scientific reports

OPEN

Check for updates

Integrated strength of osmotic potential and phosphorus to achieve grain yield of rice under water deficit by arbuscular mycorrhiza fungi

Suravoot Yooyongwech¹, Rujira Tisarum², Thapanee Samphumphuang², Muenduen Phisalaphong³ & Suriyan Cha-um²

Arbuscular mycorrhizal ecosystem provides sustainability to plant integrity under drought situations. However, host plants that survive in drought frequently lose yield. The potential of *Funneliformis mosseae* (F), *Claroideoglomus etunicatum* (C), and *Acaulospora fovaeta* (A) was assessed to evaluate in indica rice cv. Leum Pua during booting stage under 21-day water withholding. The effects of three inoculation types; (i) F, (ii) F+C (FC), and (iii) F+C+A (FCA), on physiological, biochemical, and yield traits were investigated. The three types showed an induced total chlorophyll content in the host as compared to uninoculated plants. Total soluble sugars and free proline were less regulated by FC and FCA inoculated plants than by F inoculated plants under water deficit conditions. However, the FC and FCA inoculations increased phosphorus content, particularly in the shoots of water-stressed plants. In the three inoculations, the FCA dramatically improved plant osmotic potential adaptability under water deficit stress. Furthermore, even when exposed to the water deficit condition, panicle weight, grain number, and grain maturity were maintained in FCA inoculated plants. According to the findings, the increased osmotic potential and phosphorus content of the FCA-inoculated rice plant provide a protection sign against drought stress and will benefit food security in the future.

Drought stress is occurring frequently in the present scenario of global climate change^{1,2}. In the past decade, drought stress has led to the reduction in crop yield by 70%³. Meteorological drought, agricultural drought, hydrological drought, and socio-economic drought are the various effect types of droughts that pose a threat to global food security^{4,5}. In rice crop, poor grain production due to plant growth inhibition, grain sterility and yield loss upon the exposure of drought stress have been well established^{6,7}. Proper irrigation, based on water requirement of individual crop, is one of the most common strategies used to sustain crop productivity^{8–10}. Alternatively, addition of microorganism consortium to the soil has been reported as a sustainable way to improve soil physical and chemical properties, leading to more efficient water holding capacity and drought alleviation in the targeted area^{11–14}.

Furthermore, climate change and food security are two of the world's most pressing issues³. The use of arbuscular mycorrhizal fungi (AMF) is one of the most profitable ways to improve plant development for long-term sustainability^{15,16}. *Glomus mosseae*, also known as *Funneliformis mosseae*, is one of the most common AMF species used to improve chlorophyll content, photosynthetic efficiency, mineral nutrients, and shoot–root traits in host plants under stressful conditions, such as heavy metal¹⁷ and drought stress¹⁸. Besides, even under drought stress, a single inoculation of *Funneliformis mosseae* or *Funneliformis etunicatum* has additional benefits in terms of phosphorus regulation, iron (Fe) nourishment, and growth promotion in wheat¹⁹. In the case of an AMF consortium, a combination of several AMF species is frequently used to enhance its positive effects; for example, combining of *Funneliformis* plus with *Acaulospora* species significantly improves physiological and biochemical adaptabilities in *Sorghum bicolor* for drought²⁰. Similarly, inoculation with a mixture of 25 AMF species from

¹School of Interdisciplinary Studies (Kanchanaburi Campus), Mahidol University, Kanchanaburi 71150, Thailand. ²National Center for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency (NSTDA), Pathum Thani 12120, Thailand. ³Department of Chemical Engineering, Faculty of Engineering, Chulalongkorn University, Bangkok 10330, Thailand. [⊠]email: suravoot@gmail.com four genera, including *Funneliformis*, *Acaulospora*, *Gigaspora*, and *Scutellospora*, resulted in improved plant development²¹. AMF has been implicated in some mechanisms of a plant species' drought-tolerant abilities, including increased water and nutrient uptake, superior root architecture, higher root biomass, better osmotic adjustment, and antioxidant defense system enhancement^{22–25}. However, a question of plant survival and reproductive regulation effectiveness remains in the AMF-host symbiosis under drought conditions^{26,27}.

In AMF implementation in rice, lowland paddy fields flooded with water are usually unsuitable for AMF inoculation, and the diversity of AMF consortium suitable for these fields is determined by their anaerobic root zones^{26,28,29}. In recent studies, non-flooded upland paddy cultivation was chosen as a model system to understand the AMF-plant interaction under water deficit stress^{30–33}.

To investigate the regulation of physiological and biochemical attributes in rice (upland type) during the reproductive stage under water deficit, *Funneliformis mosseae* was chosen individually and in combination with *Claroideoglomus etunicatum* and *Acaulospora fovaeta*. According to the hypothesis, the AMF consortium with *Funneliformis mosseae* may be able to regulate host plant attributive mechanisms relating to grain production efficiency under water deficit stress.

Results

Changes in sugar and proline contents.. The AMF inoculations (F, FC, and FCA) were treated to the 'Leum Pua' rice under water deficit condition (WD). Under water well condition (WW), the total soluble sugar content of F inoculated and uninoculated plants was higher than that of FC and FCA-inoculated plants. Under WD stress, the total sugar content increased to 108, 110, 79.7, and 77.6 mg g⁻¹ DW in the uninoculated, F, FC, and FCA plants, respectively (Fig. 1c). The soluble sugar content of FCA inoculated plants in the external refer-



Figure 1. Total sugar soluble content and proline content of rice cv. Leum Pua inoculated with or without AMFs (*Funneliformis mosseae*; F, with *Claroideoglomus etunicatum*; C, and *Acaulospora fovaeta*; A, in F, FC, FCA, and uninoculated control, CT) under prior water deficit on day 0 (**a** and **b**) and day 21 of well-watered and water deficit conditions (**c** and **d**). Statistical data access with a minimum of n = 4 and mean ± standard deviation. Different letters along plots represent significant difference at p = 0.05.

ence control was found to be slightly lower, reflecting the previous stage of water deficit (Fig. 1a). Similarly, the free proline content in F inoculated and uninoculated plants under WD was recorded at 41.3 and 48.3 μ mol g⁻¹ FW, respectively, whereas the free proline content in FC and FCA plants was 15.4 and 15.6 mol g⁻¹ FW, respectively (Fig. 1d). Furthermore, in all three AMF inoculations (F, FC, and FCA), the proline content in the external reference control decreased when compared to uninoculated plants (Fig. 1b).

Enrichment of chlorophyll content, and growth. In the case of photosynthetic pigments, the three AMF inoculations had a significant impact on chlorophyll *b*, which was present between chlorophyll *a* and *b*. (Fig. 2a,b). Under the experimental conditions, the effects of the three AMF treatments on total chlorophyll content in host plants were significantly improved (Fig. 2c). However, the total chlorophyll content of the F inoculated plant was higher than that of the FC and FCA-inoculated plants under the WD, with values of 209.53, 176.72, and 190.09 μ g g⁻¹ FW, respectively. In addition, plant height of the FCA inoculated plant was increased (9.53% over the control plant) under WD (Supplementary Table S1), while a reduction was demonstrated in the F and FC inoculated plants. Dry matter content (DMC) in the shoot of host plants under WD was enhanced relative to the WW condition by 29, 77, and 51% in F, FC and FCA inoculated plants, respectively. Whereas, the DMC in the root tissues under the WD was lower in the FC, FCA inoculated plants, but higher in the F inoculated plant (41% increment) (Fig. 3a,b).

Variation of phosphorus content. When compared to the uninoculated plant, phosphorus content in the root was significantly improved in the F, FC, and FCA inoculated plants under two treatments, WW and WD. Whereas the shoot phosphorus content was higher in the FC and FCA inoculated plants under the WD, at 2.20 and 2.41 mg g⁻¹ DW, respectively, compared to the F inoculated plant (1.15 mg g⁻¹ DW). On the other hand, the phosphorus content in the shoots of the FC and FCA inoculated plants increased dramatically during the booting stage when exposed to the WD (Fig. 3c,d).

Divergent osmotic potential. Principal component analysis (PCA) was performed in terms of physiology and growth, photosynthetic pigments, proline content, total soluble sugar content, osmotic potential, and plant height (Fig. 4a,d). In particular, an upregulation of osmotic potential was observed in the FCA inoculated



Figure 2. Chlorophyll, Chl *a* and *b* (**a** and **b**) and total chlorophyll content (**c**) in rice cv. Leum Pua that inoculated with or without AMFs (F, FC, FCA, and CT) on the 21st day of well-watered and water deficit conditions. Statistical data access with a minimum of n = 4 and mean \pm standard deviation. Different letters along plots represent significant difference at p = 0.05.



Figure 3. Dry matter content (DMC) of shoot and root (**a** and **b**) and phosphorus content in the shoot and root (**c** and **d**) in cv. Leum Pua inoculated with and without AMFs (F, FC, FCA, and CT) on the 21st day of well-watered and water deficit conditions. Statistical data access with a minimum of n = 4 and mean ± standard deviation. Different letters along plots represent significant difference at p = 0.05.

.....

plant prior to the WD (Fig. 4a,c). Consequently, the characteristics of the FCA inoculated plant were separated via hierarchical clustering, whereas the F and FC inoculated plants were approached more closely with the uninoculated plant (Fig. 4b). During the WD period, the osmotic potential of the FCA inoculated plant significantly increased and distinguished from the others (Fig. 4d,e). The FCA-inoculated plants had the highest osmotic potential among the three treatments, measuring -0.73 and -0.29 Mpa under the WW and WD conditions, respectively. Under WW and WD, the osmotic potential of the FC-inoculated plant was lower (-1.55 and -1.49 MPa) than that of the F inoculated plant (-1.06 and -1.15 MPa) (Fig. 4e).

Reproductive yields and harvesting quality. Reproductive parameters, such as panicle weight, panicle length, grain weight, grain number per panicle, and percentage of grain maturity per panicle were maintained in the FCA treatment upon under the WD condition (Fig. 5a). In contrast, the F and FC inoculated plants under the WD had a non-dominant panicle weight and percentage of grain maturity compared to the FCA inoculated plant under the WD (Fig. 5a-c, Supplementary Fig. S2). Furthermore, an order of these eight plant treatments within the reproductive parameters, from FC_WD to FCA_WW, in Fig. 5a appeared to correspond to the groups of score plot of these plant treatments in Fig. 4d such as group of CT_WW, FCA_WW, and FCA_WD etc.

Different of the physiological, biochemical, and reproductive responses. Summarized PCAs reduced the dimensions of parameters such as osmotic potential, phosphorus content, chlorophyll, free proline, growth performance, yield traits such as plant height, shoot and root DMC, and the reproductive traits such as panicle length and panicle weight, percentage of grain maturity, and grain number and weight. Strength of these loading vectors suggests that reproductive traits are more closely associated in the FCA inoculated plant



Figure 4. Principle component analysis, PCA, and group clustering (**a** and **b**) in rice cv. Leum Pua inoculated with or without AMFs (F, FC, FCA, and CT; the codes are included in Fig. 1) indicated by physiological, biochemical, and growth parameters, including plot of osmotic potential, data sorted by means, (**c**) under prior water limitation (0 day). The PCA (**d**) and the osmotic potential (**e**) represents on the 21st day of well-watered and water deficit conditions. Statistical data access with a minimum of n = 4 and mean \pm standard deviation. Different letters along plots represent significant difference at p = 0.05.

in the two water regimes than in the FC and F inoculated plants (Fig. 6a,b). By the way, the F inoculated plant performed well through the yield trait in the vegetative part of root DMC under the WD, when compared to the FC and FCA inoculated plants (Fig. 6b). According to the summarized PCA, the relationship between the phosphorus content and the osmotic potential within the vegetative and reproductive yields was depicted in Fig. 7. In the three AMF treatments, the phosphorus content and osmotic potential were found to not to be directly related



Figure 5. Heat map correlation of panicle and grain production/quality (**a**), and plot of panicle weight and grain maturity per panicle (**b** and **c**) of rice cv. Leum Pua inoculated with or without AMFs (F, FC, FCA, and CT) on the 21st day of well-watered and water deficit conditions. Statistical data access with n = 15 and mean ± standard deviation. Different letters along plots represent significant difference at p = 0.05.



Figure 6. Principle component analysis depicting score and loading plots of the experimental physiological and biochemical parameters, growth, panicle-grain production/quality, and dry matter content (DMC) in rice cv. Leum Pua inoculated with or without AMFs (F, FC, FCA, and CT) and on the 21st day of well-watered (WW, **a**) and water deficit (WD, **b**) conditions.



Figure 7. Clustering of separated well-watered (**a**) and water deficit (**b**) conditions in relationship of osmotic potential and phosphorus content into the reproductive parts (panicle – grain production, quality) and vegetative (dry matter content, DMC, of shoot and root) parts in rice cv. Leum Pua inoculated with or without AMFs (F, FC, FCA, and CT). PCA summary (**c**) in effect of the AMF colonization on osmotic potential, phosphorus (P) and reproductive yields. Systematic diagram jigsaws (**d**) depicting the effect of osmotic potential and phosphorus regulation by the AMF (FCA type) on rice yield maintenance under water deficit conditions.

to the reproductive yields under the WW. At the site of the WD, the osmotic potential and phosphorus content, especially in root, were closely related to panicle weight and grain number, respectively (Fig. 7a,b).

While emphasizing the influence of the AMF colonization on the osmotic potential, phosphorus content, and reproductive yields. The findings revealed a high relationship between the root phosphorus content and the colonization in the WW and WD conditions (Pearson coefficient; R > 0.8, Supplementary Table S2). Furthermore, when compared to the three inoculations in the WW and WD, the FCA inoculation showed a trend to towards the colonization influence rather than the others (Fig. 7c). It may involve a slightly high level in the percentage

of the FCA colonization under the WW and WD (Supplementary Fig. S1). However, the three inoculated plants showed the different regulation, especially in the plant physiology and yields.

Discussion

Both single and consorted inoculations of AMF have been used to promote plant survival under drought stress^{27,34}. In the present study, the water deficit stressed plants of rice cv. Leum Pua at booting stage were treated with different AMF inoculations, i.e., F (*Funneliformis mosseae*), FC (F and *Claroideoglomus etunicatum*), and FCA (F, C and *Acaulospora foveate*). Total soluble sugar content and free proline content in the F-inoculated plant under the WD were highly accumulated, among the three inoculated plants. In the case of the F inoculated plant, the adjustments of soluble sugars and proline should result in the stability of the NADP + /NADPH ratio, which acts as osmoprotectants to avoid stress in the host plant under drought stress^{21,35,36}. Furthermore, the free proline subsequence stimulates energy for root biomass accumulation, which improves water accessibility in soil³⁷. Only the F inoculated plant showed the increase in the root traits were enhanced by *Funneliformis mosseae* symbiosis in trifoliate orange under drought condition. This indicates that F inoculation could help promote root development in rice crops via catalytic function and energy regulation.

Responses of the FC and FCA inoculated plants under the WD seem to be in contrast with the F inoculated plant, due to the lower accumulation of total sugar soluble content and free proline. The total sugar soluble content of FC and FCA inoculated plants in the WD was more upregulated than in the WW condition. Lower variation in the free proline content was also found in mycorrhizal symbiosis in macadamia, and thus, total soluble sugar was suggested as the major osmolyte²¹. Furthermore, Wu et al.³⁸ announced that the osmolytes for water balance in plant cells may also originate from total non-structural carbohydrates and ions, *i.e.*, K⁺, Ca²⁺ and Mg²⁺, such as mycorrhizal inoculated citrus. In this case, the lower free proline content under drought was related to the proline biosynthesis and turnover reflected in buffer cellular redox status, resulting in maintenance of plant growth during water shortage³⁹. Besides, the low carbohydrate accumulation in plant 'source' under drought (post-anthesis stage) could result in the growth of 'sink' in their shoots, such as panicle dry weight in Sorghum⁴⁰. Between the free proline and carbohydrate content, here, the soluble sugar content might be required to generate a cellular homeostasis balance for the FC and FCA inoculated plants under the WD, involving drought susceptibility.

From the results of this study, the rice plants inoculated with the F only and those with two AMF consortiums are confirmed to have better chlorophyll pigments under both of the WW and WD. In agreement, the maize-*Funneliformis mosseae* symbiosis has been reported to lead to an increased chlorophyll content under irrigation⁴¹. Furthermore, *F. mosseae* has been proposed as a preferred partner in physiological traits in C3 plants, including photosynthetic pigments⁴². It is suggested that *F. mosseae* has the potential to be a major inoculation for chlorophyll pigment improvement in plants.

In general studies of AMF colonization and phosphate induction¹⁵, the three AMF symbiotic relationships in the WW and WD were confirmed here, and their priorities were correlated to phosphorus content in the plant host, especially in the rice root (Supplementary Table S2). However, the phosphorus content in the shoot and root of the FC and FCA inoculated plants under the WD was higher than in the F inoculated plant. In particularly, the phosphorus content in the shoot was remarkably enriched by the combinations of FC and FCA inoculation. The results are related to the report by Kobae⁴³ who reported that the mosaic of diverse mycorrhizas led to an increased ability of phosphate-uptake performance, and it is possible that the common mycorrhizal networks (CMNs) might share cellular components such as nuclease and other organelles in the coenocytic mycelia. There has been a tangible association in mycorrhizal communities between mycorrhizal functional diversity and plant nutrient stoichiometry, including phosphorus^{44,45}. As the results of the improvement of phosphorus content, the FC and FCA are thought to be more related to rice nutrient availability under the WD. Phosphorus concentration helps in regulation of shoot biomass, which is important for the production of high fertile spikes and better grain yield in winter wheat⁴⁶. Thus, the increased plant growth and yield traits are strongly correlated with the increase in phosphorus uptake in AMF inoculated plants^{47,48}. Then, the phosphorus content in both the FC and FCA inoculated plants is anticipated to be associated with shoot biomass, relating vegetative dry matter content and reproductive yield.

Despite the phosphorus replenishment, the FC inoculated plant did not improve its grain yield traits under the WD when compared to the FCA inoculated plant. This may involve in the difference of the osmotic potential in the FC and FCA plants as shown in Fig. 4. The upregulation of osmotic potential in the FCA inoculated plant should due to FCA combination, producing remarkable reproductive traits, especially those related to panicle and grain yield under the WD and WW conditions. Previous research found that osmotic potential and phosphorus content in WD were closely related to reproductive traits in *Spartina alterniflora* (when compared to WW)⁴⁹. Sawwan et al.⁵⁰ suggested that an upregulation of osmotic potential was very efficient under enhanced phosphorus content, as indicated in cell sap of an African violet. Phosphorus also induces a cell hydraulic state in the leaf tissues⁵¹. Based on the findings of the previous studies and the present study, it can be suggested that the FCA colonization should stimulate phosphorus accumulation, particularly an increase in osmotic potential during drought, which may facilitate solute allocation from root to shoot and panicle, potentially leading to drought resistance with reproductive yield, as shown in Fig. 7 in the FCA inoculated plant under the WD. The balance of the osmotic potential and phosphorus content are supposed to be important regulation for maintaining plant growth and grain development under WD stress in rice species. There is a high possibility that an effective AMF inoculation strategy might secure a reproductive rice yield during a drought situation.

Conclusions

The potential of the consortium of AMF *Funneliformis mosseae* in the upland rice, Leum Pua cultivar, under water deficit was demonstrated. Total chlorophyll content in the flag leaf of rice inoculated with F, FC, and FCA was promoted under WD. However, the responses of total soluble sugar and free proline were upregulated in the F-inoculated plant and downregulated in FC and FCA-inoculated plants, leading to drought regulations. Both FC and FCA inoculations could be used for a remarkable increase in the phosphorus level, especially in the leaf tissues of host plants under WD stress. In addition, the combination of FCA resulted in osmotic potential adaptability in the host plant under the WD condition. It is concluded that the increased osmotic potential and phosphorus content of the FCA inoculated rice plants provide protection against water deficit stress. Therefore, further development of drought-resistant crops should focus on regulatory signaling mechanisms involved in osmotic potential and phosphorus adjustment.

Materials and methods

Preparation of plant materials and AMF inoculation. Rice (*Oryza sativa* subsp. indica: cy. Leum Pua; upland black sticky rice) seeds procured through collection by the Innovative Plant Genetic and Physiology Research, National Center for Genetic Engineering and Biotechnology (BIOTEC), Thailand, were selected as the initial plant material in the present study. The common and predominant cultivar of black sticky rice used in the study is identified as glutinous rice⁵² and enriched with anthocyanins⁵³. Rice seeds were sown on the soil substrate for a month. Rice seedlings were directly transferred into $10 \times 12.5 \times 15$ cm (width × length × height) of plastic bags filled with autoclaved garden soil ($EC = 2.7 \text{ dS m}^{-1}$; pH = 5.7; total organic carbon = 12.3%; available N=0.3 mg kg⁻¹; available P=578 mg kg⁻¹; available K=3073 mg kg⁻¹; available Ca=7020 mg kg⁻¹; available Mg = 1034 mg kg⁻¹). According to arbuscular mycorrhiza fungi, the mycorrhiza powder of Funneliformis mosseae (F), Claroideoglomus etunicatum (C) and Acaulospora fovaeta (A) were obtained from Maejo University. All the three AMF have been primarily tested as single purified strains on a monocot species and the F showed the maximum benefit to their host⁴¹. On this basis individual F and consortium of FC, and FCA were used as treatments in the present study. For AMF inoculation, average 200 spores of each AMF were used. These AMFs and an organic fertilizer (Bua^{*} chicken yard manner, Charoen Pokphand Group)⁵⁴ were provided at the time of seedling transplantation. Then, the rice plants, at V_4 stage; formation of four leaf on main stem, in the plastic bags were settled in 4×2 factorials in completely randomized design (CRD) of four inoculated treatments (control, and the three of AMFs) and two conditions of water regime (water well, WW and water deficit, WD). The rice was cultivated under the greenhouse conditions until its transition to the booting stage. The temperature in the greenhouse was set at 26 ± 2 °C (nighttime)/ 32 ± 2 °C (daytime) and the relative humidity at $80 \pm 5\%$. After the booting stage, the uniform rice for 20 plants were selected at day 0 (day of starting in water withholding). The 40 bags (ten bags per each AMF treatment) were divided into a group of WW which continue watering, and a group of WD. The group of WD was withheld the water until 21 days. At the 21st day, the percentage of soil moisture content (by drying method²¹) was reported to $55.76 \pm 3.12\%$ for the WW, and $14.84 \pm 2.14\%$ for the WD. The plant samples of the two water-regimes were collected. Both sample collections of the 0 day and 21 days were used for physiological, biochemical, and morphological growth trait measurements. While, other 40 bags in the WW, and WD (re-watering after 21 days) conditions remained for yield traits at the harvesting period.

Determination of total soluble sugar and free proline content. Total soluble sugar content was determined from the flag leaf of rice. Fifty milligrams of flag leaf samples were ground with liquid nitrogen. One mL of nano-pure water was added to the sample for extraction in a 1.5 mL plastic tube that modified from Karkacier et. al.⁵⁵. After centrifugation at $10,000 \times g$ for 15 min, the extracted solution was collected and filtrated through a 0.45 µm membrane filter (VertiPure[™], Vertical^{*}). Then, the 20 µL of filtrated supernatant was analyzed by a High-Performance Liquid Chromatography (HPLC) system (WatersTM410, Massachusetts, US) equipped with a MetaCarb 87C column, using a differential refractometer detector. A mobile phase was deionized water, and the flow rate was set at 0.5 mL min⁻¹. The standard curves of sucrose, glucose, and fructose (Fluka, US) were used to calculate total soluble sugar content⁵⁵.

The analysis of free proline content was conducted with the flag leaf samples. The freeze-dried flag leaf tissues were ground to the powder using liquid nitrogen, and 50 mg of sample were mixed into 1 mL aqueous sulfosalicylic acid (3%, *w/v*). The supernatant was separated and added with the same volume of glacial acetic acid and ninhydrin reagent, then heated at 95 °C in a water bath for 1 h and kept on an ice box to terminate the reaction for 15 min. The solution was then mixed with 2 mL of toluene to precipitate the free proline, and the chromophore absorbance at 520 nm was measured on a UV–Vis spectrophotometer (HACH DR/4000; Model 48,000, HACH Company, Loveland, Colorado, US). The content of free proline was estimated against the L-proline standard curve between 0 and 500 μ M⁵⁶.

Elucidation of chlorophyll content. Analysis of chlorophyll content was performed as per the protocol modified from Shabala et al.⁵⁷ and Lichtenthaler et al.⁵⁸. The flag leaf samples were collected and chopped into small pieces. After that, 100 mg of sample were transferred to a 25 mL glass vial (Opticlear*; KIMBLE, Vineland, NJ, USA), 10 mL of 95.5% acetone was added, and the mixture was homogenized with a homogenizer (T25 basic Ultra-Turrax*; IKA, Kuala Lumpur, Malaysia). The solution was kept in the sealed glass vial with a plastic cap and incubated at 4 °C in the refrigerator for 48 h. The extracted solution was read by a UV–VIS spectrophotometer (DR/4000; Model 48000Hatch, Loveland, CO, USA) at 662 and 645 nm, and then chlorophyll *a*, *b* and total chlorophyll content were calculated.

Analysis of phosphorus content. The plants were separated into shoots and roots, and oven-dried at 80 °C for 3 days. The 0.5 g of the powdered samples was digested by a nitric acid solution using microwave digestion. The digested solutions were subjected to molybdenum blue reaction and their absorbance was read at 420 nm using spectrophotometer [DR/4000; Model 48,000, (HACH.), USA]⁵⁹.

Osmotic potential determination. Osmotic potential was investigated in according to Lanfermeijer et al.⁶⁰ method. Hundred milligrams of fresh flag leaf samples were manually crushed using glass rod in 1.5 mL plastic tube, and the 20 μ L of the extracted solution was directly dropped on a filter paper and incubated osmometer chamber (5520 Vapro*, Wescor, Utah, USA). Finally, the millimolar per kg of osmotic potential data was converted into osmotic potential in MPa in according to Fu et al.⁶¹.

AMF colonization and growth measurement. According to Brundrett et al.⁶², root samples of the rice were cleaned with tap water, followed by distilled water. In 60 percent ethanol, one centimeter of the root sample was reserved. The root was then washed three times with distilled water before being immersed in 10% KOH at 95 °C for 30 min. After that, the cleaned root was processed with 0.05% (w/v) Trypan blue for 15 min. The AMF colonization was observed under a light microscope (Zeiss, Germany) and calculated in percentage (Supplementary Fig. S1).

Plant height and dry mass per fresh mass in the rice shoot and root were measured following the IRRI protocol⁶³. At the harvesting stage, panicle length, panicle weight, total grain number per panicle, grain weight, and percentage of grain maturity by filled grains per panicle were evaluated.

Data analytics and statistical analysis. According to the information and Table 1, the data were statistically analyzed using Jamovi (v. 2.2) (https://www.jamovi.org). The physiological, biochemical, and morphological data in the present study were analyzed at least four technical replications, a replication per individual plant sample. The plant sampling was varied among each rice plant in those groups of treatments. In reproductive traits, fifteen panicles, per treatment, were used for yield traits. External references included the total soluble sugar content, proline content, and osmotic potential at the beginning (zero day; prior day of water deficit). The external reference was subjected to principal component analysis (PCA) and hierarchical clustering. Induvial box plot was shown for osmotic potential and yield traits with mean (dot), median line, at 25–75% of interquartile range with standard deviation calculation. A heat map dendrogram was created for rice production under WW and WD conditions. PCA was used to summarize the effect of water stress on physiological and biochemical factors, including parts of yield traits and colonization percentage. The dendrograms displayed phosphorus content and osmotic potential data related to rice production. Furthermore, Tukey's honestly significant difference was used in SPSS v.18 to identify a post-hoc comparison test with analysis of variance.

Complies with international, national and/or institutional guidelines. The experimental research and field studies on plants, including materials, reported here comply with all relevant institutional, national, and international guidelines and legislation.

Data availability

The data underlining this article are included in the manuscript, and the raw data will be made available to the corresponding author upon reasonable request.

Parameter test	Т	Con	T x Con
Total soluble sugar content	0.003	< 0.001	ns
Proline content	< 0.001	< 0.001	< 0.001
Chlorophyll a content	ns	ns	ns
Chlorophyll b content	< 0.001	ns	ns
Total chlorophyll content	0.013	ns	ns
Shoot dry matter content	0.020	< 0.001	< 0.001
Root dry matter content	ns	0.012	< 0.001
Shoot phosphorus	0.018	0.004	0.009
Root phosphorus	< 0.001	< 0.001	0.001
Osmotic potential	< 0.001	ns	ns
Panicle weight	< 0.001	< 0.001	ns
Grain maturity	< 0.001	< 0.001	ns

Table 1. Analyses of variance depicting the effect of inoculated treatment (T), two watering conditions (Con), and their interaction on the physiological and biochemical parameters in the 21-day water limitation and the production in harvesting state. The numbers indicate significant difference, and ns represent non-significant difference.

Received: 14 August 2022; Accepted: 10 April 2023 Published online: 12 April 2023

References

- 1. Trenberth, K. E. et al. Global warming and changes in drought. Nat. Clim. Change. 4, 17–22. https://doi.org/10.1038/nclimate20 67 (2014).
- Naumann, G. et al. Global changes in drought conditions under different levels of warming. Geophys. Res. Lett. 45, 3285–3296. https://doi.org/10.1002/2017GL076521 (2018).
- Raza, A. et al. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. Plants 8, 34. https:// doi.org/10.3390/plants8020034 (2019).
- Caball, R. & Malekpour, S. Decision making under crisis: Lessons from the millennium drought in Australia. Int. J. Disaster Risk Reduct. 34, 387–396. https://doi.org/10.1016/j.ijdrr.2018.12.008 (2019).
- Barzkar, A., Najafzadeh, M. & Homaei, F. Evaluation of drought events in various climatic conditions using data-driven models and a reliability-based probabilistic model. *Nat. Hazards.* 110, 1931–1952. https://doi.org/10.1007/s11069-021-05019-7 (2022).
- 6. Mukamuhirwa, A. *et al.* Concurrent drought and temperature stress in rice—A possible result of the predicted climate Change: Effects on yield attributes, eating characteristics, and health promoting compounds. *Int. J. Environ. Res. Public Health.* **16**, 1043. https://doi.org/10.3390/ijerph16061043 (2019).
- 7. Yu, C. Assessing the impacts of extreme agricultural droughts in China under climate and socioeco-nomic changes. *Earth's Future* 6, 689–703. https://doi.org/10.1002/2017EF000768 (2018).
- Ewaid, S. H., Abed, S. A. & Al-Ansari, N. Crop water requirements and irrigation schedules for some major crops in Southern Iraq. Water 11, 756. https://doi.org/10.3390/w11040756 (2019).
- 9. Sharma, V. Impact climate change on crop water requirement of different orchard crops for agro-climatic condition of Udaipur. Rajasthan. Indian J. Ecol. 47, 12–16 (2020).
- Zhang, P., Ma, W., Hou, L., Liu, F. & Zhang, Q. Study on the spatial and temporal distribution of irrigation water requirements for major crops in Shandong province. Water 14, 1051. https://doi.org/10.3390/w14071051 (2022).
- 11. Ahluwalia, O., Singh, P. C. & Bhatia, R. A review on drought stress in plants: Implications, mitigation and the role of plant growth promoting rhizobacteria. *Resour. Environ. Sustainabil.* **5**, 100032. https://doi.org/10.1016/j.resenv.2021.100032 (2021).
- Aslam, M. M. et al. Rhizosphere microbiomes can regulate plant drought tolerance. Pedosphere 32, 61–74. https://doi.org/10.1016/ S1002-0160(21)60061-9 (2022).
- Etesami, H., Li, Z., Maathuis, F. J. & Cooke, J. The combined use of silicon and arbuscular mycorrhizas to mitigate salinity and drought stress in rice. *Environ. Exp. Bot.* 201, 104955. https://doi.org/10.1016/j.envexpbot.2022.104955 (2022).
- Thangavel, P. et al. Arbuscular mycorrhizae: Natural modulators of plant-nutrient relation and growth in stressful environments. Arch. Microbiol. 204, 1–22. https://doi.org/10.1007/s00203-022-02882-1 (2022).
- Lanfranco, L., Fiorilli, V. & Guijahr, C. Partner communication and role of nutrients in the arbuscular mycorrhizal symbiosis. New Phytol. 220, 1031–1046. https://doi.org/10.1111/nph.15230 (2018).
- Tisarum, R., Samphumphuang, T., Yooyoungwech, S., Singh, H. P. & Cha-um, S. Arbuscular mycorrhizal fungi modulate physiological and morphological adaptations in para rubber tree (*Hevea brasiliensis*) under water deficit stress. *Biologia* 77, 1723–1736. https://doi.org/10.1007/s11756-022-01016-8 (2022).
- 17. Zhang, H. *et al.* Arbuscular mycorrhizal fungi (*Glomus mosseae*) improves growth, photosynthesis and protects photosystem II in leaves of *Lolium perenne* L. in cadmium contaminated soil. *Front. Plant Sci.* **9**, 1156. https://doi.org/10.3389/fpls.2018.01156 (2018).
- Cheng, H. Q., Zou, Y. N., Wu, Q. S. & Kuca, K. Arbuscular mycorrhizal fungi alleviate drought stress in trifoliate orange by regulating H⁺-ATPase activity and gene expression. *Front. Plant Sci.* 12, 659694. https://doi.org/10.3389/fpls.2021.659694 (2021).
- Al-Karaki, G., McMichael, B. & Zak, J. Field response of wheat to arbuscular mycorrhizal fungi and drought stress. *Mycorrhiza* 14, 263–269. https://doi.org/10.1007/s00572-003-0265-2 (2004).
- Neumann, E. & George, E. Colonisation with the arbuscular mycorrhizal fungus *Glomus mosseae* (Nicol. & Gerd.) enhanced phosphorus uptake from dry soil in *Sorghum bicolor* (L.). *Plant Soil.* 261, 245–255. https://doi.org/10.1023/B:PLSO.0000035573. 94425.60 (2004).
- Yooyongwech, S., Phaukinsang, N., Cha-um, S. & Supaibulwatana, K. Arbuscular mycorrhiza improved growth performance in Macadamia tetraphylla L. grown under water deficit stress involves soluble sugar and proline accumulation. Plant Growth Regul. 69, 285–293. https://doi.org/10.1007/s10725-012-9771-6 (2013).
- Wu, Q. S., Srivastava, A. K. & Zou, Y. N. AMF-induced tolerance to drought stress in citrus: A review. Sci. Hortic. 164, 77–87. https://doi.org/10.1016/j.scienta.2013.09.010 (2013).
- Pavithra, D. & Yapa, N. Arbuscular mycorrhizal fungi inoculation enhances drought stress tolerance of plants. Groundw. Sustain. Dev. 7, 490–494. https://doi.org/10.1016/j.gsd.2018.03.005 (2018).
- Petipas, R. H., González, J. B., Palmer, T. M. & Brody, A. K. Habitat-specific AMF symbioses enhance drought tolerance of a native Kenyan grass. Acta Oecol. 78, 71–78. https://doi.org/10.1016/j.actao.2016.12.005 (2017).
- Begum, N. et al. Improved drought tolerance by AMF inoculation in maize (Zea mays) involves physiological and biochemical implications. Plants 8, 579. https://doi.org/10.3390/plants8120579 (2019).
- Bao, X., Wang, Y. & Olsson, P. A. Arbuscular mycorrhiza under water–Carbon–phosphorus exchange between rice and arbuscular mycorrhizal fungi under different flooding regimes. *Soil Biol. Biochem.* 129, 169–177. https://doi.org/10.1016/j.soilbio.2018.11. 020 (2019).
- 27. Begum, N. *et al.* Role of arbuscular mycorrhizal fungi in plant growth regulation: Implications in abiotic stress tolerance. *Front. Plant Sci.* **10**, 1068. https://doi.org/10.3389/fpls.2019.01068 (2019).
- Sarkodee-Addo, E. et al. Arbuscular mycorrhizal fungi associated with rice (Oryza sativa L.) in Ghana: effect of regional locations and soil factors on diversity and community assembly. Agronomy 10, 559. https://doi.org/10.3390/agronomy10040559 (2020).
- Wang, Y., Bao, X. & Li, S. Effects of arbuscular mycorrhizal fungi on rice growth under different flooding and shading regimes. Front. Microbiol. 12, 756752. https://doi.org/10.3389/fmicb.2021.756752 (2021).
- Lumini, E., Vallino, M., Alguacil, M. M., Romani, M. & Bianciotto, V. Different farming and water regimes in Italian rice fields affect arbuscular mycorrhizal fungal soil communities. *Ecol. Appl.* 21, 1696–1707. https://doi.org/10.1890/10-1542.1 (2011).
- Bernaola, L. et al. Natural colonization of rice by arbuscular mycorrhizal fungi in different production areas. *Rice Sci.* 25, 169–174. https://doi.org/10.1016/j.rsci.2018.02.006 (2018).
- 32. Parvin, S. *et al.* A comparison of the arbuscular mycorrhizal fungal communities among Bangladeshi modern high yielding and traditional rice varieties. *Plant Soil.* **462**, 109–124. https://doi.org/10.1007/s11104-021-04858-4 (2021).
- Surendirakumar, K., Pandey, R. R. & Muthukumar, T. Arbuscular mycorrhizal fungi in roots and rhizosphere of black rice in terrace fields of North-East India. Proc. Natl. Acad. Sci. India Sect B Biol. Sci. 91, 277–287. https://doi.org/10.1007/s40011-020-01221-y (2021).
- 34. Boutasknit, A. *et al.* Arbuscular mycorrhizal fungi mediate drought tolerance and recovery in two contrasting carob (*Ceratonia siliqua* L.) ecotypes by regulating stomatal, water relations, and (in)organic adjustments. *Plants* **9**, 80. https://doi.org/10.3390/plant s9010080 (2020).

- Wu, H. H., Zou, Y. N., Rahman, M. M., Ni, Q. D. & Wu, Q. S. Mycorrhizas alter sucrose and proline metabolism in trifoliateorange exposed to drought stress. Sci. Rep. 7, 42389. https://doi.org/10.1038/srep42389 (2017).
- Azmat, R. & Moin, S. The remediation of drought stress under VAM inoculation through proline chemical transformation action. J. Photochem. Photobiol. B Biol. 193, 155–161. https://doi.org/10.1016/j.jphotobiol.2019.03.002 (2019).
- Kishor, P. B. K. & Sreenivasulu, N. Is proline accumulation per se correlated with stress tolerance or is proline homoeostasis a more critical issue?. *Plant Cell Environ.* 37, 300–311. https://doi.org/10.1111/pce.12157 (2014).
- Wu, Q. S. & Xia, R. X. Arbuscular mycorrhizal fungi influence growth, osmotic adjustment and photosynthesis of citrusunder well-watered and water stress conditions. J. Plant Physiol. 163, 417–425. https://doi.org/10.1016/j.jplph.2005.04.024 (2006).
- Bhaskara, G. B., Yang, T. H. & Verslues, P. E. Dynamic proline metabolism: Importance and regulation in water limited environments. *Front. Plant Sci.* 6, 484. https://doi.org/10.3389/fpls.2015.00484 (2015).
- Tovignan, T. K. *et al.* Terminal drought effect on sugar partitioning and metabolism is modulated by leaf stay-green and panicle size in the stem of sweet sorghum (*Sorghum bicolor L. Moench*). *CABI Agric. Biosci.* 1, 1–11. https://doi.org/10.1186/s43170-020-00003-w (2020).
- Yooyongwech, S. *et al.* Matching of nitrogen enhancement and photosynthetic efficiency by arbuscular mycorrhiza in maize (*Zea mays* L.) in relation to organic fertilizer type. *Plants* 11, 369. https://doi.org/10.3390/plants11030369 (2022).
- Chandrasekaran, M., Chanratana, M., Kim, K., Seshadri, S. & Sa, T. Impact of arbuscular mycorrhizal fungi on photosynthesis, water status, and gas exchange of plants under salt stress—A meta-analysis. *Front. Plant Sci.* 10, 457. https://doi.org/10.3389/fpls. 2019.00457 (2019).
- Kobae, Y. Dynamic phosphate uptake in arbuscular mycorrhizal roots under field conditions. Front. Environ. Sci. 6, 159. https:// doi.org/10.3389/fenvs.2018.00159 (2019).
- He, Y. Q., Zhu, Y. G., Smith, S. E. & Smith, F. A. Interactions between soil moisture content and phosphorus supply in spring wheat plants growth in pot culture. J. Plant. Nutri. 25, 913–925. https://doi.org/10.1081/PLN-120002969 (2002).
- Powell, J. R. & Rillig, M. C. Biodiversity of arbuscular mycorrhizal fungi and ecosystem function. New Phytol. 220, 1059–1075. https://doi.org/10.1111/nph.15119 (2018).
- 46. Chen, X. X. *et al.* Physiological and developmental traits associated with the grain yield of winter wheat as affected by phosphorus fertilizer management. *Sci. Rep.* **9**, 16580. https://doi.org/10.1038/s41598-019-53000-z (2019).
- Treseder, K. K. The extent of mycorrhizal colonization of roots and its influence on plant growth and phosphorus content. *Plant Soil.* 371, 1–13. https://doi.org/10.1007/s11104-013-1681-5 (2013).
- Mai, W., Xue, X., Feng, G. & Tian, C. Simultaneously maximizing root/mycorrhizal growth and phosphorusuptake by cotton plants by optimizing water and phosphorus management. *BMC Plant Biol.* 18, 334. https://doi.org/10.1186/s12870-018-1550-8 (2018).
- Hessini, K. et al. Effect of water stress on growth, osmotic adjustment, cell wall elasticity and water-use efficiency in Spartina alterniflora. Environ. Exp. Bot. 67, 312-319. https://doi.org/10.1016/j.envexpbot.2009.06.010 (2009).
- Sawwan, J., Shibli, R. A., Swaidat, I. & Thahat, M. Phosphorus regulates osmotic potential and growth of African violet under in vitro-induced water deficit. J. Plant Nutr. 23, 759–771. https://doi.org/10.1080/01904160009382057 (2000).
- Kaci, H. S., Chaker-Haddadj, A., Nedir-Kichou, A. & Aid, F. Act of phosphorus on cell hydraulic state, K⁺ use efficiency and induction of positive correlations between yield and vegetative traits in chickpea. *Acta Agric. Scand. Sect B Soil Plant Sci.* 72, 325–332. https://doi.org/10.1080/09064710.2021.1988138 (2022).
- Pornputtapitak, W., Pantakitcharoenkul, J., Panpakdee, R., Teeranachaideekul, V. & Sinchaipanid, N. Development of γ-oryzanol rich extract from Leum Pua glutinous rice bran loaded nanostructured lipid carriers for topical delivery. J. Oleo. Sci. 67, 125–133. https://doi.org/10.5650/jos.ess17113 (2018).
- Peanparkdee, M., Patrawart, J. & Iwamoto, S. Effect of extraction conditions on phenolic content, anthocyanin content and antioxidant activity of bran extracts from Thai rice cultivars. J. Cereal Sci. 86, 86–91. https://doi.org/10.1016/j.jcs.2019.01.011 (2019).
- Pitaktamrong, P., Kingkaew, J., Yooyongwech, S., Cha-um, S. & Phisalaphong, M. Development of arbuscular mycorrhizal fungiorganic fertilizer pellets encapsulated with alginate film. *Eng. J.* 22, 65–79. https://doi.org/10.4186/ej.2018.22.6.65 (2018).
- Karkacier, M., Erbas, M., Uslu, M. K. & Aksu, M. Comparison of different extraction and detection methods for sugars using amino-bonded phase HPLC. J. Chromatogr. Sci. 41, 331–333. https://doi.org/10.1093/chromsci/41.6.331 (2003).
- Bates, L. S., Waldren, R. P. & Teare, I. D. Rapid determination of freeproline forwater-stress studies. *Plant Soil.* 39, 205–207. https:// doi.org/10.1016/j.dental.2010.07.006 (1973).
- Shabala, S. N., Shabala, S. I., Martynenko, A. I., Babourina, O. & Newman, I. A. Salinity effect on bioelectric activity, growth, NaC accumulationand chlorophyll fluorescence of maize leaves: A comparative survey and prospects for screening. *Aust. J. Plant Physiol.* 25, 609–616. https://doi.org/10.1071/PP97146 (1998).
- Lichtenthaler, H. K. Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Method Enzymol.* 148, 350–380. https://doi.org/10.1016/0076-6879(87)48036-1 (1987).
- 59. Jackson, M. L. Soil Chemical Analysis (Prentice Hall, 1958).
- Lanfermeijer, F. C., Koerselman-Kooij, J. W. & Borstlap, A. C. Osmo-sensitivity of sucrose uptake by immature pea cotyledons disappears during development. *Plant Physiol.* 95, 832–838. https://doi.org/10.1104/pp.95.3.832 (1991).
- Fu, J., Huang, B. & Fry, J. Osmotic potential, sucrose level, and activity of sucrose metabolic enzymes in tall fescue in response to deficit irrigation. J. Am. Soc. Hort. Sci. 135, 506–510. https://doi.org/10.21273/JASHS.135.6.506 (2010).
- 62. Brundrett, M., Bougher, N., Dell, B., Grove, T. & Malajczuk, N. Working with mycorrhizas in forestry and agriculture. (ACIAR Monograph 32, 1996).
- 63. IRRI. Standard evaluation system for rice. (International Rice Research Institute, 2002).

Acknowledgements

The authors would like to thank Associate Professor Supatida Aumtong of Maejo University for providing the mycorrhizas used in the study, as well as Assistant Professor Waraporn Threeprom, Miss Naphattarakan Eaimsaard and Miss Papassara Bumroongsak for technical assistance.

Author contributions

Conceptualization, S.Y., S.C.; methodology, S.Y., S.C.; software and secondary analysis, S.Y.; formal investigation, R.T., T.S.; data curation, S.Y., S.C. writing—original manuscript draft, S.Y., S.C.; review and editing, S.Y., S.C., M.P.; consultant and supervision S.C., M.P.

Funding

This project was funded by the Thailand Research Fund (grant no. SRI5920201). The APC patron promoted by Research Management and Development Division, Central Mahidol University, and the School of Interdisciplinary Studies, Kanchanaburi campus.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at https://doi.org/ 10.1038/s41598-023-33304-x.

Correspondence and requests for materials should be addressed to S.Y.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2023