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Relating soil physical properties to other soil properties and crop yields

Upendra M. Sainju^{1⊠}, Daniel Liptzin² & Jalal D. Jabro¹

Soil physical properties can be related to other soil properties and crop yields, but their evaluations as soil health indicators relating to extensive soil properties and long-term crop yields need further exploration. We evaluated the long-term (14 and 36 year) effect of cropping systems and N fertilizations on selected soil physical properties and related them to 66 soil physical, chemical, biological, and biochemical properties and mean crop yields across years at two dryland farming sites in the semiarid region of the northern Great Plains, USA. Treatments were rotations of no-tillage and conventional tillage spring wheat (Triticum aestivum L.), barley (Hordeum vulgare L.), pea (Pisum sativum L.), and fallow with or without N fertilization. Soil samples collected in April 2019 were analyzed for physical, chemical, biological, and biochemical properties and mean crop yields were determined. The average slake aggregate (ASA), wet aggregate stability index (WASI), and intact core available water holding capacity (IAWHC) were associated with most soil physical, chemical, biological, and biochemical properties and clay concentration was associated with nutrient concentrations. These parameters were also better related to mean crop yields across years than other soil physical properties. Because of the enhanced relationship with soil properties and crop yields and simple and inexpensive measurement, ASA can be used as a potential soil health indicator in dryland cropping systems in semiarid regions.

Soil physical properties as important components of soil health influence water and nutrient movements, aeration, soil temperature, nutrient cycling, and root growth that affect crop yields and environmental quality^{1–3}. For example, increased bulk density (BD) due to increased soil compaction results in decreased pore volume that reduces water infiltration, increases aeration stress, lowers soil temperature and nutrient cycling, increases denitrification, losses mycorrhizal fungi, and reduces root growth². In contrast, increased soil aggregation enhances water and nutrient movements, reduces soil erosion, promotes C sequestration, favors microbial activity and abundance, and increases root growth and crop yields^{1,3}. Clay concentration is an important indicator of soil health that enhances the retention of soil water and nutrients⁴. While increased soil water retention enhances crop yields^{2,5}, reduced water infiltration capacity of the soil results in anaerobic condition that hampers nutrient cycling and root growth, thereby reducing crop production^{1,5}.

Some soil physical properties are related to other soil properties^{4,6,8,11,12}. Numerous researchers^{1,6,7} reported that BD was negatively correlated to soil organic C (SOC), but soil aggregation, water retention, and water infiltration capacity were positively correlated to SOC. In contrast, Nemes et al.⁸ observed that soil hydraulic conductivity (SHC) was negatively correlated to SOC because soils rich in organic matter have lower water permeability. Blanco-Canqui and Benjamin¹ concluded that the relationship between soil physical properties and SOC are affected by soil and climatic conditions and management practices. Shaver et al.⁹ found that soil aggregation was related to clay concentration. Norkaew et al.⁴ showed that water-stable aggregation (WSA) was positively related to SOC, soil total N (STN), potential N mineralization (PNM), and microbially active C (MAC), but BD was negatively related to these parameters. Shestak and Busse¹⁰ found that BD was negatively correlated to macroporosity (MAP), but was not related to microbial properties. Stott et al.¹¹ reported that soil aggregation and water infiltration capacity were positively correlated, but BD was negatively correlated, but SHC was negatively correlated, but SHC was negatively correlated to activities of N-acetyl- β -glucosamidinase (NAG) and phosphomonoesterase (PME).

Soil physical properties also vary in their relationships to crop production. Sene et al.¹⁴ found that sand concentration and SHC were positively correlated, but soil aggregation was negatively correlated to crop yield. Shang et al.¹⁵ reported that soil water content and aggregate stability (AS) were positively related, but BD was negatively

¹USDA-ARS, Northern Plains Agricultural Research Service, Sidney, MT 59270, USA. ²Soil Health Institute, Morrisville, NC 27560, USA. ^{III} email: upendra.sainju@usda.gov

Soil physical properties	Clay	ASA	WASI	BD	IAWHC	SHC		
Froid								
Clay (g kg ⁻¹)	-	0.14	0.50*	- 0.32	0.36	0.14		
ASA	0.14	-	0.66**	- 0.54*	0.52*	0.28		
WASI	0.50*	0.66**	-	- 0.38	0.51*	0.69**		
BD (Mg m ⁻³)	- 0.32	- 0.54*	- 0.38	-	- 0.71**	- 0.13		
IAWHC (cm ³ cm ⁻³)	0.36	0.52*	0.51*	- 0.71**	-	0.14		
SHC (cm h ⁻¹)	0.14	0.28	0.69**	- 0.13	0.14	-		
Sidney								
Clay (g kg ⁻¹)	-	- 0.25	- 0.24	0.05	0.34	0.17		
ASA	- 0.25	-	0.67**	- 0.22	0.38	0.11		
WASI	- 0.24	0.67**	-	- 0.36	0.46*	0.32		
BD (Mg m ⁻³)	0.05	- 0.22	- 0.36	-	0.08	0.15		
IAWHC (cm ³ cm ⁻³)	0.34	0.38	0.46*	0.08	-	0.43*		
SHC (cm h ⁻¹)	0.17	0.11	0.32	0.15	0.43*	-		

Table 1. Correlation (r) among clay concentration (Clay), average slake aggregate (ASA), wet aggregate stability index (WASI), bulk density (BD), intact core available water holding capacity (IAWHC), and saturated hydraulic conductivity (SHC) at Froid (n = 16) and Sidney (n = 24), Montana. * and ** Significant at $P \le 0.05$ and 0.01, respectively.

related to crop yield. Similarly, several researchers^{16,17} observed that BD was negatively related to crop yield. In contrast, Logsdon and Karlen² showed that BD was not related to crop yield. Maddoni et al.³ showed that AS was an important indicator of soil health that related better to corn (*Zea mays* L.) yield than labile C and N fractions.

Although some soil physical properties are related to other soil properties and crop yields, an in-depth examination of their relationships with other soil properties and long-term crop yields are needed for their proper evaluation for potential soil health indicators. We chose six soil physical properties (clay concentration, BD, average slake aggregate [ASA], wet aggregate stability index [WASI], intact core available water capacity (IAWHC], and SHC) representing particle size, soil aggregation, compaction, water retention, and water infiltration capacity that have a broad impact on soil health, such as water and nutrient movements, aeration, soil temperature, C sequestration, nutrient cycling, and microbial properties and related them to 66 other soil properties and longterm crop yields at two dryland farming sites in eastern Montana, USA. We hypothesized that these physical properties would relate to other soil properties and long-term crop yields and that some physical properties may be used as potential soil health indicators due to their close relationships with the above parameters in dryland cropping systems in the semiarid region. Our objectives were to: (1) determine if physical properties are related to other 66 other soil physical, chemical, biological, and biochemical properties, and (2) select which soil physical properties can be used as potential soil health indicators that are related to most soil properties and crop yields.

Results

Relationships among soil properties. *Soil physical properties.* At Froid, clay concentration was positively correlated to WASI (Table 1). The ASA was positively correlated to WASI and IAWHC, but negatively correlated to BD. The WASI was positively correlated to IAWHC and SHC. The BD was negatively correlated to IAWHC. At Sidney, WASI was positively correlated to ASA and IAWHC. Similarly, SHC was positively correlated to IAWHC.

The principal component analysis (PCA) indicated that soil physical properties contributed 34% of the total variability in principal component 1 (PC1) and 20% variability in principal component 2 (PC2) at Froid (Fig. 1). Clay concentration was positively associated with volumetric water content at 0.3 and 33 kPa (VWC0.3 and VWC33, respectively), volumetric water content at water saturation (VWCFS), and MAP, but negatively associated with BD. The IAWHC was positively associated with VWC33 and WSA. The ASA and WASI were positively associated with WSA, AS, and volumetric water content at the field-moist soil (VWCFM). The SHC was positively associated with VWC53 and WSA. The ASA and WASI were positively associated with VWCFM and dry aggregate stability index (DASI), but negatively associated with stone content (SC). Soil physical properties that were not associated with clay concentration, ASA, WASI, IAWHC, SHC, and BD were sand concentration, repacked core available water holding capacity (RAWHC), volumetric water content at 10 kPa (VWC10), mesoporosity (MEP), volumetric water content at 1500 kPa (VWC1500), silt concentration, and total shrinkage (TS).

At Sidney, soil physical properties contributed 33% of the total variability in PC1 and 19% of the variability in PC2 (Fig. 1). The ASA was positively associated with AS and WSA and negatively associated with VWC33, VWC10, RAWHC, and VWC1500. The WASI was positively associated with MEP, VWC0.3, and AS, but negatively associated with VWC1500. The IAWHC was positively associated with DASI. The BD was positively associated with VWC6M, but negatively associated with VWC0.3. The clay concentration was positively associated with silt concentration, but negatively associated with sand concentration. Soil physical properties that were not associated with clay concentration, ASA, WASI, IAWHC, SHC, and BD were VWCFS, TS, MAP, RAWHC, and SC.



Figure 1. Principal component analysis for the associations among average slake aggregate (ASA), bulk density (BD), clay concentration, intact core available water capacity (IAWHC), saturated hydraulic conductivity (SHC), wet aggregate stability index (WASI), and soil physical properties at Froid and Sidney, MT. At Froid, soil physical properties explained 33.9% of the total variability in the first principal component (PC1) and 20.0% in the second principal component (PC2). At Sidney, physical properties explained 32.6% of total variability in PC1 and 19.4% of the total variability in PC2. AS denotes aggregate stability; DASI, dry aggregate stability index; MAP, macro-porosity; MEP, meso-porosity; RAWHC, repacked core available water capacity; SC, stone content; TS, total shrinkage; VWC0.3, volumetric water content at 0.3 kPa; VWC10, volumetric water content at 10 kPa; VWC33, volumetric water content at 33 kPa; VWC1500, volumetric water content at 1500 kPa; VWCFM, volumetric water content in the field-moist soil; VWCFS; volumetric water content at water saturation; and WSA, water-stable aggregation.



Figure 2. Principal component analysis for the associations among average slake aggregate (ASA), bulk density (BD), clay concentration, intact core available water capacity (IAWHC), saturated hydraulic conductivity (SHC), wet aggregate stability index (WASI), and soil chemical properties at Froid and Sidney, MT. At Froid, soil chemical properties explained 30.0% of the total variability in the first principal component (PC1) and 24.1% in the second principal component (PC2). At Sidney, chemical properties explained 40.9% of total variability in PC1 and 15.4% in PC2. BpH denotes buffer pH; EC, electrical conductivity; IP, inorganic P concentration; OP, organic P concentration; and SAR, sodium-absorption ratio.

Soil chemical properties. In the PCA, soil chemical properties contributed 30% of the total variability in PC1 and 24% of the variability in PC2 at Froid (Fig. 2). The ASA and IAWHC were positively associated with Cd, inorganic P (IP), Mn, Pb, K, and Zn concentrations, but negatively associated with Na-absorption ratio (SAR). The WASI was positively associated with electrical conductivity (EC) and Zn, Mn, Pb, and K concentrations. The SHC was positively associated with Na concentration. Clay concentration was positively associated with EC and Cu, Co, B, As, Ba, and Ni concentrations, but negatively associated with organic P (OP) concentration. The BD



Figure 3. Principal component analysis for the associations among average slake aggregate (ASA), bulk density (BD), clay concentration, intact core available water capacity (IAWHC), saturated hydraulic conductivity (SHC), wet aggregate stability index (WASI), and soil biological and biochemical properties at Froid and Sidney, MT. At Froid, soil biological and biochemical properties explained 52.0% of the total variability in the first principal component (PC1) and 12.1% in the second principal component (PC2). At Sidney, biological and biochemical properties explained 34.1% of the total variability in PC1 and 16.9% in PC2. ACEP denotes autoclaved citrate-extractable protein; AST, arylsulfatase; BG, β -glucosidase; CO₂-1d, CO₂ flush at 1 d incubation; CO₂-4d, CO₂ flush at 4 d incubation; MAC, microbially active C; NAG, N-acetyl β -glucosaminidase; PLFA, phospholipid-derived fatty acid; PME, phosphomonoesterase; PNM, potentially N mineralization; POXC, KMnO₄-extractable C; SOM, soil organic matter; STC, soil total C; STN, soil total N; WEC, water-extractable C; WEON, water-extractable N; and WETN, water-extractable total N.

was positively associated with SAR, but negatively associated with Cd concentration. Soil chemical properties that were not associated with ASA, BD, Clay, IAWHC, SHC, and WASI were pH, buffer pH (BpH), and Al, Fe, Ca, Mg, and S concentrations.

At Sidney, chemical properties contributed 41% of the total variability in PC1 and 15% of the variability in PC2 (Fig. 2). The ASA was positively associated with K, IP, and OP concentrations, but negatively associated with Ni and Mg concentrations. The WASI was positively associated with K concentration, but negatively associated with Ba concentration. The BD was positively associated with Ni and Mg concentrations. Clay concentration was positively associated with Ba and Cr concentrations. Soil chemical properties that were not associated with ASA, BD, Clay, IAWHC, SHC, and WASI were pH, BpH, EC, SAR, and Pb, As, Ca, Mg, Co, B, Cu, Na, Al, MN, S, Fe, Zn, Cd, OP, and IP concentrations.

Soil biological and biochemical properties. At Froid, the PCA indicated that soil biological and biochemical properties contributed 52% of the total variability in PC1 and 12% of the variability in PC2 (Fig. 3). The ASA and WASI were positively associated with phospholipid-derived fatty acid (PLFA), water-extractable C (WEC), CO_2 flush at 4-d incubation (CO_2 -4d), KMnO_4-extractable C (POXC), soil organic matter (SOM), MAC, BG, and CO_2 flush at 1-d incubation (CO_2 -1d). The IAWHC was positively associated with water-extractable total N (WETN), PNM, water-extractable organic N (WEON), CO_2 -1d, and NO_3 -N. Clay concentration was positively associated with PLFA. The SHC was positively associated with NO₃-N and WEON. In contrast, BD was negatively associated with PME, autoclaved citrate-extractable protein (ACEP), NAG, soil total C (STC), and STN. Soil biological and biochemical properties that were not associated with ASA, BD, Clay, IAWHC, SHC, and WASI were arylsulfatase (AST), NH₄-N, PME, NAG, STC, and STN.

At Sidney, soil biological and biochemical properties contributed 34% of the total variability in PC1 and 16% of the variability in PC2 (Fig. 3). The ASA was positively associated with PNM, MAC, CO_2 -1d, NAG, and SOM. The WASI was positively associated with SOM, NAG, and STC. The IAWHC was positively associated with PLFA. The SHC was positively associated with AST. In contrast, clay concentration was negatively associated with ACEP, WEON, and BG. The BD was negatively associated with NAG, SOM, and MAC. Soil biological and biochemical properties that were not associated with ASA, BD, Clay, IAWHC, SHC, and WASI were NO₃-N, WETN, NH₄-N, WEC, STN, CO_2 -4d, and POXC.

Relationship between soil physical properties and crop yields. Clay concentration was strongly ($P \le 0.01$) related to mean crop yield across years from the combined data from Froid and Sidney, but not for the individual sites (Fig. 4). Clay concentration accounted for 24% of the variability in crop yield. An increase in clay concentration by 1 g kg⁻¹ increased crop yield by 0.03 Mg ha⁻¹. The ASA was very strongly ($P \le 0.001$) related to mean crop yield at Froid and weakly ($P \le 0.05$) related at Sidney. The ASA accounted for 79% of the variability in crop yield at Froid and 15% of the variability at Sidney. The WASI was strongly related to mean crop yield at Froid, weakly related at Sidney, and very strongly related to the combined data from Froid and Sidney. The WASI accounted



Figure 4. Relationships among clay concentration, average slake aggregate (ASA), wet aggregate stability index (WASI), and mean crop yield across years at Froid, Sidney, and the combined data from Froid and Sidney.



Figure 5. Relationships among soil bulk density (BD), intact core available water capacity (IAWHC), saturated hydraulic conductivity (SHC), and mean crop yield across years at Froid, Sidney, and the combined data from Froid and Sidney.

for 50% of the variability in crop yield at Froid, 25% of the variability at Sidney, and 30% of the variability for the combined data from Froid and Sidney.

The BD was the only soil physical property that negatively related to mean crop yield (Fig. 5). The BD was strongly related to crop yield at Froid and very strongly related to the combined data from Froid and Sidney (Fig. 5). The BD accounted for 39% of the variability in crop yield at Froid and 24% variability for the combined data from Froid and Sidney. An increase in BD by 1 Mg m⁻³ decreased crop yield by 1.85 Mg ha⁻¹ at Froid and 1.80 Mg ha⁻¹ for the combined data from Froid and Sidney. The IAWHC was weakly related to mean crop yield at Froid and strongly related to the combined data from Froid and Sidney. The IAWHC accounted for 33% of the variability in crop yield at Froid and 22% variability for the combined data from Froid and Sidney. The SHC was not related to crop yields at both sites or to the combined data from Froid and Sidney.

Discussion

Relationship among soil properties. The positive correlations among ASA, WASI, IAWHC, and SHC at Froid and Sidney suggest that well aggregated soils can also enhance soil water retention and infiltration capacities. As soil aggregation is related to SOC¹, it is possible that increased SOC in enhanced aggregated soil helped to retain more water. Also increased soil aggregation may have increased macroporosity which increased SHC. In contrast, negative correlations among BD, ASA, and IAWHC at Froid suggest that increased soil compaction can reduce slaking resistance of soil aggregates and water holding capacity. Blanco-Canqui and Benjamin¹ reported that well aggregated soils have lower BD, better consistency, and greater macropores and water retention capacity than poorly aggregated soils. Benjamin et al.¹⁸ found that soil aggregation was positively correlated to SHC, but negatively correlated to BD. Manrique and Jones¹⁹ reported that BD was negatively correlated to soil water content. Our result of positive correlation between clay concentration and WASI was similar to that observed by Shaver et al.⁹.

The positive associations among ASA, WASI, WSA, AS, VWC33, IAWHC, and VWCFM at Froid and Sidney suggest that increased soil aggregation enhanced aggregate stability, slaking resistance, and water holding capacity. Water-stable aggregates with enhanced stability have higher slaking resistance and water holding capacity and are less erodible than aggregates with reduced stability¹. The close association between IAWHC and VWC33 at Froid indicates that water holding capacity increased as soil water content increased. Similarly, the close association between SHC and VWCFM suggests that the water infiltration rate increased as the soil water content at saturation increased. The negative association between clay concentration and BD suggests that soils with high clay concentration are less dense than soils with less concentration. Similarly, the negative association between clay and sand concentrations at both sites suggests that clay concentration increased as sand concentration decreased.

The positive associations among ASA, WASI, and some nutrients at Froid and Sidney suggest that improved soil aggregation can favor retention and availability of nutrients probably by enhancing nutrient cycling. As soil aggregation is closely related to SOC^{1,6}, it is likely that increased SOC that enhanced soil aggregation also favored nutrient cycling. Similarly, the positive associations among IAWHC and some nutrients indicate that increased water holding capacity of soil also may help to retain nutrients that may be leached out of the soil profile. Increased clay concentration also may have favored the retention of nutrients, as clay concentration was positively associated with nutrients at both sites. Negatively charged clay particles may have attracted cations, helping to retain positively charged nutrients. The BD and SHC had minor associations with soil chemical properties, as soil compaction and water infiltration capacity may be less important for affecting chemical properties and nutrient levels.

A greater number of soil chemical properties were associated with ASA, BD, Clay, IAWHC, SHC, and WASI at Froid than at Sidney (Fig. 2). This suggests that the longer duration of the experiment conducted in coarse-textured soil may have altered both soil physical and chemical properties that were more related to each other than the shorter duration of the experiment conducted in fine-textured soil. This indicates that long duration experiments may be more ideal to evaluate soil physical properties as potential soil health indicators relating to chemical properties than short duration experiments in dryland cropping systems.

The positive associations among ASA, WASI, and most biological and biochemical properties reveal that soil aggregation is related to SOM, microbial abundance and activity, nitrogen mineralization, and enzyme activities. The binding agents from soil organic matter and fungal growth enhance soil aggregation¹, as fungi contribute more to soil aggregation than bacteria because of their greater abundance^{20,21}. Some researchers^{1,6,7} reported that AS was positively correlated, but BD was negatively correlated to SOM. The increased SOM associated with improved soil aggregation enhances soil microbial biomass and activity and N mineralization^{1,22}. Soil aggregation has been shown to be positively correlated to C mineralization^{23,24}, WEC²⁵, and BG activity¹¹. Norkaew et al.⁴ reported that WSA was positively related to SOM, STN, PNM, MAC, but BD was negatively related to these parameters. It is likely that soils get less dense, as SOM is increased, favoring soil biological and biochemical properties. The positive associations among IAWHC, WETN, PNM, WEON, CO₂-1d, NO₃-N, and PLFA suggest that increased soil water holding capacity can enhance N mineralization and availability by stimulating microbial biomass and activity. Rice and Havlin²⁶ reported that soil water content was linearly related to PNM. Similarly, the positive associations among SHC, NO₃-N, and WEON indicate that increased water infiltration capacity may enhance N availability by increasing N mineralization through increased aeration. In contrast, the negative associations among clay concentration and ACEP, WEON, and BG indicate that increased clay concentration might inhibit N availability by reducing enzyme activity.

Relationship between soil physical properties and crop yields. The positive relationships among ASA, WASI, IAWHC, clay concentration, and mean crop yields across years suggest that improved soil aggregation, water holding capacity, and fine-textured soils increased crop yields probably by enhancing soil water retention and availability, aeration, nutrient cycling, and root growth, as soil aggregation promotes these variables^{1,22}. Several researchers^{3,15} found that soil aggregation and stability as well as soil water content were positively related to crop yields. Our results are in contrast to those reported by Sene et al.¹⁴ who observed that soil aggregation was negatively related, but SHC was positively related to crop yield. In contrast, the negative relationship between BD and crop yield indicates that increased soil compaction reduced crop yield, likely due to reduced porosity, aeration, water and nutrient movements, and root growth. This was due to reduced BD from long-term continuous cropping systems which increased annualized crop yields. Many researchers^{15,17,27} reported that BD was negatively related to crop yield. However, Logsdon and Karlen² demonstrated that BD had no effect on crop yield.

Implications of soil physical properties. The ASA, WASI, and IAWHC were associated with most soil physical, chemical, biological, and biochemical properties, followed by clay concentration (Figs. 1, 2, and 3). The BD and SHC were minimally associated with soil properties. The ASA, WASI, and IAWHC were also weakly to very strongly related to mean crop yields across years at Froid and Sidney (Figs. 4 and 5). Clay concentration, BD, and SHC were either not related or weakly to strongly related to crop yield.

These results suggest that ASA, WASI, and IAWHC can be regarded as potential soil health indicators that were associated with most soil physical, chemical, biological, and biochemical properties and related to mean crop yields across years. However, it takes a long time to measure WASI and IAWHC. Soils need to be oven dried for 2–3 days and aggregates need to be separated in a stack of sieves in water to measure WASI. Similarly, soil water contents need to be determined at various hydraulic pressures and soils oven dried for several days to determine IAWHC. In contrast, the determination of ASA is simple, rapid, and inexpensive where the slaking resistance of aggregates is measured by a smart phone loaded with SLAKES software that calculates areas covered by aggregates before and after slaking in water for 10 min. Therefore, ASA can be considered as a potential soil health indicator under dryland cropping systems in arid and semiarid regions.

Materials and methods

Experimental details. The long-term experiments occurred at two dryland farming sites (Froid and Sidney) in eastern Montana, USA. The 36-yr-old experiment at Froid (48° 20' N, 104° 29' W, elevation 617 m) had a Dooley sandy loam (fine loamy, mixed, frigid, Typic Argiboroll) soil with sand, silt, and clay concentrations of 645, 185, and 170 g kg⁻¹, respectively, SOC 14.9 g kg⁻¹, and pH 6.2 at the 0–15 cm depth at the beginning of the experiment in 1983. The site had mean annual air temperature (30-yr average) of 8 °C and average annual precipitation of 357 mm. The 14-yr-old experiment at Sidney (48° 33' N, 104° 50' W, elevation 592 m) had a Williams loam (fine-loamy, mixed, superactive, frigid, Typic Argiustolls) soil with sand, silt, and clay concentrations of 350, 325, and 325 g kg⁻¹, respectively, pH 7.2, and SOC 13.2 g kg⁻¹ at the 0–20 cm depth at the initiation of the experiment in 2006. The site had mean annual air temperature of 8 °C and average annual precipitation of 340 mm. The distance between the two sites was 88 km.

Aase and Pikul²⁷ described the treatments and crop management for the Froid site and Sainju and Alasinrin²⁸ for the Sidney site. In short, treatments at Froid had four cropping systems which were fall and spring till continuous spring wheat (FSTCW), no-till continuous spring wheat (NTCWA), no-till spring wheat-barley (1984–1999) replaced by spring wheat-pea (2000–2019) (NTWPA), and spring till spring wheat-fallow (STWF), each replicated for four times. In each cropping system, each phase of the crop appeared in every year. Tillage in FSTCW occurred with a tandem disc in the fall and a field cultivator in the spring to a depth of 8 cm to prepare the seedbed. Similarly, tillage in STWF occurred with a field cultivator to a depth of 8 cm in the spring and during the fallow period as needed to control weeds. In other treatments, tillage was not applied. The control treatment was STWF which is a conventional cropping in the region. The size of the plot was $12 \text{ m} \times 30 \text{ m}$.

At Sidney, treatments were cropping system as the main plot and N fertilization as the split-plot factor arranged in a randomized block design with three replications. Cropping systems included conventional till barley/spring wheat-fallow (CTWF, traditional system), no-till barley/spring wheat-fallow (NTWF), no-till continuous barley/spring wheat (NTCWB), and no-till barley/spring wheat-pea (NTWPB). In each treatment, barley was grown from 2006 to 2011, which was replaced by spring wheat from 2012 to 2019. All phases of crops in the rotations occurred in each year. Tillage in CTWF was performed with a field cultivator to a depth of 8 cm to prepare seedbeds in the spring and during the fallow period to control weeds. Tillage was not applied to other treatments. Nitrogen fertilizer was broadcast to barley at 0 (N0) and 80 kg N ha⁻¹ (N1) from 2006 to 2011 and to spring wheat at 0 (N0) and 100 kg N ha⁻¹ (N1) from 2012 to 2019. At both sites, N fertilization rates were adjusted to soil residual N to a depth of 60 cm determined in the autumn of the previous year. Therefore, N fertilization rates included both soil and fertilizer N. Pea did not receive N fertilizer. The size of the split-plot was 12.0×6.0 m.

At both sites, spring wheat, barley, and pea were seeded at recommended seed rates with a no-till drill at a row spacing of 20 cm in late April of each year. A banded application of P and K fertilizers at 11 kg P ha⁻¹ and 27 kg K ha⁻¹, respectively, was done to each crop at seeding. Herbicides and pesticides were applied to each crop as needed. In mid-July to early August, crops were harvested with a combine from a swath of 11.0×1.5 m and grain yield was determined after oven drying subsamples at 65 °C for 7 days. After grain harvest, crop residues were returned to the soil.

Soil sampling to a depth of 15 cm and analysis of physical, chemical, biological, and biochemical properties were described in detail by Sainju et al.²⁹. The references for methods of analysis of soil properties and abbreviations of parameters are shown in Table 2. For determining BD, two core (3.5 cm inside diameter) samples to a depth of 7.5 cm were collected within a plot and oven dried at 110 °C for 24 h. The BD for a treatment was calculated as the mean values of two cores by dividing the weight of oven-dried soil by the volume of the core³⁰. The SHC was determined with an infiltrometer using the two ponding head method after measuring the water flow rate to a depth of 10 cm³¹. Sand, silt, and clay concentrations were determined using the pipette method³². The WASI was calculated as the mean-weight diameter of aggregates that passed through 1.00, 0.25, and 0.053 m sieves after shaking the aggregates in water for 10 min³³. For this, field-moist soil was oven-dried at 55 °C for 2-3 days and passed through a 4.75 mm sieve, out of which 19 g soil was sieved in a stack of sieves in water. The mean-weight diameter of aggregates was calculated by dividing the sum of the products of weight and diameter of each aggregate by the total weight of the aggregates. An image recognition software smartphone app "SLAKES" was used to determine ASA where non-sieved aggregates, about 4-10 mm in diameter, were dispersed in a petri dish and the picture of the area covered by the aggregates was taken by a hanging smartphone above the Petri dish³⁴. The areas covered by aggregates before and after immersing aggregates in water for 10 min were calculated by the software and then compared for measuring the slaking resistance of the aggregates. The IAWHC was

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Soil properties	Abbreviations	Reference numbers					
Soil physical properties							
Bulk density	BD	30					
Volumetric water content at field-moist condition, water saturation, and at 0.3, 10, 33, and 1500 kPA	VWCFM, VWCFS, VWC0.3, VWC10, VWC33, VWC1500	37					
Water holding capacity for intact and repacked cores	IAWHC and RAWHC	35					
Saturated hydraulic conductivity	SHC	31					
Sand, silt, and clay concentrations	Sand, silt, and clay	32					
Aggregate stability	AS	38					
Dry aggregate stability index	DASI	39					
Water-stable aggregation	WSA	39					
Wet aggregate stability index	WASI	33					
Average slake aggregate	ASA	34					
Stone content	SC	40					
Macro- and Mesoporosity	MAP and MEP	41					
Total shrinkage	TS	40					
Soil chemical properties							
pH	pH	42					
Electrical conductivity	EC	42					
Buffer pH	BpH	42					
Al, As, B, Ba, Ca, Cd, Co, Cr, Cu, Fe, K, Mg, Mn, Na, Ni, Pb, S, and Zn concentra- tions	Al, As, B, Ba, Ca, Cd, Co, Cr, Cu, Fe, K, Mg, Mn, Na, Ni, Pb, S, and Zn	43					
Inorganic and organic P concentrations	IP and OP	44					
Sodium-absorption ratio	SAR	45					
Soil biological and biochemical properties							
Soil total C and N	STC and STN	46					
Soil inorganic C	SIC	46					
Soil organic C	SOC	46					
Water-extractable C and N	WEC and WEN	47					
NH ₄ -N and NO ₃ -N concentrations	NH ₄ -N and NO ₃ -N	44					
KMnO ₄ -extrctable C	POXC	48					
CO ₂ flush at 1-d and 4-d incubations	CO ₂ -1d, CO ₂ -4d	49					
Phospholipid fatty acids	PLFA	50					
Autoclaved citrate-extractable protein	ACEP	38					
β-glucosidase	BG	51					
N-acetyl β-glucosaminidase	NAG	52					
Phosphomonoesterase	PME	53					
Arylsulfatase	AST	54					

Table 2. Determination of soil physical, chemical, biological, and biochemical properties.

determined as the difference between volumetric water contents at 33 and 1500 kPa which were determined by using the pressure plate technique³⁵.

Statistical analysis of data. The Pearson's correlation analysis was used to correlate soil physical properties. To determine the associations among soil physical, chemical, biological, and biochemical properties, a multivariate regression analysis, e.g. PCA using the PROC FACTOR procedure of SAS³⁶ was used to analyze the soil data. Data were loaded into two principal components (PC1 and PC2) that explained most variations among physical properties due to treatments. Soil properties were considered positively associated with each other when they are at acute angles, negatively associated when at right angles, and not associated when at obtuse angles. Regression analysis was used to explore the relationships between soil physical properties and mean crop yields across years for individual and combined sites. For these, soil physical properties were used as independent variables and crop yields as dependent variables. Because data for crop yields were absent during the fallow periods in alternate years and also for the latest year and yields can be affected by long-term management, it was decided to use mean crop yield across years for each treatment rather than the yield for the latest year to relate with soil physical properties. Mean crop yield was determined by dividing total yield by the number of years. Yield during the fallow period was considered zero for data analysis. Statistical significance was observed at $P \le 0.05$, unless mentioned otherwise.

Conclusions

The ASA, WASI, IAWHC, BD, clay concentration, and SHC varied in their relationships to soil properties and mean crop yields across years under dryland cropping systems at Froid and Sidney, Montana, USA. The ASA, WASI, and IAWHC were associated with most soil physical, chemical, biological, and biochemical properties and also weakly to strongly related to mean crop yields across years at both sites. The BD, clay concentration, and SHC were less associated with soil properties and varied in their relationships to crop yields. As a result, ASA, WASI, and IAWHC can be used as potential soil health indicators. However, it takes long time to measure WASI and IAWHC. Because of its simplicity, rapidity, and inexpensiveness as well as strong relationships to other soil properties and crop yields, ASA may be considered as an attractive potential soil health indicator under dryland cropping systems in arid and semiarid regions. As soil aggregation is an important parameter that enhances water infiltration, C sequestration, microbial activity, and root growth and decreases soil erosion, using ASA as a physical indicator of soil health not only reduces the cost of soil analysis, but also increases crop yields and environmental sustainability by improving the physical condition of soil, recycling nutrients, reducing soil erosion, and promoting soil C stocks that mitigate climate change.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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References

- 1. Blanco-Canqui, H., & Benjamin, J.G. Impacts of soil organic carbon on soil physical behavior. In *Quantifying and Modeling Soil Structure Dynamics* (eds. Logsdon, S. *et al.*). Soil Science Society of America 11–40 (2013).
- Logsdon, S. D. & Karlen, D. L. Bulk density as a soil health indicator during conversion to no-tillage. Soil Till. Res. 78, 143-149 (2004).
- Maddoni, G. A., Urricanet, S., Gherse, C. H. & Lavado, R. S. Assessing soil quality in the Rolling Pampa using soil properties and maize characteristics. Agron. J. 91, 280–287 (1999).
- Norkaew, S., Miles, R. J., Brandt, D. K. & Anderson, S. P. Effects of 130 years of selected cropping management systems on soil health properties for Sanborn field. Soil Sci. Soc. Am. J. 83, 1479–1490 (2019).
- Stone, L. R. & Schlegel, A. G. Tillage and crop rotation phases effects on soil physical properties in the west-central Great Plains. Agron. J. 102, 483–491 (2010).
- Blanco-Canqui, H., Mikha, M. M., Presley, D. R. & Claasen, M. M. Addition of cover crops enhances no-till potential for improving soil physical properties. Soil Sci. Soc. Am. J. 75, 1471–1482 (2011).
- McVay, K. A. et al. Management effects on soil physical properties in long-term tillage studies in Kansas. Soil Sci. Soc. Am. J. 70, 434–438 (2006).
- Nemes, A., Rawls, W. J. & Pachepsky, Y. A. Influence of organic matter on the estimation of saturated hydraulic conductivity. Soil Sci. Soc. Am. J. 69, 1330–1337 (2005).
- 9. Shaver, T. M. *et al.* Surface soil physical properties after twelve years of dryland no-till management. *Soil Sci. Soc. Am. J.* 66, 1296–1303 (2002).
- 10. Shestak, C. J. & Busse, M. D. Compaction alters physical, but not biological indices of soil health. Soil Sci. Soc. Am. J. 69, 236–246 (2005).
- Stott, D. E., Andrews, S. S., Liebig, M. A., Wienhold, B. J. & Karlen, D. L. Evaluation of β-glucosidase activity as a soil quality indicator for the Soil Management Assessment Framework. Soil Sci. Soc. Am. J. 74, 107–119 (2010).
- 12. Dick, R. P., Myrold, D. D. & Kerle, E. A. Microbial biomass and soil enzyme activities in composted and rehabilitated skid trail soils. Soil Sci. Soc. Am. J 52, 513–516 (1988).
- Martens, D. A., Johnson, J. B. & Frankenberger, W. T. Jr. Production and persistence of soil enzymes with repeated addition of organic residues. Soil Sci. 153, 53–61 (1992).
- 14. Sene, M., Vepraskas, M. J., Naderman, G. C. & Denton, H. P. Relationship of soil texture and structure to corn yield response to subsoiling. *Soil Sci. Soc. Am. J.* **49**, 422–427 (1985).
- Shang, J., Zhu, Q. & Zhang, W. Advancing soil physics for securing food, water, soil, and ecosystem services. Vadose Zone J. 17, 1–7 (2018).
- Abu-Hamdeh, N. H. Compaction and subsoiling effects on corn growth and soil bulk density. Soil Sci. Soc. Am. J. 67, 1213–1219 (2008).
- Rubio, V., Quincke, A. & Ernst, O. Deep tillage and nitrogen do not remediate cumulative soil deterioration effects of continuous cropping. Agron. J. 113, 5584–5596 (2021).
- Benjamin, J. G. et al. Cropping intensity effects on physical properties of a no-till loam soil. Soil Sci. Soc. Am. J. 71, 1160–1165 (2007).
- 19. Manrique, L. A. & Jones, C. A. Bulk density of soils in relation to soil physical and chemical properties. *Soil Sci. Soc. Am. J.* 55, 476–481 (1991).
- Hattori, T. & Hattori, Q. The physical environment in soil microbiology: An attempt to extend the principals of microbiology to soil microorganisms. Crit. Rev. Microbiol. 4, 423–461 (1976).
- Paul, E. A., & van Veen, J. A. The use of tracers to determine the dynamic nature of organic matter. In 11th Transaction International Congress of Soil Science. Edmonton, Canada 61–102 (1978).
- 22. Rivero, C. et al. Nitrogen mineralization in anaerobiosis as indicator of soil aggregate stability. Agron. J. 112, 592-607 (2020).
- Liebig, M. A., Jones, A. J., Doran, J. W. & Mielke, L. N. Potential soil respiration and relationship to soil properties in ridge tillage. Soil Sci. Soc. Am. J. 59, 1430–1435 (1995).
- van Es, H. M. & Karlen, D. L. Reanalysis validates soil health indicator sensitivity and correlation with long-term crop yields. Soil Sci. Soc. Am. J. 83, 721–733 (2019).
- Veum, K. S., Goyle, K. W., Kremer, R. & Motavalli, P. P. Relationships among water-stable aggregates and organic matter fractions under conservation management. Soil Sci. Soc. Am. J. 76, 2143–2153 (2012).
- Rice, C. W., & Havlin, J. L. Integrating mineralizable nitrogen indices into fertilizer nitrogen recommendations. In *Soil Testing Prospects for Improving Nutrient Recommendation* (eds. Havlin, J. L., & Jacobsen, J. S), vol. 40. Soil Science Society of America 1–13 (1994).

- 27. Aase, J. K. & Pikul, J. L. Jr. Crop and soil responses to long-term tillage practices in the northern Great Plains. Agron. J. 87, 652–656 (1995).
- Sainju, U. M. & Alasinrin, S. Y. Changes in soil chemical properties and crop yields with long-term cropping system and nitrogen fertilization. Agrosyst. Geosci. Environ. 3, e20019. https://doi.org/10.1002/agg2.20019 (2020).
- Sainju, U. M., Liptzin, D. & Stevens, W. B. How soil carbon fractions relate to soil properties and crop yields in dryland cropping systems?. Soil Sci. Soc. Am. J. 86, 795–809 (2021).
- Blake, G. R., & Hartge, K. H. Bulk density. In Methods of Soil Analysis. Part. 1. Physical and Mineralogical Methods (ed. Klute, A.), 2nd ed., American Society of Agronomy and Soil Science Society of America 363–382 (1986).
- 31. Reynolds, W. D. & Elrick, D. E. Ponded infiltration from a single ring: I. Analysis of steady flow. Soil Sci. Soc. Am. J. 54, 1233–1241 (1990).
- 32. Gee, G. W., & Bauder, J. W. Particle-size analysis. In *Methods of Soil Analysis. Part. 1. Physical and Mineralogical Methods*, (ed. Klute, A.), 2nd ed., American Society of Agronomy and Soil Science Society of America 383–411 (1986).
- Franzluebbers, A. J., Wright, S. F. & Studemann, J. A. Soil aggregation and glomalin under pastures in the southern Piedmont, USA. Soil Sci. Soc. Am. J. 64, 1018–1026 (2000).
- 34. Fujardo, M., McBratney, A. B., Field, D. J. & Minasty, B. Soil slaking assessment using image recognition. Soil Till. Res. 163, 119–123 (1986).
- Cassel, D. K., & Nielsen, D. R. Field capacity and available water capacity. In Methods of Soil Analysis. Part. 1. Physical and Mineralogical Methods (ed. Klute, A.) 2nd ed., American Society of Agronomy and Soil Science Society of America 901–926 (1986).
- SAS User' Guide. SAS/Share 9.4: User's guide, 2nd ed. SAS, Cary, NC. https://documentation.sas.com/doc/en/pgmsascdc/9.4_3.5/ shrref/titlepage.htm. (2019).
- Reynolds, W. D., & Topp, G. C. Soil water desorption and imbibition: Tension and pressure techniques. In Soil Sampling and Methods of Analysis (eds. Carter, M. R., & Gregorich, E. G.) 2nd ed. Canadian Society of Soil Science 981–987 (Taylor and Francis, 2008).
- Schindelbeck, R. R., Moebius-Clune, B. N., Moebius-Clune, D. J., Kuntz, K. S. & van Es, H. M. Cornell University Comprehensive Soil Health Laboratory Standard Operating Procedure (Cornell University, 2016).
- Kemper, W. D., & Rosenau, R. C. Aggregate stability and size distribution. In *Methods of Soil Analysis. Part. 1. Physical and Mineralogical Methods* (ed. Klute, A.), 2nd ed., American Society of Agronomy and Soil Science Society of America, Madison 425–442 (1986)
- Hao, X., Ball, B. C., Culley, J. L. B., Carter, M. R., & Parkin, G. W. Soil density and porosity. In Soil Sampling and Methods of Analysis, 2nd ed. Canadian Society of Soil Science 743–759 (Taylor and Francis, 2008).
- Topp, G. C., Galganev, Y. T., Ball, B. C., & Carter, M. R. Soil water desorption curves. In Soil Sampling and Methods of Analysis (ed. Carter, M. E.) 43–63 (Lewis Publishers, 1993).
- 42. Thomas, G. W. Soil pH and soil acidity. In *Methods of Soil Analysis. Part 3. Chemical Methods* (ed. Sparks, D. L.). 2nd ed. American Society of Agronomy and Soil Science Society of America 474–490 (1996).
- 43. Sikora, F. S. & Moore, K. Soil Test Methods from the Southeastern United States Southern Cooperative Series. Bulletin 419. Clemson University (2014).
- 44. Haney, R. L., Haney, E., Hossner, L. & Arnold, J. Modifications in the new soil extractant H3A-1: A multinutrient extractant. *Commun. Soil Sci. Plant Anal.* **4**, 1513–1523 (2010).
- Miller, R. O., Gavlak, R., & Horneck, D. Saturated paste extract for calcium, magnesium, sodium, and SAR. In Soil, Plant, and Water Methods for the Western Region. 4th ed. Western Coordinating Committee on Nutrient Management. Colorado State University 21–22 (2013).
- Nelson, D.W, & Sommers, L.E. Total carbon, organic carbon, and organic matter. In *Methods of Soil Analysis. Part 3. Chemical Methods* (ed. Sparks, D. L.). 2nd ed. American Society of Agronomy and Soil Science Society of America 961–1010 (1996).
- Haney, R. L. et al. Soil organic C: N vs. water extractable C:N. Open J. Soil Sci. 2, 269–274 (2012).
 Weil, R., Islam, K. R., Stine, M. A., Gruver, J. V. & Samson-Liebig, S. E. Estimating active carbon for soil quality assessment: A
- simple method for laboratory and field use. Am. J. Altern. Agric. 18, 3–17 (2003).
 49. Haney, R. L., Brinton, W. F. & Evans, E. Soil CO₂ respiration: Comparison of chemical titration, CO₂ IRGA analysis, and Solvita gel system. Renew. Agric. Food Syst. 23, 171–176 (2008).
- 50. Quideau, S. A. *et al.* Extraction and analysis of microbial phospholipid fatty acids in soils. *J. Visual. Exp.* **114**, 4360. https://doi. org/10.37991/54360 (2016).
- Tabatabai, M. A. Soil enzymes. In Methods of Soil Analysis. Part 2. Microbiological and Biochemical Properties (eds. Weaver, R. W. et al.), Soil Science Society of America 775–833 (1994).
- Deng, S., & Popova, I. Carbohydrate hydrolases. In *Methods of Soil Enzymology* (ed. Dick, R. P.), Soil Science Society of America 185-209 (2011).
- Acosta-Martinez, V., & Tabatabai, M.A. Phosphorus cycle enzymes. In *Methods of Soil Enzymology* (ed. Dick, R. P.) Soil Science Society of America 161–183 (2011).
- Klose S., Bilen, S., Tabatabai, M.A., & Dick, W.A. Sulfur cycle enzymes. In *Methods of Soil Enzymology* (ed. Dick, R. P.) Soil Science Society of America 125–151 (2011).

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U.S. and D.L., conceptualized the manuscript. D.L. acquired funding, methodology, and investigation. J.J. acquired the data and analysis. U.S. managed the experiment and wrote the original manuscript. All authors reviewed the manuscript.

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Competing interests

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Additional information

Correspondence and requests for materials should be addressed to U.M.S.

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