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## Microbial production of toluene in oxygen minimum zone waters in the Humboldt Current System off Chile

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Expansion of oxygen minimum zones in the world's oceans is likely to enhance the production of anaerobic metabolites by marine microorganisms. Here we show that toluene is present throughout the year in shelf waters of the upwelling ecosystem off Concepción (36° S), Chile, and it is a product of microbial anaerobic metabolism. The intra-annual variability in toluene concentrations is consistent with seasonal variability in the strengths of suboxic equatorial and oxygenated subantarctic water masses. Laboratory incubations of oxygen minimum zone water showed microbial production of toluene in the absence of O<sub>2</sub>. Toluene concentrations were elevated (up to 96 nM) in deeper O<sub>2</sub>-depleted waters and followed a seasonal pattern in oceanographic conditions. There is evidence to hypothesize that microbial production of toluene could be a homeostatic biochemical mechanism to thrive in the more acidic oxygen minimum zone waters. On the other hand, evidence indicates that microbial anaerobic degradation of toluene may be a source of NO<sub>2</sub><sup>-</sup> by partial denitrification, as shown for aquifer sediments. Since toluene production was not detected in incubations under aerobic conditions, we hypothesize that oxygen minimum zone waters export toluene to surrounding oxygenated waters. Expansion of hypoxia in the ocean will certainly enhance the production and export of anaerobic metabolites by marine microorganisms.

Across the world's oceans, anaerobic metabolism presently occurs in marine sediments, anoxic basins, and areas known as oxygen minimum zones (OMZ). These OMZ regions are open ocean waters<sup>1</sup>, occupying approximately 1% of the global ocean volume, but with a disproportionate influence on biogeochemical cycles through the removal of nitrogen and greenhouse gases from the ocean to the atmosphere<sup>2</sup>. Documented expansion of OMZ regions across the world's oceans<sup>3</sup> will expand anaerobic microbial metabolism in the water column and the associated production of reduced metabolites<sup>4–8</sup> readily available for further microbial degradation.

Toluene is an aromatic hydrocarbon that occurs as a minor component of gasoline and is also produced by bacteria<sup>9,10</sup> through anaerobic decarboxylation of phenylacetate derived from the catabolism of the amino acid phenylalanine<sup>10,11</sup>. Biological production of toluene has been reported in anaerobic bacterial isolates<sup>12</sup>, anaerobic sewage sludge<sup>13,14</sup>, anoxic freshwater sediments<sup>15,16</sup>, and the anoxic hypolimnion of stratified lakes<sup>17</sup>. Marine phytoplankton is also a potential source of toluene, as shown in monocultures of coccolithophores and diatoms under oxidative stress. They could be a potential source of benzenoids to the atmosphere<sup>18</sup>.

In oceanic OMZ waters, biogenic production of toluene is promoted both by low-O<sub>2</sub> conditions and by the high availability of proteinaceous organic matter (phenylalanine). As a permanent feature of the modern ocean, these OMZ waters are maintained by elevated fluxes of plankton-derived material from the photic zone, mainly in the form of protein carbon that in turn promotes oxygen depletion<sup>19</sup>. The biological production of toluene from phenylalanine has been shown to involve transamination, decarboxylation, and oxidation to form phenylacetate<sup>10</sup>, which is the substrate for a glycyl radical decarboxylase enzyme that catalyzes phenylacetate decarboxylation to toluene<sup>9,10,20</sup>, a process that is irreversibly inactivated in the presence of O<sub>2</sub><sup>10,19,21</sup>. Genomic evidence indicates that glycyl radical enzymes are widespread amongst facultative and obligate anaerobic microorganisms<sup>9,22–25</sup>.

Toluene is produced in bulk for industrial applications and consumer products and is found in concentrations of 2–4 mg/L in sewage sludge<sup>13,23</sup>. Toluene is a molecule of interest as a health hazard to humans<sup>24</sup>, and can

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account for a significant fraction of volatiles in the environment (compounds of high vapor pressure and low water solubility, with boiling points ranging between 69 and 317 °C)<sup>25</sup>. Toluene has been quantified in Narraganset Bay waters on the east coast of the US (0.2–60 nM)<sup>26</sup>, in the vicinity of a production platform in the Northwest Gulf of Mexico (21 nM)<sup>27</sup>, and Kuwait Bay (0.4–37 nM)<sup>28</sup>. These concentrations of toluene were attributed to the result of human activity and are higher than those reported in surface waters of the Caribbean Sea (0.5–4 nM)<sup>29</sup>, Resurrection Bay in the Gulf of Alaska (1–2 nM)<sup>30</sup>, Croatian coastal marine waters (2–9 nM)<sup>31</sup>, surface coastal waters of the Gulf of Mexico (0.05–4 nM)<sup>32</sup>, and the Chemotaxis Dock in Woods Hole (0.1 nM)<sup>29,33</sup>.

In the present study, we examined the microbial production of toluene in the water column of the upwelling ecosystem off Concepción in central Chile, both through laboratory incubations of OMZ waters and through monthly field oceanographic campaigns over an annual cycle. We hypothesized that toluene is a secondary metabolic product of microbial degradation of proteinaceous derived precursors under anaerobic conditions in the water column of the OMZ.

## Results

**Experimental microbial synthesis of toluene and decay of phenylalanine.** Toluene was detected both in anoxic incubations with phenylalanine and in the field by GC–MS, as evidenced by *m/z* fragments 65 ( $[M - H^+] - C_2H_2$ ), the base peak tropylium cation 91  $[M - H^+]$  and the molecular cation 92  $[M^+]$  (Fig. 2A,D). Likewise, deuterated toluene ( $C_6D_5-CH_3$ , toluene-*d*<sub>5</sub>) was detected in anoxic incubation bottles amended with deuterated phenylalanine. Mass spectra showed the diagnostics *m/z* fragments 68 ( $[M - H^+] - C_2D_2$ ), tropylium cation 96  $[M - H^+]$ , and molecular ion 97  $[M^+]$  (Fig. 2A,E). In the 5-day anoxic incubations with phenylalanine (Fig. 3), toluene net production peaked on the third day, reaching  $131 \pm 3 \mu M$  toluene and  $102 \pm 2 \mu M$  toluene-*d*<sub>5</sub> under treatment conditions for denitrification (Fig. 3A);  $26 \pm 1 \mu M$  toluene and  $35 \pm 8 \mu M$  toluene-*d*<sub>5</sub> under conditions for sulfate reduction (Fig. 3B); and  $82 \pm 7 \mu M$  toluene and  $94 \pm 9 \mu M$  toluene-*d*<sub>5</sub> in the absence of added external electron acceptors (Fig. 3C). Toluene and toluene-*d*<sub>5</sub> were below the limit of detection and quantification in oxic incubations (Fig. 3D) and in control incubations (Supplementary Fig. 1A). Net maximum production of toluene and toluene-*d*<sub>5</sub> in the presence of  $SO_4^{2-}$  was ca. 3 times lower than in incubations with  $NO_3^-$  and incubations without electron acceptors (Wilcoxon test  $p < 0.05$ ).

Most (95%) phenylalanine and phenyl-*d*<sub>5</sub>-alanine initially present decayed within 5 days of incubation under all conditions (Fig. 3), without significant differences within and between experimental treatments (Wilcoxon test,  $p > 0.05$ ). A significantly higher decay of phenylalanine and phenyl-*d*<sub>5</sub>-alanine was observed under oxic incubations (100% decay in 3 days) compared with ca. 16% remaining after 3 days under anoxic incubations (Wilcoxon test  $p < 0.05$ , Fig. 3). Decay of phenylalanine and phenyl-*d*<sub>5</sub>-alanine was not observed in abiotic controls nor in blank incubations (Supplementary Fig. 1B).

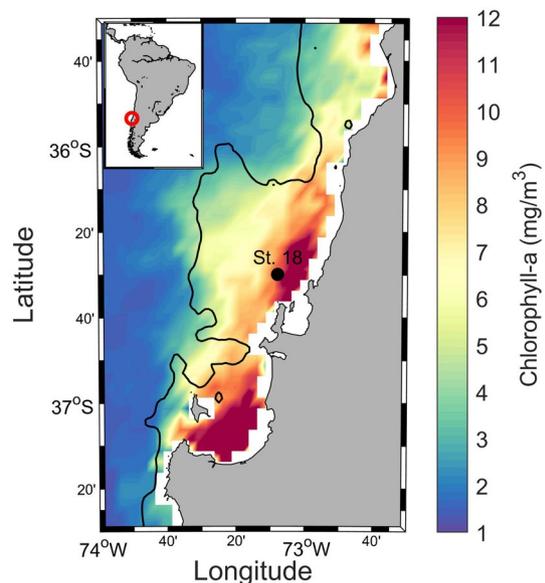
**Experimental microbial decay of toluene.** In the presence of either  $NO_3^-$  or  $SO_4^{2-}$ , about 80% of the deuterated and non-deuterated toluene produced by day three had disappeared within 1 day (Fig. 3A,B), whereas no decay was detected in the absence of electron acceptors, where toluene and toluene-*d*<sub>5</sub> remained at ca. 80  $\mu M$  until the end of the experiment (Fig. 3C, Wilcoxon  $< 0.05$ ). Evidence of sulfate reduction through production of  $HS^-$  during the incubation with  $SO_4^{2-}$  is provided by the mass spectrum (Supplementary Fig. 2).

**Year-round and depth variability of toluene in the water column.** The water column of the study area (Station 18, 90 m depth, Fig. 1) exhibits a distinct annual cycle where suboxic conditions (lower than 22  $\mu M$   $O_2$ ) below 40 m depth are observed for most of the year. However, during austral winter,  $O_2$  concentrations increase throughout the water column (Fig. 4A), as a result of the intrusion of the well-oxygenated and less saline ESPTW (Fig. 4B) on the continental shelf off Concepción. During austral summer, salinity increases due to evaporation and entrainment from below, producing summer ESPTW (Fig. 4B). This pattern is consistent with previous investigations on the dynamics of the OMZ off Concepción<sup>8,34–39</sup>.

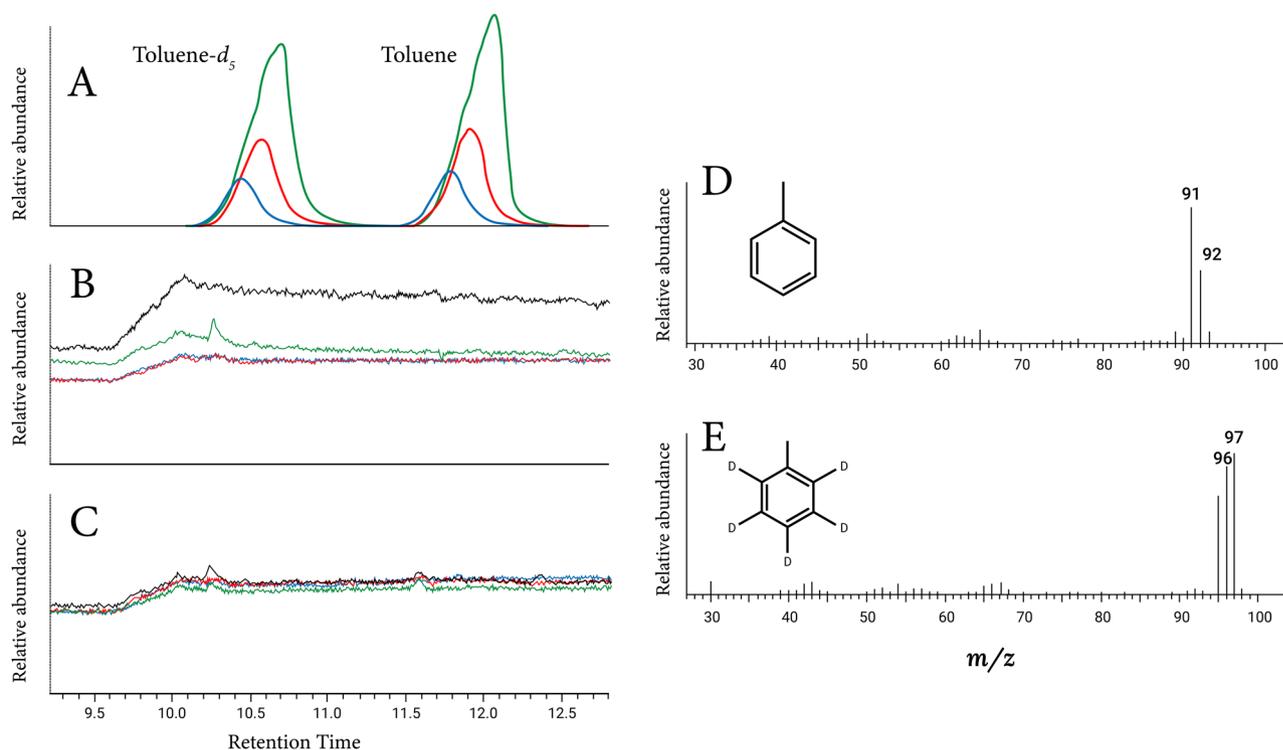
Toluene concentrations varied between 5 and 96 nM, being lower throughout the whole water column during austral autumn–winter ( $< 20$  nM), but much higher in deeper waters ( $> 40$  m) during austral spring–summer between November 2009, January–March and September–December 2010, and January 2011 (Fig. 5A), consistent with a stratified distribution linked to water mass dynamics (Fig. 5B). Thus, lower concentrations of toluene tend to be associated more with ESPTW, whereas higher concentrations are observed in ESSW (Fig. 5B).

## Discussion

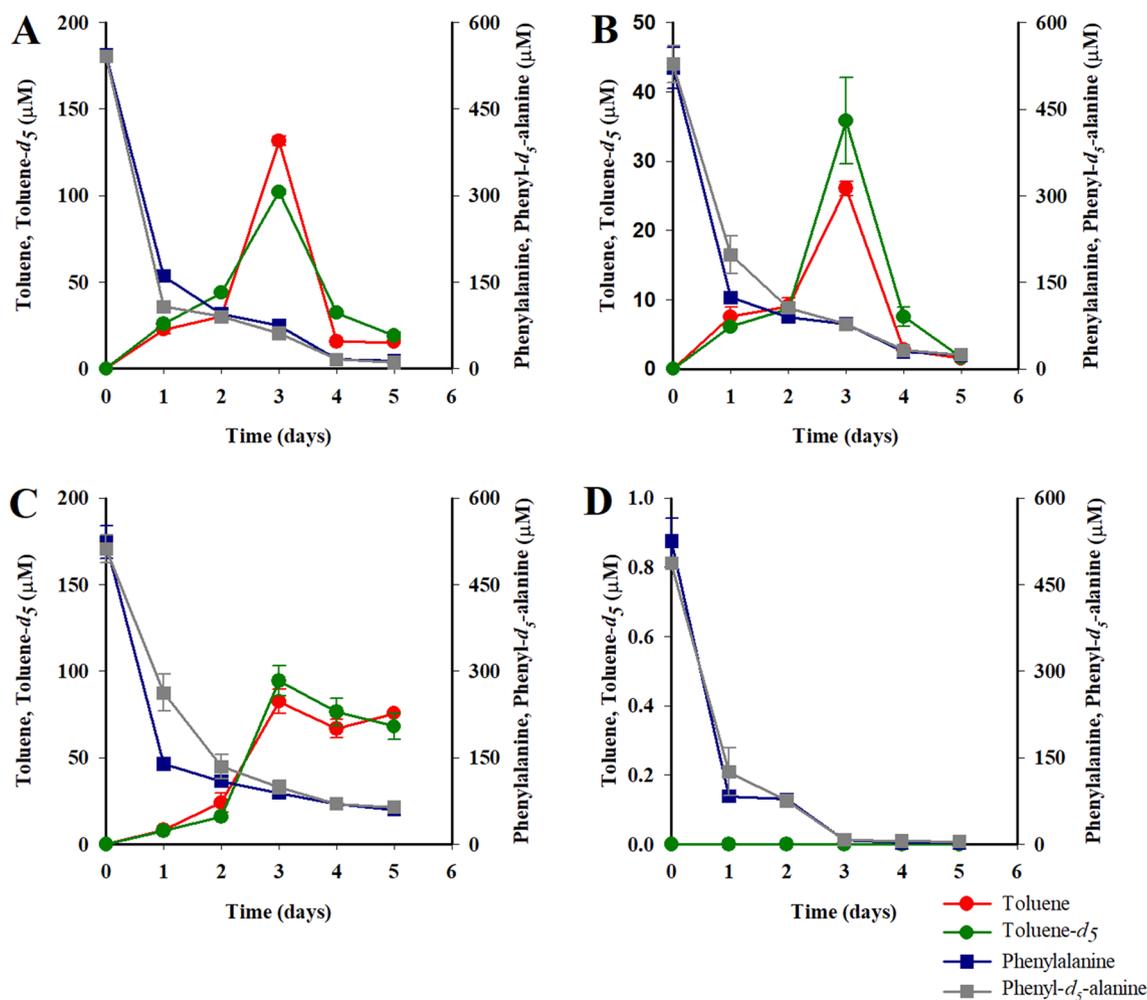
Our incubation experiments clearly demonstrated that the microbial assemblage from OMZ waters synthesizes toluene from phenylalanine as an organic substrate under anaerobic conditions (Figs. 2, 3). This corroborates previous observations in bacterial cultures of *Clostridium*<sup>12</sup> and *Tolomonas*<sup>16</sup>, in anoxic lake waters<sup>41</sup> and sediments<sup>15</sup>, during anaerobic sludge digestion<sup>13,14</sup>, and phytoplankton cultures<sup>18</sup>. Furthermore, the biosynthesis of toluene-*d*<sub>5</sub> from the deuterated aromatic ring of phenyl-*d*<sub>5</sub>-alanine shows that phenylalanine was indeed the precursor of toluene (Fig. 2A,D,E). The production of labeled toluene from the aromatic ring of the deuterated phenylalanine suggests a glycol radical toluene synthase attacks on phenylalanine or phenylacetate methylene  $\beta$ -carbon<sup>42</sup>. Glycol radical enzymes are proteins that belong to the enzymes of the Fe–S cluster, one of the oldest proteins of the planet<sup>43,44</sup>. The gene encoding for Fe–S proteins assembly was recently identified in the metagenome of Marinimicrobia bacteria (MAG-SAR406), a pervasive OMZ bacterium, detected in the Oxygen Minimum Zone of the Tropical Pacific Ocean<sup>45–47</sup>, with the capability for hydrogen and carbon monoxide oxidation, arsenic metabolism, cellulose degradation, and partial denitrification. Being a permanent feature of the modern ocean, we hypothesize that these radical enzymes are widely distributed in the microbial assemblage inhabiting the OMZ. Additionally, bacteria from the Acidobacteria phylum have been detected in OMZ's waters



**Figure 1.** Geographical location of the study site at Station 18, off Concepción in central Chile. Color scale represents the average chlorophyll-a concentration observed between November 2009 and January 2011. The black solid line shows the 200 m isobath. The map was generated using Ocean Data View<sup>40</sup>.



**Figure 2.** (A) Overlaid gas chromatograms showing retention times of toluene and toluene- $d_5$  in incubations anoxic-1 (green line), anoxic-2 (red line), and anoxic-3 (blue line). (B and C) Overlaid chromatograms from control incubations of synthetic seawater (black line) and treatments without inocula (green, red and blue lines, as in caption A) amended with phenylalanine (B) and phenyl- $d_5$ -alanine (C). (D) Mass spectrum of toluene (retention time 11.9 min) detected in both environmental samples and in anoxic incubations with phenylalanine as substrate. (E) Mass spectrum of deuterated toluene (retention time 10.2 min) produced in anoxic incubations with L-phenyl- $d_5$ -alanine.



**Figure 3.** Time course of production of toluene and toluene- $d_5$  and decay of phenylalanine and phenyl- $d_5$ -alanine in (A) anoxic-1 incubations (with excess nitrate), (B) anoxic-2 incubations (with excess sulfate), (C) anoxic-3 incubations (without electron acceptor added), and (D) oxic incubations.

of northern Chile<sup>48</sup>, and the toluene-producing enzyme phenylacetate decarboxylase (PhdB) and its cognate, a radical S-adenosylmethionine activating enzyme (PhdA), and the genes encoding for the enzyme were identified from the metaproteome and metagenome of an Acidobacteria isolated from an anaerobic sewage sludge<sup>9</sup>. Acidobacteria were found exclusively in OMZ waters compared to surface and deeper water masses in northern Chile and are therefore a potential producer of toluene; however, their abundance is lower than other pervasive bacterial groups present in these waters<sup>48</sup>.

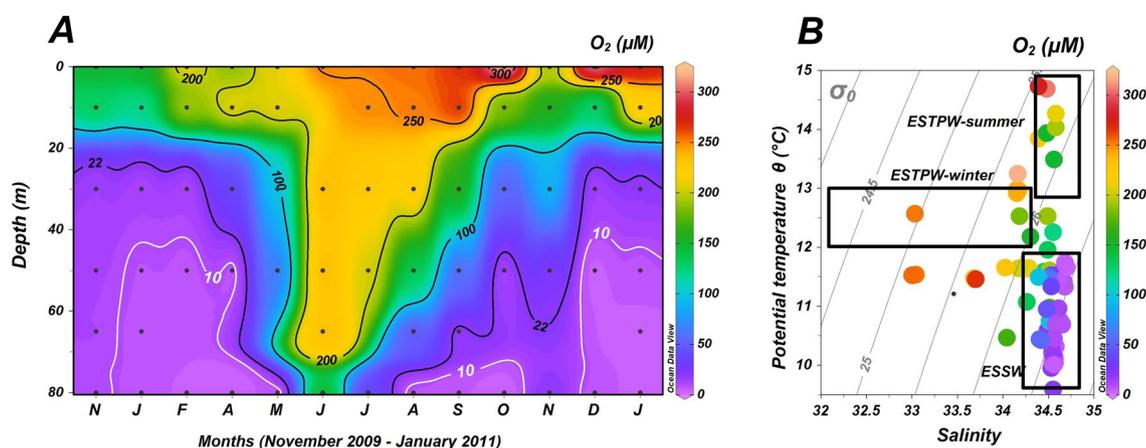
Reactions catalyzed by phenylacetate decarboxylases are endothermic; therefore, microbial production of toluene has been associated with gaining selective advantages other than energy conservation<sup>9</sup>. Toluene-producing bacteria of the OMZ could produce toluene as a mechanism of negative allelopathy against nanoheterotrophs predators abundant in OMZ waters<sup>49,50</sup>.

Toluene could also be synthesized for intracellular pH homeostasis regulation since phenylacetate decarboxylation removes protons from the cytoplasm, thus promoting cellular alkalinization<sup>9</sup>. This mechanism could represent a selective advantage for toluene-synthesizing OMZ microorganisms to thrive in the moderately acidic conditions of OMZ waters (pH 7.8) due to the intrusion of the more acidic ESSW<sup>51–53</sup>, and the co-occurrence of metabolic protonogenic reactions during glucose, acetate, and Stickland fermentation<sup>8</sup>. In addition, depletion of protons in the cytoplasm allows the development of proton-motive force that will generate metabolic energy for toluene-producers microbes<sup>54</sup>.

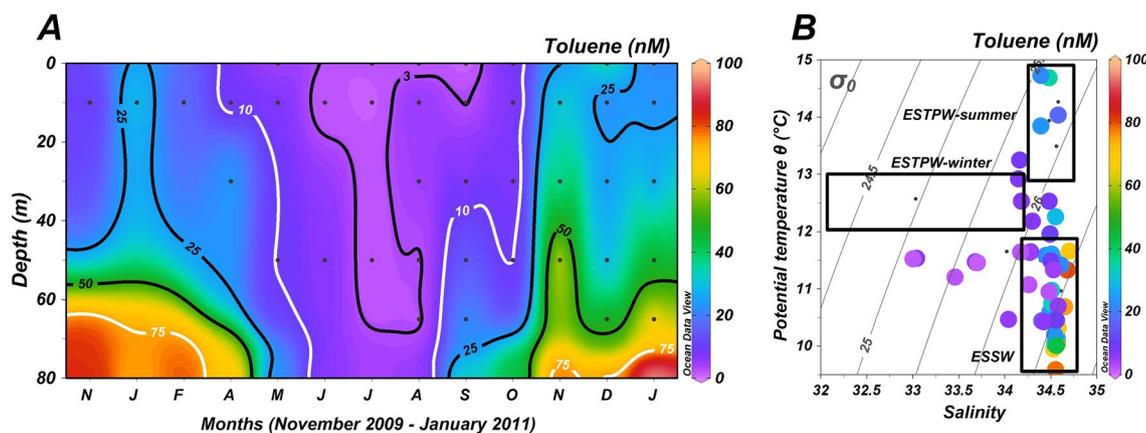
Although the experiment with deuterated phenylalanine unequivocally confirmed phenylalanine as a precursor for toluene biosynthesis, an expected 1:1 stoichiometric production of toluene from phenylalanine<sup>10</sup> was not demonstrated. The observed toluene production accounted for only 5–21% of phenylalanine decay under all anoxic treatments (Fig. 3A–C). Phenylalanine can undergo oxidation to  $\text{CO}_2$ <sup>55</sup>, thus partially explaining the relatively low toluene yield. Anaerobic catabolism of phenylalanine without production of toluene has been shown to produce phenylacetate, phenylpropionate, and phenylacetate in fermenting bacteria *Clostridium*, and producing phenylacetate in denitrifier *Thauera aromatica*<sup>11,56,57</sup>. Thus, decay of phenylalanine in our incubations

Metabolic reaction	Chemical equation	$\Delta G^0$ (kJ/mol toluene)	$\Delta G$ (kJ/mol toluene) OMZ water conditions	$\Delta G$ (kJ/mol toluene) laboratory conditions
Toluene oxidation coupled to denitrification	$C_7H_8 + 7.2NO_3^- + 0.2H^+ \rightarrow 3.6N_2 + 7HCO_3^- + 0.6H_2O$	-3395	-3454	-3530
Toluene oxidation coupled to partial denitrification	$C_7H_8 + 18NO_3^- + 3H_2O \rightarrow 18NO_2^- + 7HCO_3^- + 7H^+$	-2105	-2585	-2972
Toluene oxidation coupled to nitrite reduction	$C_7H_8 + 12NO_2^- + 5H^+ \rightarrow 6N_2 + 7HCO_3^- + 3H_2O$	-408	-337	-392
Toluene oxidation coupled to sulfate reduction	$C_7H_8 + 4.5SO_4^{2-} + 3H_2O \rightarrow 2.25H_2S + 2.25HS^- + 7HCO_3^- + 0.25H^+$	-218	-381	-350
Toluene fermentation	$C_7H_8 + 21H_2O \rightarrow 18H_2 + 7HCO_3^- + 7H^+$	+759	+293	+399

**Table 1.** Anaerobic toluene degradation reactions in laboratory incubations of suboxic waters collected in January 18, 2011 at Station 18 off Concepción at 65 m depth in the OMZ. Gibbs energy ( $\Delta G$ ) was calculated assuming substrate and product activities, temperature, and pH of the suboxic water column conditions for austral spring at the sampling site as well as for the conditions of incubations. For comparison, Standard Gibbs energy ( $\Delta G^0$ ) values are shown.



**Figure 4.** (A) Dissolved oxygen time-series data from November 2009 throughout January 2011. The white line represents the 10  $\mu M$   $O_2$  isoline. Black dots represent sampling points. (B) Potential temperature ( $\Theta$ )/salinity diagram from depth profiles taken at Station 18, off Concepción. Data points are color-coded (side color bar) for oxygen concentrations. Rectangles indicate temperature and salinity ranges for Equatorial Subsurface Water (ESSW) and Eastern South Pacific Transition Water (ESPTW) in austral winter and summer. Grey dotted lines represent isopycnals as sigma-t ( $\sigma_0$ ).



**Figure 5.** (A) Toluene time-series data from November 2009 to January 2011. White lines represent the 10 and 75  $\mu M$  toluene isolines. Black dots represent sampling points. (B) Potential temperature ( $\Theta$ )/salinity diagram from depth profiles taken at Station 18, off Concepción. Data points are color-coded (side color bar) for toluene concentrations. Rectangles indicate temperature and salinity ranges for Equatorial Subsurface Water (ESSW) and Eastern South Pacific Transition Water (ESPTW) in austral winter and summer. Grey dotted lines represent isopycnals as sigma-t ( $\sigma_0$ ).

could result from the activity of a toluene-producing glycol radical enzyme and from anaerobic catabolism mediated by benzoyl-coenzyme A<sup>11,56–58</sup>, with phenylacetate as the intermediate substrate in both pathways. Another factor that could influence the 1:1 stoichiometry in our incubations is the concurrent microbial oxidation of toluene as it is being produced, as already shown for other environments<sup>59–66</sup>. In consequence, observed changes in toluene concentrations during incubations will correspond to net production.

Toluene microbial decay by anaerobic oxidation coupled to denitrification and partial denitrification are the most thermodynamically favorable reactions ( $\Delta G > 2500 \text{ kJ mol}^{-1}$ ) compared with toluene oxidation with  $\text{NO}_2^-$  and  $\text{SO}_4^{2-}$  and fermentation (Table 1). Metagenome-assembled genomes from the eastern tropical Pacific OMZ contain genes involved in partial denitrification<sup>67</sup>, supporting the feasibility of this metabolism in OMZ waters.

In anaerobic toluene-degrading communities in aquifer sediments, variability in  $\text{NO}_2^-$  and  $\text{NO}_3^-$  concentrations determines the fate of reduction of  $\text{NO}_3^-$  ( $\text{NO}_2^-$  or  $\text{N}_2$ ) by modulating changes in the microbial community of denitrifiers that totally or partially denitrifies<sup>67</sup>. In our study site, the distribution and availability of  $\text{NO}_2^-$  showed a clear vertical and temporal pattern with the highest concentrations during austral spring and summer and restricted to the deepest layers, that resembles the vertical distribution of toluene concentrations throughout a year cycle (Supplementary Fig. 3A,C), both significantly correlated (Supplementary Table 1), suggesting the occurrence of partial denitrification. On the contrary,  $\text{NO}_3^-$  concentration decreases in the deepest layer during austral summer (Supplementary Fig. 3B), and the year-round pattern resembles the upwelling and productivity cycles in the study area, as shown by negative correlations (Supplementary Table 1) with temperature (an indicator of upwelling) and fluorescence (an indicator of phytoplankton productivity). Taken together, it suggests that partial denitrification during degradation of toluene could be occurring in these waters, apparently associated with the year cycle in nutrient availability, thus providing  $\text{NO}_2^-$  to anammox bacteria in these waters<sup>68</sup>, unveiling a novel and hitherto unknown connection of C and N cycling in OMZ waters.

The anoxic incubations also revealed that ~80% of produced toluene disappeared within 1 day in the presence of  $\text{NO}_3^-$  or  $\text{SO}_4^{2-}$ , whereas in the absence of these electron acceptors, toluene remained at  $\sim 80 \pm 4 \mu\text{M}$  on day 3 and decayed slightly to less than  $75 \pm 6 \mu\text{M}$  by day 5 (Fig. 3C). Thermodynamic considerations support this since fermentation of toluene is endergonic under both experimental and field conditions (Table 1) due to the production of  $\text{H}_2$  during its disproportionation that drives  $\Delta G$  to be  $> 0$ . However, partial pressure of  $\text{H}_2$  can be reduced by an electron-acceptor microorganism in a microbial syntrophic consortium, thus rendering an overall exothermic fermentation reaction, as shown in laboratory co-cultures<sup>66</sup>. Moreover, in a syntrophic coculture, Meckenstock et al.<sup>69</sup> also detected that syntrophic degradation is 2–3 times slower than toluene oxidation by sulfate. Our relatively short experimental incubations preclude conclusions regarding whether toluene would remain at  $80 \pm 4 \mu\text{M}$  (Fig. 3C) or would slowly decay in the absence of external electron acceptors, given sufficient microbial consortium growth.

Our study site (Station 18) is located 18 nautical miles from the coast of Concepción, a major city of Chile, and therefore input from the urban, maritime, port and industrial activities is conceivable since toluene is universally used in industry (principally as a solvent and gasoline additive) and in domestic products<sup>70</sup>. However, depth profiles of toluene in the water column (Fig. 5A and Supplementary Fig. 4) indicate natural biogeochemical processes. In austral spring, summer, and autumn, increased concentrations with depth are consistent with a deeper source for toluene in  $\text{O}_2$ -depleted waters (Figs. 4A and 5A). Despite our evidence for potential toluene production in the water column, diffusion from sediments is an additional source that cannot be ruled out, as shown in other coastal anoxic sediments<sup>70,71</sup>.

Circulation of water masses forced by atmospheric conditions represents a fundamental control of the annual cycle at the study site. Oceanographic Station 18 (Fig. 1) is at the southernmost limit of the extensive OMZ of the Humboldt Current System<sup>72</sup>. It is subject to alternate intrusions of oxygenated subantarctic waters during austral autumn–winter and of anoxic and hypoxic waters associated with Equatorial Subsurface Waters (ESSW)<sup>37</sup>. Concentrations of toluene exceeding 50 nM occurred principally below the  $10 \mu\text{M}$   $\text{O}_2$  isoline (Fig. 4A), conditions favorable for the activity of the glycol radical decarboxylase enzyme<sup>9,10</sup>, otherwise irreversibly inactivated by  $\text{O}_2$ <sup>10,19,21</sup>.

The vertical distribution of toluene in the water column appears to be inversely related to dissolved oxygen at the study site (Figs. 4A and 5A), with a significant negative correlation between the two (Spearman correlation,  $\rho = -0.6$ ,  $p < 0.05$ , Supplementary Table 1). Moreover, toluene concentrations correlate negatively with temperature and positively with salinity (Spearman correlation,  $p < 0.05$ , Supplementary Table 1), indicative of the annual cycle of upwelling of ESSW and intrusions of ESPTW (Fig. 5B), as previously observed<sup>8,37</sup>. Significant associations of toluene concentrations with  $\text{NO}_2^-$ ,  $\text{PO}_4^{3-}$ , and volatile fatty acids (VFA; Spearman correlations;  $p < 0.05$ ; Supplementary Table 1) were also observed. These are considered to be molecular indicators of fermentation<sup>8</sup> and denitrification<sup>6,73</sup> and provide the chemical environment for toluene production in the water column.

Biosynthesis of toluene from deuterated and non-deuterated phenylalanine was not detected in oxic incubations (Fig. 3D), as might be expected<sup>10</sup>. This corroborates our observations that the lowest concentrations of toluene occurred in well-oxygenated and low salinity subantarctic waters that form ESPTW in winter and summer (Fig. 5B). Because subantarctic waters are well oxygenated, we would anticipate that no production of toluene occurs in these waters, and we suggest that the relatively small inventory of toluene in ESPTW results from mixing with OMZ waters (Fig. 5). We cannot rule out that OMZ's pervasive bacterial groups SAR202 and SAR406 may degrade aromatics and high molecular weight recalcitrant dissolved organic matter<sup>47,74</sup> using monooxygenase enzymes when oxygenated waters intrude the continental shelf. Therefore, the lower concentrations of toluene observed in the most oxygenated waters of the study site (e.g. ESPTW), could be the net result of input from OMZ waters and microbial aerobic degradation as  $\text{O}_2$  availability increases during mixing (Fig. 4A,B).

The average depth-integrated toluene inventory of  $3 \text{ mmol toluene m}^{-2}$  (Supplementary Fig. 3) represents an estimated reservoir of about 2 million tons of toluene for the whole OMZ off Chile of ca.  $10 \text{ million km}^{2,72}$ , is

about 50% higher than the inventory of CH<sub>4</sub> and near 20 times lower than that of VFA (Supplementary Table 2), putative representative molecules of anaerobic metabolism. The estimated inventory of toluene is a significant quantity compared with the annual toluene production of the European Union of 1–10 million tons<sup>71</sup>.

## Conclusions

In laboratory incubations of OMZ waters off central Chile, microbial assemblages anaerobically biosynthesize toluene from phenylalanine, a finding consistent with the year-round pattern of toluene concentrations in the water column in the upwelling ecosystem of the Humboldt Current System. A fraction of this toluene is exported to surrounding oxygenated waters, namely the subantarctic Eastern South Pacific Transition Water (ESPTW).

Microbial production of toluene increases cellular pH, perhaps counteracting the external acidic media of the OMZ (pH 7.8). The field and laboratory data analysis suggest that, as shown for toluene-degrading communities in aquifer sediments, the availability of NO<sub>2</sub><sup>-</sup> to oxidize toluene could determine the fate of the reduction of NO<sub>3</sub><sup>-</sup> (NO<sub>2</sub><sup>-</sup> or N<sub>2</sub>).

Our estimate of the potential reservoir of toluene in the OMZ off Chile is in the order of 2 million tons, within the range of CH<sub>4</sub> and VFA inventories, putative representative molecules of anaerobic metabolism, as well as in the range of annual industrial production of toluene for the European Union. Warming and eutrophication of coastal waters are likely to further promote hypoxic conditions, enhance anaerobic metabolism, and potentially reveal reservoirs of various organics produced by the OMZ microbiome.

## Methods

**Sampling.** *Water samples.* Water samples were collected onboard the research vessel Kay-Kay II (Universidad de Concepción) quasi-monthly between November 2009 and January 2011 (13 sampling dates). Seawater was collected at depths of 0, 10, 30, 65, 70, and 80-m using a rosette system equipped with 10-L Niskin bottles. Subsamples of 50 mL volume were transferred onboard to pre-combusted (450 °C, 4 h) gas tight borosilicate Wheaton bottles (darkened with aluminum foil) under a N<sub>2</sub> atmosphere—using glove bags (Aldrich AtmosBag) to avoid O<sub>2</sub> contamination—then immediately poisoned with HgCl<sub>2</sub> (0.001%) to arrest microbial activity. Bottles were crimp-sealed with ultra-pure bromobutyl stoppers (Wheaton) and stored in the dark at 4 °C until further analysis of toluene in the laboratory (within 12 h). Data for temperature, salinity, O<sub>2</sub>, NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, NH<sub>4</sub><sup>+</sup> and fluorescence were provided by the COPAS Center *Time Series Oceanographic Station 18* (FONDAP COPAS) and the program *Microbial Initiative in Low Oxygen off Concepción and Oregon* (Moore Foundation [http://mi\\_loco.coas.oregonstate.edu](http://mi_loco.coas.oregonstate.edu)). Volatile fatty acid (VFA) data were taken from Srain et al.<sup>8</sup>.

**Inocula and incubation media.** Inocula of seawater for laboratory experiments were obtained on January 18, 2011, from 65 m depth (10.3 °C, 34.6 PSU, 4.0 μM O<sub>2</sub>) and transferred to 10 mL serum vacuum-tubes BD. Tubes were maintained at 10 °C in darkness until arrival at the laboratory. Inocula and reagents were manipulated in a laminar flow hood LABCONCO Class II Type IIA under a N<sub>2</sub>-saturated atmosphere (glove bags Aldrich Atmosbag) achieved by bubbling with N<sub>2</sub> (99.9% purity). O<sub>2</sub>-sensitive methylene blue (Resazurin 0.0001%, Wolfe<sup>75</sup>; McDonald et al.<sup>76</sup>) was added to anoxic incubation vessels to detect traces of O<sub>2</sub> contamination (>0.7 μM). Artificial seawater for anoxic incubations was prepared according to Lovley<sup>77</sup>.

**Experimental setup.** Experimental incubations were conducted in duplicate by transferring 30 mL of artificial seawater into darkened pre-combusted (450 °C, 4 h) borosilicate glass bottles (60 mL, Wheaton), under a N<sub>2</sub> saturated atmosphere. L-Phenylalanine in excess (500 μM, CAS 63-91-2) and D-glucose (10 μM, CAS 50-99-7), were added as carbon and nitrogen sources, and inoculated with 10% v/v of seawater collected from the OMZ. Bottles were sealed with ultra-pure bromobutyl stoppers, keeping a headspace volume of 20 mL for sampling of volatile compounds. Incubation of bottles was carried out at 10 °C in darkness, with constant orbital agitation (180 rpm). Incubations were conducted for 5 days with the headspace sampled (500 μL with a Hamilton gas-tight syringe 1750) every ca. 24 h.

Experimental treatments were (in parenthesis the initial final concentration added).

- (1) Anoxic-1 incubation under denitrifying conditions: Phenylalanine (500 μM), glucose (10 μM) and NO<sub>3</sub><sup>-</sup> (800 μM). This same concentration of NO<sub>3</sub><sup>-</sup> was subsequently added daily under a N<sub>2</sub>-saturated atmosphere using a microliter syringe Hamilton (500 μL) previously flushed with N<sub>2</sub>.
- (2) Anoxic-2 incubation under sulfate reduction conditions: Phenylalanine (500 μM), glucose (800 μM) and SO<sub>4</sub><sup>2-</sup> (450 μM), amended with 5% v/v of reducing solution (Na<sub>2</sub>O<sub>2</sub>S × 5H<sub>2</sub>O and cysteine) to promote sulfate reduction. This same concentration of SO<sub>4</sub><sup>2-</sup> was subsequently added daily under N<sub>2</sub>-saturated atmosphere using a microliter syringe Hamilton (500 μL) previously flushed with N<sub>2</sub>.
- (3) Anoxic-3 incubation under fermentative conditions: Phenylalanine (500 μM) and glucose (800 μM) without external electron acceptors.
- (4) Oxidic incubation: Phenylalanine (500 μM), glucose (800 μM) and O<sub>2</sub> (370 μM). Incubation flasks were oxygenated to saturation (8.9 mL L<sup>-1</sup> at 10 °C and 0.21 atm, Henry's Law) with synthetic air (ca. 20% O<sub>2</sub>, 80% N<sub>2</sub>, 99.99% purity). Five mL aliquots of pure synthetic air were added daily to the incubation flasks using a 1000 μL gas-tight syringe Hamilton.
- (5) Experiments Anoxic-1, Anoxic-2, Anoxic-3, and Oxidic were also repeated using deuterated phenylalanine (500 μM L-phenyl-*d*<sub>5</sub>-alanine, CAS 284664-89-7), instead of phenylalanine, to confirm that toluene synthesis came from phenylalanine.

Anoxic conditions were achieved by gently bubbling N<sub>2</sub> into incubation bottles for 15 min to displace traces of O<sub>2</sub>. Abiotic controls with substrates were incubated without OMZ inocula for each treatment.

**Analysis of toluene in ambient and experimental samples.** Analysis of toluene was carried out through Headspace Solid Phase Micro-Extraction coupled to Gas Chromatography with MSD detection (HS-SPME-GCMS). Twenty-five mL of water was removed from each sample (n = 3), under a N<sub>2</sub> saturated atmosphere, to generate a headspace. Environmental and experimental samples were placed on a thermostatic hotplate at 20 °C, with constant stirring (using an acid-cleaned and sterilized stirring bar) for 5 min to reach partition equilibrium between aqueous and gas phases. Polar and non-polar volatile organic compounds were adsorbed using a fiber of 85 µm Carboxen/PDMS Stable Flex (SUPELCO). The fiber was inserted into the insertion septum of the gas bottle and exposed to the gas phase (headspace). After adsorption of analytes, the fiber was placed in the injection port of the gas chromatograph and exposed for 5 min at 250 °C for desorption of the collected gaseous analytes. GC-MS analyses were conducted using a gas chromatograph Agilent 6890 N series coupled to a mass spectrometer Agilent 5973 Network. The mass spectrometer was operated in electron impact mode (70 eV) and spectra of toluene standards and incubation samples were acquired in full scan mode (m/z 40–600, 2.6 s<sup>-1</sup>), with spectra for environmental samples acquired using selective ion monitoring (SIM): toluene (tropylium ion m/z 91). Chromatographic separation was achieved using a HP-Plot/Q column 30 m (0.32 mm diameter, 0.20 µm film thickness), using He as carrier gas. The oven temperature program started at 100 °C (5 min), ramped to 230 °C at 50 °C min<sup>-1</sup> (held 2 min), to 250 °C at 50 °C min<sup>-1</sup> (held 7 min).

Quantification of toluene and deuterated toluene (toluene-*d*<sub>8</sub>) in both scan and selective ion monitoring (SIM) modes was determined using calibration curves (R<sup>2</sup> > 0.994) for which toluene standards (HPLC grade, ≥ 98.8% purity, Fisher) were added to artificial seawater in the range 1 nmol L<sup>-1</sup>–1 mmol L<sup>-1</sup>. Concentrations of toluene in the gas phase were calculated as  $C_{HS} = C_{AP} / (K + (V_{HS}/V_S))$  with the partition coefficient  $K = C_{AP} / C_{HS}$ .  $C_{HS}$  is the concentration in the headspace,  $C_{AP}$  is the concentration in the aqueous phase, and  $V_{HS}$  and  $V_S$  are headspace and sample volumes<sup>78</sup>, resulting in a partition coefficient of 0.9. Detection limits were calculated from the slopes and residual standard deviations derived from linear regressions of calibration curves (3 times residual error times slope)<sup>79</sup>. Limit of detection for toluene was 1 nmol L<sup>-1</sup>, whilst limit of quantification was 5 nmol L<sup>-1</sup>. Synthetic seawater was analyzed for toluene.

**Analyses of dissolved free amino acids (DFAA).** Concentrations of DFAA from incubations, and from Station 18, were quantified as OPA-derivatized adducts<sup>80</sup> with a Shimadzu LC-10AT HPLC coupled to a Shimadzu RF-10Axl fluorescence detector (set at excitation/emission of 340/450 nm), column oven CTO 10As and autosampler (Shimadzu SIL 10 ADvp). Aliquots of 0.6 mL, mixed with 0.4 mL methanol, were derivatized in the autosampler with 60 µL ortho-phthalaldehyde/2-mercaptoethanol reagent and 100 µL sodium acetate buffer 0.1 N, pH 5, and then injected (50 µL) into the HPLC. Fifteen amino acids (asp, glu, ser, his, gly, thr, arg, ala, tyr, val, met, phe, ile, leu, lys) were separated using an Alltima C18 (5 mm, 250 × 4.6 mm) column maintained at 40 °C, with a mobile phase of 5% tetrahydrofuran in 25 mM sodium acetate and methanol, and at a flow rate of 1 mL min<sup>-1</sup>. A gradient of 25–30% methanol in 35 min, 30–50% in 7 min, 50–60% in 18 min, 60–100% in 12 min was used. Initial conditions were restored within 7 min, and the column equilibrated for 10 min between injections. Amino acids were identified and quantified by comparison with chromatograms of a standard amino acid mix (Pierce 20088) run under the same conditions every 10 injections. The coefficient of variation for quantification of duplicate samples was 9.4%.

**Statistical analysis.** Since homogeneity of variances (Levene test) and normality of variables (Shapiro-Wilk test) were not fulfilled, we tested for significant differences between environmental data using the non-parametric Kruskal-Wallis ANOVA test. Statistical differences between experimental treatments were determined by using the Wilcoxon matched pairs test. Correlations were examined using Spearman R coefficients.

**Thermodynamic calculations.** Calculated Gibbs Energy values (ΔG) for proposed oxidation reactions of toluene in the suboxic water column were calculated using activities of substrates and products, temperature and pH of the water depth of inoculum collection in austral spring or from literature. The following data and sources were used: temperature (10.2 °C), NO<sub>2</sub><sup>-</sup> (5 µM) and NO<sub>3</sub><sup>-</sup> (25 µM) from the COPAS Center Oceanographic Time Series database (W. Schneider, curator), pH (7.8, this study), toluene (0.09 µM, this study), SO<sub>4</sub><sup>2-</sup> (28 mM)<sup>81</sup>, HCO<sub>3</sub><sup>-</sup> (2 mM)<sup>81</sup>, N<sub>2</sub> (389 µM)<sup>82</sup>, HS<sup>-</sup> (1 µM)<sup>83,84</sup>. Activities of N<sub>2</sub>, NO<sub>2</sub><sup>-</sup>, NO<sub>3</sub><sup>-</sup>, SO<sub>4</sub><sup>2-</sup> and HS<sup>-</sup> in experimental incubations were determined stoichiometrically from the chemical equations of the proposed reactions. Thermodynamic calculations were carried out using THERMODYN software<sup>85</sup>.

## Data availability

All data generated or analysed during this study are included in this published article and its supplementary information files.

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## References

1. Levin, L. A. *et al.* Effects of natural and human-induced hypoxia on coastal benthos. *Biogeosciences*. <https://doi.org/10.5194/bg-6-2063-2009> (2009).

2. Ulloa, O. & Pantoja, S. The oxygen minimum zone of the eastern South Pacific. *Deep. Res. Top. Part II Stud. Oceanogr.* <https://doi.org/10.1016/j.dsr2.2008.12.004> (2009).
3. Stramma, L., Johnson, G. C., Sprintall, J. & Mohrholz, V. Expanding oxygen-minimum zones in the tropical oceans. *Science* <https://doi.org/10.1126/science.1153847> (2008).
4. Levipan, H. A., Quiñones, R. A., Johansson, H. E. & Urrutia, H. Methylophilic methanogens in the water column of an upwelling zone with a strong oxygen gradient off Central Chile. *Microbes Environ.* <https://doi.org/10.1264/jsme2.22.268> (2007).
5. Gutiérrez, M. H., Pantoja, S., Tejos, E. & Quiñones, R. A. The role of fungi in processing marine organic matter in the upwelling ecosystem off Chile. *Mar. Biol.* <https://doi.org/10.1007/s00227-010-1552-z> (2011).
6. Galán, A., Thamdrup, B., Saldías, G. S. & Fariás, L. Vertical segregation among pathways mediating nitrogen loss (N<sub>2</sub> and N<sub>2</sub>O production) across the oxygen gradient in a coastal upwelling ecosystem. *Biogeosciences*. <https://doi.org/10.5194/bg-14-4795-2017> (2017).
7. Plominsky, A. M. *et al.* Metabolic potential and in situ transcriptomic profiles of previously uncharacterized key microbial groups involved in coupled carbon, nitrogen and sulfur cycling in anoxic marine zones. *Environ. Microbiol.* <https://doi.org/10.1111/1462-2920.14109> (2018).
8. Srain, B. M. *et al.* Fermentation and anaerobic oxidation of organic carbon in the oxygen minimum zone of the upwelling ecosystem off Concepción Central Chile. *Front. Mar. Sci.* <https://doi.org/10.3389/fmars.2020.00533> (2020).
9. Beller, H. R. *et al.* Discovery of enzymes for toluene synthesis from anoxic microbial communities. *Nat. Chem. Biol.* <https://doi.org/10.1038/s41589-018-0017-4> (2018).
10. Zargar, K. *et al.* In vitro characterization of phenylacetate decarboxylase, a novel enzyme catalyzing toluene biosynthesis in an anaerobic microbial community. *Sci. Rep.* <https://doi.org/10.1038/srep31362> (2016).
11. Carmona, M. *et al.* Anaerobic catabolism of aromatic compounds: A genetic and genomic view. *Microbiol. Mol. Biol. Rev.* <https://doi.org/10.1128/mbr.00021-08> (2009).
12. Pons, J. L., Rimbault, A., Darbord, J. C. & Leluan, G. Biosynthèse de toluène chez *Clostridium aerofotetium* souche WS. *Ann. l'Institut Pasteur Microbiol.* [https://doi.org/10.1016/S0769-2609\(84\)80029-0](https://doi.org/10.1016/S0769-2609(84)80029-0) (1984).
13. Mrowiec, B., Suschka, J. & Keener, T. C. Formation and biodegradation of toluene in the anaerobic sludge digestion process. *Water Environ. Res.* <https://doi.org/10.2175/106143005x41852> (2005).
14. Marczak, M., Wolska, L. & Namiesnik, J. Determination of toluene formed during fermentation of sewage sludge. *Int. J. Environ. Stud.* <https://doi.org/10.1080/00207230600662310> (2006).
15. Jüttner, F. Formation of toluene by microorganisms from anoxic freshwater sediments. *Fresenius. J. Anal. Chem.* <https://doi.org/10.1007/BF00321745> (1991).
16. Fischer-Romero, C., Tindall, B. J. & Jüttner, F. *Tolumonas auensis* gen. nov., sp. nov., a toluene-producing bacterium from anoxic sediments of a freshwater lake. *Int. J. Syst. Bacteriol.* <https://doi.org/10.1099/00207713-46-1-183> (1996).
17. Jüttner, F. & Henatsch, J. J. Anoxic hypolimnion is a significant source of biogenic toluene. *Nature* <https://doi.org/10.1038/323797a0> (1986).
18. Middelburg, J. J. & Levin, L. A. Coastal hypoxia and sediment biogeochemistry. *Biogeosciences*. <https://doi.org/10.5194/bg-6-1273-2009> (2009).
19. Heider, J., Spormann, A. M., Beller, H. R. & Widdel, F. Anaerobic bacterial metabolism of hydrocarbons. *FEMS Microbiol. Rev.* [https://doi.org/10.1016/S0168-6445\(98\)00025-4](https://doi.org/10.1016/S0168-6445(98)00025-4) (1998).
20. Rodrigues, A. V., Tantillo, D. J., Mukhopadhyay, A., Keasling, J. D. & Beller, H. R. Insight into the mechanism of phenylacetate decarboxylase (PhdB), a toluene-producing glycol radical enzyme. *ChemBioChem* <https://doi.org/10.1002/cbic.201900560> (2020).
21. Selmer, T., Pierik, A. J. & Heider, J. New glycol radical enzymes catalysing key metabolic steps in anaerobic bacteria. *Biol. Chem.* <https://doi.org/10.1515/BC.2005.114> (2005).
22. Sawers, G. Biochemistry, physiology and molecular biology of glycol radical enzymes. *FEMS Microbiol. Rev.* [https://doi.org/10.1016/S0168-6445\(98\)00020-5](https://doi.org/10.1016/S0168-6445(98)00020-5) (1998).
23. Klenk, H. P. *et al.* The complete genome sequence of the hyperthermophilic, sulphate-reducing archaeon *Archaeoglobus fulgidus*. *Nature* <https://doi.org/10.1038/37052> (1997).
24. Smith, D. R. *et al.* Complete genome sequence of *Methanobacterium thermoautotrophicum* ΔH: Functional analysis and comparative genomics. *J. Bacteriol.* <https://doi.org/10.1128/jb.179.22.7135-7155.1997> (1997).
25. Lee, C., Wakeham, S. & Arnosti, C. Particulate organic matter in the sea: The composition conundrum. *Ambio* <https://doi.org/10.1579/0044-7447-33.8.565> (2004).
26. Wakeham, S. G., Goodwin, J. T. & Davis, A. C. Distributions and fate of volatile organic compounds in Narragansett Bay, Rhode Island. *Can. J. Fish. Aquat. Sci.* <https://doi.org/10.1139/f83-336> (1983).
27. Wiesenburg, D. A., Bodennec, G. & Brooks, J. M. Volatile liquid hydrocarbons around a production platform in the Northwest Gulf of Mexico. *Bull. Environ. Contam. Toxicol.* <https://doi.org/10.1007/BF01611003> (1981).
28. Saeed, T., Khordagui, H. & Al-Bloushi, A. Distribution of volatile liquid hydrocarbons in the vicinity of power/desalination plants in Kuwait. *Water Sci. Technol.* [https://doi.org/10.1016/S0273-1223\(99\)00588-0](https://doi.org/10.1016/S0273-1223(99)00588-0) (1999).
29. Sauer, T. C. Volatile liquid hydrocarbons in waters of the Gulf of Mexico and Caribbean Sea. *Limnol. Oceanogr.* <https://doi.org/10.4319/lo.1980.25.2.0338> (1980).
30. Button, D. K., Robertson, B. R., McIntosh, D. & Jüttner, F. Interactions between marine bacteria and dissolved-phase and beached hydrocarbons after the Exxon Valdez oil spill. *Appl. Environ. Microbiol.* <https://doi.org/10.1128/aem.58.1.243-251.1992> (1992).
31. Karačonji, B., Skender, L. & Karačić, V. Benzene, toluene, ethylbenzene, and isomeric xylenes in various water samples in Croatia. *Bull. Environ. Contam. Toxicol.* <https://doi.org/10.1007/s00128-006-0943-9> (2006).
32. Sauer, T. C., Sackett, W. M. & Jeffrey, L. M. Volatile liquid hydrocarbons in the surface coastal waters of the Gulf of Mexico. *Mar. Chem.* [https://doi.org/10.1016/0304-4203\(78\)90039-7](https://doi.org/10.1016/0304-4203(78)90039-7) (1978).
33. Gschwend, P. M., Zafriou, O. C., Mantoura, R. F. C., Schwarzenbach, R. P. & Gagosian, R. B. Volatile organic compounds at a coastal site. 1. Seasonal variations. *Environ. Sci. Technol.* <https://doi.org/10.1021/es00095a010> (1982).
34. Schneider, W., Fuenzalida, R., Rodríguez-Rubio, E., Garcés-Vargas, J. & Bravo, L. Characteristics and formation of Eastern South Pacific Intermediate Water. *Geophys. Res. Lett.* <https://doi.org/10.1029/2003GL017086> (2003).
35. Schneider, W., Donoso, D., Garcés-Vargas, J. & Escribano, R. Water-column cooling and sea surface salinity increase in the upwelling region off central-south Chile driven by a poleward displacement of the South Pacific High. *Prog. Oceanogr.* <https://doi.org/10.1016/j.pocean.2016.11.004> (2017).
36. Escribano, R. & Schneider, W. The structure and functioning of the coastal upwelling system off central/southern Chile. *Prog. Oceanogr.* <https://doi.org/10.1016/j.pocean.2007.08.020> (2007).
37. Sobarzo, M., Bravo, L., Donoso, D., Garcés-Vargas, J. & Schneider, W. Coastal upwelling and seasonal cycles that influence the water column over the continental shelf off central Chile. *Prog. Oceanogr.* <https://doi.org/10.1016/j.pocean.2007.08.022> (2007).
38. Letelier, J., Pizarro, O. & Nuñez, S. Seasonal variability of coastal upwelling and the upwelling front off central Chile. *J. Geophys. Res. Ocean.* <https://doi.org/10.1029/2008JC005171> (2009).
39. Escribano, R. & Morales, C. E. Spatial and temporal scales of variability in the coastal upwelling and coastal transition zones off central-southern Chile (35–40° S). *Prog. Oceanogr.* <https://doi.org/10.1016/j.pocean.2011.07.019> (2012).
40. Schlitzer, R. Ocean Data View Version 4.3.6. (2010) Bremerhaven: Alfred Wegener Institute. <http://odv.awi.de/>

41. Niggemann, J., Ferdelman, T. G., Lomstein, B. A., Kallmeyer, J. & Schubert, C. J. How depositional conditions control input, composition, and degradation of organic matter in sediments from the Chilean coastal upwelling region. *Geochim. Cosmochim. Acta*. <https://doi.org/10.1016/j.gca.2006.12.012> (2007).
42. Blattner, F. R. *et al.* The complete genome sequence of *Escherichia coli* K-12. *Science* <https://doi.org/10.1126/science.277.5331.1453> (1997).
43. Weiss, M. C. *et al.* The physiology and habitat of the last universal common ancestor. *Nat. Microbiol.* <https://doi.org/10.1038/nmicr.2016.116> (2016).
44. Imlay, J. A. Iron-sulphur clusters and the problem with oxygen. *Mol. Microbiol.* <https://doi.org/10.1111/j.1365-2958.2006.05028.x> (2006).
45. Beman, J. M. *et al.* Biogeochemistry and hydrography shape microbial community assembly and activity in the eastern tropical North Pacific Ocean oxygen minimum zone. *Environ. Microbiol.* <https://doi.org/10.1111/1462-2920.15215> (2021).
46. Pajares, S., Varona-Cordero, F. & Hernández-Becerril, D. U. Spatial distribution patterns of bacterioplankton in the oxygen minimum zone of the Tropical Mexican Pacific. *Microb. Ecol.* <https://doi.org/10.1007/s00248-020-01508-7> (2020).
47. Bertagnolli, A. D., Padilla, C. C., Glass, J. B., Thamdrup, B. & Stewart, F. J. Metabolic potential and in situ activity of marine Marinimicrobia bacteria in an anoxic water column. *Environ. Microbiol.* <https://doi.org/10.1111/1462-2920.13879> (2017).
48. Ganesh, S., Parris, D. J., Delong, E. F. & Stewart, F. J. Metagenomic analysis of size-fractionated picoplankton in a marine oxygen minimum zone. *ISME J.* <https://doi.org/10.1038/ismej.2013.144> (2014).
49. Böttjer, D. & Morales, C. E. Nanoplanktonic assemblages in the upwelling area off Concepción (~36°S), central Chile: Abundance, biomass, and grazing potential during the annual cycle. *Prog. Oceanogr.* <https://doi.org/10.1016/j.pocean.2007.08.024> (2007).
50. Cuevas, L. A. & Morales, C. E. Nanoheterotroph grazing on bacteria and cyanobacteria inoxic and suboxic waters in coastal upwelling areas off northern Chile. *J. Plankton Res.* <https://doi.org/10.1093/plankt/fbi124> (2006).
51. Vargas, C. A. *et al.* Influences of riverine and upwelling waters on the coastal carbonate system off Central Chile and their ocean acidification implications. *J. Geophys. Res. Biogeosci.* <https://doi.org/10.1002/2015JG003213> (2016).
52. Feely, R. A., Sabine, C. L., Hernandez-Ayon, J. M., Ianson, D. & Hales, B. Evidence for upwelling of corrosive 'acidified' water onto the continental shelf. *Science (80-)*. <https://doi.org/10.1126/science.1155976> (2008).
53. Torres, R. *et al.* Air-sea CO<sub>2</sub> fluxes along the coast of Chile: From CO<sub>2</sub> outgassing in central northern upwelling waters to CO<sub>2</sub> uptake in southern Patagonian fjords. *J. Geophys. Res. Ocean.* <https://doi.org/10.1029/2010JC006344> (2011).
54. Michels, P. A. M., Michels, J. P. J., Boonstra, J. & Konings, W. N. Generation of an electrochemical proton gradient in bacteria by the excretion of metabolic end products. *FEMS Microbiol. Lett.* <https://doi.org/10.1111/j.1574-6968.1979.tb03339.x> (1979).
55. Mohamed, M. E. S., Seyfried, B., Tschek, A. & Fuchs, G. Anaerobic oxidation of phenylacetate and 4-hydroxyphenylacetate to benzoyl-coenzyme A and CO<sub>2</sub> in denitrifying *Pseudomonas* sp.—Evidence for an  $\alpha$ -oxidation mechanism. *Arch. Microbiol.* <https://doi.org/10.1007/BF00249036> (1993).
56. Barker, H. A. Amino acid degradation by anaerobic bacteria. *Annu. Rev. Biochem.* <https://doi.org/10.1146/annurev.bi.50.070181.000323> (1981).
57. Schneider, S., Mohamed, M. E. S. & Fuchs, G. Anaerobic metabolism of L-phenylalanine via benzoyl-CoA in the denitrifying bacterium *Thauera aromatica*. *Arch. Microbiol.* <https://doi.org/10.1007/s002030050504> (1997).
58. Debnar-Daumler, C., Seubert, A., Schmitt, G. & Heider, J. Simultaneous involvement of a tungsten-containing aldehyde: Ferredoxin oxidoreductase and a phenylacetaldehyde dehydrogenase in anaerobic phenylalanine metabolism. *J. Bacteriol.* <https://doi.org/10.1128/JB.00980-13> (2014).
59. Wakeham, S. G., Canuel, E. A. & Doering, P. H. Geochemistry of volatile organic compounds in seawater: Mesocosm experiments with 14C-model compounds. *Geochim. Cosmochim. Acta*. [https://doi.org/10.1016/0016-7037\(86\)90399-6](https://doi.org/10.1016/0016-7037(86)90399-6) (1986).
60. Lovley, D. R. *et al.* Oxidation of aromatic contaminants coupled to microbial iron reduction. *Nature* <https://doi.org/10.1038/339297a0> (1989).
61. Dolfing, J., Zeyer, J., Binder-Eicher, P. & Schwarzenbach, R. P. Isolation and characterization of a bacterium that mineralizes toluene in the absence of molecular oxygen. *Arch. Microbiol.* <https://doi.org/10.1007/BF00276528> (1990).
62. Evans, P. J., Mang, D. T., Kim, K. S. & Young, L. Y. Anaerobic degradation of toluene by a denitrifying bacterium. *Appl. Environ. Microbiol.* <https://doi.org/10.1128/aem.57.4.1139-1145.1991> (1991).
63. Rabus, R., Nordhaus, R., Ludwig, W. & Widdel, F. Complete oxidation of toluene under strictly anoxic conditions by a new sulfate-reducing bacterium. *Appl. Environ. Microbiol.* <https://doi.org/10.1128/aem.59.5.1444-1451.1993> (1993).
64. Beller, H. R., Spormann, A. M., Sharma, P. K., Cole, J. R. & Reinhard, M. Isolation and characterization of a novel toluene-degrading, sulfate-reducing bacterium. *Appl. Environ. Microbiol.* <https://doi.org/10.1128/aem.62.4.1188-1196.1996> (1996).
65. Harms, G., Rabus, R. & Widdel, F. Anaerobic oxidation of the aromatic plant hydrocarbon p-cymene by newly isolated denitrifying bacteria. *Arch. Microbiol.* <https://doi.org/10.1007/s002030050784> (1999).
66. Meckenstock, R. U., Krieger, R., Ensign, S., Kroneck, P. M. H. & Schink, B. Acetylene hydratase of *Pelobacter acetylenicus*: Molecular and spectroscopic properties of the tungsten iron-sulfur enzyme. *Eur. J. Biochem.* <https://doi.org/10.1046/j.1432-1327.1999.00600.x> (1999).
67. Zhu, B., Friedrich, S., Wang, Z., Táncsics, A. & Lueders, T. Availability of nitrite and nitrate as electron acceptors modulates anaerobic toluene-degrading communities in aquifer sediments. *Front. Microbiol.* <https://doi.org/10.3389/fmicb.2020.01867> (2020).
68. Sun, X. & Ward, B. B. Novel metagenome-assembled genomes involved in the nitrogen cycle from a Pacific oxygen minimum zone. *ISME Commun.* <https://doi.org/10.1038/s43705-021-00030-2> (2021).
69. Meckenstock, R. U. Fermentative toluene degradation in anaerobic defined syntrophic cocultures. *FEMS Microbiol. Lett.* [https://doi.org/10.1016/S0378-1097\(99\)00290-6](https://doi.org/10.1016/S0378-1097(99)00290-6) (1999).
70. Bravo-Linares, C. M. & Mudge, S. M. Analysis of volatile organic compounds (VOCs) in sediments using in situ SPME sampling. *J. Environ. Monit.* <https://doi.org/10.1039/b617215f> (2007).
71. Guigue, C. *et al.* Remobilization of polycyclic aromatic hydrocarbons and organic matter in seawater during sediment resuspension experiments from a polluted coastal environment: Insights from Toulon Bay (France). *Environ. Pollut.* <https://doi.org/10.1016/j.envpol.2017.06.090> (2017).
72. Fuenzalida, R., Schneider, W., Garcés-Vargas, J., Bravo, L. & Lange, C. Vertical and horizontal extension of the oxygen minimum zone in the eastern South Pacific Ocean. *Deep. Res. Part II Top. Stud. Oceanogr.* <https://doi.org/10.1016/j.dsr2.2008.11.001> (2009).
73. Galán, A., Faúndez, J., Thamdrup, B., Santibáñez, J. F. & Farías, L. Temporal dynamics of nitrogen loss in the coastal upwelling ecosystem off Central Chile: Evidence of autotrophic denitrification through sulfide oxidation. *Limnol. Oceanogr.* <https://doi.org/10.4319/lo.2014.59.6.1865> (2014).
74. Landry, Z., Swa, B. K., Herndl, G. J., Stepanauskas, R. & Giovannoni, S. J. SAR202 genomes from the dark ocean predict pathways for the oxidation of recalcitrant dissolved organic matter. *MBio* <https://doi.org/10.1128/mBio.00413-17> (2017).
75. Wolfe, R. S. Techniques for cultivating methanogens. *Methods Enzymol.* <https://doi.org/10.1016/B978-0-12-385112-3.00001-9> (2011).
76. McDonald, J. E., Rooks, D. J. & McCarthy, A. J. Methods for the isolation of cellulose-degrading microorganisms. *Methods Enzymol.* <https://doi.org/10.1016/B978-0-12-415931-0.00019-7> (2012).
77. Lovley, D. Dissimilatory Fe(III)- and Mn(IV)-reducing prokaryotes. *Prokaryotes Prokaryotic Physiol. Biochem.* [https://doi.org/10.1007/978-3-642-30141-4\\_69](https://doi.org/10.1007/978-3-642-30141-4_69) (2013).

78. Slack, G. C., Snow, N. H. & Kou, D. Extraction of volatile organic compounds from solids and liquids. *Sample Preparation Tech. Analyt. Chem.* <https://doi.org/10.1002/0471457817.ch4> (2003).
79. Shrivastava, A. & Gupta, V. Methods for the determination of limit of detection and limit of quantitation of the analytical methods. *Chronicles Young Sci.* <https://doi.org/10.4103/2229-5186.79345> (2011).
80. Lindroth, P. & Mopper, K. High performance liquid chromatographic determination of subpicomole amounts of amino acids by precolumn fluorescence derivatization with o-phthaldialdehyde. *Anal. Chem.* <https://doi.org/10.1021/ac50047a019> (1979).
81. Pilson, M. E. Q. An introduction to the chemistry of the sea. *Introduction Chem. Sea.* <https://doi.org/10.1017/cbo9781139047203> (2012).
82. Craig, H., Weiss, R. F. & Clarke, W. B. Dissolved gases in the equatorial and South Pacific Ocean. *J. Geophys. Res.* <https://doi.org/10.1029/jz072i024p06165> (1967).
83. Lavik, G. *et al.* Detoxification of sulphidic African shelf waters by blooming chemolithotrophs. *Nature* <https://doi.org/10.1038/nature07588> (2009).
84. Schunck, H. *et al.* Giant Hydrogen sulfide plume in the oxygen minimum zone off Peru supports chemolithoautotrophy. *PLoS ONE* <https://doi.org/10.1371/journal.pone.0068661> (2013).
85. Damgaard, L. R. & Hanselmann, K. Thermodyn©—A spread sheet for the calculation of free reaction energies under actual conditions. Based on K.W. Hanselmann. Microbial energetics applied to waste repositories. *Experientia* **47**, 645–687. <https://doi.org/10.1007/BF01958816> (1991).

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## Author contributions

B.M.S., designed experiments and fieldwork, conducted thermodynamic, statistical and chemical analyses and laboratory incubations, and drafted and edited the manuscript. S.P.G., co-designed experiments and fieldwork, supervised analyses, and edited the manuscript.

## Competing interests

The authors declare no competing interests.

## Additional information

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