. 45, 299–305. https://doi.org/10.1016/j.jhep.2006.01.026 (2006).
- Fabbrini, E., Sullivan, S. & Klein, S. Obesity and nonalcoholic fatty liver disease: Biochemical, metabolic, and clinical implications. *Hepatology* 51, 679–689. https://doi.org/10.1002/hep.23280 (2010).
- Marušić, M., Paić, M., Knobloch, M. & Liberati Pršo, A. M. NAFLD, insulin resistance, and diabetes mellitus type 2. Can. J. Gastroenterol. Hepatol. 2021, 6613827. https://doi.org/10.1155/2021/6613827 (2021).
- Chiefari, E. et al. Insulin resistance and cancer: In search for a causal link. Int. J. Mol. Sci. https://doi.org/10.3390/ijms222011137 (2021).
- Iyengar, N. M., Gucalp, A., Dannenberg, A. J. & Hudis, C. A. Obesity and cancer mechanisms: Tumor microenvironment and inflammation. J. Clin. Oncol. 34, 4270–4276. https://doi.org/10.1200/jco.2016.67.4283 (2016).
- Majumder, K., Gupta, A., Arora, N., Singh, P. P. & Singh, S. Premorbid obesity and mortality in patients with pancreatic cancer: A systematic review and meta-analysis. *Clin. Gastroenterol. Hepatol.* 14, 355–368.e; quiz e332. https://doi.org/10.1016/j.cgh.2015. 09.036 (2016).
- Farooki, A. & Schneider, S. H. Insulin resistance and cancer-related mortality. J. Clin. Oncol. 25, 1628–1629; author reply 1629–1630, https://doi.org/10.1200/jco.2007.10.9637 (2007).

- Raygor, V. et al. Impact of race/ethnicity on insulin resistance and hypertriglyceridaemia. Diabetes Vasc. Dis. Res. 16, 153–159. https://doi.org/10.1177/1479164118813890 (2019).
- Zhan, H., Liu, X., Piao, S., Rong, X. & Guo, J. Association between triglyceride-glucose index and bone mineral density in US adults: A cross sectional study. J. Orthop. Surg. Res. 18, 810. https://doi.org/10.1186/s13018-023-04275-6 (2023).
- Brussaard, H. E., Gevers Leuven, J. A., Frölich, M., Kluft, C. & Krans, H. M. Short-term oestrogen replacement therapy improves insulin resistance, lipids and fibrinolysis in postmenopausal women with NIDDM. *Diabetologia* 40, 843–849. https://doi.org/10. 1007/s001250050758 (1997).
- Brown, L. M. & Clegg, D. J. Central effects of estradiol in the regulation of food intake, body weight, and adiposity. J. Steroid Biochem. Mol. Biol. 122, 65–73. https://doi.org/10.1016/j.jsbmb.2009.12.005 (2010).
- Brown, L. M., Gent, L., Davis, K. & Clegg, D. J. Metabolic impact of sex hormones on obesity. *Brain Res.* 1350, 77–85. https://doi. org/10.1016/j.brainres.2010.04.056 (2010).
- Meyer, M. R., Clegg, D. J., Prossnitz, E. R. & Barton, M. Obesity, insulin resistance and diabetes: Sex differences and role of oestrogen receptors. *Acta Physiol. (Oxf.)* 203, 259–269. https://doi.org/10.1111/j.1748-1716.2010.02237.x (2011).

Author contributions

N.T. collected data. N.T., S.C., H.W.H., and J.J. organized the study and performed the statistical analysis. Z.W.L. drafted the manuscript, to which all authors contributed, and approved the final version for publication.

Funding

This work was supported by the Elderly Health Research Project of Jiangsu Commission of Health (LKZ2022008), the Natural Science Foundation of Nanjing University Of Chinese Medicine (XZR2021060), and the Foundation of The Second Affiliated Hospital of Nanjing University of Chinese Medicine (SEZ202003).

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at https://doi.org/ 10.1038/s41598-024-54192-9.

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