



OPEN

Antibacterial and in vivo toxicological studies of Bi₂O₃/CuO/GO nanocomposite synthesized via cost effective methods

Asifa Qayyum¹, Zahida Batool¹✉, Mahvish Fatima²✉, Saeed Ahmad Buzdar¹, Hafeez Ullah¹, Aalia Nazir¹, Qaiser Jabeen³, Sofia Siddique⁴ & Rimsha Imran¹

In this research work, Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposites have been synthesized via an eco-friendly green synthesis technique, solgel route and co-precipitation method respectively for the assessment of antibacterial activity as well as in vivo toxicity. The XRD patterns confirm the formation of Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposites showing monoclinic structures. Crystallite size and lattice strain are calculated by Scherrer equation, Scherrer plot and Willimson Hall plot methods. Average crystallite size measured for Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposites by Scherrer equation, Scherrer plot and WH-plot methods are (5.1, 13.9, 11.5)nm, (5.4, 14.2, 11.3)nm and (5.2, 13.5, 12.0)nm respectively. Optical properties such as absorption peaks and band-gap energies are studied by UV-vis spectroscopy. The FTIR peaks at 513 cm⁻¹, 553 cm⁻¹ and 855 cm⁻¹ confirms the successful synthesis of Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposites. The antibacterial activity of synthesized Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposites is examined against two gram-negative (*Escherichia coli* and *Pseudomonas*) as well as gram-positive bacteria (*Bacillus cereus* and *Staphylococcus aureus*) at dose 25 mg/kg and 40 mg/kg by disk diffusion technique. Zone of inhibition for Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO at dose 40 mg/kg against *E. coli* (gram - ve) are 12 mm, 17 mm and 18 mm respectively and against *Pseudomonas* (gram - ve) are 28 mm, 19 mm and 21 mm respectively. While the zone of inhibition for Bi₂O₃/GO and Bi₂O₃/CuO/GO at dose 40 mg/kg against *B. cereus* (gram + ve) are 8 mm and 8.5 mm respectively and against *S. aureus* (gram + ve) are 5 mm and 10.5 mm respectively. These amazing results reveal that Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposite as a kind of antibacterial content, have enormous potential for biomedical applications. In addition, the in vivo toxicity of synthesized Bi₂O₃/CuO/GO nanocomposite is investigated on *Swiss Albino* mice at dose of 20 mg/kg by evaluating immune response, hematology and biochemistry at the time period of 2, 7, 14 and 30 days. No severe damage is observed in mice during whole treatment. The *p* value calculated by statistical analysis of hematological and biochemistry tests is nonsignificant which ensures that synthesized nanocomposites are safe and non-toxic as they do not affect mice significantly. This study proves that Bi₂O₃/CuO/GO nanocomposites are biocompatible and can be explored further for different biomedical applications.

The field of nanoscience and nanotechnology has grown tremendously in recent years, and its development in various industries is expanding all the time¹. Integration of nanotechnology and nanomedicine have given new prospect to medicinal and therapy industries. Nanoparticles are being employed as fluorescence, antibacterial, diagnostic agents and transfectional^{2,3}. In recent years, nanomedicine has provided developing technologies for achieving essential goals including precise cancer diagnosis, therapy as well as reducing toxicities⁴.

It is vital to examine the toxicity and clearing of novel materials for potential medicinal uses⁵⁻⁷. In the breakdown of materials, clearance mechanism through kidney and liver are critical. Renal clearance may quickly clear

¹Institute of Physics, The Islamia University of Bahawalpur, Bahawalpur, Pakistan. ²Department of Physics, Deanship of Educational Services, Qassim University, P.O.Box 6595, Buraydah 51452, Saudi Arabia. ³Department of Pharmacology, Faculty of Pharmacy, The Islamia University of Bahawalpur, Bahawalpur, Pakistan. ⁴Department of Physics, University of Engineering and Technology Lahore, Lahore, Pakistan. ✉email: zahida.batool@iub.edu.pk; M.aziz@qu.edu.sa

NPs less than 5.5 nm⁸, while reticuloendothelial systems can absorb NPs of 10–15 nm range by Ist-pass extract^{9,10}. Liver metabolism can partially remove nanoparticles greater than 50 nm^{11–14}. Nanocomposites of metal oxides have gained significant interest in current history owing to unique morphological, photocatalytic, optical, physical, thermal, electric, absorption aspects^{15,16}.

Bismuth oxide (Bi₂O₃) nanoparticles have recently received a lot of attention as a semiconductor material within bismuth-based products due to its simple as well as distinctive properties¹⁷. Bi₂O₃ nanoparticles have several crystalline phases such as monoclinic, triclinic, tetragonal, cubic and BCC as Lopez et al. have identified several crystal phases of Bi₂O₃^{18,19}. Bismuth (Bi) is a substance with significant atomic number (Z = 83) and photoelectrical absorption co-efficient larger than Pt, I and Au. High X-ray absorbance makes it ideal for use in cancer treatment and as a contrasting agent^{20–24}.

GO is a novel material made up of several carbon atoms layers arranged in 2D lattice^{25–30}. GO have two unique regions, one with sp² hybrid carbon domain while second with various oxygenated groups³¹. It has promising advantages in sustainable power, electrical gadgets, transistor, photovoltaic cells and detectors among others³². Chemical and physical characteristics of any material are strengthened when Bi₂O₃ is added to it³³. Owing to literature, number of experts worked on bismuth oxide and graphene oxide composite. Das et al. prepared Bi₂O₃/GO nanocomposite sonochemically supported through hydrothermal technique for absorbing the organic dyes³³. Manavalan et al. prepared Bi₂O₃/rGO nanocomposite sonochemically to detect hormone in serum of rat and human³⁴.

Cu is an excellent material for antibacterial applications. Copper is now employed as antibiotic, antifungicide as well as antifouling agent³⁵. It was discovered that metal surfaces having copper are now the most efficient in lowering bacterial growth after examining a variety of metal surfaces^{36,37}. Copper oxide is p-type semiconductor having 1.2 eV bandgap, has received significant interest in low price, processability, wide surface area and renewability^{38,39}. CuO nanoparticles are good contender for making antimicrobial medical equipment, bandages and ointments⁴⁰. CuO nanoparticles have been used in variety of biological studies. Booshehri et al. investigate that copper oxide nanoparticles have higher antibacterial property⁴¹. CuO nanoparticles have been found to have anticancer properties in A549 lung cancer cells⁴². From historical eras copper and copper oxides have been used in numerous biomedical applications such as tissue repair, grocery bags, and dental standards and so on, owing to unique antibacterial and anticancer abilities^{43,44}. Nanocomposites are in demand in hopes of improving biological efficiency while also meeting specific requirements. As a result, GO can provide an appropriate platform for functionalizing or hosting CuO nanoparticles. CuO and GO might be a fruitful combination of two material's properties leading to a revolutionary series of nanocomposites exhibiting unique properties. As a result, we discovered these hybrid composites worth investigating in our search of materials with improved biological activity (anticancer and antibacterial property)^{45,46}.

The current work is aimed on developing an easy method for producing Bi₂O₃/GO and Bi₂O₃/CuO/GO nanocomposites to improve biomedical applications. The produced nanocomposites are analyzed using number of physical techniques including XRD, UV–vis, SEM and FTIR. Our synthesized nanocomposites are theranostic as well as antibacterial agents. Synthesized nanocomposites have many biological applications and mainly utilized as antibacterial agents. They are used for diagnostic and treatment purpose as well as used in hospitals to kill bacteria as antibacterial agents. *Escherichia coli* and *Pseudomonas* bacteria are used to investigate the antibacterial activity of synthesized nanocomposites. Main objective of this research is to determine the in vivo toxicity studies of Bi₂O₃/CuO/GO nanocomposite. Results of Hematological, biochemistry and pathological test are reported here. Research sheds new light on the in vivo toxicity of Bi₂O₃/CuO/GO nanocomposite.

Experimental

Materials. Graphene oxide (GO), Bismuth nitrate, Bismuth nitrate pentahydrate (99.9% purity), Copper nitrate trihydrate, ammonia (NH₃), Cetyltrimethylammonium bromide (CTAB) and ethanol (≥99.9% purity) were brought from sigma Aldrich. Extract of mentha leaves was used to synthesize bismuth oxide.

Synthesis of Bi₂O₃ nanoparticles. 30 g of washed mentha leaves were immersed in 300 ml of distilled water and heated for 2 h at 80 °C to prepare leaf extract for the synthesis of bismuth oxide. Leaf extract was cooled at room temperature before being filtered through Whatman filter paper. 3 g of bismuth nitrate was dissolved in 15 ml of distilled water at 80 °C and also combined with 30 ml of prepared extract at 80 °C and stirred continuously. After 20 h, Bi₂O₃ nanoparticles were obtained. Sample was washed with distilled water as well as dried. Resulted product was heated in furnace at 500 °C for 4 h to remove the impurities.

Synthesis of Bi₂O₃/GO nanocomposite. 360 mg of graphene oxide was dissolved in 225 ml of distilled water and stirred for 1 h. 5.24 mg of bismuth nitrate was mixed in above solution along with CTAB and stirred for 40 min. NH₃ was added drop wise to maintain the pH b/w 9–10. Resulted product was washed as well as dried at 95 °C and then heated at 350 °C for 4 h to get final product Bi₂O₃/GO nanocomposites.

Synthesis of Bi₂O₃/CuO/GO nanocomposite. 0.5 g of GO was dissolved in 40 ml of distilled water and stirred for 1 h. 0.5 g of Bi (NO₃)₃·5H₂O was mixed in 35 ml of ethanol and stirred for 10 min at room temperature. 5 g of copper nitrate trihydrate Cu (NO₃)₂·3H₂O was mixed in 45 ml of ethanol and stirred for 1 h. All the three solutions of GO, Bi (NO₃)₃·5H₂O and Cu (NO₃)₂·3H₂O were mixed and stirred for 1 h to produce homogenous emulsion of black color. CTAB was added followed by the addition of ammonia (NH₃) solution drop by drop to maintain the pH above 10, as a result of this, precipitation was accomplished. Precipitates were centrifuged after 4 h of stirring at room temperature and then washed with distilled water until pH was 7. Resulting Bi₂O₃/CuO/GO nanocomposite was heated overnight in an incubator at 80 °C.

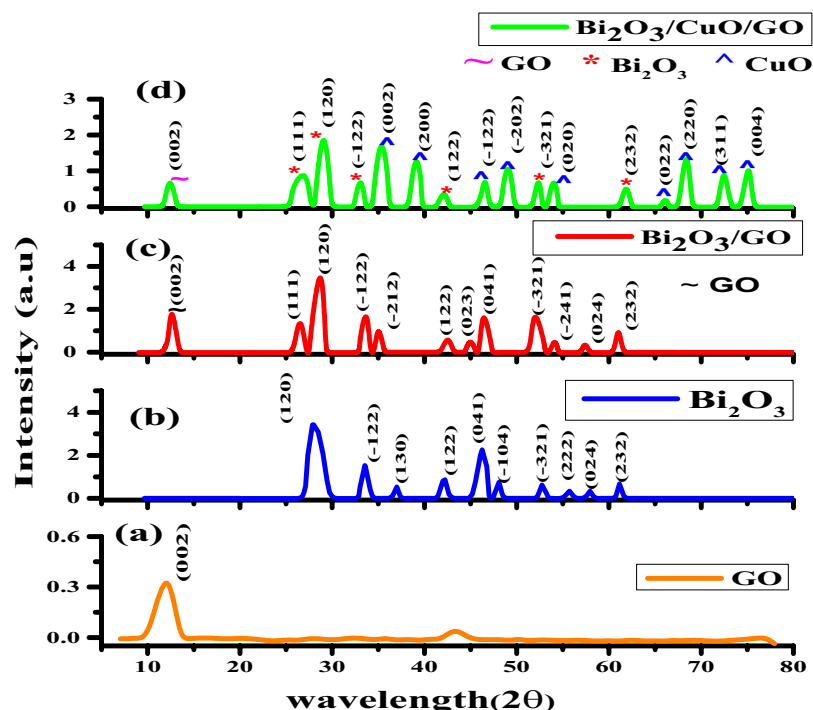


Figure 1. XRD pattern of GO (a), Bi_2O_3 (b), $\text{Bi}_2\text{O}_3/\text{GO}$ (c) and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ (d).

Experimental protocols statement. The animal studies were performed according to the ARRIVE guidelines. Additionally, All experimental protocols and animal care procedures were according to the guidelines approved by the institutional Research Ethical Committee; i.e. Pharmacy Animal Ethics committee (PAEC), under reference number PAEC/22/71.

Results and discussions

X-ray diffraction. XRD was used to investigate the crystalline structure of Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites. Figure 1a–d demonstrates the XRD patterns of GO, Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites respectively. Cu α radiations of wavelength (λ) = 1.5406 Å were used in the XRD measurements of prepared NPs. Figure 1b shows the XRD scan of Bi_2O_3 . XRD peaks of bismuth oxide revealed that nanoparticles have monoclinic structure. All the diffracted peaks with corresponding planes are confirmed by JCPDS card no. 00-041-1449. The sharp peak of bismuth oxide is observed at 27.8° with plane (120). No impurity peak is detected which indicate that the elements reached properly to form bismuth oxide phase. Figure 1c shows the XRD of $\text{Bi}_2\text{O}_3/\text{GO}$ composite prepared by sol gel method. Peak at Bragg's angle 12.5° with plane (002) correspond to the peak of GO³³. GO peak is wide due to polar–polar interaction between functional groups which contain oxygen in GO⁴⁷. Number of oxygen molecules in the composite increases when Bi_2O_3 nanoparticles are added⁴⁸. All the other peaks are in accordance with JCPDS card no. 00-041-1449 of bismuth oxide and high intensity peak occurs at 27.8° correspond to peak of Bi_2O_3 . Diffraction peaks shows that the structure of nanoparticles is monoclinic. Figure 1d shows XRD of $\text{Bi}_2\text{O}_3/\text{GO}/\text{CuO}$ nanocomposite. Diffraction peak at angle 12.4° having plane (002) correspond to peak of GO. The diffraction peaks assigned to Bi_2O_3 are detected at angle 26.7° (110), 29.0° (120), 33.0° (–122), 42.0° (122), 52.2° (–321) and 61.7° (232) have monoclinic phase according to JCPDS card 00-041-1449. The most intense peak of bismuth oxide is at 29°. Most intense diffraction peak at 35.5° with plane (002) is CuO peak according to JCPDS card no. 00-002-1041. All the diffraction peaks assigned to CuO are detected at Bragg's angle 35.3°, 39.1°, 46.5°, 49.05°, 54.09°, 66.03°, 68.4°, 72.5° and 75.1° with corresponding planes (002), (200), (–122), (–202), (020), (022), (220), (311) and (004) respectively having monoclinic phase confirmed by JCPDS card no. 00-002-1041⁴⁹. Lattice parameters, crystal size, interplanar spacing and dislocation density of synthesized materials is shown in Table 1. In ternary nanocomposite $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$, the lattice parameters a, b, c and volume is reduced in comparison to binary nanocomposite $\text{Bi}_2\text{O}_3/\text{GO}$. Similarly, the average crystallite size is also reduced 11.5 nm ($\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$) < 13.9 nm ($\text{Bi}_2\text{O}_3/\text{GO}$) respectively. The decrease in crystallite size increases the surface to volume ratio of the material which is the key factor to various novel properties of material compared to those of the corresponding bulk material. The reduction in average crystallite size is not only important for optical, electronic applications of materials⁵⁰ but also biomedical applications⁵¹ as well. The average dislocation density of ternary nanocomposite ($\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$) is decreased which is indication of reduction in grain boundaries which in turn enhance the importance of CuO presence in ternary nanocomposite as compared to binary nanocomposite $\text{Bi}_2\text{O}_3/\text{GO}$.

| Sr. no | Material | a (Å) | b (Å) | c (Å) | Volume (m ³) | Average crystal size (D) nm | Average dislocation density (σ) | Average interplanar spacing (d) |
|--------|--|-------|-------|-------|--------------------------|-----------------------------|--|---------------------------------|
| 1 | Bi ₂ O ₃ | 5.8 | 8.2 | 7.5 | 330.22 | 5.1 | 0.0726 | 0.838 |
| 2 | Bi ₂ O ₃ /GO | 5.9 | 8.1 | 7.51 | 333.95 | 13.9 | 0.014 | 0.82 |
| 3 | Bi ₂ O ₃ /CuO/GO | 5.0 | 5.6 | 5.4 | 152.7 | 11.5 | 0.009 | 0.85 |

Table 1. Lattice parameters, crystal size, inter planar spacing and dislocation density of synthesized materials.

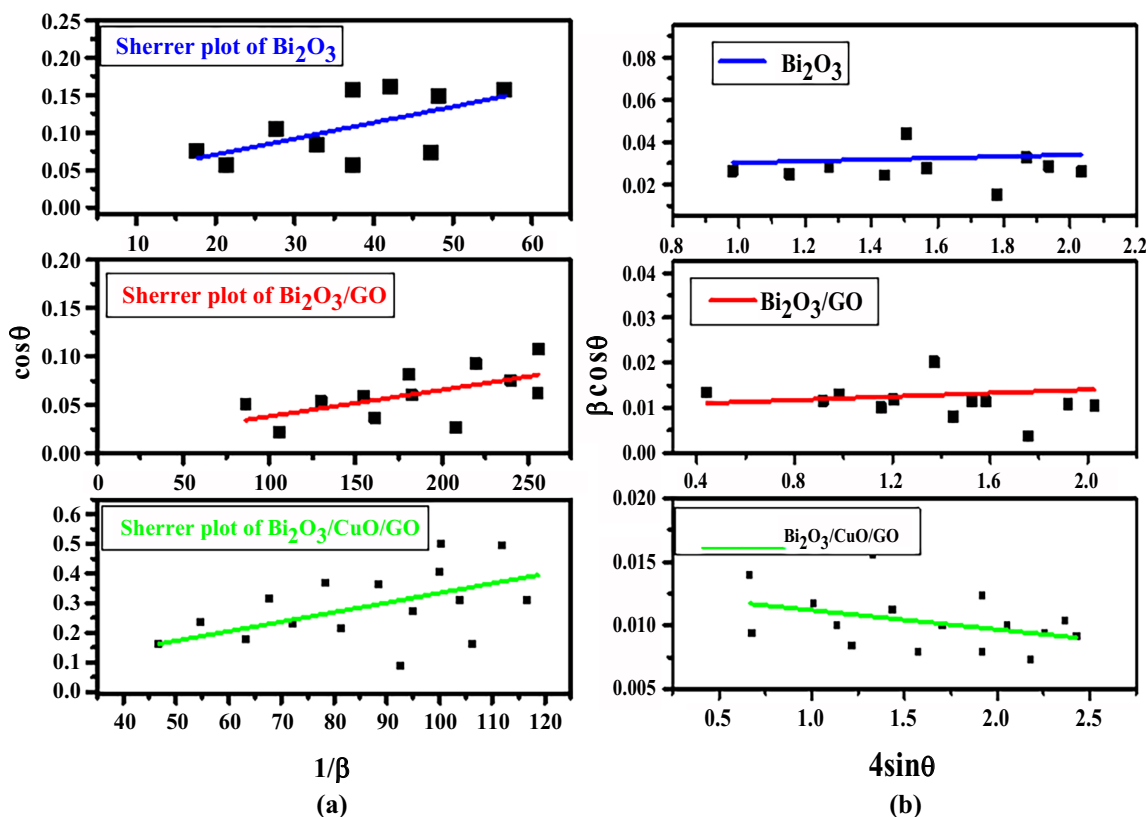


Figure 2. (a) Scherrer plot stacking (b) WH plot stacking of Bi₂O₃, Bi₂O₃/GO and Bi₂O₃/CuO/GO.

Scherrer plot method. Scherrer plot methodology was used to investigate the widening of peaks along lattice strain or crystallite size related to dislocation using XRD⁵². Bragg's peak width is equal to summation of instrumental and sample effect, as calculated by the formula:

$$\beta_{hkl} = \beta_{\text{measures}}^2 - \beta_{\text{instrumental}}^2$$

$$D = \frac{k\lambda}{\beta \cos\theta}$$

$$\text{Or } \cos\theta = \frac{k\lambda}{D} \left(\frac{1}{\beta} \right)$$

By taking $1/\beta$ along x-axis as well as $\cos\theta$ along y-axis Scherrer plot was drawn as shown in Fig. 2a. After linear fitting of data, crystal size was estimated from slope of liner line.

Williamson Hall (WH) plot method. The W–H analysis is dependent on the assumption that the estimated formulas for strain widening " β_s " as well as size broadening " β_D " change in opposite directions when Bragg's angle (θ) is taken into account. Due to crystal defects and deformation, straininduce widening can occur which is given as.

$$\varepsilon = \frac{\beta_{hkl}}{4 \tan\theta} \quad (1)$$

| Sr. no | Samples | Scherrer formula (nm) | WH plot (nm) | Scherrer plot (nm) |
|--------|--|-----------------------|--------------|--------------------|
| 1 | Bi ₂ O ₃ | 5.1 | 5.4 | 5.2 |
| 2 | Bi ₂ O ₃ /GO | 13.9 | 14.2 | 13.5 |
| 3 | Bi ₂ O ₃ /CuO/GO | 11.5 | 11.3 | 12.0 |

Table 2. Crystal size by WH plot, Scherrer formula and Scherrer plot.

Unlike Scherrer plot method, WH method is independent on $1/\cos\theta$ but dependent on $\tan\theta$. With variation in micro strain and crystal size in the crystal, the difference in 2θ allows us to distinguish b/w strain and size influences on peak widening.

$$\beta_s = 4\varepsilon \tan\theta \quad (2)$$

By adding strain and crystallite size total peak broadening is attained.

$$\beta_{hkl} = \beta_s + \beta_D \quad (3)$$

Furthermore, the uniform deformation model is used in WH analysis to assume the micro strain to be equal in all crystallographic orientations⁵². β_{hkl} for this model is

$$\beta_{hkl} = \frac{k\lambda}{D\cos\theta} + 4\varepsilon \tan\theta \quad (4)$$

By multiplying $\cos\theta$ on both sides.

$$\beta_{hkl}\cos\theta = \frac{k\lambda}{D} + 4\varepsilon \tan\theta \quad (5)$$

Here “ ε ” is strain. WH plots of are drawn by taking $4\sin\theta$ on x-axis and $\beta\cos\theta$ on y-axis as shown in Fig. 2b. Micro strain is measured from slope while crystal size is calculated from intercept of linear fitted values⁵³. Crystal size by all the three methods WH plot, Scherrer formula and Scherrer plot is shown in Table 2.

FTIR analysis. FTIR was used to study functional groups and significance of multiple kinds of functional groups within infrared spectra. FTIR analysis of NPs revealed a number of absorption peaks ranging from 4000 to 400 cm^{-1} as displayed in Fig. 3. FTIR graphs are plotted b/w wavelength (cm^{-1}) on x-axis and transmittance (a.u) on y-axis. Figure 3 is the comparison of FTIR graphs of GO (a), Bi₂O₃ (b), Bi₂O₃/GO (c) and Bi₂O₃/CuO/GO. The existence of functional groups such as hydroxyl, epoxide and carboxyl in GO is confirmed by FTIR. In graph (a), the peak observed at 3434 cm^{-1} confirms the existence of hydroxyl group (–OH). At 1723 cm^{-1} , the carbonyl group (C=O) is present. At frequency of 1395 cm^{-1} , O–H has a bending vibration^{54,55}. The presence of epoxide group (C–O) at 1104 cm^{-1} is linked with stretching vibration⁵⁶. The vibration at wavelength 1633 cm^{-1} is due to (C=C) group deposited on GO⁵⁷. In graph (b), metal oxide vibration (Bi–O) is responsible for peak at wavelength 542 cm^{-1} in Bi₂O₃. Stretching vibration of O–H is formed at 3430 cm^{-1} . In graph (c) two peaks at 543 cm^{-1} and 855 cm^{-1} with corresponding functional groups (Bi–O)⁵⁸ and (Bi–O–Bi) confirms the successful synthesis of Bi₂O₃/GO composite. Oxygen containing group (O–H) is present in Bi₂O₃/GO composite at 3434 cm^{-1} . In graph (d), main peak at 513 cm^{-1} is assigned to Cu–O stretching⁵⁹ while peak at 1601 cm^{-1} is assigned to C=O stretching^{60,61}. GO peaks at 1092 cm^{-1} , 1406 cm^{-1} and 1728 cm^{-1} with corresponding functional groups (C–O), (O–H) and (C=O) respectively as well as peaks at 855 cm^{-1} and 553 cm^{-1} having functional group (Bi–O–Bi) and (Bi–O) respectively are present in Bi₂O₃/GO nanocomposite. All these functional groups of GO and bismuth oxide are also present in the composite of Bi₂O₃/CuO/GO shown in graph (d). According to characterization the nanocomposites are successfully synthesized.

UV analysis. *Optical absorption study.* Optical absorption studies can be used to evaluate the band structure as well as bandgap energy of semiconductors, non-metallic and metallic materials. Figure 4 illustrates the absorption spectrum of (a) GO, (b) Bi₂O₃, (c) Bi₂O₃/GO and (d) Bi₂O₃/CuO/GO. In pure (a) GO and (b) Bi₂O₃, broad absorption peaks are observed at 235 nm and 290 nm respectively^{62,63}. In (c) Bi₂O₃/GO, composite peak of GO and Bi₂O₃ is present which confirms that both oxides exist in single matrix. In graph (d), CuO peak is present at 280 nm as well as GO and bismuth oxide peak is also present which confirms that three oxides exist in one matrix⁶⁴. Single oxide phase scattering caused these absorption peaks in nanocomposite. The extended tail in the absorption spectrum’s wavelength range is most likely caused by scattered radiations from mixed oxide nanoparticles.

Bandgap determination. Optical bandgap energy is estimated by this relation:

$$\alpha h\nu = A(h\nu - E_g)^n$$

A is characteristic factor; ν is frequency of incident light. Bandgap energy is estimated by plotting graph b/w $h\nu$ on x-axis and $(\alpha h\nu)^2$ on y-axis.

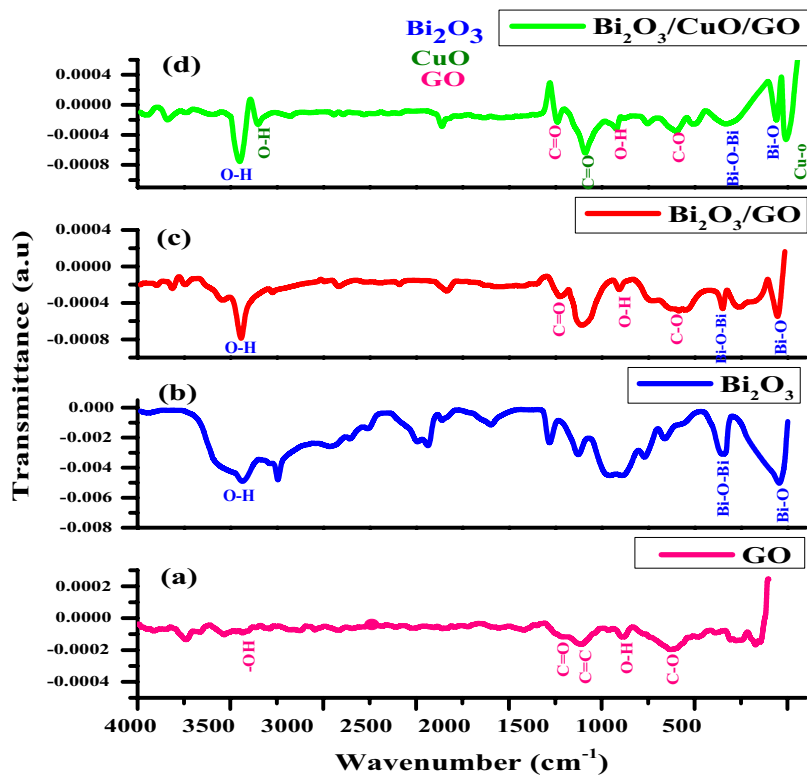


Figure 3. FTIR graphs of (a) GO, (b) Bi_2O_3 , (c) $\text{Bi}_2\text{O}_3/\text{GO}$ and (d) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$.

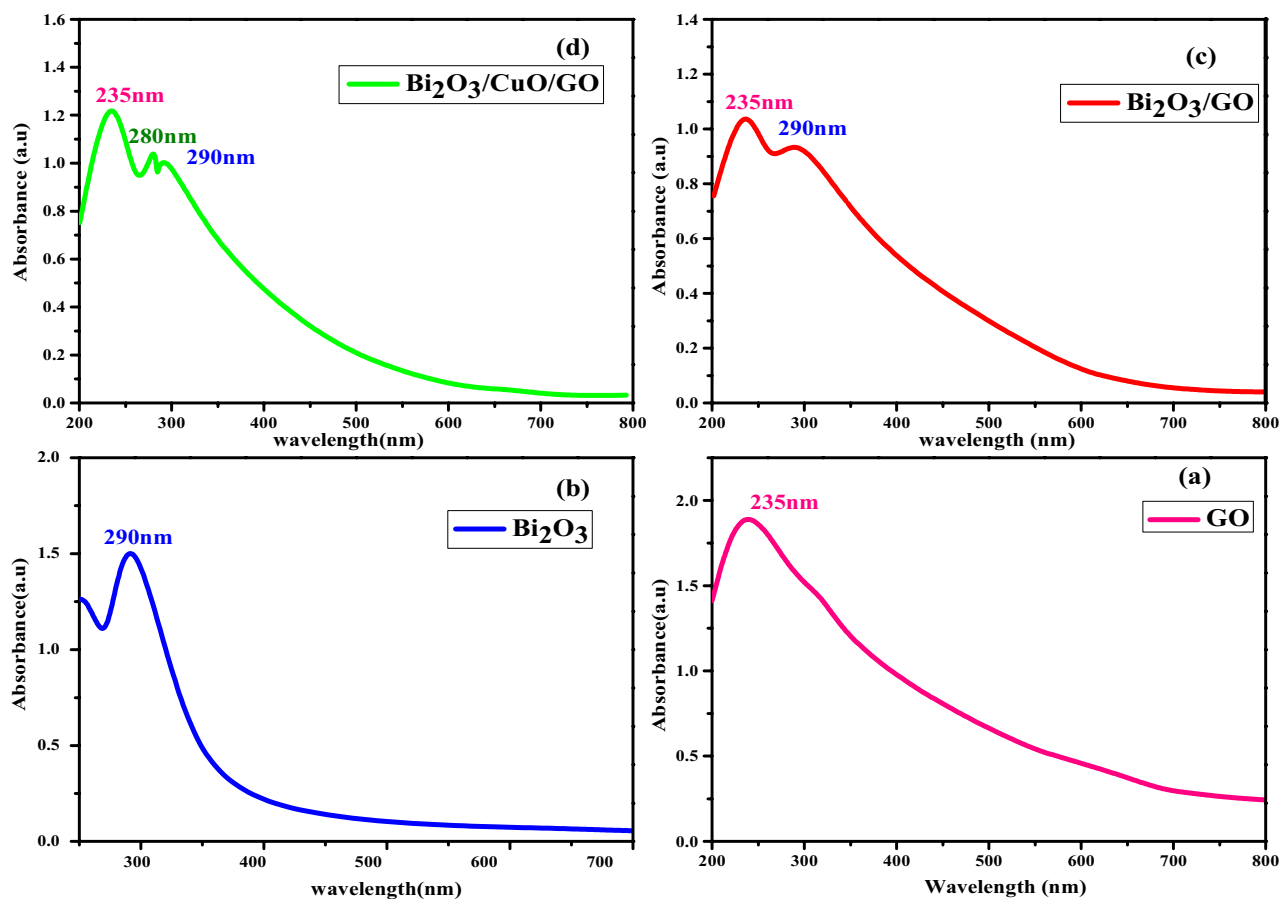


Figure 4. Illustrate the absorption spectra of (a) GO, (b) Bi_2O_3 , (c) $\text{Bi}_2\text{O}_3/\text{GO}$ and (d) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$.

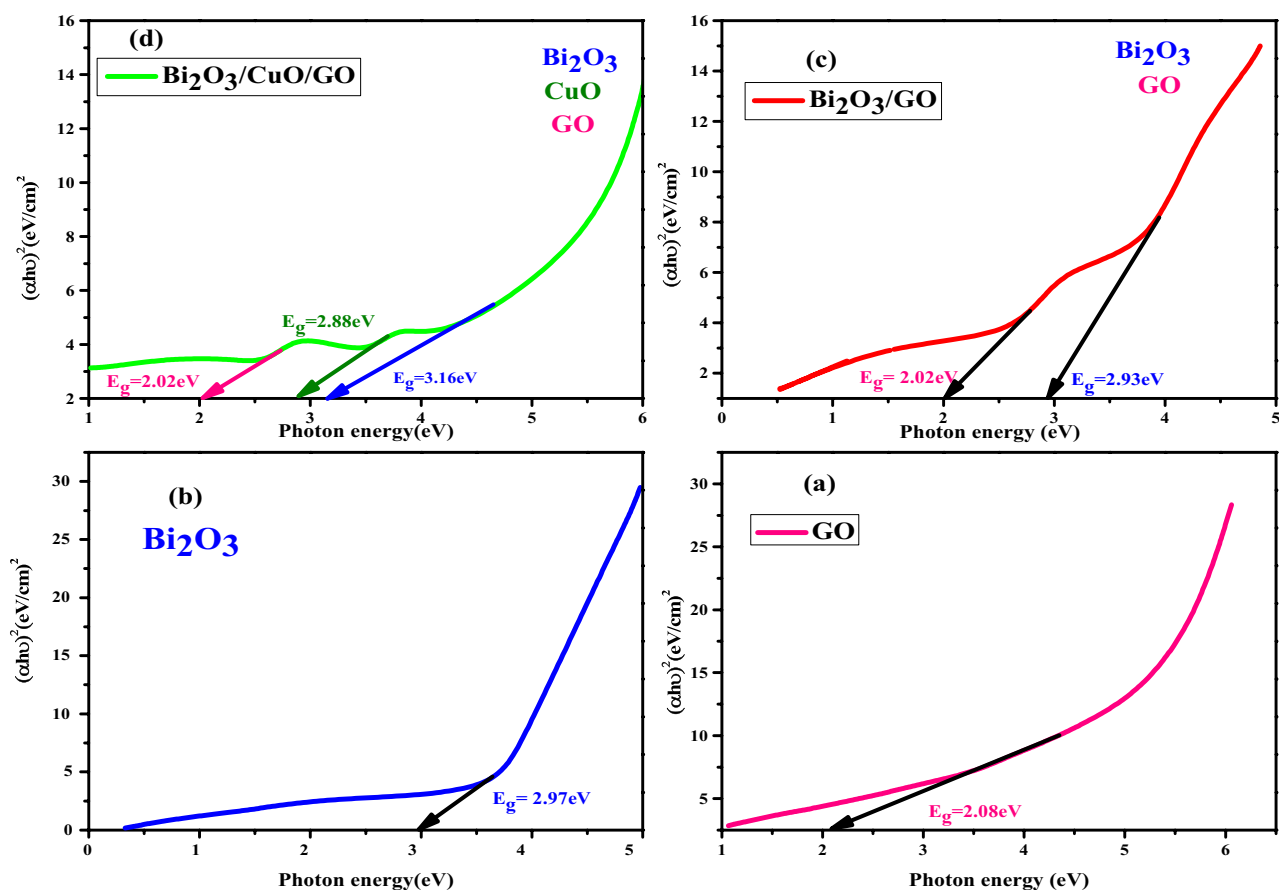


Figure 5. Band gap of (a) GO, (b) Bi_2O_3 , (c) $\text{Bi}_2\text{O}_3/\text{GO}$ and (d) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$.

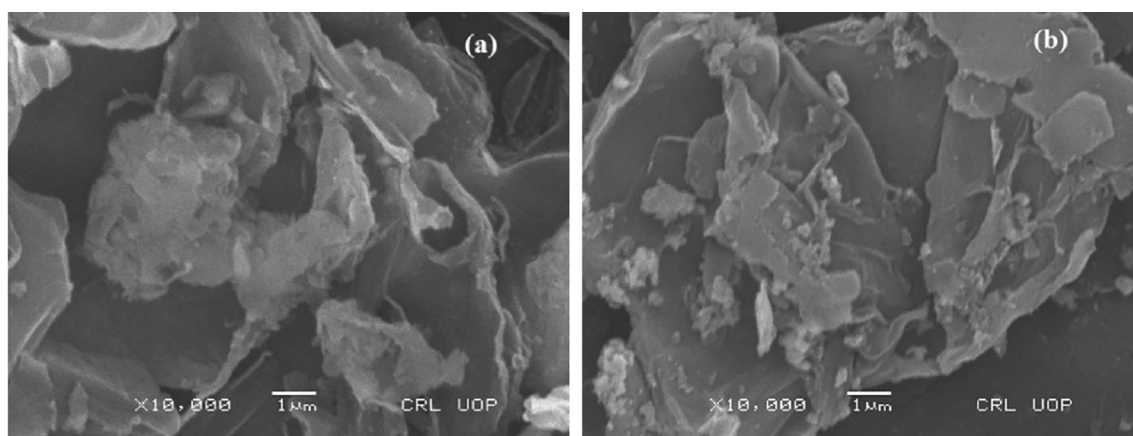


Figure 6. (a) SEM image of $\text{Bi}_2\text{O}_3/\text{GO}$ and (b) SEM image of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$.

In Fig. 5a and b, bandgap energy value of pure GO and Bi_2O_3 is 2.08 eV and 2.97 eV respectively. In graph (c), energy bandgap value of GO is 2.02 eV and Bi_2O_3 is 2.93 eV in nanocomposite. In graph (d), value of E_g for GO, CuO and Bi_2O_3 is 2.02 eV, 2.88 eV and 3.16 eV respectively in $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposite which demonstrate that composite is composed of GO, Bi_2O_3 and CuO^{62–64}.

UV analysis revealed that bandgap energy of nanocomposite may be controlled by adjusting volume fraction of the material for a variety of applications including solar cells, solid oxide and photocatalytic activity.

SEM analysis. The GO structure is sheet like and functional groups which contain oxygen may interact with GO layers and cause folding of GO sheets⁶⁵. In Fig. 6a, GO sheet like structure was observed in nanocomposite containing well-dispersed Bi_2O_3 on the surface⁶⁶. The image shows that $\text{Bi}_2\text{O}_3/\text{GO}$ nanocomposite surface is rough, may be due to development of NPs of Bi_2O_3 on GO sheets⁶⁷. In Fig. 6b, CuO NPs were distributed ran-

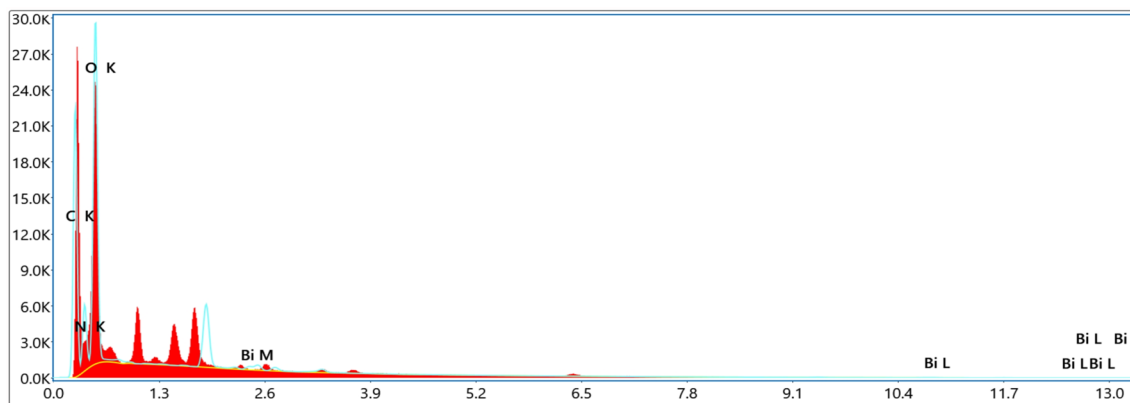


Figure 7. EDX of $\text{Bi}_2\text{O}_3/\text{GO}$.

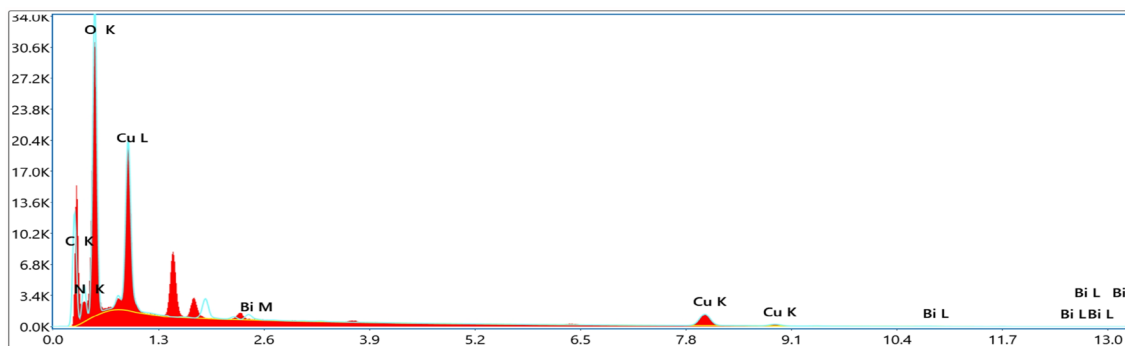


Figure 8. EDX of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$.

domly on GO sheets and data revealed that the sample $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites were grouped together and had a rough surface⁶⁸.

EDX analysis. Figure 7 shows the EDX of $\text{Bi}_2\text{O}_3/\text{GO}$ in which carbon which represents the GO is present. Bismuth is also present in the EDX of $\text{Bi}_2\text{O}_3/\text{GO}$. Similarly, in the EDX of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ bismuth and copper are present with GO which is shown in Fig. 8.

Antibacterial activity

Antibacterial activity at 40 mg/ml and 25 mg/ml concentration of Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ was performed against *E. coli* and pseudomonad (gram – ve) bacteria as well as *Bacillus cereus* and *Staphylococcus aureus* gram (+ ve) bacteria by disk diffusion technique. First of all, spread bacteria on agar plates then filter papers having synthesized samples were placed on these agar plates. These plates were placed in incubator at 37 °C for 24 h.

Results of antibacterial activity. The antibacterial results of prepared $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with concentration 40 mg/ml and 25 mg/ml are shown in Fig. 9a and b. Inhibitory zone of pseudomonas and *E. coli* (gram – ve) bacteria measured in mm is comparably higher than *B. cereus* and *S. aureus* (gram + ve) bacteria. Inhibition zone of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ composite is higher than $\text{Bi}_2\text{O}_3/\text{GO}$ nanocomposite as shown in Fig. 9a and b. Moreover, the scattering of metal oxides via GO sheets improves the antibacterial property⁶⁹. Figure 10 shows the antibacterial activity of Bi_2O_3 against *E. coli* and pseudomonas bacteria. According to Table 3 our prepared samples Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ shown better antibacterial activity than the results mentioned in literature. *Bi}_2\text{O}_3/\text{CuO}/\text{GO}* is a novel material which is not reported yet and exhibited enhanced antibacterial effects than other synthesized materials. Antibacterial activity against gram (+ ve) and gram (– ve) bacteria was performed to compare the results of both gram positive and negative bacteria. Fox 30 and Erythromycins antibiotic were used as reference against gram (– ve) and gram (+ ve) respectively.

Figure 11 shows Antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against *Bacillus cereus* (gram + ve) bacteria. Figure 12 shows antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against *Staphylococcus aureus* (gram + ve) bacteria. In Figs. 11 and 12 a disc represent the sample; B disc represent the + ve control (Erythromycin) and C shows the – ve control (distilled water). Figure 13 shows the antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml

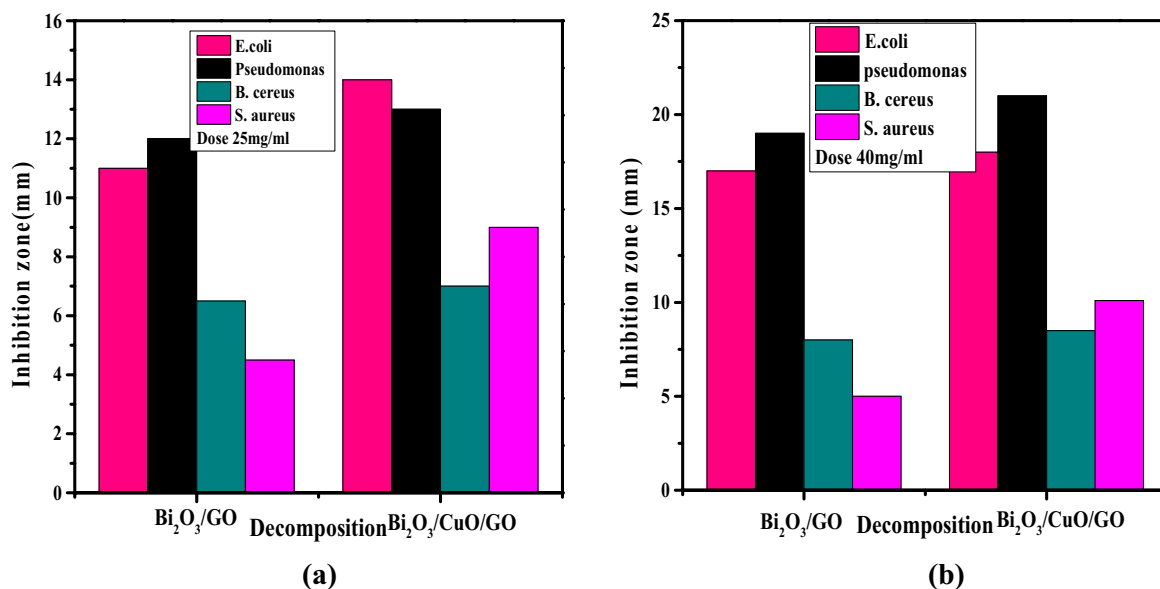


Figure 9. (a) The antibacterial activity of synthesized $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ at dose 25 mg/ml (b) the antibacterial activity of synthesized $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ at dose 40 mg/ml.

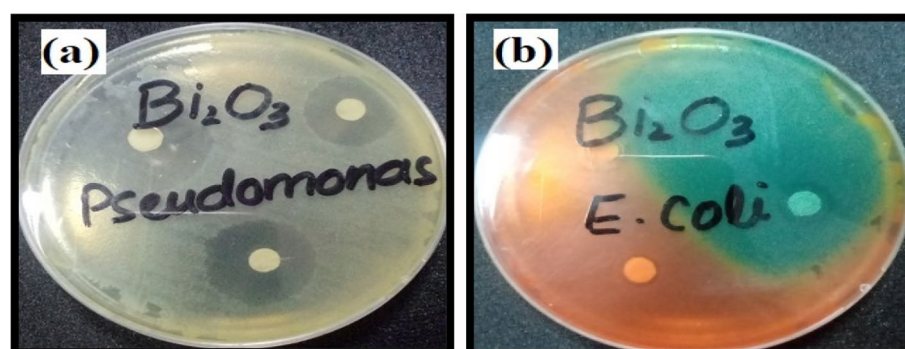


Figure 10. Antibacterial results of Bi_2O_3 on (a) *Pseudomonas* gram (–ve) (b) *E. coli* gram (–ve) bacteria.

and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against *E. coli* gram (–ve) bacteria. Figure 14 shows the antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against *pseudomonas* gram (–ve) bacteria. Results of *E. coli* gram (–ve) and *pseudomonas* gram (–ve) are better than gram (+ve) bacteria as shown in Table 4. Almost all the antibiotic discs of Fox 30 shows 18 mm zone of inhibition shown in Figs. 13a–d and 14a and c, while Fig. 14b and c shows zone of inhibition greater than 18 mm.

Figures 11 and 12 demonstrate that our sample shows better results than Erythromycin against *B. cereus* and *S. aureus* bacteria.

In vivo toxicity studies

The in vivo toxicity of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ at 20 mg/kg dose was explored on *Swiss Albino* mice (female) by analyzing body weight, acute toxicity study, hematological as well as biochemistry test at 2, 7, 14 and 30 days. Animals were divided into two groups. One is control group and second is treated group. Each group contain 4 animals. 20 mg/kg dose was administered orally to all the mice of treated group. One mouse from each group was slaughtered at 2, 7, 14 and 30 days. Blood samples and organs (liver, lungs and kidneys) were collected to perform hematological, biochemistry and pathological test. The treatment with $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ had no clear deleterious effects on the growth during 30 days period, no mortality was found and there were no significant differences in weight of body b/w $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ treated mice and control mice. The acute toxicity parameters such as Alertness, convulsions, grooming, hyperactivity, salivation, lacrimation, sweating, urination, righting reflex, gripping strength, corneal reflex, writhing reflex and pain response were investigated at 30-day time period and no meaningful difference was noticed. The animal studies were performed according to the ARRIVE guidelines. Additionally, all experimental protocols and animal care procedures were according to the guidelines approved

| Sr.no | Sample | Bacteria | Zone of inhibition (mm) | References |
|-------|--|-----------------------|-------------------------|---------------|
| 1 | Bi ₂ O ₃ | <i>E. coli</i> | 12 | Present |
| 2 | Bi ₂ O ₃ /GO | | 17 | Present |
| 3 | Bi ₂ O ₃ /CuO/GO | | 18 | Present |
| 4 | Bi ₂ O ₃ /GO | | 6.5 | ⁷⁰ |
| 5 | CuO/GO | | 11.2 | ⁷¹ |
| 6 | CuO | | 6.6 | ⁶¹ |
| 7 | CHCuO-CH | | 10 | ⁷² |
| 8 | Ag-CuO | | 17 | ⁷³ |
| 9 | Bi ₂ O ₃ | <i>Pseudomonas</i> | 28 | Present |
| 10 | Bi ₂ O ₃ /GO | | 19 | Present |
| 11 | Bi ₂ O ₃ /CuO/GO | | 21 | Present |
| 12 | CuO/GO | | 14.9 | ⁷⁴ |
| 13 | CuO | | 6.3 | ⁶¹ |
| 14 | Bilayer Wound dressing | <i>S. aureus</i> | 5.4 | ⁷⁵ |
| | | <i>E. coli</i> | 1.9 | |
| | | <i>S. epidermidis</i> | 1.0 | |
| 15 | (PU-HA)1%EEP | <i>S. aureus</i> | 2.33 | ⁷⁶ |
| | | <i>E. coli</i> | 1.96 | |
| 16 | (PU-HA)2%EEP | <i>S. aureus</i> | 5.63 | |
| | | <i>E. coli</i> | 3.18 | |
| 17 | PU-WEP | <i>S. aureus</i> | 3.89 | ⁷⁷ |
| | | <i>E. coli</i> | 3.55 | |
| 18 | CS/HA/0.5%EEP | <i>S. aureus</i> | 2.08 | ⁷⁸ |
| | | <i>E. coli</i> | 2.64 | |
| | | <i>S. epidermidis</i> | 1.02 | |

Table 3. Inhibitory zone of different samples against *E. coli* and pseudomonas.

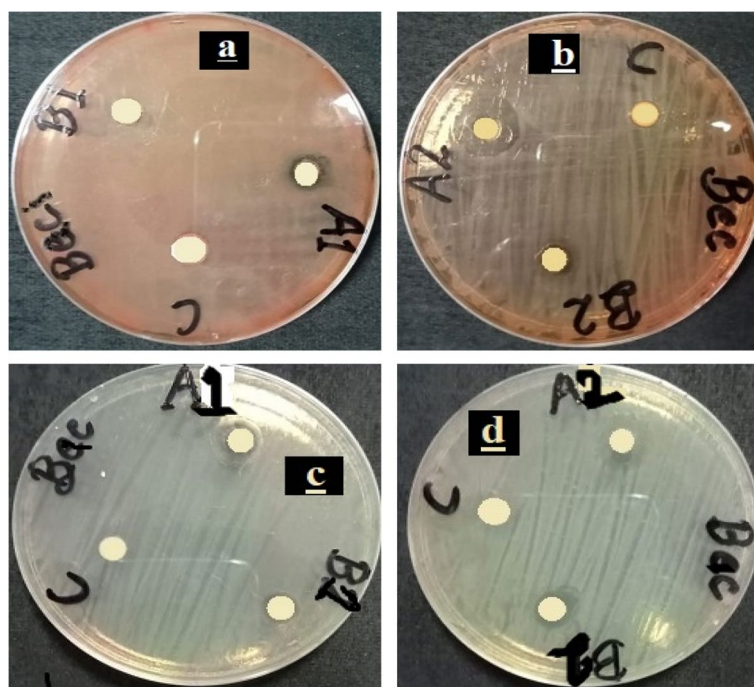


Figure 11. Antibacterial activity of (a) Bi₂O₃/CuO/GO using dose 25 mg/ml (b) Bi₂O₃/CuO/GO with dose 40 mg/ml (c) Bi₂O₃/GO with dose 25 mg/ml and (d) Bi₂O₃/GO dose 40 mg/ml against *Bacillus cereus* (gram +ve) bacteria.

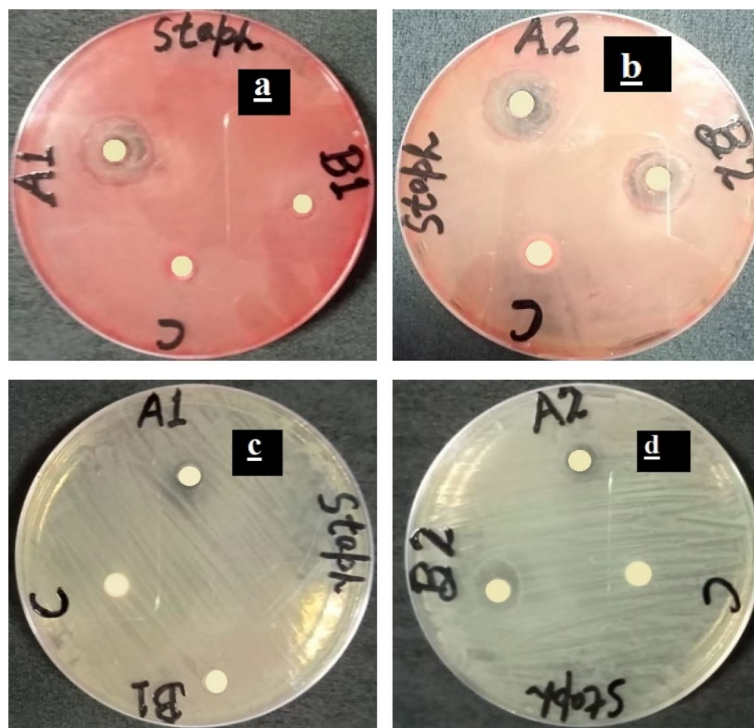


Figure 12. Antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against *Staphylococcus aureus* (gram +ve) bacteria.

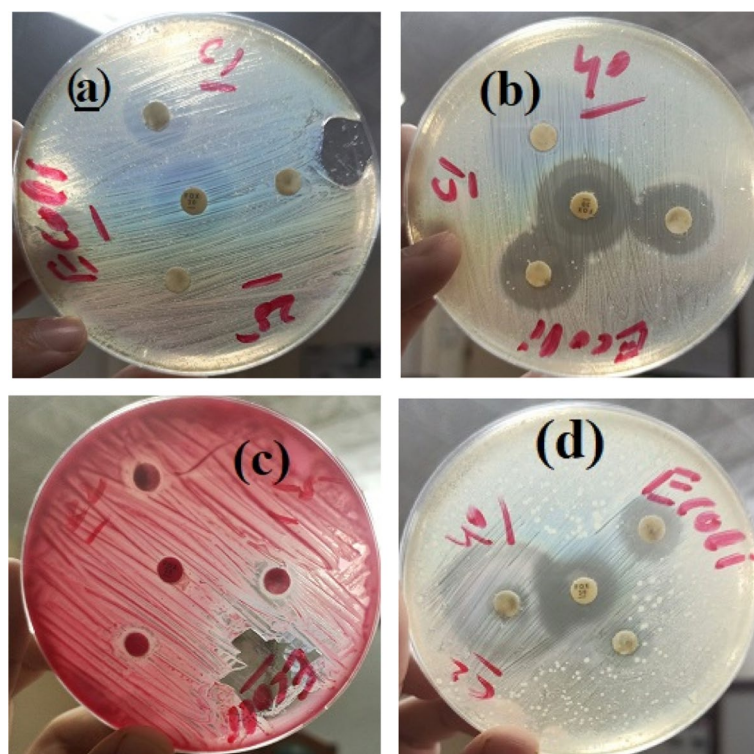


Figure 13. Antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against *E. coli* gram (-ve) bacteria.

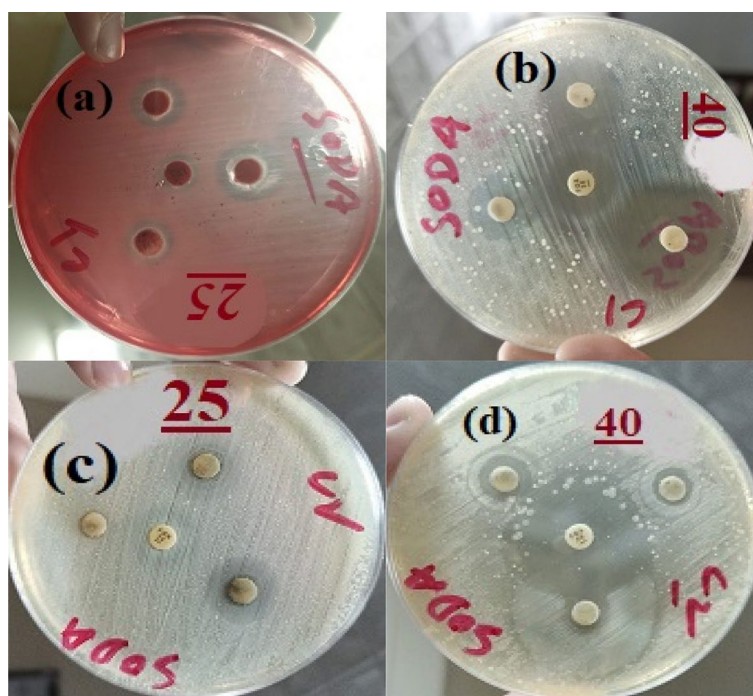


Figure 14. Antibacterial activity of (a) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ using dose 25 mg/ml (b) $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ with dose 40 mg/ml (c) $\text{Bi}_2\text{O}_3/\text{GO}$ with dose 25 mg/ml and (d) $\text{Bi}_2\text{O}_3/\text{GO}$ dose 40 mg/ml against pseudomonas gram (-ve) bacteria.

| Sr. no | Sample | Bacteria | Zone of Inhibition (mm) at dose 25 mg/ml | Zone of inhibition (mm) at dose 40 mg/ml | Zone of inhibition (mm) of control |
|--------|--|--------------------|--|--|------------------------------------|
| 1 | $\text{Bi}_2\text{O}_3/\text{GO}$ | <i>E. coli</i> | 11 | 17 | 18 |
| 2 | | <i>Pseudomonas</i> | 12 | 19 | > 18 |
| 3 | | <i>B. cereus</i> | 6.5 | 8 | 6 |
| 4 | | <i>S. aureus</i> | 4.5 | 5 | 7 |
| 5 | $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ | <i>E. coli</i> | 14 | 18 | 18 |
| 6 | | <i>Pseudomonas</i> | 13 | 21 | > 18 |
| 7 | | <i>B. cereus</i> | 7 | 8.5 | 5.5 |
| 8 | | <i>S. aureus</i> | 9 | 10.5 | 8.5 |

Table 4. Comparison of zone of inhibition against gram (+ve) and gram (-ve) bacteria.

by the institutional Research Ethical Committee; i.e., Pharmacy Animal Ethics committee (PAEC), under reference number PAEC/22/71.

Hematological test. Hematological parameters such as Hematocrit (HCT), hemoglobin (HGB), lymphocytes (LYM), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), mean corpuscular volume (MCV), mean platelet volume (MPV), clotting time (CT), platelet distribution width (PDW), platelet large cell ratio (PLCR), platelets (PLT), red blood cells (RBC), white blood cells (WBC), red cell distribution width—coefficient of variation (RDW-CV) and red cell distribution width—standard deviation (RDW-SD) were examined as shown in Fig. 15.

Two common parameters (WBC and RBC) in $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ treated mice were not different significantly from the control mice. Platelets (PLT) and PCT increased after 2 days and then gradually decreases during 30 days treatment. LYM was slightly increases after 2 days and then slightly decreases after 30 days. PLCR was decreases during 30 days of treatment. Hematological parameters such as HCT, HGB, MCH, MCHC, MCV, MPV, PDW, RDWS and RDWC did not change significantly which reveals minor biological impairment.

Biochemistry test. The basic biochemistry tests on mice treated with $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ at 2, 7, 14 and 30 days were performed shown in Fig. 16. Biochemistry test such as alanine transaminase (ALT), aspartate amino transferase (AST), albumin (ALB), blood urea nitrogen (BUN), creatinine (CREA), cholesterol, sugar, total bilirubin (TBIL), total protein, triglyceride and uric acid were evaluated. AST, ALT and CREA test were demonstrated

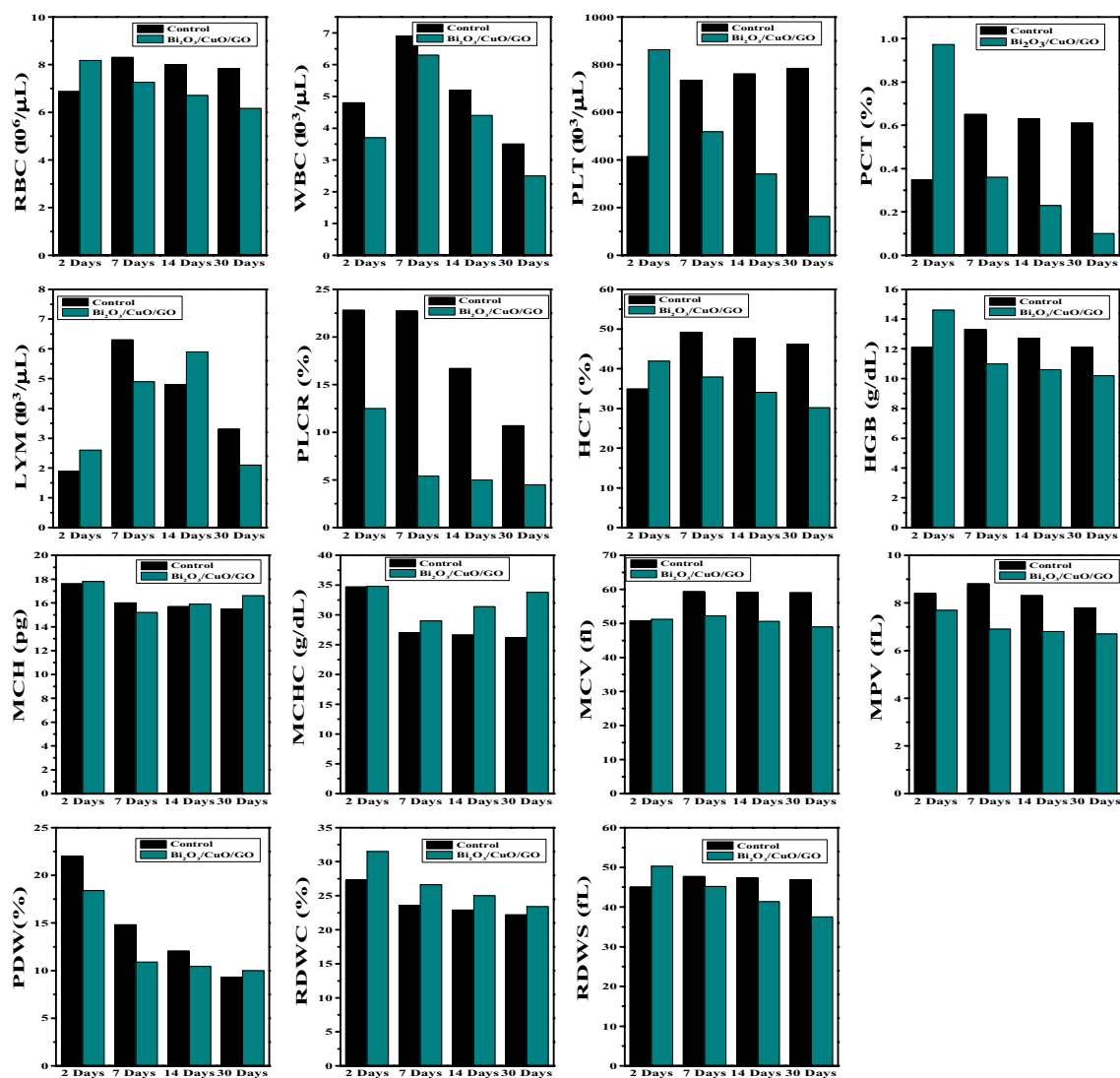


Figure 15. Hematology parameters such as RBC, WBC, PLT, PCT, LYM, PLCR, HCT, HGB, MCH, MCHC, MCV, MPV, PDW, RDWC and RDWS of mice treated with synthesized $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposite at time period of 2, 7, 14 and 30 days using 20 mg/kg dose.

as they are strongly linked to liver as well as kidney function of mice. After 2 days of treatment no significant changes were noticed in ALT and CREA but AST was decreased. After 2 days, NPs caused significant liver inflammation but did not trigger open wound. No significant change was noticed in ALT and AST after 7 days and 14 days of treatment but CREA was increased after 14 days.

ALT and AST have no significant change after 30 days as well as CREA turned to normal level, indicating that minor damage of liver caused by $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ was repaired after 30 days. This is accordance with removal and biodistribution of NPs. The accumulation of oxide-based and Au NPs in liver have been demonstrated to cause a steady increase in ALT and AST as well as substantial liver damage. Au NPs with 5 mg/kg dose level via intravenous administration cause severe injury^{79,80}. BUN was slightly increased after 30 days. ALB, cholesterol, sugar, TBIL, total protein, triglycerides and Uric acid were also investigated but no significant change was noticed after 30 days. Fe_3O_4 coated with Oleic acid and PEG at dose level 5–7.5 mg/kg caused consistent elevations in ALT, BUN and AST⁸¹. Even at dose level of 20 mg/kg, $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ demonstrates very slight liver damage. These toxicity results are comparable with Bi_2Se_3 NPs as reported in literature⁸².

Pathological tests results. We use immunohistochemistry to examine the pathogenic alterations in organs like liver, lungs and kidneys at time period of 2, 7, 14 and 30 days. We collected these organs and prepared the slides of tissues of these organs for microscopy. Throughout the whole treatment session, no damage was identified in kidneys shown in Fig. 17. There was a minor pathological change in liver and lungs was noticed. Very small dark spot was noticed in liver after 4 days but it returned to normal after 30 days. Similarly, a small dark spot was identified in lungs after 2 and 14 days but recovered in 30 days. Additional element of pathology is to analyze the removal of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ NPs qualitatively^{81,83}. Whenever NPs assemble in organs, they can accumulate and be recovered directly by using optical microscope. Mice treated with Fe_3O_4 NPs, GO and carbon nanotubes had comparable effects.

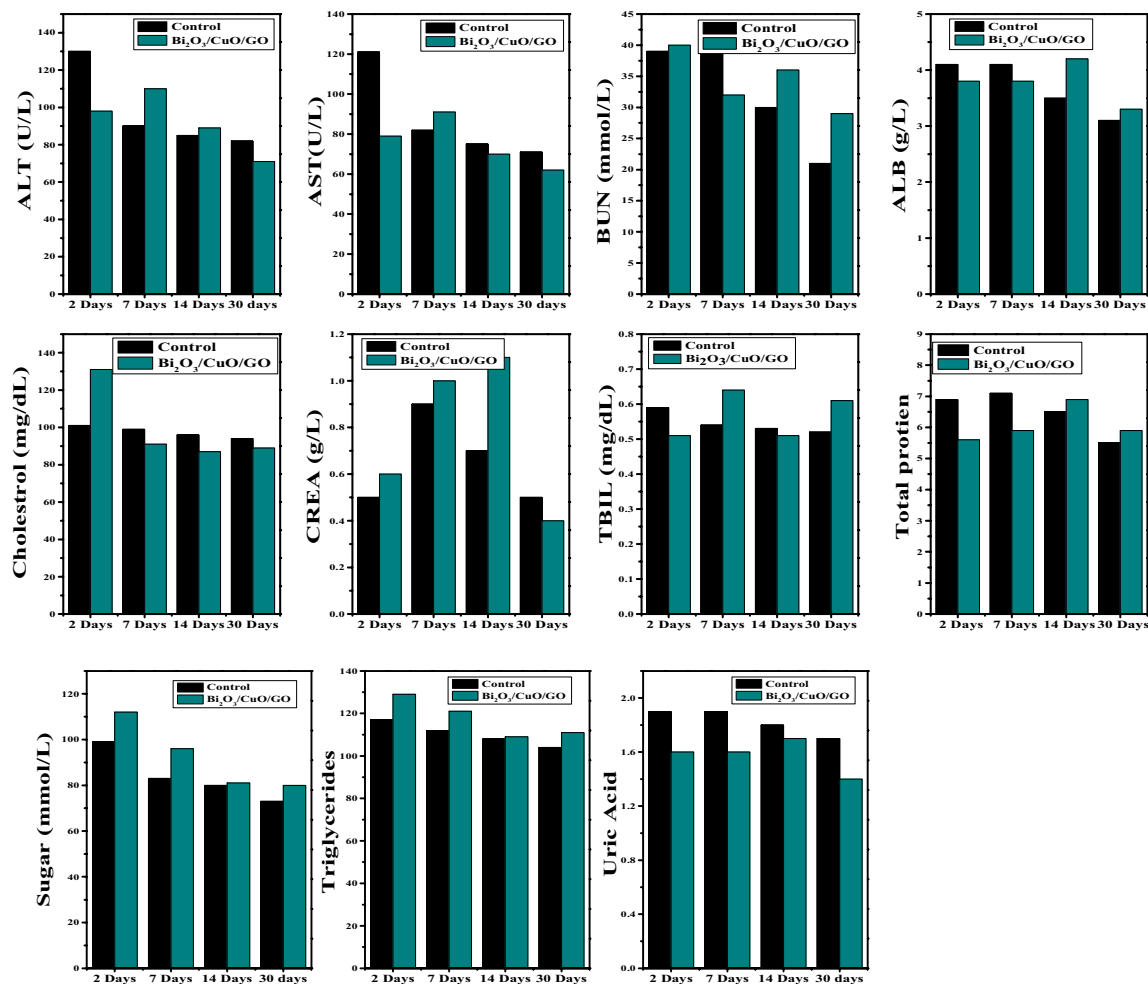


Figure 16. Biochemistry parameters such as ALT, AST, BUN, ALB, Cholesterol, CREA, TBIL, TP, Sugar, Triglyceride and Uric acid of mice treated with synthesized $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposite at time period of 2, 7, 14 and 30 days using 20 mg/kg dose.

The liver was found to have black spots, which vanished after roughly 90 days. Now days, Nanomedicine is looking for compounds which have low toxicity and show high clearance efficiency. NPs of small size are commonly believed to be removed through kidneys. But reality on the other hand is significantly more complicated than the assumptions. The clearing of Au NPs is a good example. Au NPs of size 3 nm protected with PEG were not cleared, but Au NPs of same size protected with glutathione were cleared efficiently^{79,84}. When the size of carbon materials was between 10 and 30 nm, the kidney and liver may progressively eliminate them⁸³. As a result, it is obvious that NPs clearance is influenced not just by size and morphology, but also by shape and durability. Another feasible path for the construction of metabolizable NPs is to investigate the removal of materials of large size. Our current study conclusively demonstrates that $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites may be absorbed by liver, indicating that they hold a great potential in medical uses including cancer treatment and contrast agents⁸².

Kefayat et al.⁸⁵ reported the toxicity of albumin stabilized GNPs on BALB/c mice which were injected intravenously with 10 mg/kg dose and sacrificed after 1 month. Through histopathological and biochemistry blood and biochemistry blood analysis as well as histopathological images of organs (liver, lungs, brain, spleen, heart and kidneys) it was observed that GNPs were safe and non-toxic⁸⁵. Toxicity studies of FA-AUNCs on rats with 10 mg/kg dosage injected via intra venous route for a time period of 3 days were reported by Kefayat et al.⁸⁶. By biochemistry analysis and histopathological assays of organs (kidneys, liver and spleen), they concluded that no severe damage was found⁸⁶. Ghahremani et al.⁸⁷, reported that APT-GNCs exhibit no toxic effect on BALB/c mice in a duration of 20 days when a dose of 8 mg/kg was injected intravenously to them.

Statistical analysis. Tables 5 and 6 shows the p value of hematological and biochemistry parameters respectively. GraphPad (2D scientific Graphing) software (version 8) was used to calculate p value. From p value it is very clear that synthesized nanoparticles have no significant effect on mice and our nanoparticles are safe. p Value shows non-significant results. Similarly, Table 7 shows acute toxicity study of mice at 0 h, 2 h, 6 h, 24 h and 48 h and there is no significant change occurs in behavior of mice.

Table 8 shows the in vivo toxicity comparison of synthesized $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites with other reported CuO and GO based composites.

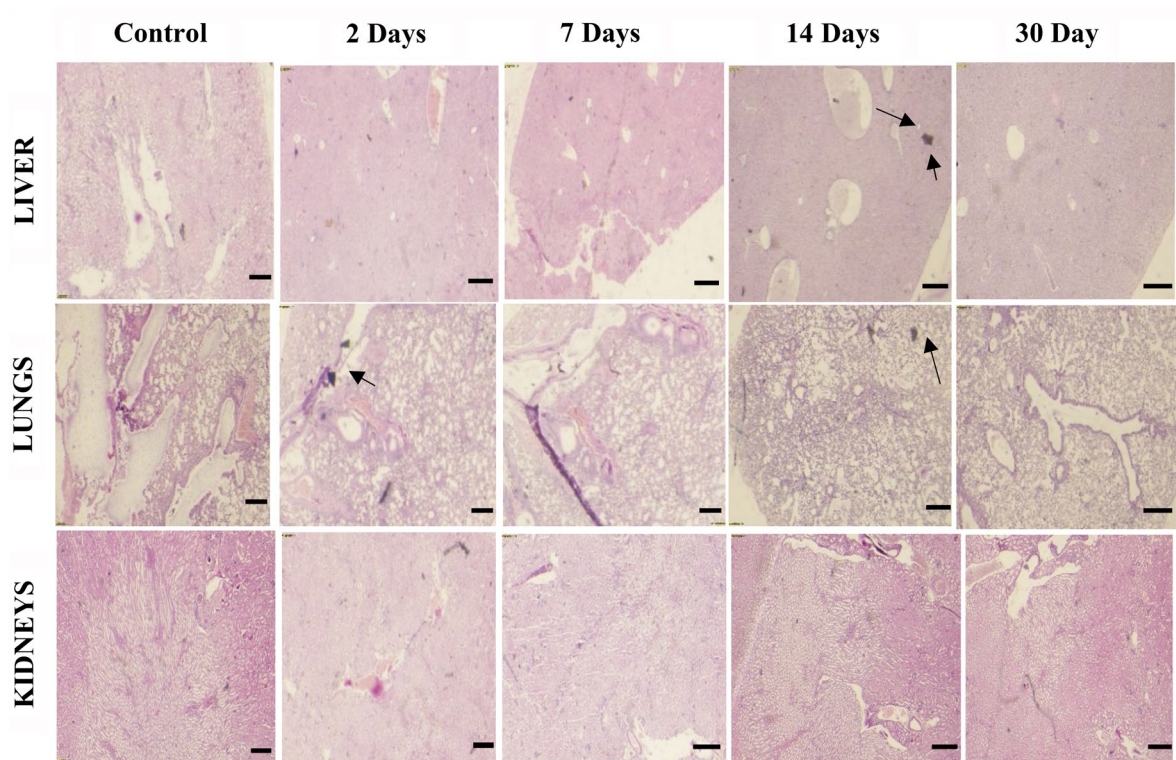


Figure 17. Pathological results of liver, kidney and lungs of mice treated with synthesized $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ NPs of dose 20 mg/kg at different time period of 2, 7, 14 and 30 days. Dimensions: 5625.6 μm (W) \times 3516 μm (H), scale bar: 87 μm .

| Parameter | Control group (Mean \pm SEM) | Treatment group ($\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposite) (Mean \pm SEM) | <i>p</i> value |
|--|--------------------------------|--|----------------|
| Hematocrit (HCT) | 44.50 \pm 6.51 | 36.01 \pm 5.02 | 0.085 |
| Hemoglobin (HGB) | 12.55 \pm 0.29 | 11.55 \pm 0.96 | 0.358 |
| Lymphocytes (LYM) | 4.07 \pm 0.95 | 3.87 \pm 0.91 | 0.884 |
| Mean corpuscular hemoglobin (MCH) | 16.20 \pm 0.48 | 16.63 \pm 0.78 | 0.658 |
| Mean corpuscular hemoglobin concentration (MCHC) | 28.63 \pm 2.03 | 32.25 \pm 1.29 | 0.183 |
| Mean corpuscular volume (MCV) | 57.06 \pm 2.08 | 50.75 \pm 0.67 | 0.028* |
| Mean platelet volume (MPV) | 8.32 \pm 0.20 | 7.02 \pm 0.22 | 0.005** |
| Clotting time (CT) | 0.56 \pm 0.07 | 0.42 \pm 0.19 | 0.511 |
| Platelet distribution width (PDW) | 14.54 \pm 2.72 | 12.44 \pm 1.99 | 0.557 |
| Platelet large cell ratio (PLCR) | 18.23 \pm 2.88 | 6.86 \pm 1.89 | 0.016* |
| Platelets (PLT) | 673.80 \pm 87.18 | 471.50 \pm 149.40 | 0.286 |
| Red blood cells (RBC) | 7.75 \pm 0.31 | 7.08 \pm 0.43 | 0.251 |
| White blood cells (WBC) | 5.10 \pm 0.70 | 4.22 \pm 0.79 | 0.440 |
| Red cell distribution width- coefficient of variation (RDW-CV) | 24.00 \pm 1.13 | 26.63 \pm 1.75 | 0.258 |
| Red cell distribution width—standard deviation (RDW-SD) | 46.73 \pm 0.60 | 43.59 \pm 2.73 | 0.305 |

Table 5. Hematological parameters of mice treated with synthesized $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposite. The values are expressed as Mean \pm SEM and statistically analyzed using t-test. The results of the treatment group are compared with those of the control group and considered non-significant if $p > 0.05$, significant if $p < 0.05$ and more significant if $p < 0.01$ ⁸⁵.

| Parameter | Control group (Mean ± SEM) | Treatment group (Bi ₂ O ₃ /CuO/GO nanocomposite) (Mean ± SEM) | p value |
|----------------------------------|----------------------------|---|---------|
| Alanine transaminase (ALT) | 96.75 ± 11.21 | 92.00 ± 8.21 | 0.744 |
| Aspartate aminotransferase (AST) | 87.25 ± 11.48 | 75.50 ± 6.22 | 0.403 |
| Albumin (ALB) | 3.70 ± 0.24 | 3.77 ± 0.18 | 0.815 |
| Blood urea nitrogen (BUN) | 32.25 ± 4.30 | 34.25 ± 2.39 | 0.699 |
| Creatinine (CREA) | 0.65 ± 0.09 | 0.77 ± 0.16 | 0.537 |
| Cholesterol | 97.50 ± 1.55 | 99.50 ± 10.53 | 0.857 |
| Sugar | 83.75 ± 5.49 | 92.25 ± 7.53 | 0.397 |
| Total bilirubin (TBIL) | 0.56 ± 0.02 | 0.57 ± 0.03 | 0.956 |
| Total protein | 6.50 ± 0.35 | 6.07 ± 0.28 | 0.386 |
| Triglyceride | 110.30 ± 2.78 | 117.50 ± 4.64 | 0.229 |
| Uric acid | 1.82 ± 0.05 | 1.57 ± 0.63 | 0.019 |

Table 6. Biochemical parameters of mice treated with synthesized Bi₂O₃/CuO/GO nanocomposite. The values are expressed as Mean ± SEM and statistically analyzed using t-test. The results of the treatment group are compared with those of the control group and considered non-significant if $p > 0$.

| Parameters | 0 h | 2 h | 6 h | 24 h | 48 h |
|-------------------|-----|-----|-----|------|------|
| Alertness | ✓ | ✓ | ✓ | ✓ | ✓ |
| Grooming | ✓ | ✓ | ✓ | ✓ | ✓ |
| Convulsions | × | × | × | × | × |
| Hyperactivity | × | × | × | × | ✓ |
| Lacrimation | × | × | × | × | × |
| Salivation | × | × | × | × | × |
| Urination | × | × | × | × | × |
| Touch Response | ✓ | ✓ | ✓ | ✓ | ✓ |
| Pain response | ✓ | ✓ | ✓ | ✓ | ✓ |
| Writhing reflex | × | × | × | × | × |
| Corneal reflex | ✓ | ✓ | ✓ | ✓ | |
| Gripping strength | ✓ | ✓ | ✓ | ✓ | ✓ |
| Righting reflex | ✓ | ✓ | ✓ | ✓ | ✓ |
| Skin color | × | × | × | × | × |

Table 7. The behavioral pattern of mice treated with synthesized Bi₂O₃/CuO/GO nanocomposite during acute toxicity study. (✓) = Present. (×) = Not present/no change.

| Type | Materials | Animal | Mechanism of exposure | Findings | Refs |
|---------|--|-------------------|---|--|---------|
| In vivo | Bi ₂ O ₃ /CuO/GO | Swiss albino mice | 20 mg/kg dose was administered orally for 30 days | Pathology shows small black spots in liver and lungs which disappear after 30 days. According to Hematological and biochemistry results there is no significant damage is found and particles are not toxic | Present |
| In vivo | rGO/Ag NC | Mice | 10 mg/kg dose was injected intraperitoneally for 7 days | According to findings, ALT, AST and creatinine increased implying a negative impact of rGO/Ag nanocomposite on liver and kidneys. Which confirms the toxic effect of green synthesized rGO/AgNC | 88 |
| In vivo | Cu NPs | Male wister rats | 50, 100 and 200 mg/kg dose administered orally for 5 days | Pathological results show that toxicity was induced in both liver and kidneys. In liver, necrosis of tissues and in kidney necrosis in proximal renal tubule as well a swelling of proximal tubule was observed | 89 |
| In vivo | CuO NPs | Male wister rats | 10, 100 and 300 mg/kg dose delivered through IP injection for 14 days | Toxicity was induced in lungs and liver with all concentration of CuO NPs. In liver, vasculature in central veins, portal triad vessels and loss of hexagonal lobules was observed. And in lungs thickening of air scars can be seen | 90 |

Table 8. Toxicity comparison of our synthesized nanocomposites with CuO ad GO based nanocomposites.

Conclusions

Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites were successfully synthesized via green method, sol–gel technique and co-precipitation method respectively. The results of characterization techniques such as X-ray diffraction, Fourier transform infrared spectroscopy, scanning electron microscopy and UV–vis spectroscopy revealed that nanocomposites were successfully synthesized. The crystal size of Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanomaterials was 5.1 nm, 13.9 nm and 11.5 nm respectively by Scherrer formula. XRD pattern confirms the monoclinic structure of all synthesized nanomaterials. Antibacterial activity demonstrates that the inhibition zone for Bi_2O_3 , $\text{Bi}_2\text{O}_3/\text{GO}$ and $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposites against gram – ve *E. coli* is 12 mm, 17 mm and 18 mm respectively and against pseudomonas is 28 mm, 19 mm and 21 mm respectively. In vivo toxicity on *Swiss albino* mice was investigated at dose of 20 mg/kg of $\text{Bi}_2\text{O}_3/\text{CuO}/\text{GO}$ nanocomposite at different time period of 2, 7, 14 and 30 days. Hematological, biochemistry and pathological results revealed that nanocomposites are less toxic and after 30 days of treatment the slight effects on liver were recovered.

Data availability

All relevant data are included in the article.

Received: 28 February 2022; Accepted: 25 July 2022

Published online: 22 August 2022

References

- Mohan, V. B. *et al.* Characterisation of reduced graphene oxide: Effects of reduction variables on electrical conductivity. *Mater. Sci. Eng. B* **193**, 49–60 (2015).
- Kumari, R. M., Goswami, R. & Nimesh, S. Application of nanotechnology in diagnosis and therapeutics. In *Nanotechnology for Energy and Environmental Engineering* (eds Ledwani, L. & Sangwai, J.) 413–440 (Springer, 2020).
- Asghar, M. A. & Asghar, M. A. Green synthesized and characterized copper nanoparticles using various new plants extracts aggravate microbial cell membrane damage after interaction with lipopolysaccharide. *Int. J. Biol. Macromol.* **160**, 1168–1176 (2020).
- Barreto, J. A. *et al.* Nanomaterials: Applications in cancer imaging and therapy. *Adv. Mater.* **23**(12), H18–H40 (2011).
- Yang, K. *et al.* Graphene in mice: Ultrahigh in vivo tumor uptake and efficient photothermal therapy. *Nano Lett.* **10**(9), 3318–3323 (2010).
- Park, J.-H. *et al.* Biodegradable luminescent porous silicon nanoparticles for in vivo applications. *Nat. Mater.* **8**(4), 331–336 (2009).
- Schipper, M. L. *et al.* A pilot toxicology study of single-walled carbon nanotubes in a small sample of mice. *Nat. Nanotechnol.* **3**(4), 216–221 (2008).
- Rosenthal, S. J. *et al.* Biocompatible quantum dots for biological applications. *Chem. Biol.* **18**(1), 10–24 (2011).
- Cho, W. S. *et al.* Acute toxicity and pharmacokinetics of 13 nm-sized PEG-coated gold nanoparticles. *Toxicol. Appl. Pharmacol.* **236**(1), 16–24 (2009).
- Choi, C. H. J. *et al.* Targeting kidney mesangium by nanoparticles of defined size. *Proc. Natl. Acad. Sci.* **108**(16), 6656–6661 (2011).
- Cho, W. S. *et al.* Size-dependent tissue kinetics of PEG-coated gold nanoparticles. *Toxicol. Appl. Pharmacol.* **245**(1), 116–123 (2010).
- Walkey, C. D. *et al.* Nanoparticle size and surface chemistry determine serum protein adsorption and macrophage uptake. *J. Am. Chem. Soc.* **134**(4), 2139–2147 (2012).
- Kairdolf, B. A. *et al.* Semiconductor quantum dots for bioimaging and biodiagnostic applications. *Annu. Rev. Anal. Chem. (Palo Alto Calif.)* **6**(1), 143–162 (2013).
- Yang, R. S. *et al.* Persistent tissue kinetics and redistribution of nanoparticles, quantum dot 705, in mice: ICP-MS quantitative assessment. *Environ. Health Perspect.* **115**(9), 1339–1343 (2007).
- Rajeshwar, K., de Tacconi, N. R. & Chenthamarakshan, C. Semiconductor-based composite materials: preparation, properties, and performance. *Chem. Mater.* **13**(9), 2765–2782 (2001).
- Camargo, P. H. C., Satyanarayana, K. G. & Wypych, F. Nanocomposites: Synthesis, structure, properties and new application opportunities. *Mater. Res.* **12**(1), 1–39 (2009).
- Park, M. *et al.* Effective formation of WO_3 nanoparticle/ Bi_2S_3 nanowire composite for improved photoelectrochemical performance. *J. Phys. Chem. C* **122**(31), 17676–17685 (2018).
- Salazar-Pérez, A. *et al.* Structural evolution of Bi_2O_3 prepared by thermal oxidation of bismuth nano-particles. *Superficies y Vacío* **18**(3), 4–8 (2005).
- Kumaraguru, S. *et al.* Enhanced texture and microhardness of the nickel surface using Bi_2O_3 particles via electrodeposition technique for engineering application. *J. Alloys Compd.* **753**, 740–747 (2018).
- Hainfeld, J. F., Slatkin, D. N. & Smilowitz, H. M. The use of gold nanoparticles to enhance radiotherapy in mice. *Phys. Med. Biol.* **49**(18), N309 (2004).
- Rahman, W. N. *et al.* Enhancement of radiation effects by gold nanoparticles for superficial radiation therapy. *Nanomedicine* **5**(2), 136–142 (2009).
- Chithrani, D. B. *et al.* Gold nanoparticles as radiation sensitizers in cancer therapy. *Radiat. Res.* **173**(6), 719–728 (2010).
- Li, Y.-J. *et al.* Gold nanoparticles as a platform for creating a multivalent poly-SUMO chain inhibitor that also augments ionizing radiation. *Proc. Natl. Acad. Sci. U.S.A.* **109**(11), 4092–4097 (2012).
- Zhang, X.-D. *et al.* Size-dependent radiosensitization of PEG-coated gold nanoparticles for cancer radiation therapy. *Biomaterials* **33**(27), 6408–6419 (2012).
- Alam, S. N., Sharma, N. & Kumar, L. Synthesis of graphene oxide (GO) by Modified hummers method and its thermal reduction to obtain reduced graphene oxide (rGO). *Graphene J. Stats* **6**(1), 1–18 (2017).
- Su, Y. *et al.* Self-limiting growth of two-dimensional palladium between graphene oxide layers. *Nano Lett.* **19**(7), 4678–4683 (2019).
- Ahmed, M. K., Mansour, S. F. & Al-Wafi, R. Nanofibrous scaffolds of ϵ -polycaprolactone containing Sr/Se-hydroxyapatite/graphene oxide for tissue engineering applications. *Biomed. Mater.* **16**(4), 045030 (2021).
- Ahmed, M. K. *et al.* Composition and design of nanofibrous scaffolds of Mg/Se-hydroxyapatite/graphene oxide @ ϵ -polycaprolactone for wound healing applications. *J. Mater. Res. Technol.* **9**(4), 7472–7485 (2020).
- Menazea, A. A. & Ahmed, M. K. Wound healing activity of chitosan/polyvinyl alcohol embedded by gold nanoparticles prepared by nanosecond laser ablation. *J. Mol. Struct.* **1217**, 128401 (2020).
- Al-Wafi, R., Mansour, S. F. & Ahmed, M. K. Mechanical, microstructural properties and cell adhesion of Sr/Se-hydroxyapatite/graphene/polycaprolactone nanofibers. *J. Thermoplast. Compos. Mater.* **34**(4), 536–556 (2020).
- Duan, F. *et al.* Heterogeneous Fenton-like degradation of 4-chlorophenol using iron/ordered mesoporous carbon catalyst. *J. Environ. Sci.* **26**(5), 1171–1179 (2014).
- Liang, F.-X. *et al.* Recent advances in the fabrication of graphene–ZnO heterojunctions for optoelectronic device applications. *J. Mater. Chem. C* **6**(15), 3815–3833 (2018).

33. Das, T. R. *et al.* Bismuth oxide decorated graphene oxide nanocomposites synthesized via sonochemical assisted hydrothermal method for adsorption of cationic organic dyes. *J. Colloid Interface Sci.* **509**, 82–93 (2018).
34. Manavalan, S. *et al.* Sonochemical synthesis of bismuth(III) oxide decorated reduced graphene oxide nanocomposite for detection of hormone (epinephrine) in human and rat serum. *Ultrason. Sonochem.* **51**, 103–110 (2019).
35. Borkow, G. & Gabbay, J. Copper as a biocidal tool. *Curr. Med. Chem.* **12**(18), 2163–2175 (2005).
36. Wilks, S. A., Michels, H. & Keevil, C. W. The survival of *Escherichia coli* O157 on a range of metal surfaces. *Int. J. Food Microbiol.* **105**(3), 445–454 (2005).
37. Noyce, J. O., Michels, H. & Keevil, C. W. Potential use of copper surfaces to reduce survival of epidemic meticillin-resistant *Staphylococcus aureus* in the healthcare environment. *J. Hosp. Infect.* **63**(3), 289–297 (2006).
38. Yin, M. *et al.* Copper oxide nanocrystals. *J. Am. Chem. Soc.* **127**(26), 9506–9511 (2005).
39. Nasrollahzadeh, M., Sajadi, S. M. & Maham, M. *Tamarix gallica* leaf extract mediated novel route for green synthesis of CuO nanoparticles and their application for N-arylation of nitrogen-containing heterocycles under ligand-free conditions. *RSC Adv.* **5**(51), 40628–40635 (2015).
40. Verma, N. & Kumar, N. Synthesis and biomedical applications of copper oxide nanoparticles: An expanding horizon. *ACS Biomater. Sci. Eng.* **5**(3), 1170–1188 (2019).
41. Yoosefi Booshehri, A., Wang, R. & Xu, R. Simple method of deposition of CuO nanoparticles on a cellulose paper and its antibacterial activity. *Chem. Eng. J.* **262**, 999–1008 (2015).
42. Sankar, R. *et al.* Anticancer activity of *Ficus religiosa* engineered copper oxide nanoparticles. *Mater. Sci. Eng. C Mater. Biol. Appl.* **44**, 234–239 (2014).
43. Kumar, K. Y., Muralidhara, H. B. & Nayaka, Y. A. Magnificent adsorption capacity of hierarchical mesoporous copper oxide nanoflakes towards mercury and cadmium ions: Determination of analyte concentration by DPASV. *Powder Technol.* **258**, 11–19 (2014).
44. Azam, A. *et al.* Antimicrobial activity of metal oxide nanoparticles against Gram-positive and Gram-negative bacteria: a comparative study. *Int. J. Nanomed.* **7**, 6003–6009 (2012).
45. Qian, Y. *et al.* Synthesis of cuprous oxide (Cu₂O) nanoparticles/graphene composite with an excellent electrocatalytic activity towards glucose. *Int. J. Electrochem. Sci.* **7**, 10063–10073 (2012).
46. Nurzulaikha, R. *et al.* Graphene/SnO₂ nanocomposite-modified electrode for electrochemical detection of dopamine. *Sens. Bio-Sens. Res.* **5**, 42–49 (2015).
47. Bahrani, S. *et al.* Ultrasound-accelerated synthesis of gold nanoparticles modified choline chloride functionalized graphene oxide as a novel sensitive bioelectrochemical sensor: Optimized meloxicam detection using CCD-RSM design and application for human plasma sample. *Ultrason. Sonochem.* **42**, 776–786 (2018).
48. Patil, S. P. *et al.* Efficient adsorption and photocatalytic degradation of Rhodamine B dye over Bi₂O₃-bentonite nanocomposites: A kinetic study. *J. Ind. Eng. Chem.* **34**, 356–363 (2016).
49. Zhu, J. *et al.* Decorating graphene oxide with CuO nanoparticles in a water–isopropanol system. *Nanoscale* **2**(6), 988–994 (2010).
50. Zahid, M. *et al.* Enhanced structural and dielectric properties of calcium and chromium based M-type hexagonal ferrites. *J. Mater. Sci. Mater. Electron.* **32**, 9183–9193 (2021).
51. Ohira, T. & Yamamoto, O. Correlation between antibacterial activity and crystallite size on ceramics. *Chem. Eng. Sci.* **68**(1), 355–361 (2012).
52. Yasmeen, S. *et al.* Synthesis, structural and optical analysis of surfactant assisted ZnO–NiO nanocomposites prepared by homogeneous precipitation method. *Ceram. Int.* **45**, 17859–17873 (2019).
53. Munawar, T. *et al.* Multi metal oxide NiO–CdO–ZnO nanocomposite–synthesis, structural, optical, electrical properties and enhanced sunlight driven photocatalytic activity. *Ceram. Int.* **46**(2), 2421–2437 (2020).
54. Zaaba, N. I. *et al.* Synthesis of graphene oxide using modified hummers method: Solvent influence. *Procedia Eng.* **184**, 469–477 (2017).
55. Chen, X. & Chen, B. Macroscopic and spectroscopic investigations of the adsorption of nitroaromatic compounds on graphene oxide, reduced graphene oxide, and graphene nanosheets. *Environ. Sci. Technol.* **49**(10), 6181–6189 (2015).
56. Novoselov, K. S. *et al.* Electronic properties of grapheme. *Physica Status Solidi (b)* **244**, 4106–4111 (2007).
57. Mekassa, B. *et al.* Synthesis, characterization, and preparation of nickel nanoparticles decorated electrochemically reduced graphene oxide modified electrode for electrochemical sensing of diclofenac. *J. Solid State Electrochem.* **22**, 3607–3619 (2018).
58. Mallahi, M. *et al.* Synthesis and characterization of bismuth oxide nanoparticles via sol–gel method. *AJER* **3**(4), 162–165 (2014).
59. Archana, S. *et al.* Development of multipurpose CuO–GO nanocomposites for heavy metals adsorption and super capacitor applications. *Energy Environ. Focus* **5**(4), 305–315 (2016).
60. Padil, V. V. T. & Černík, M. Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application. *Int. J. Nanomed.* **8**, 889 (2013).
61. Syame, S. M. *et al.* Synthesis of copper–chitosan nanocomposites and their applications in treatment of local pathogenic isolates bacteria. *Orient. J. Chem.* **33**(6), 2959–2969 (2017).
62. Bhargava, R. & Khan, S. Structural, optical and dielectric properties of graphene oxide. In *AIP Conference Proceedings* (AIP Publishing LLC, 2018).
63. Leontie, L. *et al.* Structural and optical characteristics of bismuth oxide thin films. *Surf. Sci.* **507**, 480–485 (2002).
64. Sagadevan, S., Pal, K. & Chowdhury, Z. Z. Fabrication of CuO nanoparticles for structural, optical and dielectric analysis using chemical precipitation method. *J. Mater. Sci. Mater. Electron.* **28**(17), 12591–12597 (2017).
65. Cao, W. *Semiconductor Photocatalysis: Materials, Mechanisms and Applications* (IntechOpen, 2016).
66. Mehmood, Z. *et al.* A facile approach to synthesis graphene oxide/bismuth oxide nanocomposites and their superior sunlight driven photocatalytic activity. *Optik* **197**, 163035 (2019).
67. Wang, H.-W. *et al.* Facile solvothermal synthesis of a graphene nanosheet–bismuth oxide composite and its electrochemical characteristics. *Electrochim. Acta* **55**(28), 8974–8980 (2010).
68. Banu, R., *et al.* *Visible Light Driven Photocatalytic Degradation of Brilliant Green Dye Using Graphene Oxide/Copper Oxide Binary Composite*. 2019.
69. Menazea, A. & Ahmed, M. Synthesis and antibacterial activity of graphene oxide decorated by silver and copper oxide nanoparticles. *J. Mol. Struct.* **1218**, 128536 (2020).
70. Muninathan, S. & Sivasamy, A. Bismuth oxide nanoparticles decorated Graphene layers for the degradation of Methylene blue dye under visible light irradiations and antimicrobial activities. *J. Environ. Chem. Eng.* **6**, 3745–3756 (2017).
71. Menazea, A. & Ahmed, M. K. Silver and copper oxide nanoparticles-decorated graphene oxide via pulsed laser ablation technique: Preparation, characterization, and photoactivated antibacterial activity. *Nano-Struct. Nano-Obj.* **22**, 100464 (2020).
72. Jayaramudu, T. *et al.* Chitosan capped copper oxide/copper nanoparticles encapsulated microbial resistant nanocomposite films. *Int. J. Biol. Macromol.* **128**, 499–508 (2019).
73. Khan, A. U. *et al.* A facile fabrication of silver/copper oxide nanocomposite: An innovative entry in photocatalytic and biomedical materials. *Photodiagn. Photodyn. Ther.* **31**, 101814 (2020).
74. Li, Y., Yang, D. & Cui, J. Graphene oxide loaded with copper oxide nanoparticles as an antibacterial agent against *Pseudomonas syringae* pv. tomato. *RSC Adv.* **7**(62), 38853–38860 (2017).

75. Eskandarinia, A. *et al.* A novel bilayer wound dressing composed of a dense polyurethane/propolis membrane and a biodegradable polycaprolactone/gelatin nanofibrous scaffold. *Sci. Rep.* **10**(1), 3063 (2020).
76. Eskandarinia, A. *et al.* A propolis enriched polyurethane-hyaluronic acid nanofibrous wound dressing with remarkable antibacterial and wound healing activities. *Int. J. Biol. Macromol.* **149**, 467–476 (2020).
77. Khodabakhshi, D. *et al.* In vitro and in vivo performance of a propolis-coated polyurethane wound dressing with high porosity and antibacterial efficacy. *Colloids Surf. B Biointerfaces* **178**, 177–184 (2019).
78. Eskandarinia, A. *et al.* Cornstarch-based wound dressing incorporated with hyaluronic acid and propolis: In vitro and in vivo studies. *Carbohydr. Polym.* **216**, 25–35 (2019).
79. Zhang, X.-D. *et al.* In vivo renal clearance, biodistribution, toxicity of gold nanoclusters. *Biomaterials* **33**(18), 4628–4638 (2012).
80. Zhang, X. D. *et al.* Size-dependent in vivo toxicity of PEG-coated gold nanoparticles. *Int. J. Nanomed.* **6**, 2071–2081 (2011).
81. Gu, L. *et al.* In vivo clearance and toxicity of monodisperse iron oxide nanocrystals. *ACS Nano* **6**(6), 4947–4954 (2012).
82. Zhang, X. D. *et al.* Metabolizable Bi₂Se₃ nanoplates: Biodistribution, toxicity, and uses for cancer radiation therapy and imaging. *Adv. Funct. Mater.* **24**(12), 1718–1729 (2014).
83. Yang, K. *et al.* In vivo pharmacokinetics, long-term biodistribution, and toxicology of PEGylated graphene in mice. *ACS Nano* **5**(1), 516–522 (2011).
84. Zhou, C. *et al.* Near-infrared emitting radioactive gold nanoparticles with molecular pharmacokinetics. *Angew. Chem. Int. Ed. Engl.* **51**(40), 10118–10122 (2012).
85. Kefayat, A. *et al.* Investigation of different targeting decorations effect on the radiosensitizing efficacy of albumin-stabilized gold nanoparticles for breast cancer radiation therapy. *Eur. J. Pharm. Sci.* **130**, 225–233 (2019).
86. Kefayat, A. *et al.* Ultra-small but ultra-effective: Folic acid-targeted gold nanoclusters for enhancement of intracranial glioma tumors' radiation therapy efficacy. *Nanomed. Nanotechnol. Biol. Med.* **16**, 173–184 (2019).
87. Ghahremani, F. *et al.* AS1411 aptamer-targeted gold nanoclusters effect on the enhancement of radiation therapy efficacy in breast tumor-bearing mice. *Nanomedicine (Lond)* **13**(20), 2563–2578 (2018).
88. El-Zahed, M. M. *et al.* In vivo toxicity and antitumor activity of newly green synthesized reduced graphene oxide/silver nanocomposites. *Bioresour. Bioprocess.* **8**(1), 44 (2021).
89. Lei, R. *et al.* Integrated metabolomic analysis of the nano-sized copper particle-induced hepatotoxicity and nephrotoxicity in rats: A rapid in vivo screening method for nanotoxicity. *Toxicol. Appl. Pharmacol.* **232**(2), 292–301 (2008).
90. Doudi, M. & Setorki, M. Acute effect of nano-copper on liver tissue and function in rat. *Nanomed. J.* **1**, 331–338 (2014).

Acknowledgements

The researchers would like to thank the Deanship of Scientific Research, Qassim University for funding the publication of this project.

Author contributions

A.Q.: Methodology, laboratory experiments, analysis, writing, Z.B.: Conceptualization, supervision, writing—review and editing, M.F.: Validation, revision, statistical analysis, EDX measurements S.A.B.: Validation, visualization, H.U.: Fruitful discussion on pathological test and analysis, A.N.: Writing—review and editing, visualization, Q.J.: Supervising the laboratory experiments for immunotoxicological studies on mice, S.S.: Writing—review and editing, R.I.: review and editing.

Competing interests

The authors declare no competing interests

Additional information

Correspondence and requests for materials should be addressed to Z.B. or M.F.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022