



OPEN Waterlogging increases greenhouse gas release and decreases yield in winter rapeseed (*Brassica napus* L.) seedlings

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A sustainable future depends on increasing agricultural carbon (C) and nitrogen (N) sequestration. Winter rapeseeds are facing severe yield loss after waterlogging due to the effects of extreme rainfall, especially in the seedling stage, where rainfall is most sensitive. Uncertainty exists over the farming greenhouse gas (GHG) release of rapeseed seedlings following the onset of waterlogging. The effect of waterlogging on GHG release and leaf gas exchange in winter rapeseed was examined in a pot experiment. The experiment included waterlogging treatments lasting 7-day and 21-day and normal irrigation as a control treatment. According to our findings, (1) The ecosystem of rapeseed seedlings released methane (CH₄) and nitrous oxide (N₂O) in a clear up change that was impacted by ongoing waterlogging. Among them, N₂O release had a transient rise during the early stages under the effect of seedling fertilizer. (2) The net photosynthetic rate, transpiration rate, stomatal conductance, plant height, soil moisture, and soil oxidation–reduction potential of rapeseed all significantly decreased due to the ongoing waterlogging. However, rapeseed leaves showed a significant increase in intercellular carbon dioxide (CO₂) concentration and leaf chlorophyll content values after waterlogging. Additionally, the findings demonstrated an extremely significant increase in the sustained-flux global warming potential of the sum CO₂-eq of CH₄ and N₂O throughout the entire waterlogging stress period. Therefore, continuous waterlogging can increase C and N release from rapeseed seedlings ecosystem and decrease yield. Therefore, we suggest increasing drainage techniques to decrease the release of agricultural GHGs and promote sustainable crop production.

One of the significant challenges that humanity is facing is climate change. In the Paris climate agreement, the UN member states formalized the world's commitment to reduce greenhouse gas (GHG) emissions to hold global warming to well below 2 °C and possibly below 1.5 °C in response to global warming¹. Carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O) are the three gases that contribute 66, 20, and 10%, respectively, to global warming^{2,3}. Since the pre-industrial era (280 ppm), atmospheric CO₂ concentrations have increased; by 2021, they will reach 415.7 ppm⁴. It will reach 1000 ppm this century⁵. CH₄ and N₂O are among the most significant GHGs in the atmosphere after CO₂ because of their significantly greater radiative forcing effects⁶.

It is known that agricultural activities account for 30% of the world's anthropogenic GHG release⁵. Agricultural soil has been the main source of N₂O and CH₄ emissions, accounting for approximately 66% and 50% of total emissions, respectively^{7,8}. Currently, methanogens are known to produce CH₄ through hydrogenotrophic, acetoclastic, and methylotrophic methanogenesis⁹. Although N₂O can be produced under both anaerobic conditions (via denitrification) and aerobic conditions (via nitrification), the majority of N₂O production occurs in waterlogged soils¹⁰.

Rapeseed (*Brassica napus* L.) is one of the most significant oilseed crops in the world. Rapeseed is cultivated on 67 million hectares in China yearly, yielding 4.5 million tons of seeds¹¹. The Yangtze River Basin is the largest rapeseed-producing region in China¹². Extreme rainfall has recently emerged as an issue for rapeseed production

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in the Yangtze River Basin¹³. Rapeseed, particularly, experienced waterlogging at the seedling and flowering stages, which negatively impacted grain yield¹⁴.

The seedling stage of a plant's life cycle is when most plants are most sensitive to environmental change. Seedlings in such environments may sustain hypoxia and anoxia damages due to a notable reduction in gas diffusion in floodwaters after waterlogging¹⁵. Additionally, under anaerobic conditions, fermentation converts pyruvate into lactate or ethanol, and metabolic processes like photosynthesis, respiration, and ion transport are significantly impaired, slowing growth^{16–19}. Rapeseed seedlings have significantly lowered plant height, leaf area, total root length, and dry matter^{14,20,21}. The overall result was a decrease in the yield of rapeseed grains²². Studies demonstrate that after waterlogging, rapeseed seedlings exhibited growth retardation and delayed development²³. Additionally, Froelking, et al.²⁴ have suggested that flooded rice fields release methane, while during the transition from flooded to drained state, nitrous oxide is emitted. It has been reported that adopting alternate wetting and drying (AWD) technique can effectively reduce methane emissions under field conditions in rice fields²⁵. However, far less is known about GHGs releases in rapeseed ecosystems, especially when subjected to waterlogging during the seedling stage. In the seedling stage of rapeseed, we hypothesized that waterlogging could increase CH₄ and N₂O emissions. Our objectives were as follows: (1) to monitor the water content and redox potential of the soil in the root zone, simultaneously to measure agronomic traits such as plant height, leaf chlorophyll content (SPAD) value, and grain yield; (2) to compare leaf photosynthetic properties; (3) to evaluate the potential impacts of the CH₄ and N₂O emissions on global warming.

Materials and methods

Experimental design

The experiment was designed as follows. From November 2021 to June 2022, a pot experiment was carried out in the experimental farm of East China Normal University, Shanghai, China (N31° 02' 10", E121° 26' 55"). However, abundant rainfall in the region is unevenly distributed throughout the seasons due to the subtropical monsoon climate. About 16 °C is the average annual temperature.

The soil (pH 7.3) included 18.60 g kg⁻¹ of soil organic matter, 1.12 g kg⁻¹ of total nitrogen, 24.64 mg kg⁻¹ of available phosphorus, and 108.08 mg kg⁻¹ of available potassium. A single-factor, entirely randomized design was used in this experiment to investigate the effects of various waterlogging times on rapeseed. On October 28, 2021, rapeseed seeds were sown. After 80 days from sowing (January 14, 2022), the waterlogging was conducted. In the test, three treatments—waterlogging lasting 7-day and 21-day and one control treatment (normal irrigation, CK)—was established. Hunan Agricultural University provided the rapeseed cultivar used for oilseeds, Zhongyou 821. Rows were 0.15 m apart, and plants were spaced 0.1 m apart. Seeds were manually spread by direct seeding in November, and harvested in May. Only the healthiest seedling per location was kept within 20d of germination. The pot's dimensions were 0.635 × 0.425 × 0.400 m. Each treatment was repeated four times.

The soil sample for the test was collected from the plow layer of a rice-rapeseed rotation field at Wujing farm, Minghang, Shanghai. After the soil was air-dried, thoroughly mixed, and 5.0 mm sieved, 75 kg of soil was added to each plastic pot. Organic manure (100 g pot⁻¹, November 2, 2021) was applied and homogenized by hand mixing as base fertilizers. Each pot received an equal amount (20 g) of seedling fertilizer (nitrogen: phosphorus: potassium = 15: 15: 15, January 20, 2022). The experiment was carried out in a partially controlled environment by growing the rapeseed seedling in a rainout shelter, stimulating controlled irrigation, and neglecting the impact of unpredictable rainfall. A consistent water level of at least 3 cm was maintained throughout the waterlogging, which lasted for 7d or 21d. After the waterlogging treatment, the water in the pot was released, restoring normal water management when soil moisture in the plastic basin reached 20–30% of the field's water capacity. The other cultivation and management measures were the same as the regular field management measures, except for water control.

Sampling and analytical methods

Collecting plant samples Plant height and SPAD (A portable chlorophyll meter, SPAD 502 Plus, Konica Minolta Optics, Japan) were measured on the 5th, 21st, and 42nd days Midway through May, when the seeds reached maturity (mid-May), they were harvested from each pot and sun-dried to a set weight.

Key environmental parameters After field sampling, the soil moisture was measured by Decagon EC-5 sensors (Decagon Devices, Pullman, WA, USA). An oxidation–reduction potential (ORP) meter (FJA-6, Nanjing, China) was used to measure soil ORP.

Leaf gas exchange measurements The Li-6800R portable photosynthesis system (Lincoln, NE, USA) was used to measure the leaf gas exchange measurements of net photosynthetic rate (A , $\mu\text{mol m}^{-2} \text{s}^{-1}$), stomatal conductance (G_{sw} , $\text{mol m}^{-2} \text{s}^{-1}$), intercellular CO₂ concentration (C_i , $\text{mol m}^{-2} \text{s}^{-1}$), and transpiration rate (E , $\text{mol m}^{-2} \text{s}^{-1}$). The following are the precise operations: The light-saturated photosynthetic photon flux was 1200 $\mu\text{mol m}^{-2} \text{s}^{-1}$, the reference CO₂ concentration was 400 $\mu\text{mol mol}^{-1}$, and the airflow rate into the leaf cuvette was 500 mL min⁻¹. The temperature (15 °C) and humidity (60%) were kept constant. On February 26, 2022 (42nd days after exposure to water stress), measurements were taken from 8:30 to 11:30 AM. Each seedling was periodically sampled over time, with the second or third most recently matured leaf taken from the apical meristem. Each treatment was replicated four times for a total of 20 leaves per pot.

Collecting gas samples In situ CO₂, CH₄, and N₂O fluxes were monitored with a static chamber during the waterlogging using a high-precision GHG analyzer (CO₂/CH₄/N₂O/H₂O Analyzer; Picarro-G2508, Picarro Inc., USA). Poly methyl methacrylate (PMMA) was used to create the static chamber with a thickness of 0.30 cm. The chamber measured 0.2 m in length, 0.15 m in width, and 0.45 m in height. The gas samples were collected in real-time on 5th (January 19, 2022), 7th (January 21, 2022), 10th (January 24, 2022), 16th (January 30, 2022), and 42nd (February 26, 2022) days after exposure to water stress. All gas samples were collected in the field

from 9:00 to 11:00 AM. In addition, within a sampling date, the gas sampling time interval was 5 min and the measurement frequency was 1 Hz.

GHG fluxes and sustained-flux global warming potential calculations

GHG fluxes were calculated using the following formula:

$$F = \frac{dc}{dt} \times \frac{PV}{RAT} \quad (1)$$

where F is the GHG flux ($\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$); dc/dt is the rate of change of GHG concentration (ppm) with time t (s); P is the air pressure, the standard is 101.2237×10^3 (Pa); V is the effective volume (m^3) of the static closed chamber; R is the gas constant, defaulted to 8.3144 ($\text{J mol}^{-1} \text{K}^{-1}$); A is the chamber coverage (m^2); and T is the average soil temperature ($T = 273.15$ °C)²⁶.

This study estimated the dynamics of total radioactive forcing using the sustained-flux global warming potential (SGWP). The total emission of CH_4 and N_2O was calculated in mass CO_2 equivalents (CH_4 : 45; N_2O : 270) over a time horizon of 100 years²⁷. We only calculated daytime (8–18 h) SGWP scaled to a day due to a lack of nighttime flux measurement. The following formula was used to determine how much CH_4 and N_2O emissions contribute to SGWP:

$$SGWP_{Ratio} = \frac{SGWP_{(CH_4+N_2O)}}{SGWP} = \frac{(45 \times F_{CH_4}) + (270 \times F_{N_2O})}{[F_{CO_2} + (45 \times F_{CH_4}) + (270 \times F_{N_2O})]} \quad (2)$$

where F_{CO_2} , F_{CH_4} , and F_{N_2O} are mass flux in units (e.g., $\mu\text{g CO}_2 \text{ m}^{-2} \text{ s}^{-1}$); $SGWP_{(CH_4+N_2O)}$, expressed as CO_2 equivalents, are 45 and 270, respectively, multiplied by their respective flux values ($\mu\text{g CO}_2 \text{ m}^{-2} \text{ s}^{-1}$); $SGWP$ represents the total greenhouse gas warming potential expressed as CO_2 equivalents ($\mu\text{g CO}_2 \text{ m}^{-2} \text{ s}^{-1}$). The $SGWP_{ratio}$ is the ratio of $SGWP_{(CH_4+N_2O)}$ to the total $SGWP$.

Statistical analysis

Microsoft Excel 2018 for Windows was used to tabulate the data. Data analysis was performed using SPSS (IBM SPSS 23.0, SPSS Inc). One-way analysis of variance (ANOVA) was used to determine significant intergroup differences of each parameter. GraphPad Prism 8 software was used to create the graphics.

Results

Effects of waterlogging stress on plant and soil characteristics in rapeseed seedlings

The variation in SPAD, plant height, soil moisture, and ORP values during waterlogging is depicted in Fig. 1. Plant height revealed growth retardation as waterlogging stress treatment time increased. Compared to CK, the waterlogged plant's height was significantly lowered ($P < 0.05$, Fig. 1A) after waterlogging 21-day treatment. Plant height decreased significantly ($P < 0.05$, Fig. 1A) on waterlogging 7-day and 21-day treatments after waterlogging 21st days (February 4, 2022). The ORP in all the waterlogged soils exhibited similar trends, with declining values over time and lower average values in the waterlogging 21-day treatment ($P > 0.05$, Fig. 1C). The SPAD value of the leaves increased ($P > 0.05$, Fig. 1B) after waterlogging during the seedling stage. Significant differences between waterlogging 7-day (or 21-day) treatments and CK were observed in terms of the soil moisture ($P > 0.05$, Fig. 1A). After waterlogging, the soil moisture gradually reduced.

Effects of waterlogging stress on the gas exchange in rapeseed leaves

A , E , G_{sw} , and C_i variations during waterlogging are shown in Fig. 2. On waterlogging 7-day and 21-day treatments, A was significantly ($P < 0.05$, Fig. 2A) decreased compared to the control. A was at its lowest point at waterlogging 21-day treatment, although there was no significant difference between waterlogging 7-day and waterlogging 21-day (Fig. 2A). Furthermore, waterlogging 21-day treatment showed a significant decrease in E compared to the control, although waterlogging 7-day treatment showed no significant difference ($P > 0.05$, Fig. 2B) from the control, with the waterlogging 21-day treatment mark seeing a significant increase. C_i in the control was lower than in the 7-day and 21-day waterlogging treatments, with the waterlogging 21-day treatment mark significantly increasing ($P > 0.05$, Fig. 2C). All waterlogging treatments demonstrated the opposite tendency of the C_i , with a significant decrease in G_{sw} at waterlogging 21-day treatment compared to the control ($P > 0.05$, Fig. 2D).

Effects of waterlogging stress on GHGs release in the rapeseed ecosystem

The effects of waterlogging on total GHG emissions are shown in Fig. 3. CO_2 flux increased initially and then decreased as the waterlogging period increased (Fig. 3A). Throughout the waterlogging period, no statistically significant differences in CH_4 flux were found across all groups ($P > 0.05$, Fig. 3B). At the end of waterlogging, the CH_4 flux was a trend for higher waterlogging 7-day and 21-day treatments, it did reach significance ($P < 0.05$, Fig. 3B). However, the rapeseed field in the waterlogging 7-day and 21-day treatments increased significantly N_2O emissions ($P < 0.05$, Fig. 3C) compared to the control. The N_2O fluxes of all groups had a wave change trend of initially increasing, then decreasing, and then increased during the observation period. In contrast, the overall treatments N_2O fluxes increased after fertilization on the 6th days (January 20, 2022), and treatments for waterlogging 7-day and 21-day treatments increased more quickly than the control. Subsequently, the N_2O fluxes of both waterlogging 7-day and 21-day treatments drastically increased compared to the control, reaching significant levels after waterlogging lasting 10th days (January 24, 2022). However, after waterlogging developed, the combined CO_2 -eq emissions from CH_4 and N_2O played a role in providing negative feedback. $SGWP_{ratio}$

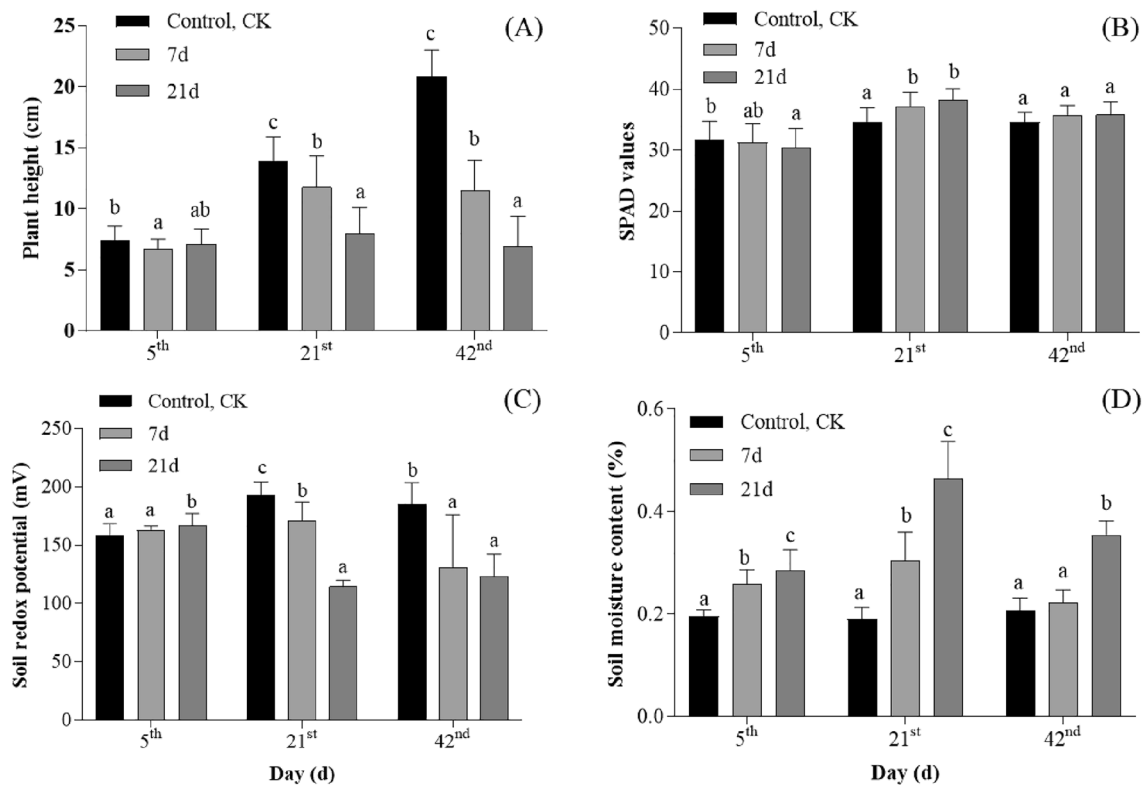


Figure 1. Effects of waterlogging stress on plant and soil characteristics in rapeseed seedlings. Treatment consisted of one control treatment, normal irrigation (control, CK), and two treatments: waterlogging lasting 7-day and 21-day. Significant differences among sets at the maximum points are denoted by lowercase letters above each bar ($P < 0.05$). (A) Represent plant height, (B) represent SPAD values, (C) represent soil redox potential, (D) represent soil moisture. The x-axis denotes the duration in days post-waterlogging exposure. Bars and error bars represent mean and standard error.

considerably decreased ($P < 0.001$, Fig. 3D) on waterlogging 7-day and waterlogging 21-day compared to the control (not significantly different from day 0d to 7d).

Effects of waterlogging stress on yield in the rapeseed ecosystem

The yield of rapeseed variations during waterlogging are shown in Fig. 4. The yield of rapeseed significantly decreased ($P < 0.05$) after 7-day and 21-day of waterlogging when compared to the control. Compared to the waterlogging 7-day treatment, the rapeseed yield significantly decreased in the waterlogging 21-day treatment ($P < 0.05$).

Discussion

Effects of waterlogging stress on agronomy traits and soil characteristics in rapeseed

Crop growth and development are directly reflected in plant height and yield. According to earlier research, waterlogging significantly affected plant height²⁸. In this study, shorter average plant height and decreased rapeseed yield were expected during waterlogging stress periods. Rapeseed plant heights were specifically in the range of normal irrigation > 7-day of waterlogging > 21-day of waterlogging as the waterlogging period increased ($P < 0.05$). Evidence that soil moisture increases quickly after waterlogging was obtained through field investigations (Fig. 1D). Meanwhile, waterlogging results in a lowered redox potential (Fig. 1C), which is statistically significantly different from controls ($P < 0.05$) using normal irrigation. Shabala et al.²⁹ agreed with this view as well. The observed decrease in plant height and yield during the waterlogging stress periods can be attributed to several factors. Firstly, waterlogging leads to reduced oxygen availability in the root zone, resulting in hypoxic conditions for the plant roots³⁰. This oxygen deficiency can negatively affect root respiration and nutrient uptake, ultimately leading to stunted plant growth³¹ and reduced crop yield. Moreover, the decrease in redox potential observed in the waterlogged conditions indicates a shift towards anaerobic conditions in the soil (Fig. 1C). Under anaerobic conditions, there is limited availability of essential nutrients such as nitrogen and phosphorus, which are crucial for plant growth and development³². The nutrient imbalance caused by waterlogging can further impair the overall growth and vigour of rapeseed plants.

In addition, the association between leaf chlorophyll concentration and leaf surface photosynthetic efficiency³³. Measurements with the SPAD were used in this study to assess the relative chlorophyll content³⁴. Our findings indicated that the SPAD values in the 7-day or 21-day waterlogging treatments were higher than

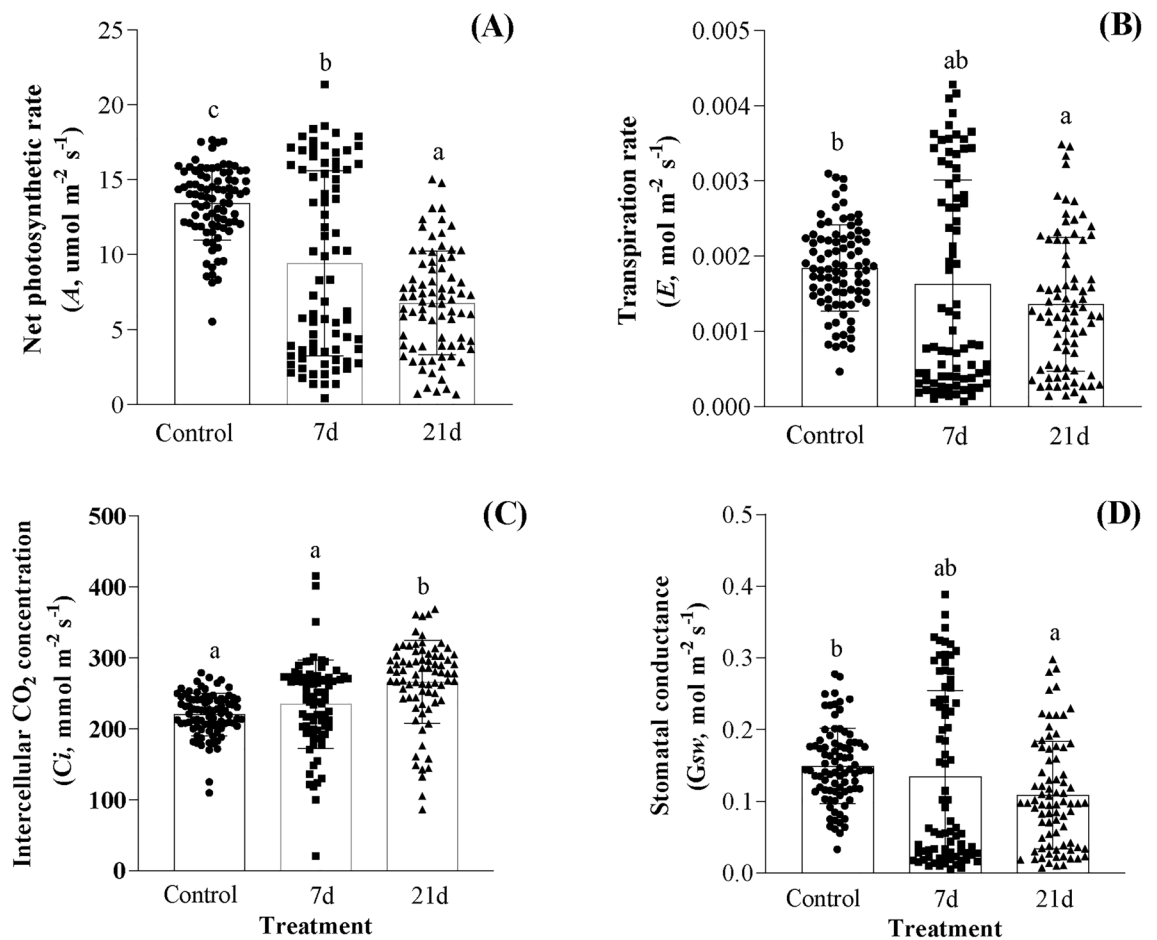


Figure 2. Effects of waterlogging stress on the gas exchange in rapeseed leaves. Treatment consists of one control treatment, normal irrigation (control, CK), and two treatments: waterlogging lasting 7-day and 21-day. Significant variations among sets at the maximal points are denoted by lowercase letters above each bar ($P < 0.05$). (A) Represent net photosynthetic rate, A ; (B) represent transpiration rate, E ; (C) represent intercellular CO_2 concentration, C_i ; (D) represent stomatal conductance, G_{sw} . Bars and error bars represent mean and standard error. Each black symbol (circle, square, triangle) represents a single sample of control, 7d and 21d, respectively, and the distance between the samples represents the difference in composition of the samples.

those in the normal irrigation (Fig. 1B). However, previous studies have shown that waterlogging stress dramatically decreased the total leaf chlorophyll concentration of rapeseed seedlings^{14,35}. This finding is in contrast to the results of this study. After waterlogging, an increase in SPAD values in rapeseed plants may be attributed to multiple factors. First, waterlogging restricts oxygen supply to the roots and reduces nutrient availability, resulting in hypoxia and nutrient limitation in rapeseed plants^{31,36}. Under such conditions, plants may respond by enhancing chlorophyll synthesis to adapt to the changing environment, improve photosynthetic efficiency, and sustain growth vigor, thereby increasing SPAD values³⁷. Second, waterlogging stress can alter leaf anatomy and cellular morphology, affecting the accumulation and distribution of chlorophyll within the leaves, consequently leading to an increase in SPAD values¹⁷. Studies have suggested that under waterlogging conditions, adjustments in leaf anatomy, such as increased stomatal density and reduced leaf thickness, may promote chlorophyll accumulation and enhance light energy utilization efficiency. It is important to note that while an increase in SPAD values may indicate an elevation in chlorophyll content, it does not necessarily translate to an increase in plant height in rapeseed. Other factors including stomatal limitations, nutrient imbalances, and metabolic abnormalities could also influence plant growth and yield under waterlogging stress³⁸.

Effects of waterlogging stress on the gas exchange in rapeseed leaves

Water stress is one of the main factors limiting the output of crops, which also affects plant physiology. According to Kuai, et al.³⁹, under waterlogging stress, the photosynthetic leaf rate of rapeseed decreased at the seedling and bolting stages. As observed in the present study, when compared to the control, A was significantly decreased ($P < 0.05$) on waterlogging 7d and waterlogging 21d (Fig. 2A). According to Pandey, et al.⁴⁰, waterlogging stress decreased A rate and Rubisco activity. This was accompanied by a decrease in G_{sw} (Fig. 2D), which led to less starch in the leaves. Chlorophyll concentration is a well-known key indicator of the photosynthetic ability of

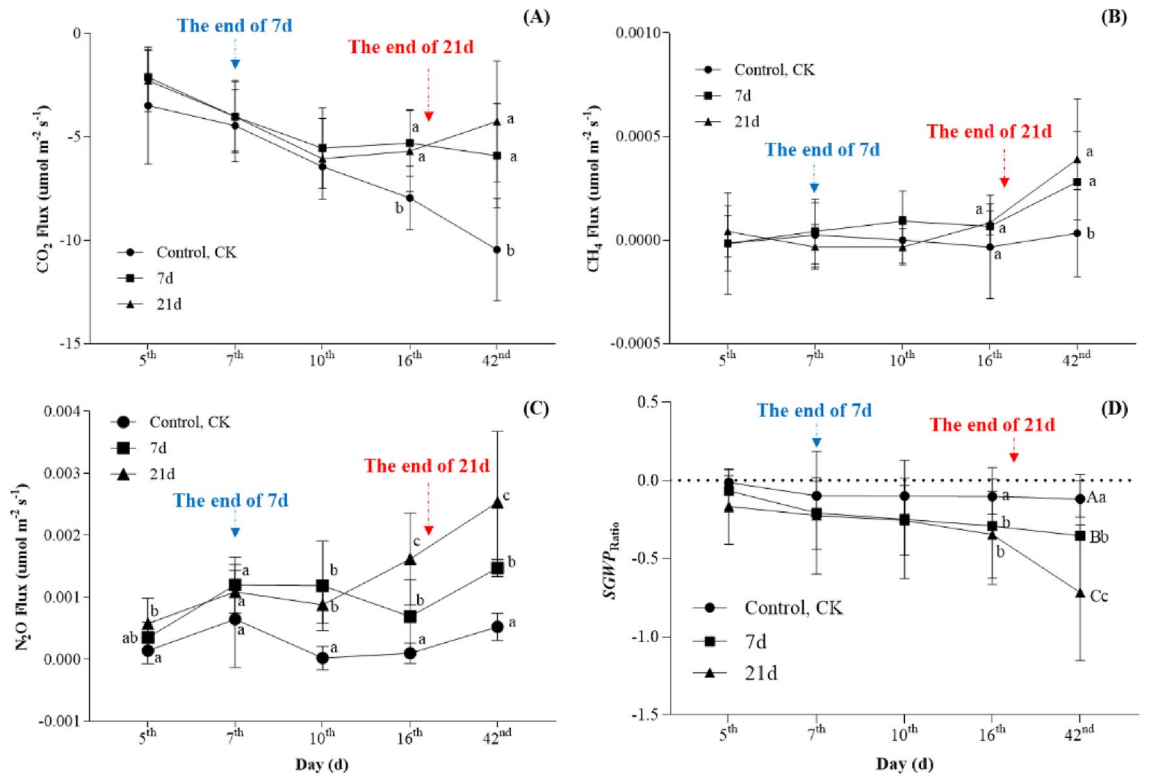


Figure 3. Effects of waterlogging stress on GHGs release in the rapeseed ecosystem. Treatment consists of one control treatment, normal irrigation (control, CK), and two treatments: waterlogging lasting 7-day and 21-day. Different uppercase letters represent highly significant ($P < 0.001$), different lowercase letters represent significant ($P < 0.05$), and the same letters represent no significant ($P > 0.05$). (A) Represent CO₂ Flux, (B) represent CH₄ Flux, (C) represent N₂O Flux, (D) represent SGWP_{ratio}. The x-axis denotes the duration in days post-waterlogging exposure. The end of 7d (or 21d) represents the time point when the waterlogging ends on the 7th day (or 21st day).

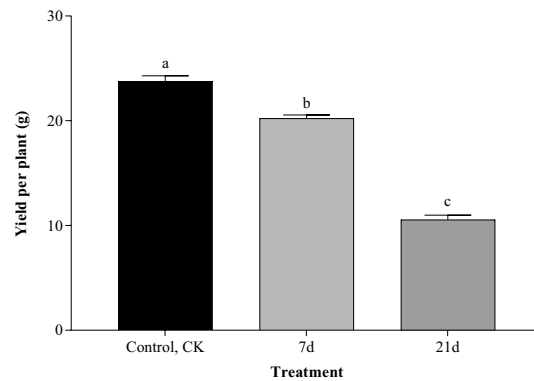


Figure 4. Effects of waterlogging stress on yield in the rapeseed ecosystem. Treatment consists of one control treatment, normal irrigation (control, CK), and two treatments: waterlogging lasting 7-day and 21-day. Bars and error bars represent mean and standard error. Lowercase letters above each bar indicate significant differences among sets at the maximal points ($P < 0.05$).

plants. However, in our experiment, waterlogging treatments increased the SPAD values of rapeseed leaves (Fig. 1B), which led to a decrease in the photosynthetic rate of rapeseed leaves (Fig. 2A). Further analysis of Fig. 2A revealed that the control had a more concentrated photosynthetic rate than the waterlogging treatment, which had a more scattered rate. This difference is probably caused by the various methodologies that were used. Additionally, we hypothesized that the rapeseed leaves under waterlogging stress quickly accumulated large quantities of carbohydrates and energy. Rapeseed under stress conditions require significant energy to maintain their growth and development through self-repairing behavior, but this needs further research. According to

Rao, et al.⁴¹, the photosynthetic system of mulberry seedling leaves can self-repair in a flooded environment. This compensatory mechanism for self-regulation has been proposed by Zhang, et al.⁴². Additionally, in this study, waterlogging 21d displayed a significant decrease ($P < 0.05$) in E compared to the control, and waterlogging 7d displayed no significant difference ($P > 0.05$) in E compared to the control (Fig. 2B). Stomata are the “windows” for exchanging gases between plant leaves and their surroundings. The condition of stomatal conductance (G_{sw}) has a direct impact on the photosynthetic and transpiration ability of plants (Fig. 2D). G_{sw} decreased at 7d and 21d following waterlogging compared to the control, with a significant increase observed at 21d ($P < 0.05$). The C_i in all waterlogging treatments showed the opposite trend compared to the C_i , with a significant increase at 21d compared to the control ($P < 0.05$, Fig. 2C).

Effects of waterlogging stress on GHGs release and yield in rapeseed ecosystem

Waterlogging is an important source of anthropogenic greenhouse gas (GHG) emissions. We previously hypothesize that waterlogging increases CH_4 emissions while decreases N_2O emissions. In this study, as waterlogging occurs, we observe an elevated CH_4 emission rate in the rapeseed ecosystem, which aligns with our expected results. The primary reason for this increase is the excessive soil moisture resulting from waterlogging, creating an anaerobic environment favorable for the growth and reproduction of methanogenic bacteria, which release a significant amount of CH_4 . This finding is consistent with previous comprehensive reviews on methanogenesis and methanotrophy in soil⁴³. Moreover, in this study, we also observe a significant increase in N_2O emissions. From our analysis, we attribute this increase to the effect of soil moisture on soil N mineralization, as it partially regulates the temporal fluctuations of soil N mineralization^{44,45}. Multiple studies demonstrate that water content is a crucial controlling factor influencing soil N_2O emissions. Specifically, the amount of soil pore water content determines the magnitude of N_2O emissions during soil rewetting⁴⁶. Hence, under short-term waterlogging conditions, there is an increased flux of N_2O ⁴⁷, further corroborating our research findings.

There are multiple potential causes for waterlogging in rapeseed fields, which can be broadly categorized into two main groups. One possible cause is excessive precipitation, leading to excessive soil moisture. This may be attributed to frequent or extreme rainfall events resulting from climate change⁴⁸. Another potential cause is poor soil drainage, which hinders the rapid removal of excess water. This may be due to factors such as poor soil structure, high soil density, or inadequate surface drainage systems^{49,50}. Regardless of the reasons, based on the findings of this study, whenever waterlogging occurs, the emissions of CH_4 and N_2O greenhouse gases in rapeseed fields increase, along with an increase in the $SGWP_{ratio}$ (Fig. 3). This not only directly affects the yield of rapeseed in the current year but also exacerbates the global warming potential, subsequently impacting the next year's rapeseed production. Therefore, in addition to addressing the drainage issues in rapeseed fields, reducing greenhouse gas emissions is equally essential in addressing waterlogging problems. This series of measures contributes to the sustainable development of rapeseed production.

Conclusions

We discovered that rapeseed exposed to waterlogging stress released more GHGs than they would under normal irrigation. When compared to normal irrigation, waterlogging for 7-day or 21-day resulted in decreased yield, plant height, soil moisture, soil ORP, A , E , and G_{sw} . Compared to normal irrigation, waterlogging for 7-day or 21-day increased SPAD values and C_i . The $SGWP_{ratio}$ of the sum CO_2 -eq of CH_4 and N_2O significantly increased compared to normal irrigation during the entire flooding stress period. We concluded that continuous waterlogging over a short period could decrease rapeseed yield and increase seedling rapeseed C and N release, further contributing to global warming. Therefore, we suggest increasing drainage techniques to decrease the release of agricultural GHGs and promote sustainable crop production.

Data availability

The datasets used and/or analysed during the current study available from the corresponding author on reasonable request.

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References

- Rogelj, J. et al. Paris agreement climate proposals need a boost to keep warming well below 2 °C. *Nature* **534**, 631–639. <https://doi.org/10.1038/nature18307> (2016).
- Barba, J., Poyatos, R. & Vargas, R. Automated measurements of greenhouse gases fluxes from tree stems and soils: magnitudes, patterns and drivers. *Sci. Rep.* **9**, 4005. <https://doi.org/10.1038/s41598-019-39663-8> (2019).
- Yu, G. R., Hao, T. X. & Xing, Z. J. Discussion on action strategies of China's carbon peak and carbon neutrality. *Bull. Chin. Acad. Sci.* **37**, 423–434 (2022).
- WMO. *More bad news for the planet: greenhouse gas levels hit new highs*. <https://public.wmo.int/en/media/press-release/more-bad-news-planet-greenhouse-gas-levels-hit-new-highs> (2022).
- Climate Change, I. P. C. C. *Climate Change 2014: Synthesis Report 2015* (Intergovernmental Panel on Climate Change, 2014).
- Richardson, A. D. et al. Six years of ecosystem-atmosphere greenhouse gas fluxes measured in a sub-boreal forest. *Sci. Data* **6**, 117. <https://doi.org/10.1038/s41597-019-0119-1> (2019).
- Smith, K. A. et al. Exchange of greenhouse gases between soil and atmosphere: Interactions of soil physical factors and biological processes. *Eur. J. Soil Sci.* **54**, 779–791. <https://doi.org/10.1046/j.1351-0754.2003.0567.x> (2003).
- Thauer, R. K. Biochemistry of methanogenesis: A tribute to Marjory Stephenson. 1998 Marjory Stephenson Prize Lecture. *Microbiology (Reading)* **144**(9), 2377–2406. <https://doi.org/10.1099/00221287-144-9-2377> (1998).
- Ragab, A., Shaw, D. R., Katuri, K. P. & Saikaly, P. E. Effects of set cathode potentials on microbial electrosynthesis system performance and biocathode methanogen function at a metatranscriptional level. *Sci. Rep.* **10**, 19824. <https://doi.org/10.1038/s41598-020-76229-5> (2020).

10. Oertel, C., Matschullat, J., Zurba, K., Zimmermann, F. & Erasmi, S. Greenhouse gas emissions from soils—A review. *Geochemistry* **76**, 327–352. <https://doi.org/10.1016/j.chemer.2016.04.002> (2016).
11. Fan, C. M. *et al.* Advances of oilseed rape breeding. *J. Plant Genet. Resour.* **19**, 447–454. <https://doi.org/10.13430/j.cnki.jpgr.2018.03.009> (2018).
12. Li, X. *et al.* Responses of plant development, biomass and seed production of direct sown oilseed rape (*Brassica napus*) to nitrogen application at different stages in Yangtze River Basin. *Field Crops Res.* **194**, 12–20. <https://doi.org/10.1016/j.fcr.2016.04.024> (2016).
13. Guo, R., Zhu, Y. & Liu, Y. A comparison study of precipitation in the Poyang and the Dongting Lake Basins from 1960–2015. *Sci. Rep.* **10**, 3381. <https://doi.org/10.1038/s41598-020-60243-8> (2020).
14. Men, S. *et al.* Effects of supplemental nitrogen application on physiological characteristics, dry matter and nitrogen accumulation of winter rapeseed (*Brassica napus* L.) under waterlogging stress. *Sci. Rep.* **10**, 1–10. <https://doi.org/10.1038/s41598-020-67260-7> (2020).
15. Yuan, L. B. *et al.* The anaerobic product ethanol promotes autophagy-dependent submergence tolerance in Arabidopsis. *Int. J. Mol. Sci.* <https://doi.org/10.3390/ijms21197361> (2020).
16. Irfan, M., Hayat, S., Hayat, Q., Afroz, S. & Ahmad, A. Physiological and biochemical changes in plants under waterlogging. *Protoplasma* **241**, 3–17. <https://doi.org/10.1007/s00709-009-0098-8> (2010).
17. Pan, J., Sharif, R., Xu, X. & Chen, X. Mechanisms of waterlogging tolerance in plants: Research progress and prospects. *Front. Plant Sci.* **11**, 627331. <https://doi.org/10.3389/fpls.2020.627331> (2021).
18. Sharma, P. C., Kumar, A. & Vineeth, T. V. in *Abiotic Stress Management for Resilient Agriculture* (eds Minhas, P. S., Rane, J. & Pasala, R. K.) 177–220 (Springer, 2017).
19. Zhou, W. *et al.* Plant waterlogging/flooding stress responses: From seed germination to maturation. *Plant Physiol. Biochem.* **148**, 228–236. <https://doi.org/10.1016/j.plaphy.2020.01.020> (2020).
20. Kuai, J. *et al.* Leaf characteristics at recovery stage affect seed oil and protein content under the interactive effects of nitrogen and waterlogging in rapeseed. *Agriculture* **10**, 207. <https://doi.org/10.3390/agriculture10060207> (2020).
21. Nabloussi, A. *et al.* Assessment of a set of rapeseed (*Brassica napus* L.) varieties under waterlogging stress at different plant growth stages. *OCL* **26**, 36. <https://doi.org/10.1051/ocl/2019033> (2019).
22. Zhou, W. & Lin, X. Effects of waterlogging at different growth stages on physiological characteristics and seed yield of winter rape (*Brassica napus* L.). *Field Crops Res.* **44**, 103–110 (1995).
23. Liu, K. *et al.* The state of the art in modeling waterlogging impacts on plants: what do we know and what do we need to know. *Earth's Future* **8**, e2020EF001801. <https://doi.org/10.1029/2020EF001801> (2020).
24. Frolking, S., Li, C., Braswell, R. & Fuglestvedt, J. Short- and long-term greenhouse gas and radiative forcing impacts of changing water management in Asian rice paddies. *Glob. Change Biol.* **10**, 1180–1196. <https://doi.org/10.1111/j.1529-8817.2003.00798.x> (2004).
25. Runkle, B. *et al.* Methane emission reductions from the alternate wetting and drying of rice fields detected using the eddy covariance method. *Environ. Sci. Technol.* **53**, 671–681. <https://doi.org/10.1021/acs.est.8b05535> (2019).
26. Zhang, L. *et al.* Ratoon rice with direct seeding improves soil carbon sequestration in rice fields and increases grain quality. *J. Environ. Manag.* **317**, 115374. <https://doi.org/10.1016/j.jenvman.2022.115374> (2022).
27. Neubauer, S. C. & Megonigal, J. P. Moving beyond global warming potentials to quantify the climatic role of ecosystems. *Ecosystems* **18**, 1000–1013. <https://doi.org/10.1007/s10021-015-9879-4> (2015).
28. Li, H. J. *et al.* Consistency of different indices in rapeseed (*Brassica napus*) may predict the waterlogging tolerance. *Int. J. Agric. Biol.* **18**, 61–67 (2015).
29. Shabala, S. Physiological and cellular aspects of phytotoxicity tolerance in plants: The role of membrane transporters and implications for crop breeding for waterlogging tolerance. *New Phytol.* **190**, 289–298. <https://doi.org/10.1111/j.1469-8137.2010.03575.x> (2011).
30. Kaur, G. *et al.* Impacts and management strategies for crop production in waterlogged or flooded soils: A review. *Agron. J.* **112**, 1475–1501. <https://doi.org/10.1002/agj2.20093> (2020).
31. Gu, C. *et al.* Soil enzyme activity in soils subjected to flooding and the effect on nitrogen and phosphorus uptake by oilseed rape. *Front. Plant Sci.* **10**, 368. <https://doi.org/10.3389/fpls.2019.00368> (2019).
32. Bansal, R. & Srivastava, J. P. Effect of waterlogging on photosynthetic and biochemical parameters in pigeonpea. *Russ. J. Plant Physiol.* **62**, 322–327. <https://doi.org/10.1134/S1021443715030036> (2015).
33. Azhar, A., Makihara, D., Naito, H. & Ehara, H. Evaluating sago palm (*Metroxylon sagu* Rottb) photosynthetic performance in waterlogged conditions: Utilizing pulse-amplitude-modulated (PAM) fluorometry as a waterlogging stress indicator. *J. Saudi Soc. Agric. Sci.* **19**, 37–42. <https://doi.org/10.1016/j.jssas.2018.05.004> (2020).
34. Shibaeva, T. G., Mamaev, A. V. & Sherudilo, E. G. Evaluation of a SPAD-502 plus chlorophyll meter to estimate chlorophyll content in leaves with interveinal chlorosis. *Russ. J. Plant Physiol.* **67**, 690–696. <https://doi.org/10.1134/S1021443720040160> (2020).
35. Habibzadeh, F., Sorooshzadeh, A., Pirdashti, H., Mohammad, S. & Sanavy, M. Effect of nitrogen compounds and tricyclazole on some biochemical and morphological characteristics of waterlogged-canola. *Int. Res. J. Appl. Basic Sci.* **3**, 77–84 (2012).
36. Wang, Z. *et al.* Calcium peroxide alleviates the waterlogging stress of rapeseed by improving root growth status in a rice-rape rotation field. *Front. Plant Sci.* <https://doi.org/10.3389/fpls.2022.1048227> (2022).
37. Wu, X. *et al.* Individual and combined effects of soil waterlogging and compaction on physiological characteristics of wheat in southwestern China. *Field Crops Res.* **215**, 163–172. <https://doi.org/10.1016/j.fcr.2017.10.016> (2018).
38. Mitra, A. *et al.* in *Plant Growth and Stress Physiology* (eds Gupta, D. K. & Palma, J. M.) 1–22 (Springer, 2021).
39. Kuai, J. *et al.* Leaf carbohydrates assimilation and metabolism affect seed yield of rapeseed with different waterlogging tolerance under the interactive effects of nitrogen and waterlogging. *J. Agron. Crop Sci.* **206**, 823–836. <https://doi.org/10.1111/jac.12430> (2020).
40. Pandey, D. M., Goswami, C. L., Kumar, B. & Jain, S. Hormonal regulation of photosynthetic enzymes in cotton under water stress. *Photosynthetica* **38**, 403–407. <https://doi.org/10.1023/a:1010925604941> (2000).
41. Rao, L., Li, S. & Cui, X. Leaf morphology and chlorophyll fluorescence characteristics of mulberry seedlings under waterlogging stress. *Sci. Rep.* **11**, 13379. <https://doi.org/10.1038/s41598-021-92782-z> (2021).
42. Zhang, Y., Liu, G., Dong, H. & Li, C. Waterlogging stress in cotton: Damage, adaptability, alleviation strategies, and mechanisms. *Crop J.* **9**, 257–270. <https://doi.org/10.1016/j.cj.2020.08.005> (2021).
43. Serrano-Silva, N., Sarria-Guzmán, Y., Dendooven, L. & Luna-Guido, M. Methanogenesis and methanotrophy in soil: A review. *Pedosphere* **24**, 291–307. [https://doi.org/10.1016/S1002-0160\(14\)60016-3](https://doi.org/10.1016/S1002-0160(14)60016-3) (2014).
44. Barton, L. *et al.* Nitrous oxide emissions from a cropped soil in a semi-arid climate. *Glob. Change Biol.* **14**, 177–192. <https://doi.org/10.1111/j.1365-2486.2007.01474.x> (2008).
45. Ma, L. N. *et al.* The effects of warming and nitrogen addition on soil nitrogen cycling in a temperate grassland, northeastern China. *PLoS ONE* **6**, e27645. <https://doi.org/10.1371/journal.pone.0027645> (2011).
46. Barrat, H. A. *et al.* The impact of drought and rewetting on N₂O emissions from soil in temperate and Mediterranean climates. *Eur. J. Soil Sci.* **72**, 2504–2516. <https://doi.org/10.1111/ejss.13015> (2021).
47. Chen, Z. *et al.* Increased N₂O emissions during soil drying after waterlogging and spring thaw in a record wet year. *Soil Biol. Biochem.* **101**, 152–164. <https://doi.org/10.1016/j.soilbio.2016.07.016> (2016).

48. Tabari, H. Climate change impact on flood and extreme precipitation increases with water availability. *Sci. Rep.* **10**, 13768. <https://doi.org/10.1038/s41598-020-70816-2> (2020).
49. Bronick, C. J. & Lal, R. Soil structure and management: A review. *Geoderma* **124**, 3–22. <https://doi.org/10.1016/j.geoderma.2004.03.005> (2005).
50. Singh, A. Poor-drainage-induced salinization of agricultural lands: Management through structural measures. *Land Use Policy* **82**, 457–463. <https://doi.org/10.1016/j.landusepol.2018.12.032> (2019).

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Author contributions

L.L. and L.Z. wrote the main manuscript text and L.Z. prepared Figs. 1–4. All authors reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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