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Prognosis value and positive association of Rab1A/IL4Rα aberrant expression in gastric cancer

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Gastric cancer (GC) is the most common gastrointestinal cancer and the leading cause of worldwide cancer-associated mortality. Several GC patients are diagnosed at the advanced stage with an unsatisfactory 5-year survival rate. Rab1A was significantly associated with IL4R α expression in non-small cell lung cancer. However, their potential correlation in expression and prognosis remains largely unknown in GC. In this study, Rab1A/IL-4R α was significantly increased in GC than in para-cancerous tissues, and Rab1A/IL-4R α overexpression caused poor prognosis among GC patients. Rab1A expression was significantly correlated with IL-4R α expression in GC tissues, as determined by IHC analysis. In addition, the mRNA expression of Rab1A was closely linked with the IL-4R α mRNA expression in GC tissue expressed by *qPCR*. Furthermore, the Kaplan–Meier analysis demonstrated that the group with negative Rab1A and IL-4R α expression had longer 5-year survival rates than the other group. Besides, the group with positive Rab1A and IL-4R α expression had a worse prognosis than the other group. Finally, nomograms revealed the overall 3 and 5-year survival determined crucial roles of Rab1A/IL-4R α expression in predicting the prognosis of GC patients. Therefore, Rab1A/IL-4R α is vital in GC, providing a novel perspective on targeted GC therapy.

Gastric cancer (GC) is the most common gastrointestinal cancer with the leading cause of cancer-associated global mortality^{1,2}. Although diverse and systematic treatments have been employed for GC, including surgical treatment, chemoradiotherapy, and targeted therapy, the current 5-year survival rate remains unsatisfactory^{3,4}. Unfortunately, most GC patients are diagnosed at an advanced stage, with distant organ metastasis, leading to poor prognosis⁵. Moreover, long-term use of chemotherapy drugs causes tumor cell drug resistance, and insensitivity to tumor drugs limits the chemotherapy efficacy⁶. Therefore, finding novel therapeutic targets to promote the efficacy of GC treatment is an urgent requirement.

Rab proteins are members of the GTP enzyme superfamily and are recognized as housekeeping proteins in intracellular membrane dynamics involving various membrane transport processes. Rab1A, a member of the Rab family⁷, is a GTP enzyme controlling vesicular transport from the ER to the Golgi apparatus^{8,9}. Recent studies indicated Rab1A regulates signal transduction¹⁰, cell migration¹¹, and cell autophagy¹². Furthermore, abnormal Rab1A expression is associated with some clinical disease occurrences, including Parkinson's disease¹³ and primary cardiomyopathy¹⁴. Previous studies reported that Rab1A was aberrantly expressed in many cancers, leading to poor prognoses, including breast¹⁵, lung¹⁶, and liver¹⁷ cancers. Moreover, Rab1A behaved as an oncogene in various cancers, enhancing growth, metastasis, and cancer cell drug resistance^{18,19}.

Interleukin-4 (IL-4) and its receptors (IL-4R) are crucial in cancer cell proliferation and other biological behaviors like migration and invasion^{20,21}. Additionally, IL-4R promoted cancer cell resistance to chemotherapy drugs in colorectal cancer²². One of the receptor chains of IL-4Ra was highly expressed in solid cancers and was closely associated with locally advanced tumor staging, promoting poor prognosis^{23,24}.

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Results

Expression levels of Rab1A in GC and paired adjacent tissues. Firstly, IHC staining helped investigate the Rab1A expression in 115 GC patients (Fig. 1A). The results indicated that Rab1A was significantly expressed in GC tissues than in matched normal ones (P < 0.001) (Fig. 1B). Moreover, subgroup analysis was performed based on the lymph node metastases (LNM) and tumor-lymph node-metastasis (TNM) staging state. Rab1A expression was higher in the LNM group than in the non-LNM group (P < 0.001) (Fig. 1C). Furthermore, the TNM III group showed a significantly higher Rab1A expression than the other group (P = 0.004) (Fig. 1D).

Relationship between Rab1A and the clinic-pathological factors in GC patients. The relationship between Rab1A and clinic-pathological factors was analyzed using the Chi-square or Fisher's tests. The results are displayed in Table 1, indicating Rab1A expression was closely related to LNM (P < 0.001), degree of differentiation (P = 0.012), Venous invasion (P = 0.046), and TNM staging (P = 0.026). There was no significant difference between Rab1A expression and the left clinic-pathological factors.

The influence of Rab1A overexpression on prognosis in GC patients. The Kaplan–Meier analysis was used to explore the influence of Rab1A overexpression on prognosis. Our results showed that the Rab1A positive group had a worse prognosis than the negative group (P < 0.001) (Fig. 1E). Moreover, we performed subgroup analysis



Figure 1. Rab1A expression and Kaplan–Meier analysis among 115 GC patients. (**A**) Rab1A expression using IHC staining in 115 GC and para-cancer tissues (scale bar 100um). (**B**–**D**) The IHC score of Rab1A in (**B**) GC and para-cancer tissues, (**C**) GC with or without lymph node metastasis, (**D**) GC with TNM I–II or TNM III staging. (**E**–**G**) Kaplan–Meier analysis of Rab1A positive vs. Rab1A negative in (**E**) 115 GC patients, (**F**) GC patients with TMN I–II staging, and (**G**) GC patients with TMN III staging.

	Rab1A							
Variables	Negative	Positive	P value					
Age (years)								
<65	29	30	0.928					
≥65	28	28						
Gender								
Male	38	42	0.503					
Female	19	16						
Tumor size (cm)								
< 5	42	42	0.878					
≥5	15	16						
Depth of tumor invasion								
T1-2	23	17	0.214					
T3-4	34	41						
Lymph node metastasis								
No	31	11	< 0.001°					
Yes	26	47						
Degree of differentiation								
Well	23	11	0.012 ^a					
Poor	34	47						
Venous invasion								
Negative	38	28	0.046ª					
Positive	19	30						
Neural invasion								
Negative	32	24	0.113					
Positive	25	34						
TNM staging								
I–II	41	30	0.026ª					
III	16	28						

Table 1. Relationship between Rab1A and clinic-pathological factors in 115 GC patients. ${}^{a}P < 0.05$, ${}^{b}P < 0.01$, ${}^{c}P < 0.001$.

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on prognosis depending on the various TNM states using the Kaplan–Meier analysis. Rab1A overexpression group showed a poorer 5-year survival rate than the low-expression group in TNM I–II staging (P < 0.001) (Fig. 1F). However, no significant difference was observed in TNM III staging (P = 0.222) (Fig. 1G). Univariate analysis indicated that differentiation degree, venous invasion, neural invasion, depth of tumor invasion, LNM, TNM stage, and Rab1A expression were closely related to poor prognosis (P < 0.05) (Table 2). Moreover, multivariate analysis showed that Rab1A expression, TNM stage, LNM, and neural invasion were critical prognostic factors in GC (Table 2).

	Univariate analysis			Multivariate analysis		
Varieties	HR	95% CI	Р	HR	95% CI	Р
Age (≤60 or>60 years)	0.931	0.563-1.479	0.711			
Gender (male/female)		0.777-2.345	0.288			
Size of tumor (\leq 5 or > 5 cm)		0.424-1.214	0.216			
Degree of differentiation (moderate-well/poor)		0.292-0.929	0.027 ^a	0.745	0.407-1.362	0.339
Venous invasion (negative/positive)		0.260-0.691	0.001 ^b	0.852	0.489-1.485	0.572
Neural invasion (negative / positive)		0.262-0.716	0.001 ^b	0.647	0.362-1.164	0.147
Depth of tumor invasion (T1-2/T3-4)		0.201-0.645	0.001 ^b	0.790	0.396-1.691	0.544
Lymph node metastasis (negative/positive)		0.146-0.482	< 0.001 ^c	0.555	0.258-1.196	0.133
TNM stage (I-II/III)		0.183-0.491	< 0.001 ^c	0.608	0.319-1.160	0.131
Rab1A expression (negative/positive)		0.224-0.618	< 0.001 ^c	0.587	0.338-1.018	0.058

Table 2. Results of univariate and multivariate analyses of postoperative patients' survival by Cox's proportional hazard model. ${}^{a}P < 0.05$, ${}^{b}P < 0.01$, ${}^{c}P < 0.001$.

Overexpression of IL-4Ra led to poor prognosis in various TNM staging. The IHC staining was performed to investigate the IL-4Ra expression level in 115 GC patients to explore its influence on prognosis (Fig. 2A). IL-4Ra was significantly increased in GC tissues than in para-cancerous tissues (P<0.001) (Fig. 2B). Moreover, subgroup analysis helped examine the IL-4Ra expression level according to different LNM and TNM staging. Thus, IL-4Ra expression in the LNM group was significantly higher than the non-LNM group (P<0.001) (Fig. 2C). Additionally, the TNM III group showed higher IL-4Ra expression than the other group (P<0.001) (Fig. 2D).

The Kaplan–Meier analysis indicated IL-4R α overexpression caused poor prognosis (P < 0.001) (Fig. 2E). Moreover, subgroup analysis revealed that the group with increased IL-4R α expression had shorter 5-year survival rates in TNM I–II and TNM III stages (P < 0.001, P = 0.001) (Fig. 2F,G).

Positive correlation of Rab1A expression with the IL-4Ra expression in GC. We searched the TCGA database through the GEPIA platform to investigate the association of Rab1A and IL-4Ra expression. The results indicated Rab1A expression was closely related to the L-4Ra expression in GC tissues. In contrast, no significant correlation was observed on expression in para-cancer tissues (P=0.005) (Fig. 3A), (P=0.72) (Fig. 3B). Interestingly, Rab1A was significantly associated with IL-4Ra expression (P<0.001) (Fig. 3C). Then the IHC staining helped validate the association of Rab1A/IL-4Ra in 115 GC patients. The outcome indicated that Rab1A expression was significantly associated with IL-4Ra expression in GC tissues (P<0.001) (Fig. 3D). However, no significant difference could be found in para-cancer tissues (P=0.109) (Fig. 3E). The subgroup analysis across various TNM stages had a positive correlation of Rab1A/IL-4Ra expression in TNM I–II and TNM III staging (P=0.010, P=0.034) (Fig. 3F,G).

Rab1A/ IL-4Ra expression was significantly elevated in GC tissues than in para-cancer. Moreover, Rab1A expression was mainly related to IL-4Ra expression in GC tissues. Thus, a cluster analysis was performed



Figure 2. IL-4R α expression and Kaplan–Meier analysis of 115 GC patients. (A) IL-4R α expression using IHC staining in 115 GC and para-cancer tissues (scale bar 100um). (B–D) The IHC score of IL-4R α in (B) GC and para-cancer tissues, (C) GC with or without lymph node metastasis, (D) GC with TNM I–II or TNM III staging. (E–G) Kaplan–Meier analysis of IL-4R α positive vs. IL-4R α negative in (E) 115 GC patients, (F) GC patients with TMN I–II staging, and (G) GC patients with TMN III staging.



Figure 3. Association of Rab1A/IL-4R α expression in GC. (**A**,**B**) the association between Rab1A/IL-4R α expression in TCGA datasets through the GEPIA platform in GC tissues(**A**) and normal tissues (**B**). (**C**) Constituent ratio depicting the correlation between Rab1A and IL-4R α expression in GC. (**D**–**G**) The association of Rab1A with IL-4R α expression using the IHC score in (**D**) GC tissues, (**E**) normal tissue, (**F**) GC with stage TNM I–II staging, and (**G**) GC with TNM III staging. (**H**) Cluster analysis of Rab1A and IL-4R α IHC score based on GC tumor and paired normal tissues.

depending on the IHC scores of Rab1A/IL-4Ra in GC and normal gastric tissues (Fig. 3H). The PERMANOVA analysis revealed that the IHC scores of Rab1A and IL-4Ra significantly differed between tumor and normal tissues (P = 0.001). Therefore, Rab1A and IL-4Ra levels can be differentiated between GC and normal tissues.

Finally, the *qPCR* helped explore the Rab1A/IL-4Ra mRNA expression in 24 tumor and normal tissue cases, displaying heatmap (Fig. 4A). The results indicated that the expression of Rab1A/IL-4Ra mRNA was significantly elevated in GC tissues than in normal tissues (P < 0.001) (Fig. 4B–E). Additionally, the mRNA expression of Rab1A was closely associated with that of IL-4Ra in GC tumor tissues (P < 0.001) (Fig. 4F). Furthermore, cluster analysis based on the mRNA level of Rab1A/IL-4Ra in GC and normal gastric tissues were performed (Fig. 4G), showing 25% tumor and 75% normal tissues within Cluster 1 (Fig. 4H).

The influence of Rab1A/IL-4R\alpha overexpression on GC patient prognosis. Initially, the Kaplan-Meier analysis helped explore the influence of Rab1A/IL-4R α overexpression on GC patient prognosis. Therefore, both negative expressions of Rab1A and IL-4R α group showed longer 5-year survival rates than in the other group (P < 0.001) (Fig. 5A). Besides, positive Rab1A and IL-4R α group expressions revealed a worse prognosis than the other group (P < 0.001) (Fig. 5B). Moreover, the nomograms helped predict the 3 and 5-year overall GC patient survival (Fig. 5C). The total points for each prognostic factor on the nomogram point scale predicted the survival rate. The outcome indicated Rab1A/IL-4R α expression was crucial in predicting the 3 and 5-year overall GC patient survival.



Figure 4. The correlation between Rab1A and IL-4R α expression in 24 GC and paired normal tissues using *qPCR*. (**A**) the heatmap exhibits the correlation between Rab1A and IL-4R α expression in GC and para-cancer normal tissues. (**B**,**C**) Rab1A expression in GC and para-cancer tissues. (**D**,**E**) IL-4R α expression in GC and para-cancer tissues. (**F**) The correlation between Rab1A and IL-4R α expression in 20 GC tissues. (**G**) Cluster analysis for normal and tumor tissues depending on the mRNA level of Rab1A/IL-4R α in GC and normal gastric tissues and (**H**) percentage of tumor and normal tissues in each cluster.

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Discussion

GC is the fifth most common cancer worldwide and the fourth primary cause of cancer-related mortality^{26,27}. Although the development of various diagnostic and treatment strategies for GC, the 5-year survival rate is < 30%. This is because most GC patients are diagnosed as an advanced stage or even have distant organ metastasis. Secondly, the resistance to chemotherapy drugs is also an important reason for the poor prognosis of patients with advanced GC^{28} . Therefore, early diagnosis and finding novel therapeutic targets are urgent.

Rab1A is a RAB family member and a small GTPase studied frequently⁷. Rab1A is anchored to the endoplasmic reticulum and Golgi membrane while mainly expressing itself in the endoplasmic system^{8,9}. Previous studies established that Rab1A was widely expressed in the ER with a deep exploration of Rab1A, increasing the pressure of the ER and promoting cell apoptosis²⁹. Rab1A transports substances between membrane systems and is vital in transmitting transmembrane signals between cells¹⁰.

Recent studies indicated Rab1A, an oncogene, is aberrantly elevated in various cancers. Furthermore, Rab1A overexpression led to poor prognoses, including lung cancer¹⁶, liver cancer¹⁷, and colorectal cancer³⁰. A previous study reported that Rab1A and IL4Ra expression is highly associated with lung cancer tissues²⁵. However, the potential correlation of Rab1A and IL4Ra remains largely unknown in GC. Our study mainly investigates



Figure 5. Nomograms predict 3 and 5-year overall survival of GC patients. The nomogram point scale helped predict the 3 and 5-year overall survival rates predicted using the total points associated with each prognostic factor.

the correlation between Rab1A and IL4R α expression and the effect of Rab1A/IL4R α on patient prognosis. This study first performed the IHC staining to assess Rab1A expression and determine the association between Rab1A expression and clinic-pathological factors. Rab1A was more highly expressed in GC tissues than in matched normal tissues. Furthermore, the subgroup analysis depicted that Rab1A expression in the LNM group was significantly higher than in the non-LNM group. Moreover, the TNM III group had a considerably higher Rab1A expression than the other group. Besides, the results indicated that Rab1A expression was closely associated with LNM. No significant difference was observed between Rab1A expression and tumor size or T staging. Thus, Rab1A could promote the ability to migrate.

A previous study has reported that Rab1A overexpression caused a poor prognosis in CRC^{9,30,31}, GC³², and intrahepatic cholangiocarcinoma¹⁹. The Rab1A positive group had a worse prognosis than the negative group in GC patients. Moreover, the subgroup analysis revealed Rab1A overexpression group showed a poorer 5-year survival rate than the low-expression group in TNM I–II staging. Simultaneously, no significant difference was obtained in TNM III staging, indicating that Rab1A is crucial in prognosis, particularly in TMN I–II staging. In GC patients with TNM stage III, the prognosis is generally poor. Moreover, Rab1A expression cannot affect the prognosis of these patients.

A previous study reported that the expression of Rab1A and IL4Ra is highly correlated in lung cancer tissues, including the expression region and degree²⁵. Our research first reported that IL-4Ra significantly increased

in GC tissues than in para-cancerous tissues. Moreover, based on different LNM and TNM staging, subgroup analysis indicated that IL-4Ra expression in the LNM group was significantly higher than in the no-LNM group. The TNM III group showed a higher IL-4Ra expression than the other group. Then Kaplan–Meier analysis determined the effect of IL-4Ra overexpression on prognosis. The results indicated that IL-4Ra overexpression led to a poor prognosis. Moreover, subgroup analysis based on different TNM stages suggested that the group with high IL-4Ra expression led to shorter 5-year survival rates in TNM I-II and TNM III.

We first checked TCGA database using the GEPIA platform to assess the association of Rab1A and IL-4Ra expression. Rab1A expression was closely associated with the L-4Ra expression in GC tissues. In contrast, no significant correlation with expression was observed among para-cancer tissues. Then the IHC staining indicated that Rab1A expression was significantly associated with IL-4Ra expression in GC tissues. However, no significant difference could be seen in para-cancer tissues, consistent with the TGGA database. Finally, *qPCR* helped investigate the Rab1A/IL-4Ra mRNA expression in 24 tumor and normal tissue cases. Our results showed that the mRNA expression of Rab1A was closely related to the IL-4Ra mRNA expression in GC tumor tissues. Additionally, the Kaplan–Meier analysis helped explore the influence of Rab1A/IL-4Ra overexpression on GC patient prognosis. The results showed that both negative expressions of Rab1A and IL-4Ra group had longer 5-year survival rates than the other group. Besides, positive Rab1A and IL-4Ra group expressions had a worse prognosis than the other group. Finally, the nomograms predicting 3 and 5-year overall survival indicated that Rab1A/IL-4Ra expression had crucial roles in predicting the GC patient prognosis. Thus, the Rab1A/IL-4Ra could predict patient prognosis and be tractable as novel targets in individualized GC therapy.

Conclusion

In summary, Rab1A/IL-4Ra was significantly elevated in GC tissues than in para-cancerous tissues, and its overexpression caused poor prognosis in GC patients. Rab1A expression was significantly related to IL-4Ra expression in 115 GC tissues using the IHC staining score analysis. Besides, the Kaplan–Meier analysis revealed that the group with negative Rab1A and IL-4Ra expression had longer 5-year survival rates than the other group. Therefore, Rab1A/IL-4Ra is vital in GC, providing a novel vision for targeted GC therapy.

Materials and methods

Patients and tissue specimens. One hundred fifteen cases of GC and para-cancer tissues were obtained from 2015 to 2017 from the Department of General Surgery, the First Affiliated Hospital of Wannan Medical College. Twenty-four cases of fresh GC and adjacent normal tissues were recruited in 2022 from the same department. These GC patients had not undergone preoperative chemoradiotherapy before surgery. The study was approved by the Independent Ethics Committee (IEC) of the First Affiliated Hospital of Wannan Medical College (IRB number 202248). All the patients provided their written informed consent.

Immunohistochemistry (IHC). The immunohistochemical staining (IHC) helped detect the Rab1A/ IL4Ra expression in 115 GC and para-cancer tissues. These GC tissues were fixed in formalin, embedded in paraffin, cut into 5um, and stained using IHC, based on our previous study ³². Sections were incubated for two hours using the anti-Rab1A and anti-IL4Ra at 1:100 dilution at room temperature. The process was visualized using the tissue staining kit (Zhongshan Biotechnology, Beijing, China). The staining score was calculated based on our previous study. Five regions were randomly selected for staining evaluation, and the IHC score was determined by multiplying the staining intensity (0, negative; 1, weak; 2, moderate; and 3, strong) and extent (0, 0–5%; 1, 6–25%; 2, 26–50%; 3, 51–75%; and 4, >75%). We considered 0 as –, 1 ~ 4 as +, 5 ~ 8 as ++, and 9 ~ 12 as +++, for the final staining score. Our study regarded ++ or +++ as a high expression and – or + as a low expression. We used Anti-Rab1A (1:100, Abcam, ab302545) and anti-IL-4Ra (1:100, Abcam, ab203398) antibodies for immunohistochemistry.

RNA isolation and quantitative real-time PCR (*qPCR***).** According to the manufacturer's protocol, we extracted total RNA from the 24 matched cases of fresh GC and adjacent normal tissue using TRIzol reagent (Invitrogen, Life Technologies, USA). Following the treatment of DNAse I (Thermo Fisher Scientific, USA) to remove genomic DNA, 1 μ g RNA was reverse transcribed with a RevertAid First Strand cDNA Synthesis Kit (Thermo Fisher Scientific, USA). qRT-PCR was performed with Power SYBR* Green PCR Master Mix (ABI, USA) on the 7500 real-time PCR system (ABI, USA) based on the manufacturer's instructions. We used the following primers: Rab1A sense (ACA GTG GCT GCA GGA AAT AGA) and antisense (AGC AAA TTC CTT CGC TGT TG); IL4Ra sense (CCG CCT CGT GGC TAT AAT AA) and antisense (CAG GGC AAG AGC TTG GTA AG). The 2^{- $\Delta\Delta C$}T method determined the fold changes relative to β-actin (internal control).

Statistical analysis. The total data were expressed as means ± SEM. The statistical analyses were performed using the SPSS 22.0 software (SPSS Inc., Chicago, IL, USA), GraphPad Prism 8, and R program (version 3.6.1 for Windows, http://cran.r-project.org/). The t-test (unpaired, two-tailed) or Mann–Whitney U test helped compare the means between the groups. The Chi-square or Fisher's tests were used to explore the IHC results. Univariate and multivariate analyses were performed using SPSS 22.0 following relevant guidelines and regulations, with P < 0.05 being considered a significant difference. The authors have confirmed that all methods were carried out in accordance with relevant guidelines and regulations.

Data availability

The datasets used and/or analyzed during the current study available from the corresponding author on reasonable request.

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Author contributions

H.L. and Z.C. conducted the research, analyzed the data, and wrote the manuscript. B.J. contributed to data collection and analysis. X.S. and M.X. designed the study and wrote the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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