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Fe3O4@nano‑almondshell/ OPEN Si(CH2)3/2‑(1‑piperazinyl) ethylamine as an efective magnetite almond shell‑based nanocatalyst for the synthesis of dihydropyrano[3,2‑*c***]chromene and tetrahydrobenzo[***b***]pyran derivatives**

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The preparation and design of nano-catalysts based on magnetic biopolymers as green and biocompatible nano-catalysts have made many advances. This paper deals with the preparation of magnetite biopolymer-based Brønsted base nano-catalyst from a nano-almond (*Prunus dulcis***) shell. This magnetite biopolymer-based nano-catalyst was obtained through a simple process based on the core-shelling of nano-almond shell and Fe3O4 NPs and then the immobilization of 3-chloropropyltrimethoxysilane as linker and 2-aminoethylpiperazine as a basic section. Structural and morphological analysis of this magnetite biopolymer-based nano-catalyst were done using Fourier transform infrared spectroscopy, feld emission scanning electron microscopy, X-ray difraction, Thermogravimetric analysis, Vibrating sample magnetization, Energy-dispersive X-ray spectroscopy, Brunauer–Emmett–Teller, and Transmission electron microscopy techniques. The performance of the synthesized Fe3O4@nano-almondshell/Si(CH2)3/2-(1-piperazinyl)ethylamine as a novel magnetite biopolymer-based nano-catalyst for the synthesis of dihydropyrano[3,2-***c***]chromene** and tetrahydrobenzo[b]pyran was investigated and showed excellent efficiency.

In recent years, extensive research has been conducted on new polymer-based nano-catalysts. New polymers known as biopolymers will be synthesized from non-edible and highly available plants as well as agricultural and industrial wastes^{1-[3](#page-13-1)}. In addition, some biopolymers can be obtained from renewable sources. These biopolymers include polysaccharides (cellulose, dextrin, chitosan, etc.), protein polymers (gluten, ovalbumin, soy protein, collagen, etc.), bacterial protein (3-hydroxybutyrate), and other polymers^{[4](#page-13-2)}. Among these biopolymers, cellulose, and derivatives become important due to their high fexibility, abundance, chemical inertness, high strength, and ability to modify surface chemistry⁵⁻⁸. The almond (Prunus dulcis) shell is a highly efficient biomass shell and is generally disposed of or incinerated as waste, which causes environmental pollution^{[9](#page-13-5)}. Almond shells make up about 35–75% of the total weight of the fruit. Tis volume of the shell has a high practical potential that has attracted a lot of attention in recent year[s10,](#page-13-6)[11.](#page-13-7) Senturk et al. used the almond shell as an adsorbent to remove rhodamine dye from aqueous solutions^{[12](#page-13-8)}. Mohan et al. have prepared magnetically activated carbon from almond shells to remove 2,4,6-trinitrophenol from wate[r13.](#page-13-9) Cellulose is one of the main components of the almond shell, which turns this waste into suitable material for preparing nano-catalysts 14,15 14,15 14,15 .

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Benzopyran or chromene is an organic bicyclic heterocyclic compound consisting of benzene and pyran ring[s16](#page-14-0)[,17.](#page-14-1) Chromene derivatives have various biological and medicinal properties and therapeutic applications that have been considered by pharmaceutical and organic chemist[s18.](#page-14-2) Chromenes have shown a variety of biologi-cal properties such as antimicrobial¹⁹, antibacterial^{[20](#page-14-4)}, anticancer^{[21](#page-14-5)}, anti-HIV^{[22](#page-14-6)}, and sex pheromone²³. Therefore, due to the biological and therapeutic properties and the great importance of chromenes, several pathways for the synthesis of these compounds have been reported, including one-step or multi-step methods^{[24](#page-14-8)}. One of the best attractive methods for the synthesis of chromenes is based on multi-component reactions $(MCRs)^{25,26}$ $(MCRs)^{25,26}$ $(MCRs)^{25,26}$. Multicomponent reactions are one of the most successful methods in the feld of increasing structural diversity and molecular complexity using a simple process. Tis method, as a developing process for the preparation of organic compounds, allows the development of many chemical compounds, with more structural diversity. Also, these reactions are considered a useful and efective tool for the synthesis of organic compounds and generally show good selectivity along with the reduction of by-products compared to the classical step-by-step preparation^{27,[28](#page-14-12)}. Higher efficiency, simplicity, saving time, and materials are some of the advantages of this category of reactio[n29](#page-14-13). Dihydropyrano[3,2-*c*]chromene and tetrahydrobenzo[*b*]pyran are heterocyclic organic compounds containing oxygen and are very attractive. For this reason, so far, many catalysts including ZnO NPs^{30} , t-ZrO₂ NPs^{31} , SB-DABCO@eosin³², Fe₃O₄@GO-NH₂^{[33](#page-14-17)}, [PEMIM][OH]³⁴, [(EMIM)Ac]³⁵, L-Proline³⁶, Chitosan-ZnO^{[37](#page-14-21)}, CESA^{[38](#page-14-22)}, Glycine³⁹, rGO@Fe₃O₄@ZrCp₂Cl₂^{[40](#page-14-24)}, Fe₃O₄@G.tea/Cu⁴¹, etc. have been used for the synthesis of this class of compounds.

In this work, Fe₃O₄@nano-almondshell/Si(CH₂)₃/2-(1-piperazinyl)ethylamine, abbreviated FNASiPPEA, as magnetite almond shell-based nano-catalyst was prepared and identifed using FT-IR, FESEM, XRD, TGA, VSM, EDS-map, BET, TEM techniques, and then use it in the synthesis of the dihydropyrano[3,2-*c*]chromene and tetrahydrobenzo[*b*]pyran under optimized conditions (Fig. [1\)](#page-1-0).

Results and discussions

In this paper, the FNASiPPEA, a magnetite biopolymer-based nano-catalyst, was used as an environmentally friendly basic nano-catalyst for the synthesis of dihydropyrano[3,2-*c*]chromene (DHPC) and tetrahydrobenzo[*b*] pyran (THBP) derivatives through a multicomponent reaction under optimized conditions. The FNASiPPEA was first prepared by preparing Fe₃O₄@nanoalmondshell according to previously reported methods⁴². After

Figure 1. Schematic representation of FNASiPPEA, dihydropyrano[3,2-*c*]chromene, and tetrahydrobenzo[*b*] pyran.

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that, FNASiPPEA was prepared by immobilization of 3-chloropropyltrimethoxysilane (CPTMS) and fnally 2-aminoethylpiperazine (AEP) (as a base agent) on the surface of the nano-catalyst.

FT-IR of FNASiPPEA. FT-IR spectra of Fe₃O₄@nanoalmondshell, AEP, and FNASiPPEA are shown in Fig. [2](#page-2-0). FT-IR spectrum of nano-almondshell (Fig. 2a) shows distinct peaks at 3428 cm^{-1} , 2920 cm^{-1} , and 1122 cm-1, which are related to O–H, C–H, and C–O vibrational stretching, respectively. In the FNASiPPEA spectrum (Fig. [2](#page-2-0)c), a distinct peak at 588 cm⁻¹ is attributed to the Fe–O stretching vibration. Also, the broad peak at the range of 3400 cm⁻¹ is attributed to the stretching vibration of N–H, which overlaps with the stretching vibration of the O–H group. CPTMS immobilization on Fe_3O_4 @nanoalmondshell is confirmed by a characteristic peak at 1111 cm⁻¹, which corresponds to the Si-O stretching vibration. The characteristic peak at 1451 cm⁻¹ is related to C–N stretching vibration.

FESEM and TEM of FNASiPPEA. The surface morphology and detailed structure of the FNASiPPEA nano-catalyst were investigated using FESEM (Fig. [3\)](#page-3-0). Figures [3](#page-3-0)a and b show the average particle size of the catalyst (11–43 nm) which appeared as nanospheres with pseudo-spherical morphology. The intrinsic structure was characterized using TEM measurements (Fig. [3](#page-3-0)c) which show core–shell nanoparticles.

PXRD (Powder X-ray diffraction) of FNASiPPEA. Figure [4](#page-3-1) shows the XRD patterns of Fe₃O₄ NPs and magnetite biopolymer-based FNASiPPEA nanoparticles. All the diffraction peaks appearing at $2\theta = 31^\circ$, 35°, 43°, 54°, 57°, and 63° in the spectrum (4a) can be indexed as centered cubic Fe₃O₄, which agrees well with the reported data correspond^{[43](#page-14-27)}. In the XRD pattern (4b), a new peak appears at $2\theta = 23^\circ$ and a broad peak at 2*θ*=20–30°, which is due to the presence of nano-almondshell and amorphous silica, respectively.

TGA of FNASIPPEA. Figure [5](#page-4-0) shows the TGA and DTA curves of magnetite FNASIPPEA. The nano-catalyst shows a small initial mass reduction at a temperature lower than 100 °C due to the removal of absorbed water and other organic solvents. At temperatures higher than 100 °C, (180–370 °C) the highest weight loss is observed in the TGA curve, which was probably due to the decomposition of nano-almondshell and organic parts (amine groups and methoxy groups) from the catalyst.

VSM of FNASIPPEA. The magnetic properties of the FNASIPPEA were evaluated in Fig. [6.](#page-4-1) The magnetic curve shows no remanence and coercivity, indicating the nanocatalyst's superparamagnetic behavior. The saturation magnetization value of FNASiPPEA (33 emu/g) is lower than that of Fe₃O₄ (47 emu/g). The low magnetization of the catalyst is attributed to the non-magnetic functionalized nano-almond shell coating on Fe₃O₄ NPs. However, the magnetic susceptibility of FNASiPPEA is strong enough to be separable by an external magnet from the reaction medium.

EDX and EDS-map of FNASiPPEA. The elemental composition of the FNASiPPEA nano-catalyst was determined by EDX. As shown in Fig. [7](#page-4-2), Fe, O, C, Si, and N signals are related to respectively, Fe₃O₄, and functionalized nano-almondshell, which appears in the EDX spectrum. The percentage composition of Fe, C, O, N, Cl, and Si elements is 30.39, 25.50, 15.39, 19.55, 9.08, and 0.48% respectively. According to the results of the EDSmapping analysis Fig. [8](#page-5-0), the distribution of these elements is homogeneously on the surface of the nano-catalyst.

Figure 2. FT-IR spectra of (**a**) nano-almond shell, (**b**) AEP, and (**c**) FNASiPPEA.

Figure 3. FESEM images of FNASiPPEA (**a** and **b**), and (**c**) TEM of FNASiPPEA.

Figure 4. XRD patterns of (**a**) Fe3O4 NPs, and (**b**) FNASiPPEA.

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Figure 5. TGA/DTA curves of FNASiPPEA.

Figure 6. VSM analysis of (**a**) Fe₃O₄ NPs, (**b**) FNASiPPEA.

Figure 7. EDS diagram of FNASiPPEA.

Figure 8. Maps of FNASiPPEA.

BET of FNASIPPEA. The BET (Brunauer–Emmett–Teller) surface area of the prepared nano-catalyst was obtained by nitrogen adsorption and desorption measurements (Fig. [9](#page-6-0)). The N_2 isotherms related to the type IV isotherm in the IUPAC classification have shown H_3 type rings, which can indicate the existence of mesopores and also have non-hard pores. As shown in Table [1](#page-6-1) a₀, BJH (Barrett–Joyner–Halenda), and pore diameter were 7.0116 m² g⁻¹, 0.050029 cm³ g⁻¹, and 28.206 nm respectively.

All the above results confrm the successful synthesis of magnetite biopolymer-based FNASiPPEA. Afer the detailed description of the prepared nano-catalyst, its catalytic performance was investigated for DHPC synthesis. Therefore, different reaction conditions such as the amount of catalyst, solvent, and temperature were investigated for a model reaction between 4-nitrobenzaldehyde, 4-hydroxycoumarin, and malononitrile (Table [2](#page-7-0)). While screening the model reaction in different solvents such as H₂O, EtOH, and H₂O/EtOH, the best result was obtained in the EtOH solvent (Table [2,](#page-7-0) entry 10).

Afer optimizing the reaction conditions, to determine the application range of FNASiPPEA, various alde-hydes were used in the reaction. The results are summarized in Table [3.](#page-8-0)

Then, optimization of the reaction conditions for the synthesis of THBP was carried out. Hence, the reaction between 4-nitrobenzaldehyde, dimedone, and malononitrile in the presence of 0.02 g of catalyst at 50 °C in solvent-free conditions has been adopted (Table [4](#page-9-0), entry 9). The results are summarized in Table [4.](#page-9-0)

We used various aldehydes in the reaction to investigate the application range of FNASiPPEA as a magnetite biopolymer-based nano-catalyst. The result is presented in Table [5](#page-9-1).

To compare the efficiency of this magnetite biopolymer-based nano-catalyst with other catalysts for the synthesis of DHPC and THBP derivatives, a summary of the results was collected in Tables [6](#page-10-0) and [7.](#page-10-1) As can be seen in Tables [6](#page-10-0) and [7](#page-10-1), the reaction efficiency of this catalyst is better than other catalysts and the reaction time is shorter than others.

Hot fltration test. Because the FNASiPPEA is a heterogeneous nano-catalyst, a heterogeneity test called hot fltration was performed. In this way, frst, the reaction was allowed to continue in the presence of the FNASiPPEA nano-catalyst, and then afer half the time, the catalyst was removed from the reaction mixture and continued the reaction, as can be seen in Fig. [10a](#page-11-0), no reaction progress was observed in the absence of the nano-catalyst, which indicates no leakage of solid catalyst into the reaction mixture. Therefore, the FNASiPPEA nano-catalyst is heterogeneous and suitable for DHPC and THBP synthesis reactions without any leaching.

Reusability of FNASiPPEA. To check the recyclability of the catalyst, after the completion of the reaction, the catalyst can be separated from the reaction mixture with a magnet, and afer washing with chloroform (CHCl3) and drying at the ambient temperature, and then can be reused for the synthesis of DHPC and THBP, which includes aldehyde (1 mmol), 1,3-diketone (4-hydroxycoumarin, dimedone), (1 mmol), and malononitrile (1.5 mmol) under optimized conditions. Terefore, the reusability of the catalyst for the model reaction was evaluated for the synthesis of DHPC and THBP (Figs. [11](#page-12-0) and [12](#page-12-1)). FT-IR, XRD, VSM, and FESEM analyses of the nano-catalyst recovered afer the 3rd run was also performed. According to Figs. [10](#page-11-0)b, c, d, and e, the matching

BET	
V_m	1.611 [cm ³ (STP) g^{-1}]
$a_{s,BET}$	7.0116 [$m^2 g^{-1}$]
C	45.307
Total pore volume($p/p_0 = 0.990$)	0.049443 [cm ³ g ⁻¹]
Mean pore diameter	28.206 [nm]
Langmuir plot	
Vm	1.9705 [cm ³ (STP) g ⁻¹]
$a_{s,Lang}$	7.0116 [$m^2 g^{-1}$]
B	0.3498
t plot	
Plot data	Adsorption branch
a ₁	5.4877 [$m^2 g^{-1}$]
V_{1}	$0 \ [\text{cm}^3 \text{ g}^{-1}]$
BJH plot	
Plot data	Adsorption branch
V_{p}	0.050029 [cm ³ g ⁻¹]
$r_{p,peak}(Area)$	7.99 [nm]
$\mathbf{a}_{\rm p}$	9.0803 [$m^2 g^{-1}$]

Table 1. Parameters obtained from porosity analysis.

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Table 2. Optimization of reaction conditions for the synthesis of DHPC.^{a a}Conditions: 4-nitrobenzaldehyde (1 mmol), 4-hydroxycoumarin (1 mmol), and malononitrile (1.5 mmol), Solvent (10 ml). b Isolated Yield.

of FT-IR, XRD, VSM, and SEM patterns obtained afer the third run with the primary nano-catalyst confrmed the preservation of the catalyst structure.

Proposed mechanism for synthesis of dihydropyrano[3,2‑*c***]chromene and tetrahydrobenzo[***b***]**

pyran. Figure [13](#page-13-12) shows the possible mechanism for the synthesis of dihydropyrano[3,2-*c*]chromene and tetrahydrobenzo[*b*]pyran derivatives using FNASiPPEA as a magnetite Brønsted base nano-catalyst. First, the Knoevenagel condensation between malononitrile and aldehyde is followed by loss of water to form an intermediate (a). Then the Michael addition between the intermediates (a) and (b) (dimedone, 4-hydroxycoumarin) and then intramolecular cyclization and tautomerization in the presence of the catalyst leads to the production of the corresponding product.

Experimental section

Materials and methods. Chemicals were purchased from Merck, Fluka, and Aldrich Chemical Companies. 1 H NMR and 13C NMR spectra were recorded at 400 and 100 MHz, respectively. Fourier transform infrared (FT-IR) measurements (in KBr pellets or ATR) were recorded on a Brucker spectrometer. Melting points were determined on a Büchi B-540 apparatus. The X-ray diffraction (XRD) pattern was obtained by a Philips Xpert MPD difractometer equipped with a Cu Kα anode (*k*=1.54 Å) in the 2*θ* range from 10° to 80°. Field Emission Scanning Electron Microscopy (FESEM) was obtained on a Mira 3-XMU. VSM measurements were performed by using a vibrating sample magnetometer (Meghnatis Daghigh Kavir Co. Kashan Kavir, Iran). Energy-dispersive X-ray spectroscopy (EDS) of nano-catalyst was measured by an EDS instrument and Phenom pro X. The EDX-MAP micrographs were obtained on the MIRA II detector SAMX (France). Thermal gravimetric analysis (TGA) was conducted using the "STA 504" instrument. Transmission electron microscopy (TEM) was obtained using a Philips CM120 with a LaB6 cathode and an accelerating voltage of 120 kV. BELSORP MINI II nitrogen adsorption apparatus (Japan) for recording Brunauer–Emmett–Teller (BET) of nano-catalyst at 77 K.

Preparation of nano-almondshell. To prepare the nano-almondshell, the almondshell was heated in boiling water for 30 min, dried, and powdered. After that, was treated with a 17.5 w/v NaOH solution at 90 °C for 24 h under refux conditions. Subsequently, the almondshell was fltered and washed with distilled water until the alkali was eliminated. Then, bleached with 100 mL of 1:1 aqueous dilution of 3.5% w/v sodium hypochlorite (NaOCl) at 80 °C for 3 h under reflux conditions. The resulting almondshell particles were hydrolyzed partially using the 35% sulfuric acid (H₂SO₄) aqueous solution with an almondshell-to-acid weight ratio of 1–10 at 45 °C. After 3 h, the obtained suspension was diluted with water five-fold to stop the hydrolysis reaction. The suspension was centrifuged at 4000 rpm to separate the nano-almondshell from the acidic medium (yield 60%).

Table 3. FNASiPPEA catalyzed the synthesis of DHPC.^{a a}Conditions: aldehyde (1 mmol), 4-hydroxycoumarin (1 mmol), malononitrile (1.5 mmol), 80 °C, EtOH Solvent, catalyst (0.02 g) ^bIsolated yield.

Preparation of Fe₃O₄@nano-almondshell. In a 250 mL flask, 3 g of nano-almondshell and 100 mL acetic acid (CH₃COOH) of 0.05 M were added. After that, FeCl₃.6H₂O (3.51 g, 13 mmol) and FeCl₂.4H₂O (1.29 g, 6.5 mmol) were added and stirred for 6 h at 80 °C. Then, 8 mL of NH₄OH (25%), was added dropwise and stirred for 45 min. The precipitated brown products were isolated from the solution by a magnet, washed 3 times with distilled water, and dried in an oven at 80 °C for 4 h. The weight of the Fe₃O₄@nano-almondshell obtained is 4.141 g.

Synthesis of Fe3O4@nano‑almondshell/Si(CH2)3Cl (FNASiP‑Cl). In a 100 mL fask, 1 g of dried Fe_3O_4 @nano-almondshell was dispersed in the mixture of 10 mL of chloroform (CHCl₃), and 3.4 mL of 3-chloropropyltrimethoxysilane was added dropwise. The mixture was sonicated at 25 °C for 20 min and then, the mixture was carried out under refux conditions for 4 h. Finally, the result was collected using a magnet and washed three times with chloroform.

Preparation of magnetite FNASiPPEA. The FNASiP-Cl $(0.5 g)$ was dispersed in ethanol by ultrasonic for 20 min at room temperature and then dried. The next, 0.5 g of dried FNASiP-Cl and 2-(1-piperazinyl)ethylamine (AEP) (0.129 g, 1 mmol) was heated in 10 mL *N, N*-dimethylformamide (DMF) for 24 h at 80 °C. The resulting precipitates were cooled, washed with dichloromethane (CH_2Cl_2) , and dried.

General synthesis of dihydropyrano[3,2‑*c***]chromene and tetrahydrobenzo[***b***]pyran deriva‑ tives.** For the synthesis of dihydropyrano[3,2-*c*]chromene, in a 50 ml round bottom fask, 4-hydroxycoumarin (1 mmol, 0.162 g), 4-nitrobenzaldehyde (1 mmol, 0.151 g), malononitrile (1.5 mmol, 0.099 g), FNASiP-PEA (0.02 g), and 10 ml EtOH was added. The reaction mixture was refluxed at 80 °C and stirred for appropriate periods as shown in Table [2.](#page-7-0) Afer the end of the reaction (TLC, n-hexane: ethyl acetate 6:4), the catalyst FNA-SiPPEA was separated from the reaction mixture by an external magnet, the solvent was removed under reduced pressure, and the precipitate was washed with methanol and recrystallized with chloroform for further purifcation.

Table 4. Optimization of the reaction conditions for the synthesis of THBP.^{a a}conditions: 4-nitrobenzaldehyde (1 mmol), dimedone (1 mmol), malononitrile (1.5 mmol), Solvent (10 ml). b Isolated Yield.

Table 5. Synthesis of THBP derivatives catalyzed by FNASiPPEA.^a ^aConditithons: aldehyde (1 mmol), dimedone (1 mmol), malononitrile (1.5 mmol), 50 °C, EtOH as the solvent, catalyst (0.02 g), ^bIsolated yield.

Table 6. Comparison of FNASiPPEA with other catalysts for the synthesis of DHPC.

Table 7. Comparison activity of FNASiPPEA with another catalyst for the synthesis of THBP.

For the synthesis of tetrahydrobenzo[*b*]pyran, the reaction of dimedone (1 mmol, 0.140 g), 4-nitrobenzaldehyde (1 mmol, 0.151 g), malononitrile (1.5 mmol, 0.099 g), and magnetite nano-catalyst FNASiPPEA (0.02 g) was carried out in 10 ml of EtOH (Table [4\)](#page-9-0). Afer the completion of the reaction (TLC, *n*-hexane: ethyl acetate 7:3), the magnetite catalyst was removed from the reaction mixture by a magnet, the solvent was removed under reduced pressure, and the product was obtained afer washing and recrystallization with chloroform.

Conclusion

In summary, the magnetite almondshell-based nano-catalyst was prepared, characterized, and used for the synthesis of DHPC and THBP. The prepared nano-catalyst FNASiPPEA shows high catalytic activity and good reusability. Meanwhile, this method is non-toxic and biodegradable, it may be used to prepare other biopolymerbased nano-catalysts for more interesting reactions.

Figure 11. Reusability of FNASiPPEA for the synthesis of DHPC.

Figure 12. Recyclability of FNASiPPEA for the synthesis of THBP.

Figure 13. Proposed mechanism for synthesis of DHPC and THBP.

Data availability

The datasets generated and/or analysed during the current study are available in this article and its supplementary information fles.

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Author contributions

D.M. is a Ph.D. student. She has worked on the subject of manuscripts in the laboratory with A.H.B. and B.F.M. helps as a supervisor. D.M. wrote the manuscript, and A.H.B. and B.F.M. edit the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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