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OPEN Design, synthesis, and in silico studies of quinoline-based-benzo[d] imidazole bearing different acetamide derivatives as potent α-glucosidase inhibitors

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In this study, 18 novel quinoline-based-benzo[d]imidazole derivatives were synthesized and screened for their α -glucosidase inhibitory potential. All compounds in the series except 9q showed a significant α -glucosidase inhibition with IC₅₀ values in the range of 3.2 ± 0.3–185.0 ± 0.3 μ M, as compared to the standard drug acarbose (IC₅₀ = 750.0 \pm 5.0 μ M). A kinetic study indicated that compound 9d as the most potent derivative against α-glucosidase was a competitive type inhibitor. Furthermore, the molecular docking study revealed the effective binding interactions of 9d with the active site of the α -glucosidase enzyme. The results indicate that the designed compounds have the potential to be further studied as new anti-diabetic agents.

Diabetes mellitus (DM) is a chronic metabolic disease characterized by hyperglycemia, with the disorder in carbohydrate, fat, and protein metabolism in the body¹. DM is known as an important public health threat with around 450 million cases worldwide in 2019. This number is expected to rise to 700 million by 2045 worldwide confirming further action is required in this field^{2,3}. Long-term DM can increase the risk of various health complications including blindness, renal failure, foot amputation, as well as cardiovascular, retinopathy, and renal diseases⁴. Type 2 diabetes mellitus (T2DM) with around 90% of all cases is categorized as a major sub-type of DM. It was considered that glycemic control could be effective prevention and treatment for T2DM⁵⁻⁷

a-Glucosidase (EC 3.2.1.20) is a catalytic hydrolase enzyme present on the brush border of the small intestine which hydrolyzes oligosaccharides, trisaccharides, and disaccharides to glucose and other monosaccharides at their non-reducing ends⁷⁻¹⁰. The produced monosaccharides especially glucose enter the bloodstream, resulting in postprandial hyperglycemia thus causing diabetes¹¹⁻¹³. Therefore, the inhibition of α -glucosidase might reduce carbohydrate digestion, delay glucose uptake, and consequently, decrease blood sugar levels^{14,15}. The α -glucosidase enzyme can be inhibited by acarbose, voglibose, and miglitol with sub-optimal efficacy¹⁶. Also, long-term administration of mentioned inhibitor may cause several side effects, such as abdominal pain, diarrhea, and flatulence. As a result, a need of effective inhibitors to target α -glucosidase is highly needed¹⁷⁻¹⁹.

In the last few decades, different synthetic small molecules including benzo[d]imidazole²⁰, isatin²¹ benzo[b] thiophene²² pyrimidine²³, xanthone²⁴, chromene⁶, azole^{18,25} against α -glucosidase attracted increasing attention.

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Figure 1. Schematic illustration of previously reported α -glucosidase inhibitors and newly designed compound.

Regarding promising anti-diabetic properties of quinolone heterocyclic scaffold and benzo[d]imidazole moiety, in this study, the novel series of quinoline-based-benzo[d]imidazole bearing different acetamide derivatives were synthesized, and evaluated for their inhibition potential against the α -glucosidase enzyme. Also, kinetic as well as molecular docking studies of the most potent compound were performed to evaluate their inhibition pattern against α -glucosidase.

Results and discussion

Design of quinoline-based-benzo[d]imidazole derivatives. During the last years, several nonsugar-based α -glucosidase inhibitors were identified. The random screening of the in-house library resulted in introducing compound **A** (Fig. 1) bearing benzo[d]imidazole moiety with good potency against α -glucosidase²⁰. The follow-up structural optimization of **A** resulted in a series of novel 2-phenyl-1H-benzo[d]imidazole derivatives (compound **B** and **C**) with IC₅₀ values in the range of 0.71 to > 100 μ M compared to the acarbose as a positive control with an IC₅₀ value of 258.53 ± 1.27 μ M. Preliminary structure–activity relationships (SARs) study revealed that the benzo[d]imidazole core played key role in the inhibition of α -glucosidase activity¹⁷.

Also, recent studies demonstrated the α -glucosidase inhibitory activity of quinoline-containing compounds. The preliminary bioassay results revealed that compounds **D** and **E** (Fig. 1) had significant inhibitory potency compared to acarbose (IC₅₀=66.5±1.5 µg/mL)²⁶. To further improve the α -glucosidase inhibitory activity of quinolone derivatives, the structural modification was carried out. These analogs exhibited inhibitory potential with IC₅₀ values in the ranges between 2.60 and 102.12 µM (Compound **F**, Fig. 1)²⁷.

Furthermore, it was reported that methyl-thioacetamide moiety (compounds **G** and **H**) can not only improve α -glucosidase inhibition through generating optimum structure to effectively participate within the active site but also provide a suitable site for derivatization²⁸⁻³⁰.

In this context, molecular hybridization as a powerful tool for drug designing was applied so that benzo[d] imidazole and quinoline as potent heterocyclic pharmacophores were conjugated to different acetamide derivatives. Novel designed compounds were synthesized and evaluated for their α -glucosidase inhibitory activities. Preliminary SAR studies were conducted. Further, kinetic study plus in silico assessments were performed to evaluate the binding of the active compound to the enzyme.

Chemistry. The synthesis of compounds 9a-r is schematically shown in Fig. 2. Briefly, phosphoryl chloride in *N*,*N*-dimethylformamide (DMF) was added dropwise to the cold *N*-phenylacetamide (1) under reflux conditions for 15 h to obtain 2-chloroquinoline-3-carbaldehyde (2)³¹. Compound 2 and sodium sulfide were then dissolved in DMF and stirred at room temperature for 2 h to achieve 3-formyl-2-mercaptoquinoline (3)³². Then,



the reaction of O-phenylenediamines (4) and 3-formyl-2-mercaptoquinoline (3) in the presence of sodium metabisulfite in DMF at 150 °C for 2 h afforded the target compound 5^{33} . Synthesis of desired compounds **8a-r** was performed through the reaction of aniline derivatives (**6a-r**) with chloroacethylchloride (7) in DMF³⁴. Finally, the reaction of compounds **8a-r** and compound **5** in acetone in presence of K₂CO₃ led to the formation of products **9a-r**³⁵. The structure of all compounds was confirmed using NMR and IR spectroscopy as well as elemental

Structure–activity relationship (SAR) exploration. The results of the α -glucosidase inhibitory assay are displayed in Table 1. In general, all compounds showed significant α -glucosidase inhibition with IC₅₀ values in the range of 3.2 ± 0.3 to $185.0 \pm 0.3 \mu$ M in comparison to acarbose with an IC₅₀ value of $750.0 \pm 5.0 \mu$ M. The exception come back to **9q** which showed IC₅₀>750.

As can be seen in Table 1, benzimidazole-thioquinoline structure bearing phenylacetamide exhibited good inhibitory activities against α -glucosidase (**9a**, IC₅₀ = 30.2 ± 0.4 μ M). The incorporation of a fluorine atom at the *ortho* position of phenylacetamide (**9b**) resulted in an around the twofold loss of potency compared to **9a**. Furthermore, changing the position from *ortho* to *para* in compound **9c** (IC₅₀ = 13.5 ± 0.6 μ M) resulted in the second potent derivative in the halogen-substituted set.

Impotently, the introduction of 3-chlorophenyl at R position, compounds **9d**, displayed a significant α -glucosidase inhibition (IC₅₀=3.2±0.3 μ M) with around 250-fold improvement in the potency compared to the positive control, acarbose. Indeed, compound **9e** (R=4-chlorophenyl, IC₅₀=110.4±0.2 μ M) had inferior activity compared to **9d**. Replacement of chlorine substitution with bromine resulted in compounds **9f** and **9g**. Compound **9f** (R=2-Bromophenyl) was another potent derivative in halogen-substituted set (IC₅₀=23.4±0.2 μ M). However converting 2-Br substitution to 4-Br was not favorable (**9g**, IC₅₀=185.0±0.3 μ M) compared to **9f**. Additionally, **9h** as the multi-substituted chlorine derivative with inferior activities compared with **9c**, still exhibited promising potency compared to acarbose.

Overall, the mono-electron withdrawing group (EWG) at *para* position had a destructive effect against α -glucosidase while *ortho* and *meta* position seems more favorable. The exception in this trend came back to **9c** bearing 4-fluorine. This could be due to the smaller size and better electronegativity compared to the rest of halogen derivatives.

The evaluations on 9j-m as the mono electron-donating-substituted group (EDG) showed overall improvement in the potency so that 9l (R=4-methoxyphenyl) with an IC₅₀ of $5.7 \pm 0.3 \mu$ M was categorized as the top potent inhibitor in this group and second top potent entry among all derivatives followed by 9k (R=4-methyl phenyl) and 9j (R=2-methyl phenyl). Next, the assessment of compounds 9n and 9o were performed and,

analysis.

Compound	R	IC ₅₀ (µM) ^a	Concentrations of precipitation (µM)					
9a	Phenyl	30.2 ± 0.4	≥200					
9b	2-Fluorophenyl	61.3 ± 0.4	≥200					
9c	4-Fluorophenyl	13.5 ± 0.6	≥200					
9d	3-Chlorophenyl	3.2 ± 0.3	≥200					
9e	4-Chlorophenyl	110.4 ± 0.2	≥200					
9f	2-Bromophenyl	23.4 ± 0.2	≥200					
9g	4-Bromophenyl	185.0 ± 0.3	≥200					
9h	2,6-Dichlorophenyl	100.8 ± 0.1	≥200					
9i	4-Nitrophenyl	19.7±0.2	≥200					
9j	2-Methylphenyl	16.5 ± 0.4	≥200					
9k	4-Methylphenyl	12.3 ± 0.2	≥200					
91	4-Methoxyphenyl	5.7 ± 0.3	≥200					
9m	4-Ethylphenyl	55.6 ± 0.2	≥200					
9n	2,3-Dimethylphenyl	9.8 ± 0.5	≥200					
90	2,6-Dimethylphenyl	147.0 ± 0.2	≥200					
9p	Naphthalene	17.7 ± 0.8	≥200					
9q	Benzyl	750<	≥200					
9r	4-Fluorobenzyl	33.0±0.1	≥200					
Acarbose	-	750.0 ± 5.0	-					

Table 1. α -Glucosidase inhibitory activity of compounds **9a-r**. ^aData represented in terms of mean ± SD.

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disappointingly, **90** bearing symmetric multi-substituted moiety (R = 2,6-diCH₃, IC₅₀ = 147.0 μ M) recorded the reduction in the activity compared to **9n**. However, **9o** derivative still demonstrated around eightfold improvement in the potency compared to acarbose with IC₅₀ of 750.0 μ M.

Precise assessments on the **9j**-**o** derivatives also indicated that the position of substitutions seems to have the most dominant role in the inhibition compared to the lipophilicity of moiety.

Ring substitution assessments were also performed in which phenyl (9a) was replaced with naphthyl (9p). An improvement in the activity showed that a bulk structure is more favorable.

Next, the investigation of SAR indicated that nitro (**9i**, $IC_{50} = 19.7 \pm 0.2 \mu M$) and methoxy (**9l**, $IC_{50} = 5.7 \pm 0.3 \mu M$) moieties were optimal substituents at the *para* position of phenylacetamide which improved the α -glucosidase inhibition. These results suggested that such substitution may probably enhance the ligand–protein interaction with the α -glucosidase active site.

9q and 9r were also synthesized to evaluate the role of elongation of the linker between aryl substitutions and thioacteamide moiety. Compound 9q with benzyl substitution exhibited dramatically reduction in the α -glucosidase inhibition compared to 9a which exhibited the destructive effect of elongation of the linker in the unsubstituted derivatives. Also, there was a similar trend in the potency in 9r bearing 4-fluorobenzyl compared to 9c (R=4-Fluorophenyl).

The summary of the SARs to improve α -glucosidase inhibitory activity was depicted in Fig. 3. Overall, it can be understood that the most potent derivative (9d) exhibited better inhibitory activity against a-glucosidase compared to lead compounds including A to G reported in Fig. 1 concerning their positive control.

Enzyme kinetic studies. To gain insight into the mechanism of action of **9d** as the most potent α -glucosidase inhibitor, kinetic measurements were performed. According to Fig. 4a, the Lineweaver–Burk plot showed that the K_m gradually increased and V_{max} remained unchanged with increasing inhibitor concentration indicating a competitive inhibition. The results show **9d** bonded to the active site on the enzyme and compete with the substrate for binding to the active site. Furthermore, the plot of the K_m versus different concentrations of inhibitor gave an estimate of the inhibition constant, K_i of 3.2 μ M (Fig. 4b).

Docking analyses. To identify the accuracy and validation of docking procedures, the self-docking of acarbose (as a crystallographic ligand) was performed through induced fit docking of Schrödinger software. Alignment of the best pose of acarbose in the active site of α -glucosidase and crystallographic ligand recorded an RMSD value of 1.73 Å (RMSD should be less than 2 Å) which confirms the accuracy of docking. Next, the same

Bulk EWG at ortho and meta position seems more favorable



Figure 3. Summary of the SARs.



-2

-4



4

2

0

[DVM 2] (µM)

docking procedure was repeated with all derivatives and their binding to α -glucosidase was analyzed. Results are summarized in Table 2.

The in silico studies showed the binding energy of acarbose was -6.14 kcal/mol while the glide score value of **9a-r** ranges from -4.65 to -6.92 kcal/mol. As can be seen, the most potent derivative in in vitro assay was **9d** (IC₅₀= 3.2 ± 0.3) > **9l** (IC₅₀= 5.7 ± 0.3 μ M) > **9n** (IC₅₀= 9.8 ± 0.5 μ M) > **9k** (IC₅₀= 12.3 ± 0.2 μ M) exhibited the best in silico results with glide score value of -6.92, -6.33, -6.90 and -6.72 kcal/mol, respectively. Assessments on lest potent derivatives, **9q** (IC₅₀=750), **9g** (IC₅₀= 185.0 ± 0.3) and **9o** (IC₅₀= 147.0 ± 0.2) reveal low binding interaction with the targeted enzyme with binding energy of -4.30, -4.65 and -4.99 value.

The docking results between α -glucosidase and compound **9d** was shown in Fig. 5. Compound **9d** was well inserted into the active site and recorded a Glide score of – 6.92. Compound **9d** established critical hydrogen bond interaction with Trp481 and benzimidazole. Also, benzimidazole participated in pi–cation interaction with Arg600. On the other side of the molecule, 3-chlorophenylacetamide established H-bound interaction with Asp616 and halogen-bound interaction with Leu677. Notably, in most derivatives, the designed scaffold participated in the critical interactions within the active site of the enzyme and showed similar kinds of interactions to the native ligand.

Conclusion

In this study, a series of novel quinoline-based-benzo[d]imidazole bearing different acetamide derivatives were designed, synthesized and their inhibitory activity against α -glucosidase was performed. Most of these derivatives showed increased activity compared to acarbose as the positive control. The analysis of the SAR indicated that *meta*-chlorine substitution, as well as polar group with potential hydrogen interactions at the R position, was beneficial to α -glucosidase inhibition. The most potent candidate in this series **9d** (IC₅₀=3.2±0.3 µM) was chosen for further biological evaluation. The enzyme kinetics assessments indicated that compound **9d** inhibited α -glucosidase in a competitive inhibition manner (*Ki*=3.2 µM). According to the docking study, compound **9d** was well fitted in the active site of α -glucosidase through both hydrophobic and hydrogen interactions. Overall, it can be understood that the most potent derivative (**9d**) exhibited better inhibitory activity against a-glucosidase compared to lead compounds including **A**, to **G** reported compared to positive control reported in Fig. 1. In silico assessments confirmed the critical role of benzimidazole and aryl-acetamides to participate in interactions with the binding site of an enzyme.

Regarding that T2DM is public health concern nowadays, the inhibition of α -glucosidase is considered an efficient approach to target T2DM. It was shown that quinoline-based-benzo[d]imidazole bearing different acetamides constructed a new nucleus which provided a significant role for α -glucosidase inhibition. However, to better extract the SARs of this set of compounds, in the future project, heteroaryl or aliphatic substituents at the R position will be synthesized. Also, bioisosteric replacement of benzo[d]imidazole with other heteroaromatic rings will increase our insight into the design of more potent α -glucosidase inhibitors.

Experimental

Chemistry. All the reagents were purchased from commercial sources. ¹H and ¹³C NMR spectra were determined by a Bruker FT-400 MHz spectrometer in DMSO- d_6 . All the chemical shifts were reported as (δ) values ppm. The MS spectra were recorded using an Agilent 7890A spectrometer at 70 eV. CHNOS analysis was performed using ECS4010 Costech Company. IR spectra were obtained with a Nicolet, FR -IR Magna 550. Meltingpoint were also recorded using Kofler hot-stage apparatus.

Synthesis of 2-chloroquinoline-3-carbaldehyde $(2)^{31}$. To N, N-dimethylformamide (70.0 mmol) in the roundbottomed flask, phosphorus oxychloride (120.0 mmol) was added dropwise and the reaction mixture was stirred for 1 h at 0–5 °C. To this flask, N-phenylacetamide (30.0 mmol) was added and stirred for an extra 30 min followed by refluxing for 5–4 h under N₂ atmosphere. After the reaction was completed (TLC monitoring), the mixture was poured into crushed ice under constant stirring. The precipitate obtained was vacuum filtered, washed with water, air-dried, and recrystallized from EtOAc to give the 2-chloroquinoline-3-carbaldehyde.

Synthesis of 2-mercaptoquinoline-3-carbaldehyde $(3)^{32}$. The reaction was initiated by stirring the mixture of 2-chloroquinoline-3-carbaldehyde 2 (1 mmol) and sodium sulfide (1 mmol) for 2 h at room temperature in dry DMF (50 mL). Then, the reaction mixture was poured into crushed ice and made acidic with acetic acid. The product was filtered off, washed with water, and dried to give the desired 2-mercaptoquinoline-3-carbaldehyde that was further purified by recrystallization in ethanol.

Synthesis of 3-(1H-benzo[d]imidazol-2-yl)quinoline-2-thiol (5)³³. 2-Mercaptoquinoline-3-carbaldehyde (1 mmol) and o-phenylenediamine (1.2 mmol) were dissolved in 2 mL DMF. Under stirring at room temperature, 1 mmol of sodium metabisulfite is added and allowed to react at 120 °C for about 4 h. After completion of the reaction, the mixture was precipitated in ice water, filtered, and dried at room temperature.

Synthesis of 2-chloro-N-phenylacetamide derivatives $(8a-r)^{34}$. To a solution of aniline derivatives (1 mmol) in DMF (4 mL), chloroacetylchloride was added at 0 °C. The mixture was stirred at room temperature for 5 h and poured into water and then filtered to get the **8a-r**. The obtained solids were then filtered, dried, and recrystallized from ethanol.

Compound	R	Glide score	Amino acid	Type of interaction
		-6.43	Asp616	H-bound
9a	Phenyl		Leu678	H-bound
			Phe649	Pi-pi stacking
a1		- 5.81	Asp616	H-bound
90	2-Fluorophenyl		Asp282	H-bound
	4-Fluorophenyl		Trp376	Pi–pi stacking
		- 6.59	Trp481	Pi–pi stacking
0.			Leu677	H-bound
9c			Asp616	H-bound
			Arg600	Pi-cation
			Asp518	Salt bridge
	3-Chlorophenyl		Arg600	Pi-cation
		- 6.92	Trp481	H-bound
9d			Asp616	H-bound
			Asp616	H-bound
			Leu677	Halogen bound
	4-Chlorophenyl	- 5.39	Arg600	Pi-cation
9e			Asp282	H-bound
			Trp481	H-bound
	2-Bromophenyl		Arg600	Pi-cation
9f		-6.14	Asp282	H-bound
			Trp481	H-bound
			Asp282	H-bound
9g	4-Bromophenyl	-4.65	Trp481	Pi-pi stacking
			Phe649	Pi-pi stacking
			Trp481	Pi–pi stacking
	2,6-Dichlorophenyl	- 4.99	Trp481	Pi–pi stacking
9h			Trp481	Pi–pi stacking
			Phe649	Pi–pi stacking
			Phe649	Pi–pi stacking
	4-Nitrophenyl	-6.31	Asp616	H-bound
9i			Asp282	H-bound
			Arg281	Pi-cation
	2-Methylphenyl	-6.13	Asp616	H-bound
9j			Asp282	H-bound
			Phe525	Pi-pi stacking
9k	4-Methylphenyl	- 6.72	Phe649	Pi-pi stacking
			Phe649	Pi-pi stacking
			Asp616	H-bound
			Ser676	H-bound
	4-Methoxyphenyl	-6.33	Leu677	H-bound
91			Asp616	H-bound
			Phe649	Pi-pi stacking
			Trp481	Pi–pi stacking
	4-Ethylphenyl	-6.54	Asp282	H-bound
9m			Asp282	H-bound
			Trp481	H-bound
9n	2,3-Dimethylphenyl	- 6.900	Asp616	H-bound
			Trp481	H-bound
			Phe649	Pi–pi stacking
90	2.6-Dimethylphanyl	- 5 53	Asp616	H-bound
		5.55	Asp282	H-bound
Continued				

Compound	R	Glide score	Amino acid	Type of interaction
9p	Naphthalene	-6.25	Asp616	H-bound
			Asp616	H-bound
			Trp481	H-bound
			Phe525	Pi-pi stacking
			Trp376	Pi-pi stacking
9q	Benzyl	-4.30	Trp376	Pi-pi stacking
			Phe525	Pi-pi stacking
9r	4-Fluorobenzyl	- 5.90	Asp282	H-bound
			Asp282	H-bound
			Trp376	Pi-pi stacking
			Trp481	Pi-pi stacking
			Trp481	Pi-pi stacking
			Trp481	Pi-pi stacking
Acarbose	-	-6.14	Asp616	H-bound
			Asp616	Salt bridge
			Asp518	H-bound
			Phe525	H-bound

Table 2. Docking scores and interactions of compounds against the α-glucosidase (PDB ID: 5NN8).



Figure 5. 3D and 2D proposed binding modes of compounds 9d (blue color) with α -glucosidase.

General method for synthesis of 2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-phenylacetamide derivatives $(9a-r)^{35}$. A mixture of 3-(1H-benzo[d]imidazol-2-yl)quinoline-2-thiol (1 mmol) and potassium carbonate (1.5 mmol) in DMF were stirred at room temperature for 15–20 min. Afterward, N-chloroacetyl-aniline (1.2 mmol) was added to the above reaction mixture and stirred for an extra 4–5 h. After completion of the reaction, ice-cold water was added to the reaction mixture and stirred for 20 min. The obtained solid was filtered and washed with cold water several times. The acquired crude solid was purified by recrystallization from ethanol.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-*N*-phenylacetamide (9a). Brown solid; Yield: 93%; MP = 180–182 °C; IR (KBr, v_{max}) 3310 (NH), 3025 (C–H Aromatic), 2970 (CH₂ Aliphatic), 1675 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 10.71 (s, 1H, NH), 9.07 (s, 1H, H₃), 8.47 (s, 1H, H₄), 8.19 (d, *J*=8.00 Hz, 1H, H₈), 8.10 (d, *J*=8.60 Hz, 1H, H₅), 7.86 (t, *J*=8.40 Hz, 1H, H₇), 7.81(d, *J*=7.6 Hz, 2H, H₂, H₆), 7.65 (t, *J*=8.00 Hz 1H, H₄), 7.41 (t, *J*=7.90 Hz, 2H, H₃, H₅), 7.16 (t, *J*=7.40 Hz, 1H, H₃) ppm. ¹³C NMR (100 MHz, DMSO- d_6): δ 167.80, 157.98, 148.94, 147.10, 143.98, 139.79, 136.69, 135.03, 131.63, 129.24, 128.79, 127.65, 126.76, 125.08,

123.74, 123.63, 123.20, 123.40, 119.78, 119.48, 112.01, 36.55 ppm; ESI-MS ($C_{24}H_{18}N_4OS$): calculated m/z 410.15 [M+H]⁺, observed m/z 410.10 [M+H]⁺ Anal. Calcd. For $C_{24}H_{18}N_4OS$ C, 70.22; H, 4.42; N, 13.65; Found: C, 70.39; H, 4.60; N, 13.76;

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(2-fluorophenyl)acetamide (9b). Brown solid; Yield: 87%; MP = 183–185 °C; IR (KBr, v_{max}) 3325 (NH), 3060 (C–H Aromatic), 2950 (CH₂ Aliphatic), 1680 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.15 (s, 1H), 10.21 (s, 1H), 8.79 (s, 1H), 8.03 (d, *J*=7.9 Hz, 1H), 7.98 (d, *J*=8.4 Hz, 1H), 7.96–7.88 (m, 1H), 7.82 (t, *J*=8.2 Hz, 1H), 7.61 (t, *J*=7.4 Hz, 1H), 7.35–7.21 (m, 4H), 7.15–7.00 (m, 3H), 4.23 (s, 2H) ppm. ¹³C NMR (100 MHz, DMSO- d_6): δ 168.45, 157.90, 154.95, 152.52 (CF, ¹*J*_{CF}=243 Hz), 148.91, 147.12, 136.82, 131.63, 128.77, 127.64, 126.90, 126.84, 126.78, 125.48, 125.40, 125.14, 124.86, 124.04, 123.22, 116.00, 115.81, 35.95 ppm; ESI–MS (C₂₄H₁₇FN₄OS): calculated m/z 428.11 [M+H]⁺, observed m/z 428.20 [M+H]⁺, Anal. Calcd. for C₂₄H₁₇FN₄OS: C, 67.27; H, 4.00; N, 13.08; Found: C, 67.45; H, 4.18; N, 13.24;

2-((3-(1H-benzo[d]imidazol-2-yl)naphthalen-2-yl)thio)-N-(4-fluorophenyl)acetamide (9c). Brown solid; Yield: 89%; MP = 189–191 °C; IR (KBr, v_{max}) 3330 (NH), 3025 (C–H Aromatic), 2915 (CH₂ Aliphatic),1640 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.70 (s, 1H), 10.49 (s, 1H), 8.78 (s, 1H), 8.01 (d, *J* = 8.00 Hz, 1H), 7.92 (d, *J* = 8.30 Hz, 1H), 7.79 (t, *J* = 7.70 Hz, 1H), 7.73–7.56 (m, 5H), 7.33–7.26 (m, 2H), 7.15 (t, *J* = 8.8, 2H), 4.17 (s, 2H), ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ 159.57, 157.95 (CF, ^{*I*}*J*_{CF} = 238.1 Hz), 157.19, 148.96, 147.09, 136.68, 136.21, 131.63, 128.79, 127.3, 126.76, 125.08, 123.19, 121.25, 121.17, 115.92, 115.70, 36.49 ppm; ESI–MS (C₂₄H₁₇FN₄OS): calculated m/z 428.11 [M + H]⁺, observed m/z 428.30 [M + H]⁺, Anal. Calcd. for C₂₅H₁₈FN₃OS: C, 70.24; H, 4.24; N, 9.83; Found: C, 70.41; H, 4.47; N, 9.99.

 $\begin{array}{l} 2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(3-chlorophenyl)acetamide (9d). & Brown solid; Yield: \\ 89\%; MP = 190-192°; IR (KBr, v_{max}) 3310 (NH), 3015 (C-H Aromatic), 2880 (CH₂ Aliphatic), 1645 (C=O) Cm⁻¹; \\ ^{1}H NMR (400 MHz, DMSO-$ *d* $_6) & 13.6 (s, 1H), 10.65 (s, 1H), 8.79 (s, 1H), 8.01 (d,$ *J*= 8.10 Hz, 1H), 7.93-7.77 (m, 4H), 7.66-7.50 (m, 3H), 7.10 (d,*J* $= 8.00 Hz, 1H), 4.18 (s, 3H) ppm. \\ ^{13}C NMR (100 MHz, DMSO-$ *d* $_6): & 168.42, 167.60, 166.09, 162.75, 157.90, 150.04, 149.10, 147.06, 143.03, 141.29, 136.63, 136.21, 133.61, 131.56, 130.39, 128.76, 127.59, 126.73, 125.12, 123.23, 123.24, 123.02, 118.97, 117.86, 31.22 ppm; ESI-MS (C₂₄H₁₇ClN₄OS): calculated m/z 444.08, [M + H]⁺, observed m/z 444.20, [M + H]⁺; Anal. Calcd. C₂₄H₁₇ClN₄OS, C, 64.79; H, 3.85; N, 12.59; Found: C, 64.95; H, 4.02; N, 12.77. \\ \end{array}$

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(4-chlorophenyl)acetamide (9e). Brown solid; Yield: 90%; MP = 185–187 °C IR (KBr, v_{max}) 3275 (NH), 3030 (C–H Aromatic), 2980 (CH₂ Aliphatic), 1680 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.14 (s, 1H), 10.57 (s, 1H), 8.78 (s, 1H), 8.01 (d, *J*=8.00 Hz, 1H), 7.89 (d, *J*=8.40 Hz, 1H), 7.78 (t, *J*=7.70 Hz, 2H), 7.68 (d, *J*=8.50 Hz, 3H), 7.59 (t, *J*=7.50 Hz, 1H), 7.37 (d, *J*=8.50 Hz, 2H), 7.29 (d, *J*=7.20 Hz, 2H) ppm. ESI–MS (C₂₄H₁₇ClN₄OS): calculated m/z 444.08, [M+H]⁺, observed m/z 444.20,[M+H]⁺, ¹³C NMR (100 MHz, DMSO- d_6): δ 168.06, 157.91, 148.93, 147.07, 138.77, 136.66, 131.64, 129.16, 128.79, 127.59, 127.14, 126.78, 125.05, 123.15, 120.99, 36.58 ppm; Anal. Calcd. for C₂₄H₁₇ClN₄OS; C, 64.79; H, 3.85; N, 12.59; Found: C, 64.95; H, 3.99; N, 12.79.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(2-bromophenyl)acetamide (9f). Brown solid; Yield: 93%; MP = 181–183 °C; IR (KBr, v_{max}) 3300(NH), 3030 (C–H Aromatic), 2965 (CH₂ Aliphatic), 1655 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.14 (s, 1H), 10.43 (s, 1H), 8.77 (s, 1H), 8.01 (d, *J* = 7.90 Hz, 1H), 7.93 (d, *J* = 8.40 Hz, 1H), 7.80 (d, *J* = 5.50 Hz, 1H), 7.78 (d, *J* = 6.1 Hz, 1H), 7.67–7.56 (m, 3H), 7.35–7.24 (m, 4H), 7.04 (t, *J* = 7.3 Hz, 1H) ppm. ESI–MS (C₂₄H₁₇BrN₄OS): calculated m/z 490.03, [M + H]⁺, observed m/z 488.10, [M + H]⁺, ¹³C NMR (100 MHz, DMSO- d_6): δ 168.25, 157.53, 149.29, 147.09, 136.81, 136.69, 133.05, 131.53, 128.75, 128.52, 127.87, 126.91, 126.84, 125.93, 125.25, 123.60, 122.90, 116.69, 115.96, 35.84 ppm; Anal. Calcd. for C₂₄H₁₇BrN₄OS C, 58.90; H, 3.50; N, 11.45; Found: C, 59.11; H, 3.69; N, 11.75.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(4-bromophenyl)acetamide (9g). Brown solid; Yield: 95%; MP = 185–187°; IR (KBr, v_{max}) 3340 (NH), 3025 (C–H Aromatic), 2870 (CH₂ Aliphatic), 1640 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.15 (s, 1H), 10.58 (s, 1H), 8.01 (d, *J*=7.90 Hz, 1H), 7.90 (d, *J*=8.10 Hz, 1H), 7.78 (t, *J*=7.70 Hz, 2H), 7.64 (d, *J*=8.50 Hz, 3H), 7.60–7.56 (m, 1H), 7.49 (d, *J*=8.50 Hz 2H), 7.35–7.25 (m, 2H), 4.17 (s, 2H) ppm. ESI–MS (C₂₄H₁₇BrN₄OS): calculated m/z 490.03, [M+H]⁺, observed m/z 488.10, [M+H]⁺, ¹³C NMR (100 MHz, DMSO-*d*₆): δ 168.10, 157.91, 148.94, 147.08, 143.98, 139.19, 136.66, 135.04, 132.07. 131.63, 128.78, 127.60, 126.77, 125.08, 123.76, 123.16, 122.41, 121.49, 121.40, 119.78, 115.18, 112.02, 36.62 ppm; Anal. Calcd. for C₂₄H₁₇BrN₄OS; C, 58.90; H, 3.50; N, 11.45; Found: C, 59.05; H, 3.70; N, 11.62.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(2,6-dichlorophenyl)acetamide (9h). Brown solid; Yield: 86%; MP = 188–190 °C; IR (KBr, v_{max}) 3340 (NH), 3035 (C–H Aromatic), 2960 (CH₂ Aliphatic), 1675 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.15 (s, 1H), 9.92 (s, 1H), 8.81 (s, 1H), 8.03 (d, *J* = 8.00 Hz, 1H), 7.97 (d, *J* = 8.40 Hz, 1H), 7.86 (d, *J* = 8.80 Hz, 1H), 7.81 (t, *J* = 7.30 Hz, 1H), 7.75–7.57 (m, 4H), 7.37 (d, *J* = 8.80 Hz, 1H), 7.33–7.25 (m, 2H) 4.26 (s, 2H) ppm. ESI–MS (C₂₄H₁₇Cl₂N₄OS): calculated m/z 478.10, [M + H]⁺, observed m/z 444.20, [M + H]⁺, ¹³C NMR (100 MHz, DMSO- d_6): δ 168.85, 157.60, 148.88, 147.10, 136.88, 134.69, 131.65, 129.30, 129.25, 128.79, 128.08, 127.77, 126.91, 126.45, 126.23, 125.19, 123.24, 35.80 ppm; Anal. Calcd. for C₂₄H₁₆Cl₂N₄OS C, 60.13; H, 3.36; N, 11.69; Found: C, 60.24; H, 3.49; N, 11.85.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(4-nitrophenyl)acetamide (9j). Pale yellow solid; Yield: 93%; MP = 180–182 °C; IR (KBr, v_{max}) 3320 (NH), 3020 (C–H Aromatic), 2965 (CH₂ Aliphatic), 1670 (C=O), 1555–1350 (NO₂) Cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.60 (s, 1H), 11.80 (s, 1H), 8.79(s, 1H), 8.24 (d, *J*=9.30 Hz, 1H), 8.01 (d, *J*=7.70 Hz, 1H), 7.91 (d, *J*=9.30, 2H), 7.86–7.74 (m, 3H), 7.63 (d, *J*=7.70 Hz, 1H), 7.57 (t, *J*=8.00 Hz, 1H), 7.35–7.25 (m, 2H), 4.21 (s, 2H) ppm. ESI–MS (C₂₄H₁₇N₅O₃S): calculated m/z 455.11, [M+H]⁺, observed m/z 455.20, [M+H]⁺, ¹³C NMR (100 MHz, DMSO-*d*₆): δ 169.12, 157.79, 148.91, 147.01, 145.99, 143.96, 142.54, 136.61, 135.04, 131.67, 128.80, 127.49, 126.81, 125.60, 125.08, 123.79, 123.04, 122.43, 119.79, 119.08, 112.02, 36.80 ppm; ESI–MS (C₂₄H₁₇N₅O₃S C): calculated m/z 455.11 [M+H]⁺, observed m/z 455.20 [M+H]⁺; Anal. Calcd. for C₂₄H₁₇N₅O₃S C, 63.29; H, 3.76; N, 15.38; N, 7.31; Found: C, 63.49; H, 3.96; N, 15.55.

2-((3-(1H-benzo[d]imidazol-2-yl)naphthalen-2-yl)thio)-N-(o-tolyl)acetamide (9k). Brown solid; Yield: 86%; MP = 179–181 °C; IR (KBr, v_{max}) 3360 (NH), 3070 (C–H Aromatic), 2980 (CH₂ Aliphatic), 1680 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.15 (s, 1H), 10.34 (s, 1H), 8.77 (s, 1H), 8.01 (d, *J*=6.60 Hz, 1H), 7.93 (d, *J*=7.10 Hz, 1H), 7.80 (d, *J*=7.40 Hz, 2H), 7.66–7.46 (m, 4H), 7.30 (t, *J*=9.50 Hz, 2H), 7.10 (d, *J*=8.10 Hz, 2H), 4.15 (s, 2H), 2.20 (s, 3H), ppm. ¹³C NMR (100 MHz, DMSO- d_6): δ 167.54, 158.01, 148.96, 147.11, 137.30, 136.70, 132.52, 131.61, 129.60, 128.77, 127.66, 126.74, 125.08, 123.23, 119.77, 119.51, 112.02, 36.54, 20.91 ppm; Anal. Calcd. for C₂₆H₂₁N₃OS: C, 73.73; H, 5.00; N, 9.92; Found: C, 73.92; H, 5.19; N, 10.11.

2-((3-(1H-benzo[d]imidazol-2-yl)naphthalen-2-yl)thio)-N-(p-tolyl)acetamide (9l). Cream solid; Yield: 91%; MP = 181–183 °C; IR (KBr, v_{max}) 3345 (NH), 3040 (C–H Aromatic), 2900 (CH-Aliphatic), 1670 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.15 (s, 1H), 10.34 (s, 1H), 8.77 (s, 1H), 8.01 (d, *J*=7.30 Hz, 1H), 7.93 (d, *J*=7.70 Hz, 1H), 7.79 (t, *J*=7.70 Hz, 2H), 7.63–7.48 (m, 4H), 7.29 (s, 2H), 7.10 (d, *J*=8.20 Hz, 2H), 4.16 (s, 2H), 2.23 (s, 3H) ppm. ESI–MS ($C_{26}H_{21}N_3OS$): calculated m/z 424.14, [M+H]⁺, observed m/z 424.10, [M+H]⁺¹³C NMR (100 MHz, DMSO- d_6): δ 162.6, 160.2, 147.4, 141.2, 140.1, 136.5, 133.7, 132.2, 131.0, 129.4, 128.4, 128.3, 126.2, 126.0, 124.0, 120.9, 28.1, 16.1 ppm; Anal. Calcd. for $C_{26}H_{21}N_3OS$ C, 73.73; H, 5.00; N, 9.92; Found: C, 73.82; H, 5.14; N, 9.99.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(4-methoxyphenyl)acetamide (9m). Cream solid; Yield: 93%; MP = 191–193 °C; IR (KBr, v_{max}) 3340 (NH), 3030 (C–H Aromatic), 2910 (CH-Aliphatic), 1680 (C=O) Cm⁻¹; ¹H NMR (400 MHz DMSO-*d*₆) δ 13.14 (s, 1H), 10.27 (s, 1H), 8.77 (s, 1H), 8.02 (d, *J*=7.70 Hz, 1H), 7.95 (d, *J*=8.40 Hz, 1H), 7.80 (t, *J*=8.30 Hz, 1H), 7.59 (t, *J*=7.90 Hz, 1H), 7.54 (d, *J*=9.00 Hz, 2H), 7.34–7.25 (m, 2H), 6.88 (d, *J*=9.10 Hz, 1H), 4.15 (s, 2H), 3.71 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ 167.23, 158.02, 155.60, 148.96, 147.12, 136.71, 132.94, 131.62, 128.78, 127.68, 126.75, 125.08, 123.24, 121.03, 114.33, 55.58, 36.43 ppm; ESI–MS (C₂₅H₂₀N₄O₂S): calculated m/z 424.14 [M+H]⁺, observed m/z 424.10 [M+H]⁺; Anal. Calcd. for C₂₅H₂₀N₄O₂S; C, 68.16; H, 4.58; N, 12.72; Found C, 68.35; H, 4.76; N, 12.90.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(4-ethylphenyl)acetamide (9n). Brown solid;Yield:93%; MP = 178–180 °C; IR (KBr, v_{max}) 3300 (NH), 3020 (C–H Aromatic), 2975(CH₂ Aliphatic), 1670 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.12 (s, 1H), 10.34 (s, 1H), 8.78 (s, 1H), 8.01 (d, *J*=8.00 Hz, 1H), 7.95 (d, *J*=8.40 Hz, 1H), 7.79 (t, *J*=7.30 Hz, 1H), 7.75–7.65 (m, 2H), 7.58 (t, *J*=7.60 Hz, 1H), 7.54 (d, *J*=8.40 Hz, 2H), 7.30 (d, *J*=6.10 Hz, 2H), 7.13 (d, *J*=8.30 Hz, 2H), 4.17 (s, 2H), 2.53 (d, *J*=7.70 Hz, 2H), 1.14 (t, *J*=7.30 Hz, 3H) ppm. ESI–MS (C₂₆H₂₂N₄OS): calculated m/z 438.55, [M+H]⁺, observed m/z 438.10,[M+H]⁺, ¹³C NMR (100 MHz, DMSO- d_6): δ 167.55, 158.00, 148.97, 147.11, 139.01, 137.47, 136.70, 131.61, 128.78, 128.41, 127.68, 126.75, 125.08, 123.24, 119.61, 36.52, 28.06, 16.18, 16.12 ppm; Anal. Calcd. for C₂₆H₂₂N₄OS; C, 71.21; H, 5.06; N, 12.78; Found: C, 71.39; H, 5.26; N, 12.97.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(2,3-dimethylphenyl)acetamide (90). Brown solid; Yield: 93%; MP = 185–187 °C IR; (KBr, v_{max}) 3300 (NH), 3020 (C–H Aromatic), 2975 (CH₂ Aliphatic) 1675 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.6 (s, 1H), 9.76 (s, 1H), 8.79 (s, 1H), 8.03 (d, *J* = 8.90 Hz, 1H), 8.01 (d, *J* = 7.90 Hz, 1H), 7.83 (t, *J* = 8.30 Hz, 1H), 7.61 (t, *J* = 7.90 Hz, 1H), 7.30 (d, *J* = 6.60, 2H), 7.15 (d, *J* = 7.50 Hz, 1H), 7.06–6.92 (m, 4H), 4.22 (s, 1H), 2.20 (s, 3H), 2.02 (s, 3H), ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ 167.73, 157.98, 149.00, 147.19, 137.38, 136.87, 136.66, 131.60, 131.57, 128.81, 127.77, 127.33, 126.77, 125.62, 125.15, 123.78, 123.39, 35.74, 20.59, 14.47 ppm; ESI–MS (C₂₆H₂₂N₄OS): calculated m/z 438.15 [M + H]⁺, observed m/z 438.10 [M + H]⁺; Anal. Calcd. for C₂₆H₂₂N₄OS C, 71.21; H, 5.06; N, 12.78; Found C, 71.39; H, 5.26; N, 12.91.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(2,6-dimethylphenyl)acetamide (9p). Brown solid; Yield: 93%; MP = 185–187 °C; IR (KBr, v_{max}) 3325 (NH), 3045 (C–H Aromatic), 2980(CH₂ Aliphatic) 1665 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.11 (s, 1H), 9.57 (s, 1H), 8.77 (s, 1H), 8.03 (t, *J*=8.40 Hz, 2H), 7.84 (t, *J*=7.70 Hz, 1H), 7.62 (t, *J*=7.20 Hz, 2H), 7.35–7.25 (m, 2H), 7.07–6.98 (m, 4H), 4.24 (s, 1H), 2.05 (s, 6H) ppm. ESI–MS (C₂₆H₂₂N₄OS): calculated m/z 438.15 [M+H]⁺, observed m/z 438.10 [M+H]⁺; ¹³C NMR (100 MHz, DMSO- d_6): δ 162.6, 161.6, 147.4, 140.9, 139.5, 133.7, 132.7, 132.3, 131.0, 129.4, 128.9, 128.3, 127.9, 127.5, 126.3, 126.0, 123.3, 43.3 ppm; Anal. Calcd. for C₂₆H₂₂N₄OS C, 71.21; H, 5.06; N, 12.78; Found C, 71.39; H, 5.26; N, 12.91.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(naphthalen-2-yl)acetamide (9q). Cream solid; Yield: 91%; MP=182-184° C; IR (KBr, v_{max}) 3340 (NH), 3030 (C-H Aromatic), 2900 (CH-Aliphatic), 1670

 $(C=O) Cm^{-1}; {}^{1}H NMR (400 MHz, DMSO-$ *d* $_6) \delta 13.16 (s, 1H), 10.33 (s, 1H), 8.80 (s, 1H), 8.10 (d,$ *J*=8.30 Hz, 1H), 8.05 (d,*J*=7.90 Hz, 2H), 7.92 (d,*J*=7.8 Hz, 1H), 7.86–7.80 (m, 1H), 7.79–7.66 (m, 3H), 7.63 (d,*J*=7.70 Hz, 2H), 7.53–7.44 (m, 2H), 7.37 (t,*J* $=7.60 Hz, 1H), 7.34–7.24 (m, 2H), 4.36 (s, 2H), ppm. ESI–MS (C₂₈H₂₀N₄OS): calculated m/z 460.14 [M+H]⁺, observed m/z 460.10 [M+H]⁺; {}^{13}C NMR (100 MHz, DMSO-$ *d* $_6): \delta 168.50, 149.01, 147.21, 136.87, 134.23, 134.13, 131.61, 128.84, 128.54, 128.40, 127.79, 126.80, 126.46, 126.10, 126.06, 125.83, 125.18, 123.40, 122.18, 35.94 ppm; Anal. Calcd. for C₂₈H₂₀N₄OS; C, 73.02; H, 4.38; N, 12.17; Found: C, 73.32; H, 4.55; N, 12.34.$

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-benzylacetamide (9r). Brown solid; Yield: 94%; MP = 183–185 °C; IR (KBr, v_{max}) 3310(NH) , 3045(C-H Aromatic), 2975 (CH₂ Aliphatic) 1655 (C=O) Cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 13.11 (s, 1H), 8.76 (s, 1H), 8.71 (t, *J*=6.00 Hz, 1H), 8.03 (d, *J*=8.00 Hz, 1H), 7.90 (d, *J*=8.5 Hz, 1H), 7.81 (t, *J*=7.70 Hz 1H), 7.62 (t, *J*=7.60 Hz, 2H), 7.34–7.25 (m, 2H), 7.21–7.13 (m, 6H), 4.31 (d, *J*=6.00 Hz, 2H), 4.06 (s,1H) ppm. ESI–MS (C₂₆H₂₂N₄OS): calculated m/z 424.14 [M+H]⁺, observed m/z 424.10 [M+H]⁺; ¹³C NMR (100 MHz, DMSO- d_6): δ 168.67, 157.83, 149.00, 147.15, 139.83, 136.79, 131.51, 128.73, 128.55, 127.90, 127.45, 127.05, 126.73, 125.10, 123.39, 42.83, 35.17 ppm; Anal. Calcd. for C₂₅H₂₀N₄OS: C, 70.73; H, 4.75; N, 13.20; Found: C, 70.92; H, 4.90; N, 13.38.

2-((3-(1H-benzo[d]imidazol-2-yl)quinolin-2-yl)thio)-N-(4-fluorobenzyl)acetamide (9s). Brown solid;Yield:89%; MP = 186–188 °C; IR (KBr, v_{max}) 3350 (NH), 3060 (C–H Aromatic), 2975 (CH₂ Aliphatic), 1670 (C=O) cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 8.74 (d, *J* = 14.20 Hz, 2H), 8.01 (d, *J* = 8.00 Hz, 1H), 7.90–7.75 (m, 2H), 7.70 (s, 2H), 7.60 (s, 1H), 7.39–7.17 (m, 4H), 6.95 (t, *J* = 8.9, 2H), 4.29 (s, 2H), 4.04 (s, 2H), ppm. ESI–MS (C₂₅H₁₉FN₄OS): calculated m/z 442.14 [M+H]⁺, observed m/z 442.10 [M+H]⁺; ¹³C NMR (100 MHz, DMSO- d_6): δ 168.75, 162.67, 160.27 (CF, ¹*J*_{CF} = 248.00 Hz), 157.81, 149.10, 147.10, 136.74, 136.04, 131.43, 129.49, 129.41, 128.71, 127.84, 126.71, 125.10, 123.45, 122.98, 115.30, 115.10, 42.20, 35.21 ppm; Anal. Calcd. for C₂₅H₁₉FN₄OS: C, 67.86; H, 4.33; N, 12.66; Found: C, 68.04; H, 4.52; N, 12.81.

 α -Glucosidase inhibitory assay. The α -glucosidase inhibitory activities of all synthesized derivatives were assayed according to the previously reported procedure^{9,36}.

Enzyme kinetic studies. The mode of inhibition of the most active compound (**9c**), identified with the lowest IC₅₀, was investigated against α -glucosidase at different concentrations of *p*-nitrophenyl α -D-glucopyranoside (4–16 mM) as substrate in the absence and presence of **9c** at different concentrations (0, 0.8, 1.6, and 3.2 μ M). A Lineweaver–Burk plot was generated to identify the type of inhibition and the Michaelis–Menten constant (K_m) value was determined from the plot between reciprocal of the substrate concentration (1/[S]) and reciprocal of enzyme rate (1/V) over various inhibitor concentrations. The experimental inhibitor constant (K_i) value was constructed by secondary plots of the inhibitor concentration [I] versus K_m^{-6} .

Molecular docking. To perform the molecular modeling investigations, the Maestro Molecular Modeling platform (version 10.5) by Schrödinger, LLC was used. The X-ray crystal structure of the receptor was downloaded from the PDB database (PDB ID: 5NN8). The protein is then prepared using a protein preparation wizard. At this point, all water molecules and co-crystallized ligands were removed, the missing side chains and loops were filled using the prime tool, and PROPKA assigned H-bonds at pH 7.4. To prepare the ligands, the 2D structures of the ligands were drawn in ChemDraw (ver. 16) and converted into SDF files, which were used further by the ligprep module. Ligands were prepared by OPLS_2005 force field using EPIK at a target pH of 7.0 ± 2 . The grid box was generated for each binding site using entries with a box size of 25 Å, all derivatives were docked on binding sites using induced-fit docking, reporting 10 poses per ligand to form the final complex^{9,37}.

Data availability

All data generated or analyzed during this study are included in this published article and its Supplementary Information files.

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Author contributions

M.N. synthesized compounds. A.D. synthesized compounds. A.I. performed docking study and contributed to the preparation of the manuscript. M.A. performed chemical analysis. N.D. and M.A. contributed to the design and characterization of compounds. M.K. performed the biological assay. M.M.K. synthesized compounds. S.M. contributed to the design and characterization of compounds. M.D. supervised the biological tests. M.A.F. supervised the biological tests. B.L. contributed to the design and characterization of compounds. M.A. supervised all phases of the study. M.M. supervised all phases of the study. All authors reviewed the manuscript.

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Competing interests

The authors declare no competing interests.

Additional information

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