scientific reports



OPEN Elevated serum levels of bone morphogenetic protein-9 are associated with better outcome in AQP4-IgG seropositive NMOSD

Hiroki Masuda¹, Masahiro Mori¹, Akiyuki Uzawa¹, Tomohiko Uchida¹, Mayumi Muto^{1,2}, Ryohei Ohtani^{1,3}, Reiji Aoki¹ & Satoshi Kuwabara¹

Lymphatic drainage in the central nervous system is regulated by meningeal lymphatic vasculature, and recurrent neuroinflammation alters lymphatic vessel remodeling. Patients with aquaporin-4 antibody-positive neuromyelitis optica spectrum disorder (AQP4 + NMOSD) were reported to demonstrate worse outcomes compared with patients with anti-myelin oligodendrocyte glycoproteinassociated disorders (MOGAD). This study aimed to investigate the serum cytokines relevant to vascular remodeling after attacks and their prognostic role in patients with AQP4 + NMOSD. This study measured the serum levels of 12 cytokines relevant to vascular remodeling, including bone morphogenetic protein-9 (BMP-9) and leptin, in 20 patients with AQP4+ NMOSD and 17 healthy controls (HCs). Disease controls included 18 patients with MOGAD. Serum and cerebrospinal fluid interleukin-6 levels were also measured. Clinical severity was evaluated with Kurtzke's Expanded Disability Status Scale (EDSS). Compared with HCs, patients with AQP4 + NMOSD showed higher BMP-9 (median; 127 vs. 80.7 pg/mL; P = 0.0499) and leptin levels (median; 16,081 vs. 6770 pg/mL; P = 0.0224), but not those with MOGAD. Better improvement in EDSS at 6 months was associated with baseline BMP-9 levels in patients with AQP4 + NMOSD (Spearman's rho = -0.47; P = 0.037). Serum BMP-9 is upregulated at relapse and may contribute to vascular remodeling in AQP4 + NMOSD. Serum BMP-9 levels could predict clinical recovery 6 months after the attack.

Neuromyelitis optica spectrum disorder (NMOSD), an inflammatory disorder in the central nervous system (CNS), is considered a subtype of multiple sclerosis (MS), and NMO-IgG findings differentiated NMOSD from MS¹. The positivity of anti-aquaporin-4 antibodies (AQP4-IgG) was one of the NMOSD features², but recent studies reported some patients with AQP4-IgG-negative NMOSD having positive anti-myelin oligodendrocyte glycoprotein (MOG-IgG)^{3,4}. Additionally, similar clinical features and laboratory findings, including increased cerebrospinal fluid (CSF) interleukin-6 (IL-6) levels, were reported in AQP4-IgG-positive NMOSD (AQP4+NMOSD) and MOG-IgG-associated disorders (MOGAD)⁵⁻⁷. Favorable clinical outcomes were reported in MOGAD compared with AQP4 + NMOSD even in the first optic neuritis^{8,9}. However, the mechanism behind milder clinical outcomes in MOGAD compared with AQP4+NMOSD was not fully elucidated. A recent study reported the different patterns and extent of helper T cell profiles between AQP4+NMOSD and MOGAD¹⁰. Therefore, AQP4 + NMOSD and MOGAD have distinct pathophysiological differences.

Meanwhile, the meningeal lymphatic vasculature regulated CNS lymphatic drainage and neuroinflammation¹¹. Surgical and pharmacological blockade of lymphatic function attenuated experimental autoimmune encephalomyelitis (EAE), which is the animal model of MS. Inflammation has induced neuro-lymphatic protein expression in MS brain vasculature¹². Recurrent inflammation regulated lymphatic vessel remodeling¹³, and vascular remodeling is relevant to blood-brain barrier (BBB) remodeling. Therefore, we hypothesized that cytokines relevant to vascular regeneration as a short-term prognostic factor after attacks in AQP4 + NMOSD. This study investigated vascular regeneration cytokine profiles after attacks and their prognostic factor in AQP4 + NMOSD.

¹Department of Neurology, Graduate School of Medicine, Chiba University, 1-8-1, Inohana, Chuo-Ku, Chiba-Shi 260-8670, Japan. ²Department of Neurology, Chiba Rosai Hospital, 2-16, Tatsumidai-Higashi, Ichihara-Shi 290-0003, Japan. ³Department of Neurology, Kimitsu Chuo Hospital, 1010, Sakurai, Kisarazu-Shi 292-8535, Japan. [⊠]email: hiroki_masuda@chiba-u.jp

Materials and methods

Standard protocol approvals and patient consent. The study procedure was approved by the ethics committee of the Chiba University School of Medicine (Nos. 842 and 1937) and Sannou Hospital. All patients provided written informed consent. The methods used in this study comply with the Declaration of Helsinki and its subsequent amendments, and were performed in accordance with the relevant guidelines and regulations.

Participants and samples. This study recruited 22 patients with AQP4+NMOSD, 20 healthy controls (HCs), and 20 patients with MOGAD as disease controls. All patients with AQP4+NMOSD fulfilled the 2015 international diagnostic criteria for NMOSD². Anti-MOG-IgG-positive disorders with CNS involvement were defined as MOGAD¹⁰. The presence of AQP4-IgG and MOG-IgG antibodies was confirmed by a cell-based assay measured as previously described¹⁴.

Serum samples of HCs were obtained as previously reported. Briefly, 196 HCs who underwent a complete medical check-up at Sannou Hospital provided serum samples as volunteers^{15,16}, of whom 20 age- and sex-matched samples with MOGAD were included in the study.

Serum and CSF samples in the acute phase were obtained before giving any attack treatment, including steroids and plasma exchange, in patients with AQP4 + NMOSD and MOGAD.

Demographic characteristics, including sex ratio and age at sampling, and clinical features, including disease duration to sampling, the percentage of the first attack, and Kurtzke's Expanded Disability Status Scale (EDSS) before the attack, at sampling, and 6 months after the attack, were investigated. The prognostic factor at 6 months after the attack was analyzed by investigating the correlation between Δ EDSS (6 M-pre), which is EDSS at 6 months minus EDSS before the attack, and clinical items or cytokine levels in the acute phase. Laboratory findings, including CSF cell count, CSF protein concentration, the quotient of albumin (Qalb), IgG index, and the percentage with positive oligoclonal IgG bands, and baseline treatment at sampling and treatment in the acute phase were compared. The effects of treatments on Δ EDSS (6 M-pre) or cytokine levels relevant to better improvement in EDSS at 6 months after the attack were also investigated.

Cytokine measurements. All serum samples, just after centrifugation at 3000 rpm for 10 min, and all CSF samples were immediately stored at – 80 °C until cytokine analysis, other than IL-6. The serum and CSF IL-6 levels were measured on a single detection immediately after centrifugation at room temperature using the electrochemiluminescence immunoassay according to the manufacturer's instruction (Roche Diagnostics K.K., Tokyo, Japan). The serum cytokine concentrations relevant to the blood vessel regeneration were measured using the MILLIPLEX* (Merck Millipore, Darmstadt, Germany) human angiogenesis/growth factor magnetic bead panel 1 with a single detection, according to the manufacturer's instruction. Fluorescence intensity from the immunoassay was acquired and analyzed using xPONENT 4.2 Software (Luminex Corporation, Austin, TX, USA). The measured cytokines relevant to the vascular remodeling included epidermal growth factor, angiopoietin 2, granulocyte-colony stimulating factor (G-CSF), bone morphogenetic protein-9 (BMP-9), endoglin, leptin, hepatocyte growth factor (HGF), placental growth factor, vascular endothelial growth factor (VEGF)-C, VEGF-D, fibroblast growth factor-2 (FGF-2), and VEGF-A. Values under the dynamic range were replaced by half of the lower sensitivity limit.

Statistical analysis. JMP pro version 15.0.0 (SAS Institute Inc, Cary, NC, USA) was used for statistical analysis. Continuous data were compared using the Mann–Whitney *U* test or Steel test HCs as control. The Steel–Dwass test was performed on the items with statistical differences. Categorical outcomes were evaluated using Fisher's exact test. Spearman's rank test was performed to analyze correlations between elevated cytokines compared with HCs and clinical items. Correlations between clinical items and serum or CSF IL-6 levels were investigated by Spearman's rank test. The *p*-value was considered significant at 0.05. Since prednisolone could affect the cytokine levels including BMP-9¹⁷, an analysis of covariance (ANCOVA) was added to evaluate the effect of prednisolone on the cytokines which were shown to be related to prognosis using significant different items with or without prednisolone as covariates.

Ehical approval. The study procedure was approved by the ethics committee of the Chiba University School of Medicine (No. 842 and 1937) and Sannou Hospital.

Informed consent. All patients provided written informed consent.

Results

Demographics and clinical characteristics in patients and HCs. Table 1 shows the demographic and clinical characteristics of patients. This study excluded 2 patients with AQP4+NMOSD, 4 with MOGAD, and 3 HCs because of the shortage of included beads for measuring cytokines. Finally, 20 patients with AQP4+NMOSD, 18 patients with MOGAD, and 17 HCs were included.

The female-to-male ratio was not different among the three groups. The percentages of the female were 85.0, 66.7, and 76.5% for AQP4+NMOSD, MOGAD, and HCs, respectively. The median age was 51.5, 45.0, and 44.0 for AQP4+NMOSD, MOGAD, and HCs, respectively (interquartile range; 15.3, 37.5, and 6.0, respectively). Ages were not different among AQP4+NMOSD, MOGAD, and HCs, and disease duration was not different between AQP4+NMOSD and MOGAD. EDSS before the attack and at 6 months was higher in patients with AQP4+NMOSD compared to those with MOGAD (median; 1.5 vs. 0.0 and 2.0 vs. 1.9, P=0.0324 and 0.0013, respectively). The percentage of patients with the first attack was higher in patients with MOGAD (61.1%)

		MOGAD	HCs	P value				
	(N=20)	(N=18)	(N=17)	AQP4+NMOSD vs MOGAD				
Demographic and clinical features								
Female (%)	17/20 (85.0)	12/18 (66.7)	13/17 (76.5)	0.2603				
Age (years)	51.5 (15.3)	45.0 (37.5)	44.0 (6.0)	0.7536				
Disease duration (years)	2.0 (5.5)	0.0 (1.8)		0.0220				
Days from attack to sampling in acute phase	14.5 (17.5)	14.0 (18.3)		0.8262				
Days from attack to treatment in acute phase	15.0 (17.0)	12.5 (19.0)		0.6666				
EDSS before the attack	1.5 (2.0)	0.0 (1.0)		0.0324*				
EDSS at sampling in acute phase	4.0 (2.8)	5.0 (3.0)		0.5837				
EDSS at 6 months after attack	2.0 (3.3)	1.0 (2.0)		0.0013*				
ΔEDSS (6 M-pre)	0.75 (2.4)	0.0 (1.5)		0.2406				
First attack (%)	4/20 (20.0)	11/18 (61.1)		0.0189*				
Laboratory findings								
CSF cell count (/µL)	5.8 (10.0)	3.2 (8.0)		0.1784				
CSF protein concentration (mg/dl)	44.5 (18.8)	32.0 (38.5)		0.2852				
Qalb (*10 ⁻³)	5.8 (3.0)	4.3 (5.2)		0.3495				
IgG index	0.67 (0.19)	0.62 (0.19)		0.2360				
Positive oligoclonal IgG bands (%)	6/16 (37.5)	2/18 (11.1)		0.1131				
Baseline treatments								
Prednisolone only	10	3						
Prednisolone plus immunosuppressant	1	1						
Prednisolone plus regular plasma exchange	1	0						
None	8	14						
Treatments in the acute phase								
mPSL pulse	11	12						
mPSL pulse plus plasma exchange	8	4						
Oral prednisolone	0	0						
None	1	2						

Table 1. The demographic and clinical characteristics of patients with AQP4+NMOSD, MOGAD, and HCs. Data are presented as median [interquartile range] or number (%). *P<0.05. Δ EDSS (6 M-pre) = EDSS at six months after the attack minus EDSS before the attack. Immunosuppressant includes azathioprine and tacrolimus. AQP4+NMOSD: anti-aquaporin-4-IgG positive neuromyelitis optica, CSF: cerebrospinal fluid, EDSS: Kurtzke's Expanded Disability Status Scale, MOGAD: anti-myelin oligodendrocyte glycoprotein IgG associated disorders, mPSL: methyl prednisolone, Qalb: quotient albumin.

than in patients with AQP4+NMOSD (61.1% vs. 20.0%, respectively, P < 0.001). Laboratory findings revealed no difference between CSF cell count, CSF protein concentration, Qalb, IgG index, and oligoclonal IgG bands positivity and AQP4+NMOSD and MOGAD.

The median days from attack to sampling in the acute phase were 14.5 and 14.0 in patients with AQP4+NMOSD and MOGAD, respectively (interquartile range: 17.5 and 18.3, range: 2–68 days and 1–81 days, respectively).

Cytokine profiles in patients with AQP4+NMOSD and MOGAD in the acute phase and HCs. Table 2 shows the cytokine profiles in patients with AQP4+NMOSD and MOGAD in the acute phase and those in HCs. Patients with AQP4+NMOSD showed higher BMP-9 and leptin compared with HCs. FGF-2 was elevated in patients with MOGAD compared with HCs. HGF levels were increased in AQP4+NMOSD and MOGAD. Figure 1 shows the elevated cytokines compared with HCs. Leptin levels in patients with AQP4+NMOSD and MOGAD remained higher compared with HCs after the Steel–Dwass test for BMP-9 and leptin (P=0.0224). HGF was higher in AQP4+NMOSD and MOGAD compared with HCs after the Steel–Dwass test (P=0.0006 and 0.0033, respectively). Vascular regeneration-related cytokine and IL-6 levels were not different between AQP4+NMOSD and MOGAD.

Correlations between elevated cytokines and clinical items or IL-6 levels in patients with AQP4+NMOSD. Table 3 shows the correlations between elevated cytokines compared with HCs and clinical items. Positive correlations were found between BMP-9 levels and disease duration or EDSS in patients with AQP4+NMOSD before the attack. Conversely, BMP-9 levels negatively correlated with Δ EDSS (6 M-pre) (rho = -0.4690, *P* = 0.0370, Fig. 2A). BMP-9 levels showed no correlations with other clinical items, including age, CSF cell count, CSF protein, Qalb, IgG index, EDSS at sampling, and EDSS 6 months after the attack. BMP-9 levels suggested the negative correlation with CSF IL-6 levels (rho = -0.4823, *P* = 0.0567) and exhib-

				P value				
	HCs (N = 17)	AQP4+NMOSD (N=20)	MOGAD (N=18)	AQP4+NMOSD vs HCs	MOGAD vs HCs	AQP4+NMOSD vs MOGAD		
Serum								
IL-6 (pg/mL)	N.A	3.2 (4.3) (N = 15)	1.7 (2.3) (N=14)			0.2171		
EGF (pg/mL)	24.9 (82.6)	25.9 (58.5)	39.7 (61.8)	0.9335	0.7422			
Angiopoietin 2 (pg/mL)	833 (1169)	807 (549)	543 (572)	0.9792	0.2861			
G-CSF (pg/mL)	13.7 (45.7)	20.8 (17.8)	18.5 (11.6)	0.7918	0.2688			
BMP-9 (pg/mL)	80.7 (119)	127 (71.7)	134 (68.9)	0.0499*	0.2273			
Endoglin (pg/mL)	936 (524)	993 (359)	1094 (267)	0.5449	0.2706			
Leptin (pg/mL)	6770 (10,859)	16,081 (23,260)	8829 (9188)	0.0224*	0.8989			
HGF (pg/mL)	66.7 (81.6)	163 (125)	186 (127)	< 0.001*	0.0022*			
PLGF (pg/mL)	2.2 (4.5)	3.9 (4.2)	3.0 (3.8)	0.5033	0.9430			
VEGF-C (pg/mL)	836 (1447)	513 (913)	1305 (1053)	0.9129	0.1776			
VEGF-D (pg/mL)	117 (655)	55.0 (46.7)	59.1 (56.0)	0.0836	0.1234			
FGF-2 (pg/mL)	20.3 (54.2)	46.8 (15.9)	58.4 (49.1)	0.0779	0.0070*			
VEGF-A (pg/mL)	175 (179)	274 (418)	259 (194)	0.1236	0.6048			
CSF								
IL-6 (pg/mL)	N.A	12.4 (16.8) (N = 16)	9.9 (22.6) (N=17)			0.8009		

Table 2. Cytokine profile in patients with AQP4+NMOSD and MOGAD in the acute phase and those in HCs. Data are presented as median [interquartile range]. **P*<0.05. AQP4+NMOSD: anti-aquaporin-4-IgG positive neuromyelitis optica spectrum disorder, BMP-9: bone morphogenetic protein-9, CSF: cerebrospinal fluid, EGF: epidermal growth factor, FGF-2: fibroblast growth factor-2, G-CSF: granulocyte-colony stimulating factor, HCs: healthy controls, HGF: hepatocyte growth factor, IL-6: interleukin-6, MOGAD: anti-myelin oligodendrocyte glycoprotein IgG associated disorders, N.A.: not acquired, PLGF: placental growth factor, VEGF: vascular endothelial growth factor.



Figure 1. Elevated cytokine levels in AQP4+NMOSD, MOGAD, and HCs. Box plots are demonstrated for each group. *Statistically significant after the Steel test HCs as control. AQP4+NMOSD: anti-aquaporin-4-IgG positive neuromyelitis optica spectrum disorder, HCs: healthy controls, MOGAD: anti-myelin oligodendrocyte glycoprotein IgG associated disorders.

	Age	Disease duration	CSF cell count	CSF protein	Qalb	IgG index	EDSS before attack	EDSS in the acute phase	EDSS six months after attack	ΔEDSS (6 M-pre)
AQP4+NMOSD										
BMP-9 (pg/	0.1387	0.5444*	-0.4197	0.1911	0.2316	-0.4228	0.4927*	-0.1667	0.1507	-0.4690*
mL)	(<i>P</i> =0.5599)	(P=0.0131)	(<i>P</i> =0.0654)	(<i>P</i> =0.4196)	(P=0.3259)	(<i>P</i> =0.0713)	(P=0.0273)	(<i>P</i> =0.4825)	(<i>P</i> =0.5260)	(P=0.0370)
Leptin (pg/	0.0256	0.1575	0.2231	0.4266	0.4586*	-0.1526	0.2274	-0.2546	0.2295	-0.0894
mL)	(P=0.9146)	(P=0.5072)	(P=0.3445)	(<i>P</i> =0.0607)	(P=0.0420)	(P=0.5328)	(P=0.3349)	(<i>P</i> =0.2788)	(P=0.3304)	(P=0.7077)
HGF (pg/mL)	0.1713	0.4110*	0.0085	0.2169	0.2968	-0.0567	0.4014*	0.0600	0.3834*	0.1139
	(P=2112)	(P=0.0104)	(P=0.9597)	(P=0.1909)	(P=0.0704)	(P=0.7389)	(P=0.0125)	(P=0.7204)	(P=0.0175)	(P=0.4959)
Serum IL-6	-0.0585	-0.4130	0.5553*	0.0510	0.0946	-0.1309	-0.5078	0.36232	0.4605	0.8312*
(pg/mL)	(P=0.8359)	(P=0.1260)	(P=0.0316)	(P=0.8569)	(P=0.7375)	(P=0.6419)	(P=0.0533)	(P=0.1844)	(P=0.0841)	(P=0.0001)
CSF IL-6 (pg/	0.2762	-0.0391	0.4709	0.3444	0.2559	0.5353*	0.0923	0.5015*	0.1665	0.1648
mL)	(P=0.3004)	(P=0.8856)	(P=0.0656)	(<i>P</i> =0.1915)	(P=0.3388)	(P=0.0326)	(P=0.7339)	(P=0.0478)	(<i>P</i> =0.5377)	(P=0.5418)
MOGAD										
HGF (pg/mL)	0.0884	0.5410	-0.0451	0.0415	0.0950	0.1632	0.5234*	-0.0550	0.4347	0.1883
	(<i>P</i> =0.7272)	(P=0.0204)	(P=0.8590)	(<i>P</i> =0.8703)	(P=0.7076)	(<i>P</i> =0.5175)	(P=0.0258)	(P=0.8284)	(<i>P</i> =0.0715)	(<i>P</i> =0.4544)
FGF-2 (pg/	-0.2444	0.1655	-0.3588	-0.0842	0.0290	-0.3926	0.3018	-0.2735	0.3644	0.2646
mL)	(P=0.3283)	(<i>P</i> =0.5116)	(P=0.1437)	($P=0.7399$)	(P=0.9089)	(P=0.1071)	(P=0.2236)	(<i>P</i> =0.2721)	(<i>P</i> =0.1371)	(<i>P</i> =0.2886)
Serum IL-6	0.1456	-0.1377	0.1469	-0.2949	-0.2974	0.3477	-0.5359^{*}	-0.0657	-0.0159	0.2269
(pg/mL)	(P=0.6195)	(<i>P</i> =0.6387)	(P=0.6163)	(P=0.3060)	(P=0.3018)	(P=0.2232)	(P=0.0482)	(<i>P</i> =0.8235)	(P=0.9569)	(P=0.4352)
CSF IL-6 (pg/	0.1141	0.0618	0.3567	0.3247	0.3186	0.4779	0.1400	0.3397	0.6799^{*}	0.7211*
mL)	(<i>P</i> =0.6628)	(P=0.8138)	(<i>P</i> =0.1599)	(<i>P</i> =0.2035)	(<i>P</i> =0.2126)	(<i>P</i> =0.0523)	(P=0.5919)	(<i>P</i> =0.1822)	(P=0.0027)	(P=0.0011)

Table 3. Spearman's rank correlation coefficient (rho) of the correlation between elevated cytokines or IL-6levels and clinical items in AQP4 + NMOSD and MOGAD. Data are presented as median [interquartile range].*P < 0.05. AQP4 + NMOSD: anti-aquaporin-4-IgG positive neuromyelitis optica spectrum disorder, BMP-9:bone morphogenetic protein-9, CSF: cerebrospinal fluid, FGF-2: fibroblast growth factor-2, HGF: hepatocytegrowth factor, IL-6: interleukin-6, MOGAD: anti-myelin oligodendrocyte glycoprotein IgG associateddisorders.

ited no correlation with serum IL-6 levels (P=0.7277). Meanwhile, leptin levels positively correlated with Qalb (rho=0.4586, P=0.0420). No correlations were found between leptin levels and other clinical items, including age, disease duration, CSF cell count, CSF protein, Qalb, IgG index, EDSS before the attack, EDSS at sampling, EDSS 6 months after the attack, and Δ EDSS (6 M-pre). HGF positively correlated with disease duration (rho=0.4110, P=0.0104), EDSS before the attack (rho=0.4014, P=0.0125), and EDSS 6 months after the attack (rho=0.3834, P=0.0175). HGF levels showed no correlations with serum or CSF IL-6 levels.

Treatment effects on \DeltaEDSS (6 M-pre) and BMP-9 levels in patients with AQP4+NMOSD. In patients with AQP4+NMOSD, Δ EDSS (6 M-pre) and BMP-9 levels showed no difference between methyl-prednisolone pulse therapy and methylprednisolone pulse plus plasma exchange in the acute phase (*P*=0.3449 and 0.4828, respectively). Patients with AQP4+NMOSD who received only prednisolone as a baseline treatment showed lower Δ EDSS (6 M-pre) compared with those without baseline treatments at the attack (median; 0.0 vs 2.25, interquartile range; 1.25 vs 3.0, *P*=0.0184). BMP-9 levels were not different between patients with AQP4+NMOSD who received only prednisolone as a baseline treatments (*P*=0.0832).

BMP-9 levels were not different with or without prednisolone after ANCOVA in patients with AQP4+NMOSD. To exclude the prednisolone effects on BMP-9 expression, the demographic and clinical characteristics were compared with or without prednisolone in patients with AQP4+NMOSD. The results showed the longer disease duration (median; 5.0 years vs 0.5 years, interquartile range; 11.3 vs 1.0, P=0.0206) and days from attack to sampling in acute phase (median; 7.0 vs 27.0, interquartile range; 14.5 vs 30.8, P=0.0339) in patients with prednisolone compared with those without prednisolone. Age, sex, days from attack to treatment in the acute phase, EDSS before the attack, and EDSS at the sampling in the acute phase were not different between the two groups. ANCOVA showed no difference in BMP-9 levels with or without prednisolone when disease duration and days from attack to sampling in the acute phase were used as covariates (P=0.151).

Correlations between elevated cytokines and clinical items or IL-6 levels in patients with MOGAD. HGF showed the positive correlation with EDSS before the attack in patients with MOGAD (rho=0.5234, P=0.0258). No correlations were found between HGF levels and clinical items other than EDSS before the attack. FGF-2 demonstrated no correlations with all clinical items. Further, HGF and FGF-2 showed no correlation with serum or CSF IL-6 levels.

Correlations between serum or CSF IL-6 levels and clinical items in patients with AQP4 + NMOSD and MOGAD. Serum IL-6 levels in patients with AQP4 + NMOSD showed the positive correlation with CSF cell count (rho = 0.5553, P = 0.0316) and Δ EDSS (6 M-pre) (rho = 0.8312, P = 0.0001, Fig. 2B). CSF IL-6 levels posi-



Figure 2. Correlations between elevated cytokine levels compared with HCs and clinical items in patients with AQP4 + NMOSD. (A) Negative correlation between Δ EDSS (6 M-pre) and BMP-9. (B) Positive correlation between Δ EDSS (6 M-pre) and serum IL-6 levels. AQP4 + NMOSD: anti-aquaporin-4-IgG positive neuromyelitis optica spectrum disorders, CSF: cerebrospinal fluid, BMP-9: bone morphogenetic protein-9, EDSS; Kurtzke's Expanded Disability Status Scale, IL-6: interleukin-6. Δ EDSS (6 M-pre) = EDSS at 6 months after the attack minus EDSS before the attack.

tively correlated with IgG index (rho = 0.5353, P = 0.0326) and EDSS at sampling in the acute phase (rho = 0.5015, P = 0.0478). Meanwhile, CSF IL-6 levels in patients with MOGAD positively correlated with Δ EDSS (6 M-pre) (rho = 0.7211, P = 0.0011) and EDSS 6 months after the attack (rho = 0.6799, P = 0.0027). Serum IL-6 levels negatively correlated with EDSS before the attack (rho = -0.5359, P = 0.0482).

Serum IL-6 levels were not different with or without prednisolone after ANCOVA in patients with AQP4+NMOSD. No statistical difference was found in serum IL-6 levels with or without prednisolone in patients with AQP4+NMOSD after ANCOVA was performed with disease duration and days from attack to sampling in the acute phase as covariates (P=0.564).

Discussion

This study revealed elevated BMP-9, leptin, and HGF levels in patients with AQP4+NMOSD and FGF-2 and HGF levels in patients with MOGAD in the acute phase compared with HCs. Δ EDSS (6M-pre) in patients with AQP4+NMOSD showed a negative correlation with BMP-9 levels and a positive correlation with serum IL-6 levels. Leptin levels positively correlated with Qalb in patients with AQP4+NMOSD. Lower Δ EDSS (6M-pre) was observed in patients with AQP4+NMOSD who received only prednisolone as a baseline treatment compared with those without baseline treatments.

BMP-9 is produced by hepatic stellate cells in the liver and is a differentiating factor for cholinergic CNS neurons^{18,19}. Additionally, BMP-9 prevents vascular permeability by VEGF-receptor-2 signaling and vascular endothelial (VE)-cadherin internalization and occludin expression promotion¹⁸. BMP-9 administration in neurological diseases improves memory and alleviates the pathology in the animal model of Alzheimer's disease^{20,21}. BMP-9 overexpression decreases cell death and improves cell viability in astrocytes in the cerebral ischemia–reperfusion rat model²². AQP4-IgG was reported to target astrocytes^{2,23,24}. Severe astrocytic damage in neuromyelitis optoca was also demonstrated²⁵. Therefore, decreasing astrocyte death by BMP-9 overexpression could lead to improve EDSS 6 months after the attack in patients with AQP4 + NMOSD. Our study indicates higher serum BMP-9 in the acute phase as a good prognostic factor to predict disability severity 6 months after the attack in patients with AQP4 + NMOSD. To our best knowledge, no other study about BMP-9 was performed in the field of AQP4 + NMOSD.

The results demonstrated a positive correlation between serum IL-6 levels in patients with AQP4+NMOSD and Δ EDSS (6 M-pre). CSF IL-6 levels were negatively correlated with BMP-9 levels in patients with AQP4+NMOSD. IL-6 plays an important role in NMOSD pathogenesis^{5,26,27}. Increased serum and CSF IL-6 levels were demonstrated in the acute phase in patients with NMOSD⁶. Additionally, CSF IL-6 levels were elevated in the acute phase in patients with NMOSD and MOGAD⁷. Two recent randomized controlled trials confirmed that IL-6 receptor inhibition reduced NMOSD relapses^{28,29}. Hence, our study suggests serum IL-6 levels in patients with AQP4+NMOSD as a short-term prognostic factor in those diseases.

Our study revealed elevated leptin levels and a positive correlation between leptin levels and Qalb in patients with AQP4 + NMOSD. A previous study reported increased serum leptin levels in patients with NMO and MS³⁰. Leptin was reported to protect the brain from ischemic injury by reducing neuronal cell death^{31–33}. A recent study revealed that leptin protected the brain from ischemia by stabilizing the BBB³⁴. Therefore, our result may be explained by the positive feedback of leptin in stabilizing the BBB in the acute AQP4 + NMOSD phase.

Our study revealed higher HGF levels in patients with AQP4+NMOSD and MOGAD compared with HCs. Previous studies reported that HGF alleviates EAE severity and limits cytotoxic T-cell generation and its effector functionsyy^{35,36}. However, our study revealed no correlation between HGF and Δ EDSS (6 M-pre). To our best knowledge, this is the first report to show HGF elevation in patients with AQP4+NMOSD. Therefore, further investigation is required to conclude HGF function in AQP4+NMOSD pathogenesis.

Our study has some limitations. First, the percentage of the first attack was higher in patients with MOGAD compared to those with AQP4 + NMOSD, which could lead to baseline treatment differences, thereby affecting the cytokine values, particularly in the acute phase in patients with AQP4 + NMOSD. Second, body mass index was not obtained in our study. As leptin is associated with adipose tisse, difference in body mass index among the groups could affect the results. Finally, each group had a small sample size. Hence, a prospective study with increased samples and adjusted baseline treatment is required in the future.

In conclusion, our study demonstrated that serum BMP-9 levels could predict clinical recovery 6 months after an attack in patients with AQP4+NMOSD. However, prospective studies with larger sample size are required to confirm our results.

Data availability

The data that support our findings are available from the corresponding author, upon reasonable request.

Received: 15 December 2022; Accepted: 27 February 2023 Published online: 02 March 2023

References

- Lennon, V. A. *et al.* A serum autoantibody marker of neuromyelitis optica: distinction from multiple sclerosis. *Lancet (Lond. England)* 364, 2106–2112. https://doi.org/10.1016/s0140-6736(04)17551-x (2004).
- Wingerchuk, D. M. et al. International consensus diagnostic criteria for neuromyelitis optica spectrum disorders. Neurology 85, 177–189. https://doi.org/10.1212/wnl.00000000001729 (2015).
- Jurynczyk, M. et al. Clinical presentation and prognosis in MOG-antibody disease: A UK study. Brain 140, 3128–3138. https:// doi.org/10.1093/brain/awx276 (2017).
- Zamvil, S. S. & Slavin, A. J. (2015) Does MOG Ig-positive AQP4-seronegative opticospinal inflammatory disease justify a diagnosis of NMO spectrum disorder? *Neurol.(R) Neuroimmunol. Neuroinflam.* 2, e62. https://doi.org/10.1212/nxi.00000000000062.
- Uzawa, A. et al. Markedly increased CSF interleukin-6 levels in neuromyelitis optica, but not in multiple sclerosis. J. Neurol. 256, 2082–2084. https://doi.org/10.1007/s00415-009-5274-4 (2009).
- 6. Uzawa, A. *et al.* Cytokine and chemokine profiles in neuromyelitis optica: significance of interleukin-6. *Mult Scler.* **16**, 1443–1452. https://doi.org/10.1177/1352458510379247 (2010).
- Kaneko, K. *et al.* CSF cytokine profile in MOG-IgG+ neurological disease is similar to AQP4-IgG+ NMOSD but distinct from MS: a cross-sectional study and potential therapeutic implications. *J. Neurol. Neurosurg. Psychiatry* 89, 927–936. https://doi.org/ 10.1136/jnnp-2018-317969 (2018).
- Nagireddy, R. B. R. et al. Clinicoradiological comparative study of Aquaporin-4-IgG seropositive neuromyelitis optica spectrum disorder (NMOSD) and MOG antibody associated disease (MOGAD): A prospective observational study and review of literature. J. Neuroimmunol. 361, 577742. https://doi.org/10.1016/j.jneuroim.2021.577742 (2021).
- 9. Masuda, H. *et al.* Clinical difference after the first optic neuritis between aquaporin-4-IgG-associated and myelin oligodendrocyte glycoprotein-IgG-associated disorders. *J. Neurol.* **269**, 1996–2003. https://doi.org/10.1007/s00415-021-10764-7 (2022).
- Liu, J. et al. Peripheral blood helper T cell profiles and their clinical relevance in MOG-IgG-associated and AQP4-IgG-associated disorders and MS. J. Neurol. Neurosurg. Psychiatry 91, 132–139. https://doi.org/10.1136/jnnp-2019-321988 (2020).
- Louveau, A. *et al.* CNS lymphatic drainage and neuroinflammation are regulated by meningeal lymphatic vasculature. *Nat. Neurosci.* 21, 1380–1391. https://doi.org/10.1038/s41593-018-0227-9 (2018).
- Chaitanya, G. V. et al. Inflammation induces neuro-lymphatic protein expression in multiple sclerosis brain neurovasculature. J. Neuroinflam. 10, 125. https://doi.org/10.1186/1742-2094-10-125 (2013).

- Kelley, P. M., Connor, A. L. & Tempero, R. M. Lymphatic vessel memory stimulated by recurrent inflammation. Am. J. Pathol. 182, 2418–2428. https://doi.org/10.1016/j.ajpath.2013.02.025 (2013).
- Sugimoto, K. *et al.* The accuracy of flow cytometric cell-based assay to detect anti-myelin oligodendrocyte glycoprotein (MOG) antibodies determining the optimal method for positivity judgement. J. Neuroimmunol. 336, 577021. https://doi.org/10.1016/j. jneuroim.2019.577021 (2019).
- 15. Masuda, H. *et al.* Validation of the Japanese version of the modified fatigue impact scale and assessment of the effect of pain on scale responses in patients with multiple sclerosis. *Clin. Exp. Neuroimmunol.* **6**, 409–412 (2015).
- Masuda, H. *et al.* Validation of the Modified Fatigue Impact Scale and the relationships among fatigue, pain and serum interleukin-6 levels in patients with neuromyelitis optica spectrum disorder. J. Neurol. Sci. 385, 64–68. https://doi.org/10.1016/j.jns.2017.11.041 (2018).
- Caperuto, L. C. *et al.* Modulation of bone morphogenetic protein-9 expression and processing by insulin, glucose, and glucocorticoids: Possible candidate for hepatic insulin-sensitizing substance. *Endocrinology* 149, 6326–6335. https://doi.org/10.1210/en. 2008-0655 (2008).
- Desroches-Castan, A., Tillet, E., Bouvard, C. & Bailly, S. BMP9 and BMP10: Two close vascular quiescence partners that stand out. Dev. Dyn. 251, 178–197. https://doi.org/10.1002/dvdy.395 (2022).
- López-Coviella, I., Berse, B., Krauss, R., Thies, R. S. & Blusztajn, J. K. Induction and maintenance of the neuronal cholinergic phenotype in the central nervous system by BMP-9. Science 289, 313–316. https://doi.org/10.1126/science.289.5477.313 (2000).
- Wang, Z. et al. Intranasal BMP9 ameliorates Alzheimer disease-like pathology and cognitive deficits in APP/PS1 transgenic mice. Front. Mol. Neurosci. 10, 32. https://doi.org/10.3389/fnmol.2017.00032 (2017).
- Adams, S. L. *et al.* Immunohistochemical analysis of activin receptor-like kinase 1 (ACVRL1/ALK1) expression in the rat and human hippocampus: Decline in CA3 during progression of Alzheimer's disease. *J. Alzheimers Dis.* 63, 1433–1443. https://doi. org/10.3233/jad-171065 (2018).
- Feng, Y. & Hu, Y. Bone morphogenetic protein 9 serves a protective role in response to ischemic-reperfusion in the brain by promoting ERK activation. Mol. Med. Rep. 17, 2845–2852. https://doi.org/10.3892/mmr.2017.8253 (2018).
- 23. Paul, F. et al. Antibody to aquaporin 4 in the diagnosis of neuromyelitis optica. PLoS Med. 4, e133. https://doi.org/10.1371/journ al.pmed.0040133 (2007).
- Asseyer, S., Cooper, G. & Paul, F. Pain in NMOSD and MOGAD: A systematic literature review of pathophysiology, symptoms, and current treatment strategies. *Front. Neurol.* 11, 778. https://doi.org/10.3389/fneur.2020.00778 (2020).
- Fujihara, K. Neuromyelitis optica and astrocytic damage in its pathogenesis. J. Neurol. Sci. 306, 183–187. https://doi.org/10.1016/j. jns.2011.02.018 (2011).
- Chihara, N. et al. Interleukin 6 signaling promotes anti-aquaporin 4 autoantibody production from plasmablasts in neuromyelitis optica. Proc. Natl. Acad. Sci. USA 108, 3701–3706. https://doi.org/10.1073/pnas.1017385108 (2011).
- Fujihara, K. et al. Interleukin-6 in neuromyelitis optica spectrum disorder pathophysiology. Neurol. (R) Neuroimmunol. Neuroinflam. 7, 841. https://doi.org/10.1212/nxi.00000000000841 (2020).
- Yamamura, T. et al. Trial of satralizumab in neuromyelitis optica spectrum disorder. N. Engl. J. Med. 381, 2114–2124. https://doi. org/10.1056/NEJMoa1901747 (2019).
- Traboulsee, A. *et al.* Safety and efficacy of satralizumab monotherapy in neuromyelitis optica spectrum disorder: A randomised, double-blind, multicentre, placebo-controlled phase 3 trial. *Lancet Neurol.* 19, 402–412. https://doi.org/10.1016/s1474-4422(20) 30078-8 (2020).
- Bahrami, E. et al. Leptin hormone level in serum of opticospinal, neuromyelitisoptica and multiple sclerosis patients. Clin. Exp. Neuroimmunol. 5, 77–83. https://doi.org/10.1111/cen3.12092 (2014).
- Hu, S. *et al.* Leptin attenuates cerebral ischemic injury in rats by modulating the mitochondrial electron transport chain via the mitochondrial STAT3 pathway. *Brain Behav.* 9, e01200. https://doi.org/10.1002/brb3.1200 (2019).
- Zhang, W. F. et al. Protective effects of leptin against cerebral ischemia/reperfusion injury. Exp. Ther. Med. 17, 3282–3290. https:// doi.org/10.3892/etm.2019.7377 (2019).
- Zhang, W., Jin, Y., Wang, D. & Cui, J. Neuroprotective effects of leptin on cerebral ischemia through JAK2/STAT3/PGC-1-mediated mitochondrial function modulation. *Brain Res Bull* 156, 118–130. https://doi.org/10.1016/j.brainresbull.2020.01.002 (2020).
- Hung, W. T. *et al.* Leptin protects brain from ischemia/reperfusion-induced infarction by stabilizing the blood-brain barrier to block brain infiltration by the blood-borne neutrophils. *Eur J Neurosci* 52, 4890–4907. https://doi.org/10.1111/ejn.14896 (2020).
- Bai, L. et al. Hepatocyte growth factor mediates mesenchymal stem cell-induced recovery in multiple sclerosis models. Nat Neurosci 15, 862–870. https://doi.org/10.1038/nn.3109 (2012).
- Benkhoucha, M., Molnarfi, N., Schneiter, G., Walker, P. R. & Lalive, P. H. The neurotrophic hepatocyte growth factor attenuates CD8+ cytotoxic T-lymphocyte activity. J. Neuroinflam. 10, 154. https://doi.org/10.1186/1742-2094-10-154 (2013).

Acknowledgements

We appreciate Dr. Takahiro Kageyama, Dr. Manami Kato, and Kazusa Miyachi for helping us to measure cytokines.

Author contributions

H.M. drafted the first manuscript. M.M. and S.K. revised the manuscript and gave final approval of the current submission. H.M., M.M., and S.K. contributed to the conception and design of the study; H.M., M.M., A.U., T.U., M.M., R.O., and R.A. contributed to the acquisition and analysis of data; H.M. measured the cytokines and analysis. H.M. drafted the text and prepared the figures.

Competing interests

A. Uzawa has received honoraria from Alexion Pharmaceuticals and Argenx. Other authors report no competing interests.

Additional information

Correspondence and requests for materials should be addressed to H.M.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2023