scientific reports



OPEN Radular force performance of stylommatophoran gastropods (Mollusca) with distinct body masses

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The forces exerted by the animal's food processing structures can be important parameters when studying trophic specializations to specific food spectra. Even though molluscs represent the second largest animal phylum, exhibiting an incredible biodiversity accompanied by the establishment of distinct ecological niches including the foraging on a variety of ingesta types, only few studies focused on the biomechanical performance of their feeding organs. To lay a keystone for future research in this direction, we investigated the in vivo forces exerted by the molluscan food gathering and processing structure, the radula, for five stylommatophoran species (Gastropoda). The chosen species and individuals have a similar radular morphology and motion, but as they represent different body mass classes, we were enabled to relate the forces to body mass. Radular forces were measured along two axes using force transducers which allowed us to correlate forces with the distinct phases of radular motion. A radular force quotient, AFQ = mean Absolute Force/bodymass^{0.67}, of 4.3 could be determined which can be used further for the prediction of forces generated in Gastropoda. Additionally, some specimens were dissected and the radular musculature mass as well as the radular mass and dimensions were documented. Our results depict the positive correlation between body mass, radular musculature mass, and exerted force. Additionally, it was clearly observed that the radular motion phases, exerting the highest forces during feeding, changed with regard to the ingesta size: all smaller gastropods rather approached the food by a horizontal, sawing-like radular motion leading to the consumption of rather small food particles, whereas larger gastropods rather pulled the ingesta in vertical direction by radula and jaw resulting in the tearing of larger pieces.

The typical force exerted by feeding organs is a useful parameter indicating specializations to distinct food types, as it correlates with the food spectrum (see e.g.¹; for a review for stress-related puncture mechanics, see²). This topic had been studied quite intensively in vertebrates (for a summary of the relevant literature, see³): bite force analyses had been performed on mammals (e.g.^{4–8}), reptiles (e.g.^{9–12}), fish (e.g.^{13–15}), and birds (e.g.^{16,17}). Even though the majority of animal species belong to the invertebrates, unfortunately fewer work focused on the forces exerted by their structures involved in either gathering or acquiring food due to the difficulties of an experimental set-up for studies of small structures. Exceptions and pioneers in this field are studies performed on representatives of Arthropoda: spiders, crustaceans, scorpions, and insects¹⁸⁻²⁴.

For molluscs, even though they represent the second specious animal group²⁵ with around 80,000 recent species only within the Gastropoda²⁶, only a few studies approached the forces exerted by their feeding organ, the radula. Since the species belonging to this animal phylum occupy almost any marine, freshwater or terrestrial environment and established extremely varied ecological niches²⁷, accompanied with feeding on a wide range of food sources with various mechanical properties, mollusc trophic specializations are of very high interest for evolutionary biologists. Their radula, one important molluscan autapomorphy and the interface between

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Figure 1. Mollusc species used in this study. (A) Mature *Lissachatina fulica*. (B) *Helix pomatia*. (C) Immature *Lissachatina fulica*. (D) *Arion vulgaris*. (E) *Cepaea nemoralis*. Scale bar = 4 cm.

the organism and its ingesta (food, minerals), is highly diverse and offers an immense opportunity to study the structural adaptations enabling feeding on distinct ingesta types²⁸.

The radula consists of a thin, chitinous membrane with rows of embedded, sometimes mineralized teeth which is supported by thick, underlying odontophoral cartilages and moved by numerous muscles of the buccal mass. The sometimes highly complex radular motion (see e.g.^{29–34}) brings the tooth cusps in contact with the ingesta leading to the tearing, cutting, and gathering of food.

As the teeth are involved as actual organism-ingesta interface, usually the shape of teeth or overall morphology of the radula had been examined and related to the ingesta (e.g.^{35–49}; also examined for the tooth anchorage:⁵⁰). In the context of phenotypic plasticity, different shapes of radular teeth as an answer to shifts of the ingesta have also been studied^{40,51–61}. Sometimes these analyses are complemented by material property estimations of teeth^{33,62–73}. Additionally, ingesta consumption, grazing activity, food choice experiments, and fecal analyses for diverse gastropod species have been investigated relating the gastropods with their preferred food, the abundance of food or other parameters of the microhabitat (^{74–80}; for comprehensive reviews on diet of Heterobranchia, see^{81,82}).

The majority of previous studies have focused on the radular teeth themselves, but the forces exerted by this organ or its biomechanical performance have unfortunately only been investigated in a few papers devoted (1) to the feeding force calculations (⁸³; or force calculations for radula-inspired gripping devices:⁸⁴), (2) to the experiments revealing the forces needed to remove algae^{85,86}, or (3) to the first in vivo experiment performed on a single mollusc species⁸⁷.

Here, before the broad topic of trophic specialization in molluscs can be approached and, to lay a keystone for further studies, we investigated the in vivo radular forces while foraging for five different stylommatophoran species using force transducers following the protocol of⁸⁷. As the radular motion or radular morphology could potentially influence the forces exerted by the feeding organ we first selected species with a similar radular type (isodont) and similar radular motion to get a good impression on radular forces without being confronted with overflowing radular diversity, which could make results less comparable or prone to artefacts. Additionally, since the chosen gastropods however represent distinct body mass classes, we tested if and to which extend the chosen stylommatophoran specimens follow common laws and predictions for scaling of force and body mass.

Material and methods

For force measurements we have chosen five stylommatophoran species (Gastropoda: Heterobranchia) that were easy to obtain, as they are either often kept as pets or could be collected easily around Hamburg. Additionally, all these species possess a similar radular type with numerous, similar shaped (isodont), and small teeth, thus the direct influence of the tooth morphology on the forces produced is probably rather small. Also, stylommatophoran gastropods show a rather similar radular foraging motion, which can be described as a licking motion (for details on motion and radular type see e.g.^{34,87–93}). There are surely differences in motion between species, because the arrangement or thickness of radular muscles might differ. Additionally, individual gastropods of the same species might also prefer a slightly different feeding motion, but the broad cycles of the pro- and retraction are comparable.

We have chosen the following gastropods, since they can be sorted to three distinct body mass classes (see Fig. 1 and below): mature *Lissachatina fulica* (Bowdich, 1822), mature *Cepaea nemoralis* (Linnaeus, 1758), mature *Cepaea hortensis* (Müller, 1774), mature *Helix pomatia* Linnaeus, 1758, and mature *Arion vulgaris* Moquin-Tandon, 1855. Since no adult stylommatophoran species with a body mass between mature *Helix* and mature



Figure 2. Experimental set-up (drawn with Adobe Illustrator CS 6 and modified from⁸⁷) and characteristic radular force measurements. (**A**) Gastropods were placed on an acrylic platform with a hole of 2 mm diameter. The sliced food (e.g. carrot), glued to a needle, was firmly mounted to a force transducer connected with an amplifier and computer-based data acquisition and processing system. The food was stuck through the hole, so that animals could feed on it without involving their foot. (**B**) Image of the mouth opening taken through the glass platform, scale bar = 1 cm. (**C**) Characteristic radular force measurement curves, mN, of mature *Lissachatina fulica*, (above) and *Arion vulgaris* (below); left side: vertical direction (positive peaks = pulling up, negative peaks = pushing down), right side: horizontal direction (positive peaks = posterior direction, negative peaks = anterior direction). JA = jaw, FL = flour, FO = foot, LI = lip, RA = radula.

Lissachatina fulica is easily obtainable, we selected immature *Lissachatina fulica* (one juvenile stage) for experiments. Overall, 24 individuals were used; individuals of *L. fulica* were received from private animal breeders, the other species were collected in Hamburg, Germany, in May 2020. Species identification is based on the relevant literature.

The selected gastropods species were assigned to three different body mass classes: (A) large sized animals (whole body mass: 75-105 g) = mature *L. fulica* [N = 5 individuals]; (B) medium sized animals (whole body mass: 10-38 g) = *H. pomatia* and immature *L. fulica* [N = 10 individuals]; (C) small sized animals (whole body mass: 0.7-3.6 g) = *C. nemoralis*, *C. hortensis*, *A. vulgaris* [N = 9 individuals]; and cohorts (mature *L. fulica* [N = 5 individuals], immature *L. fulica* [N = 9 individuals], mature *L. fulica* [N = 5 individuals], mature *L. fulica* [N = 9 individuals], mature *L. fulica* [N = 6 individuals]. Before each experiment, individuals were weighed (body mass with shell) (see Supplementary Table 1).

Forces exerted by the radula were measured following the protocol of⁸⁷. Snails were placed on an acrylic platform with a small hole of 4 mm diameter. The platform was attached to a laboratory jack so that the height could be adjusted. Food stripes (sliced to pieces of 3 [width] × 2 [length] × 20 or 40 [height] mm; either carrot (for all species except *A. vulgaris*) or fresh strawberry (for *A. vulgaris*), depending on the specific preference of the species, were glued to a needle, which was mounted onto a force transducer FORT-10 (World Precision Instruments, Sarasota, FL, USA) and stuck through the hole so that the snail could feed on it, but without the involvement of the foot (Fig. 2). A 1000 g sensor was used for mature *L. fulica*, for all other individuals a 25 g sensor was used. Forces could only be measured in either vertical or horizontal direction, but not in both simultaneously. Thus the experimental set-up was remodeled to receive data for both directions (see also⁸⁷). The force transducers were connected to an amplifier (Biopac System, Inc., CA, USA) and a computer-based data acquisition and processing system (AcqKnowledge[™], Biopac Systems, Inc., v3.7.0.0, World Precision Instruments, Sarasota, FL,

USA). Not all force peaks were analyzed in AcqKnowledge[™] due to the large sample size, but about 30 maximal and minimal force peaks were evaluated per feeding unit, which is the time needed for eating up the part of the carrot or strawberry that was not coated with glue. For the detailed quantity of evaluated force peaks per species, individual, and direction see Supplementary Table 1; overall, 4407 force peaks were analyzed (see Fig. 2C for characteristic force peaks). Forces were either sorted to direction (negative force values = pushing, positive force values = pulling) or the absolute values summarized as Absolute Force (thus, regardless of the direction). Force values were corrected for mean body mass with shell to receive Relative Force I (force/mean body mass with shell, mN/g). Mean and standard deviations were calculated and all statistical analyses were performed with the program JMP Pro 14 (SAS Institute Inc., Cary, NC, 1989-2007) comparing the exerted forces between (a) the distinct body mass classes, (b) the cohorts, (c) the directions, (d) the individual animals. A Shapiro-Wilk test to test distribution was carried out and since data is non parametric a Kruskal-Wallis test was executed. For linear regression of Absolute Force and Relative Force I versus whole body mass, the mean and the values were displayed on logarithmic axes with excel 2013 (Microsoft Corporation, Redmond, USA) and trend lines were generated. A Radular Force Quotient, AFQ = mean Absolute Force/Bodymass^{0.67} and a Relative Force Quotient, RFQ = mean Relative Force I/Bodymass^{-0.33}, were determined. Additionally, such quotients were also determined for Forces sorted to direction/Bodymass^{-0.33} and for mean Relative Force I sorted to direction/Bodymass^{-0.33}.

Detailed radular motion, while feeding on a flat surface was documented with a Keyence VHX-500 digital microscope (KEYENCE, Neu-Isenburg, Germany) by placing the individual on an acrylic platform, providing flour paste as food (see also^{34,87}). The behavior, while foraging on a carrot, was documented with an iPad Pro (11 zoll; Apple Inc., Cupertino, USA) equipped with a 12 megapixel wide angle lens with 30 frames per second (see Supplementary Videos 1 and 2). Videos were cut and cropped with Adobe Premiere Pro 2020 (Adobe Inc., San Jose, USA).

Some animals (see Supplementary Fig. 8) were either killed by brief boiling (shelled animals) or by placing them in carbonated water (slugs). They were preserved in 70% EtOH and inventoried in the malacological collection of the Zoological Museum Hamburg (ZMH) of the Centrum für Naturkunde (CeNak); *Cepaea nemoralis*: ZMH 154748/8, *Helix pomatia*: ZMH 154749/1, *Lissachatina fulica*: ZMH 154751/2, *Cepaea hortensis*: ZMH 154754/1, *Arion vulgaris*: ZMH 154747/12.

The shells of dead specimens were removed and the soft parts were weighed to receive body mass without shell. Forces were corrected for body mass without shell to receive Relative Force II (force/body mass without shell, mN/g). To estimate the mass of the entire buccal mass (BRJ), the radula and jaw (RJ), and the buccal mass musculature (B) these specimens were dissected which was documented with a Keyence VHX-500 digital microscope (KEYENCE, Neu-Isenburg, Germany). The BRJ was first extracted, freed from surrounding tissue (see Supplementary Fig. 7) and weighed in wet condition with an accuracy weighing machine (Sartorius Cubis, MSE, Sartorius AG, Göttingen, Germany). Subsequently the radula and jaw (RJ) were separated from the buccal mass musculature (B) manually; RJ and B were weighed in wet condition. B and RJ were then dried for one week and weighed again to obtain data on dry mass. Forces were corrected for dry B to receive Relative Force III (force/ dry B mass, mN/mg) and for dry RJ to receive Relative Force IV (force/dry RJ mass, mN/mg).

For scanning electron microscope (SEM) images radulae and jaws were rewetted and cleaned with proteinase K digesting food particles according to the protocol of⁹⁴, followed by a short ultrasonic bath. Structures were mounted on SEM stubs, coated with palladium and visualized with a Zeiss LEO 1525 (One Zeiss Drive, Thornwood, USA). Radular length, width, and area could be calculated. Forces were corrected for radular area to receive Relative Force V (force/radular area, mN/mm²).

Results

Radula and its teeth. All analyzed species possess an isodont radula (Supplementary Figs. 3–6). *Lissa-chatina fulica* displays ~ 84 teeth per row (no central tooth, 16 lateral and 26 marginal teeth on each side), *Helix pomatia* ~ 87 teeth per row (one central tooth, 23 lateral and 20 marginal teeth on each side), *Arion vulgaris* ~ 75 teeth per row (one central tooth, 16 lateral and 21 marginal teeth on each side), *Cepaea nemoralis* and *C. hort-ensis* ~ 51 teeth per row (one central tooth, 12 lateral and 13 marginal teeth on each side). The jaws of all species are thick, curved, chitinous plates with ribs (i.e. odontognathous).

Radular motion. Video footage reveals the radular motion and feeding behaviour (see Supplementary Videos 1 and 2). While feeding, the radula is pushed simultaneously in ventral (vertical down) and anterior (horizontal anterior) direction, before the organ is finally pulled in dorsal (vertical up) and posterior (horizontal posterior) direction and the mouth is closed (see also⁸⁷). With the first part of the motion the radula loosens food items from the ground and collects particles, which are transported into the mouth opening in the latter phase of the feeding action. When feeding on larger ingesta (e.g. a piece of carrot; see Supplementary Video 1), the anterior part of the radula and the jaw act in concert as counter bearing squeezing and pulling the ingesta. When comparing the feeding behaviours of different individuals we can see that the large sized individuals can completely enclose the carrot piece with their lips, resulting in a dragging on the carrot in vertical direction, tearing large pieces, whereas the small and medium sized individuals are not as comfortable with this due to the small dimension of their mouth. These individuals usually nibble on edges of the item, cutting and slicing smaller pieces in rather horizontal direction employing their radula like a saw, sometimes involving additionally the foot as a clamp (Supplementary Video 2). They also drag the food in ventral direction, but this behaviour is not as pronounced and is not as forcefully as in the smaller specimens (Fig. 4). All gastropods were able to consume the food items offered, but small sized individuals needed to invest approximately 800–900% and medium sized ones 400-500% more time to consume the similar sized food items than large sized gastropods.



Figure 3. Absolute Force (blue boxplots) and Relative Force I (red boxplots) for distinct body mass classes (large-, medium-, and small-sized gastropods). (**A**) Absolute Force (regardless of the direction of measurement). (**B**) Relative Force I (regardless of the direction of measurement). (**C**) Absolute Force sorted to directions) (**D**) Relative Force I sorted to directions. Pink = pushing of radula, grey = pulling of radula.

Force measurements. Overall, 4407 individual force peaks were evaluated. For the quantity of evaluated peaks per species, individual, and direction see Supplementary Table 1. For the Absolute Forces exerted by each body mass class see Fig. 3; for the Absolute Forces exerted by each cohort see Fig. 4 and Supplementary Table 2; for the Absolute Forces of individual gastropods see Supplementary Figs. 1, 2, and Supplementary Tables 3, 4.

Force measurements of body mass classes. Comparing the Absolute Forces regardless of the direction between the three body mass classes of animals (Fig. 3A) we detect significant differences (p < 0.0001, ChiSquare: 995.6664, df: 2). Large sized individuals are capable of exerting highest forces, followed by the medium sized, and finally small sized individuals (see Table 1). When comparing the Relative Force I regardless of the direction between the three body mass classes of animals (Fig. 3B) we detect significant differences between groups (p < 0.0001, ChiSquare: 409.7297, df: 2). Here, small sized individuals exhibited the highest Relative Force I, followed by the medium sized, and finally large sized animals (see Table 1).

Absolute Forces sorted to direction (Fig. 3C) also differed significantly between body mass classes (see Table 1; for statistics see Table 2). For horizontal anterior direction, the highest forces were, however, exerted by the medium sized class, followed by the large, and finally small sized gastropods. For horizontal posterior direction, the highest forces were again exerted by the medium sized class, followed by the small sized gastropods, and finally large ones. In the direction vertical down, the large sized gastropods showed highest forces, followed by



Figure 4. Absolute Force (blue boxplots) and Relative Force I (red boxplots) for distinct cohorts. (**A**) Absolute Force (regardless of the direction of measurement). (**B**) Relative Force I (regardless of the direction of measurement). (**C**) Absolute Force sorted to directions. (**D**) Relative Force I sorted to directions. For values of C and D see Supplementary Table 2. Pink=pushing of radula, grey=pulling of radula.

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the medium, and finally small gastropods. The same is found for vertical up: large gastropods, medium, and finally small ones.

When comparing the Relative Force I sorted to direction (Fig. 3D) we again detect significant differences between body mass classes (see Table 1; for statistics see Table 2). Here, for horizontal anterior direction, the highest Relative Force I was exerted by the small sized gastropods, followed by the medium, and finally large ones. For horizontal posterior direction, the highest Relative Force I was again exerted by the small, followed by

		Absolute mN	force,		Relative I, mN/g	e force		Quantity of evaluated	
Body mass classes	Direction	Mean	±SD	Kruskal–Wallis test	Mean	±SD	Kruskal–Wallis test	force measurements	
Large sized individuals	All	73.96	62.49		0.84	0.74		1025	
	Vertical up	147.35	41.35	<i>p</i> < 0.0001, ChiSquare: 744.5827, df: 3	1.69	0.54	<i>p</i> < 0.0001, ChiSquare: 750.5548, df: 3	372	
	Vertical down	- 22.77	17.79		0.25	0.21		107	
	Horizontal posterior	38.82	19.49		0.43	0.20		323	
	Horizontal anterior	- 26.97	2.91		0.30	0.13		223	
Medium sized individuals	All	41.44	49.18		2.47	2.94		2581	
	Vertical up	14.72	10.40	<i>p</i> < 0.0001, ChiSquare:	0.92	0.65	<i>p</i> < 0.0001, ChiSquare: 1855.5616, df: 3	988	
	Vertical down	- 3.94	2.30		0.24	0.17		440	
	Horizontal posterior	74.52	49.76	1928.4320, df: 3	4.28	2.65		536	
	Horizontal anterior	- 82.22	55.10		4.98	3.59		617	
Small sized individuals	All	14.46	24.49		6.01	8.40		801	
	Vertical up	1.48	0.72	<i>p</i> < 0.0001, ChiSquare: 636.3539, df: 3	0.85	0.51	<i>p</i> < 0.0001, ChiSquare: 625.8867, df: 3	200	
	Vertical down	- 0.86	0.44		0.50	0.28		210	
	Horizontal posterior	39.56	35.37		13.63	8.67		211	
	Horizontal anterior	- 15.34	7.87		9.26	9.08		180	

Table 1. For body mass classes: absolute force, mean \pm SD (mN), obtained during measurements in bothdirections (vertical up, vertical down, horizontal posterior, horizontal anterior), and relative force I, mean \pm SD(mN/g), with results from Kruskal–Wallis test, and quantity of evaluated radular force measurements (see alsoFig. 4). SD, standard deviation.

	Direction	p	ChiSquare	df					
Body mass classes									
	Horizontal anterior			11					
Ab a shut famaaa	Horizontal posterior	<0.0001	3569.9093						
Absolut loices	Vertical down	< 0.0001							
	Vertical up								
	Horizontal anterior			11					
Deletine ferrer I	Horizontal posterior	<0.0001	3455.7572						
Relative force I	Vertical down	< 0.0001							
	Vertical up								
Cohorts									
	Horizontal anterior			23					
Ab a shut famaaa	Horizontal posterior	<0.0001	2674 4421						
Absolut forces	Vertical down	< 0.0001	30/4.4431						
	Vertical up								
	Horizontal anterior			23					
Relative force I	Horizontal posterior	< 0.0001	2610 2024						
Relative lorce I	Vertical down	< 0.0001	5019.3924						
	Vertical up	1							

 Table 2.
 For body mass classes and cohorts: results from Kruskal–Wallis test for absolute force and relative force I.

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the medium, and finally large sized class individuals. In the direction vertical down, the small sized gastropods showed highest Relative Force I, followed by the large, and finally medium gastropods. For vertical up direction, the large gastropods exerted highest Relative Force I, followed by the medium, and finally small ones.

Force measurements of cohorts. When comparing Absolute Forces regardless of the direction between cohorts (Fig. 4A) we found significant differences (p < 0.0001, ChiSquare: 1026.4480, df: 5). Highest forces were exerted by mature *L. fulica*, followed by *H. pomatia*, immature *L. fulica*, *C. nemoralis*, *A. vulgaris*, and finally *C. hortensis* (see Supplementary Table 2). When comparing Relative Force I regardless of the direction between cohorts (Fig. 4B) we found significant differences (p < 0.0001, ChiSquare: 499.1690, df: 5). Highest Relative Force I was exerted by *C. hortensis*, followed by *C. nemoralis*, *A. vulgaris*, immature *L. fulica*, *H. pomatia*, and finally mature *L. fulica* (see Supplementary Table 2).

Comparing the Absolute Forces sorted to direction between cohorts we found significant differences (Fig. 4B; for values see Supplementary Table 2; for statistics see Table 2). For horizontal anterior direction, the highest forces were exerted by *H. pomatia*, followed by the immature *L. fulica*, mature *L. fulica*, *A. vulgaris*, *C. hortensis*, and finally *C. nemoralis*. For horizontal posterior direction, the highest forces were exerted by *H. pomatia*, followed by *C. nemoralis*, the immature *L. fulica*, *A. vulgaris*, and finally *C. hortensis*. For vertical down direction, the highest forces were exerted by mature *L. fulica*, *A. vulgaris*, and finally *C. hortensis*. For vertical down direction, the highest forces were exerted by mature *L. fulica*, followed by *H. pomatia*, the immature *L. fulica*, *A. vulgaris*, *C. nemoralis*, and finally *C. hortensis*. For vertical up direction, the highest forces were exerted by mature *L. fulica*, followed by *H. pomatia*, the immature *L. fulica*, *A. vulgaris*, *C. nemoralis*, and finally *C. hortensis*. For vertical up direction, the highest forces were exerted by mature *L. fulica*, followed by the immature *L. fulica*, *H. pomatia*, *C. nemoralis*, and finally *C. hortensis*. For vertical up direction, the highest forces were exerted by mature *L. fulica*, followed by the immature *L. fulica*, *H. pomatia*, *C. nemoralis*, and finally *C. hortensis*.

When comparing the Relative Force I sorted to direction between cohorts we found significant differences (Fig. 4C; for values see Supplementary Table 2; for statistics see Table 2). For horizontal anterior, the highest Relative Force I was exerted by *C. hortensis*, followed by *A. vulgaris*, the immature *L. fulica*, *C. nemoralis*, *H. pomatia*, and finally mature *L. fulica*. For horizontal posterior direction, the highest Relative Force I was exerted by *C. hortensis*, *A. vulgaris*, the immature *L. fulica*, *H. pomatia*, and finally mature *L. fulica*. For horizontal posterior direction, the highest Relative Force I was exerted by *C. nemoralis*, followed by *C. hortensis*, *A. vulgaris*, the immature *L. fulica*, *H. pomatia*, and finally mature *L. fulica*. For vertical down direction, the highest Relative Force I was exerted by *C. hortensis*, followed by *A. vulgaris*, the immature *L. fulica*, mature *L. fulica*, *C. nemoralis*, and finally *H. pomatia*. For vertical up direction, the highest Relative Force I was exerted by mature *L. fulica*, followed by *C. hortensis*, the immature *L. fulica*, *A. vulgaris*, *c. nemoralis*, and finally *H. pomatia*. For vertical up direction, the highest Relative Force I was exerted by mature *L. fulica*, followed by *C. hortensis*, the immature *L. fulica*, *A. vulgaris*, *C. nemoralis*, and finally *H. pomatia*.

Masses of body, radula, buccal mass musculature, and radular sizes. Highest whole body mass (see Supplementary Table 1) was measured for *L. fulica* mature number (no.) 1, followed by *H. pomatia*, L. *fulica* immature no. 9, *C. nemoralis* no. 1, *A. vulgaris* no. 2, 4, 5, *C. nemoralis* no. 2, *A. vulgaris* no. 1, and finally *C. hortensis*.

Overall, we found that the body mass (with and without shell) relates in proportion to the masses of the whole buccal mass (wet; BRJ), the radular musculature (buccal mass musculature, wet and dry; B), and the radula and jaw (wet and dry; RJ). When individuals were heavier, they usually possessed higher muscle mass, a heavier radula and jaw (see Supplementary Fig. 8 and Supplementary Table 6). Exceptions were: *L. fulica* immature no. 9 (18 g body mass and 205.01 mg whole buccal mass) and *H. pomatia* (38 g body mass and 163.30 mg whole buccal mass), *C. nemoralis* no. 1 (3.60 g body mass and 27.20 mg whole buccal mass), and *A. vulgaris* no. 2 (3.50 g body mass and 56.66 mg whole buccal mass). Comparing mature (mature 1: 78.00 g body mass and 286.80 mg BRJ) and immature *L. fulica* (immature no. 9: 18.00 g body mass and 205.01 mg BRJ) we found that the body mass increases for the factor ~4 and BRJ increases for the factor 1.4.

We found that smaller gastropods are capable of exerting higher forces per whole buccal mass, radular muscle, and radula and jaw mass, dry as well as wet, (see Supplementary Fig. 9 and Supplementary Table 6). *Cepaea hortensis* exerted the highest force per radular musculature mass, followed by *C. nemoralis* no. 1, *A. vulgaris* no. 4 and 5, *C. nemoralis* no. 2, *H. pomatia*, *L. fulica* mature no. 1, *L. fulica* immature no. 9, and finally *A. vulgaris* no. 1 and 2. The same sequence was also found for force per body mass without shell. With the exception of *A. vulgaris* no. 4 and 5, exerting the highest forces per radula and jaw mass, we detected the same order for this parameter.

The radular length and width (see Supplementary Fig. 8 and Supplementary Table 5) do not consistently correlate with body mass, the highest radular width was measured for *L. fulica* mature no. 1, followed by *A. vulgaris* no. 5, *L. fulica* immature no. 9, *H. pomatia*, *A. vulgaris* no. 4, *A. vulgaris* no. 1, *A. vulgaris* no. 2, *C. nemoralis* no. 2, *C. nemoralis* no. 1, and finally *C. hortensis*. Both gastropods with the largest width possessed shorter radulae than all other specimens. *L. fulica* immature no. 9 possessed the largest radular area, followed by *H. pomatia*, *L. fulica* mature no. 1, *A. vulgaris* no. 4, *A. vulgaris* no. 2, *A. vulgaris* no. 5, *C. nemoralis* no. 1, *A. vulgaris* no. 1, *L. fulica* mature no. 1, *A. vulgaris* no. 4, *A. vulgaris* no. 2, *A. vulgaris* no. 5, *C. nemoralis* no. 1, *A. vulgaris* no. 1, *C. nemoralis* no. 2, and finally *C. hortensis*. The radular area again did not consistently correlate with the whole body mass.

Force quotients. We found that the stylommatophoran species perfectly follow the predictions for scaling of force and body mass as the mean Absolute radular Force (regardless of the direction) scales to body mass^{0.67} with the quotient 4.25 and the mean Relative Force I (regardless of the direction) scales to body mass^{-0.33} with the quotient 4.35 (Fig. 5A,B). When the variance of the Absolute Force (regardless of the direction) is scaled to body mass^{0.67} and the variance of the Relative Force I (regardless of the direction) is scaled to body mass^{-0.33} we receive quotients of 3 (Fig. 5C,D). When the forces are sorted to directions the picture is rather puzzling: for horizontal anterior a quotient of 27 (Fig. 6A,B), for horizontal posterior a quotient of 31–32 (Fig. 6C,D), for vertical down a quotient of 2.4 (Fig. 6E,F), and for vertical up a quotient of 0.54 (Fig. 6G,H) is calculated.

Discussion

Force output is often referred to as proportional to muscle mass^{0.67}, the muscle cross-sectional area^{95,96}, or to body mass^{0.6797} whereas the forces, corrected for body mass (force/mass), are referred to be proportional to body mass^{-0.3397}. The stylommatophoran species examined here, follow the predictions for scaling of force and body mass for mean values. Following previous studies^{22,98} we here experimentally determined a Radular Force Quotient AFQ, AFQ = mean Absolute Force/Bodymass^{0.67}, of 4.25 and a Relative Force Quotient RFQ, RFQ = mean Relative Force I/Bodymass^{-0.33} of 4.35 which can be further used for predictions of forces in Gastropoda. However, when the forces are sorted to directions the picture becomes puzzling, indicating the need for further studies.

The here measured Absolute feeding Forces for *Helix pomatia* are in a similar range as to those documented by⁸⁷ for *Cornu aspersum*, both gastropod groups have comparable body mass. We detected that gastropods with a higher body mass and a larger body size were capable of exerting higher radular forces, which is not surprising.



Figure 5. Linear regression, displayed on logarithmic axes, with trend lines (black = real trend line; red = calculated trend line for the factors 4.25 and 4.35), regardless of the direction. (**A**) Mean Absolute Force *versus* mean whole body mass. A radular force quotient, AFQ = mean Absolute Force/Bodymass^{0.67}, of 4.25 was determined. (**B**) Mean Relative Force I *versus* mean whole body mass. A Relative Force I *versus* mean whole body mass. A Relative Force I/Bodymass^{-0.33}, of 4.35 was determined. (**C**) Variance of Absolute Force *versus* mean whole body mass. (**D**) Variance of Relative Force I *versus* mean whole body mass. For C and D a force quotient of 3.00 was determined.

However, the relationship between forces generated by the feeding organ and body size is well documented for vertebrates, but not for molluscs. When forces are corrected for body size or mass, ecological adaptations related with this parameter usually became more pronounced (e.g.^{4,7,16,99,100}; for hypotheses about body size related evolution of bite force, see also^{101,102}). When determining the radular force per body mass (termed Relative Force I), we see that smaller gastropods were capable of exerting the highest forces, followed by the medium, and finally large ones, similar to e.g. carnivorans in placental mammals⁷. This is also not surprising, because of different scaling of body mass and cross-sectional area of muscles^{103,104}. Usually, the sampled gastropod specimens exhibiting a higher body mass also possess a proportionally higher mass of buccal mass (correlation between buccal mass size and gastropod body size was also previously described by⁵¹). However, the immature and adult *Lissachatina* fulica, exhibiting strong differences in body mass and size, but having almost similar buccal mass sizes and masses, are an exception to this rule. In most experiments mature L. fulica exerted higher radular forces than the immature gastropods, but some measurements (outliers) revealed that immature L. fulica are also capable of generating the same feeding forces as the mature ones. This could be an indication that the ingesta type does not change during ontogeny; however, this aspect awaits further investigation. But our analyses of Relative Force I (Fig. 4) reveal distinct radular force patterns for cohorts, both Cepaea species have a wide range of exerted forces, followed by Arion, Lissachatina immature, and finally, with the smallest range, Lissachatina mature. This could be an indication that species have specific radular forces. Cepaea potentially feeds naturally on a broader food spectrum whereas Lissachatina is more restricted, but this also awaits further investigations. Arion was fed with fresh strawberries, since it did not want to feed on carrots. This could have influenced the forces generated; potentially Arion is capable of exerting higher forces. It should be additionally stated that here the whole body masses were related to the radular forces generated. But, as we experimented with four snail species and one slug species (Arion vulgaris), whole body masses are not directly comparable, because the mass of the shell could cause artefacts. In future studies the body mass without shell should be determined persistently to detect a potentially more accurate relationship or even correlation.

In past studies, it has been shown that the forces generated usually correlate with the muscle mass, muscle size, or muscle diameter (for vertebrates: e.g.^{8,16,105,106}; for invertebrates:¹⁰⁷). This is congruent to the here observed patterns in gastropods. However, for precise interpretation of the relationship between buccal mass size and the mass of the radular muscles (buccal mass musculature) knowledge about the functional role of each feeding muscle must be available, which is not the case for the taxa studied here. There are some detailed studies analyzing the in vivo buccal mass movement and buccal mass muscle function in *Aplysia*^{89,90,92,108}. But since *Aplysia* belongs to the Opisthobranchia and is not closely related to the taxa involved in this experiment



Figure 6. Linear regression, displayed on logarithmic axes, with trend lines (black = real trend line; red = calculated trend line for the factors 4.25 and 4.35), for each direction (**A**–**B**: horizontal anterior, **C**–**D**: horizontal posterior, **E**–**F**: vertical down, **G**–**H**: vertical up). Left side (**A**, **C**, **E**, **G**). Variance of Absolute Force *versus* mean whole body mass. Right side (**B**, **D**, **F**, **H**). Variance of Relative Force I *versus* mean whole body mass.

the function of the buccal mass muscles for these stylommatophoran taxa cannot be assigned yet. However, to deeply understand the relationship between measured radular forces involved in specific radular motions, the function of each muscle, their work in concert, as well as the muscle and fiber size, length, or diameter need to be investigated. As there are many different parameters that relate with feeding force (for vertebrates: e.g. the skull geometry and size:^{10,17,99,109-111}; or muscular development:¹⁰⁵), it is very difficult to produce reliable models for molluscs due to the lack of solid experimental data. Additionally, studies on invertebrates reveal that muscle stress varies considerably depending on the muscle¹¹²⁻¹¹⁴. This could also be the case for molluscs as the forces exerted per radular muscular mass differ extremely between the analyzed stylommatophoran individuals (see Supplementary Fig. 9 and Supplementary Table 6). This again shows that pure anatomy-based studies on the muscle systems of invertebrates do not necessarily provide data on physiology (see also²⁴). Additionally, the area of the radula used for foraging could influence results, but the working area could so far only be determined by involving sandpapers of different roughness¹¹⁵. Since sandpaper is a rather flat surface in comparison to a slice of carrot we cannot directly translate our past findings to the experiments here.

Our results clearly indicate that larger and heavier animals exerted higher forces in the vertical directions, whereas medium and small individuals exerted higher forces in horizontal directions. This is additionally supported by the analyses of the video footage showing that larger animals rather pull with radula and jaw, whereas smaller individuals use their radular often like a saw in anterior–posterior direction when approaching the food item (see Supplementary Video 1 and 2). This shift in feeding behavior seems to depend on the ingesta size in relation to the mouth opening size. Smaller and medium sized gastropods—even though capable of embracing the whole item with the lip—seem to prefer this alternative feeding pattern. An ingesta-depending shift in feeding pattern, i.e. dynamics of swallowing, had been documented for other gastropod taxa when altering parameters of the ingesta¹⁰⁸, e.g. its hardness¹¹⁶, its load and width¹¹⁷, or its size^{118–120}. Additionally, gastropods of different sizes have been found to feed on different ingesta types, possibly correlating with ontogenetic size changes of the mouthparts¹²¹.

As already stated above, heavier gastropods were usually capable of exerting higher radular forces than smaller ones (except for immature and adult *L. fulica*). This indicates that larger gastropods might forage on a broader spectrum of ingesta, as it had been previously described for turtles¹⁰⁹. However, the smaller individuals were able to exert higher forces per body mass and additionally show a distinct radular motion pattern resulting in a distinct feeding pattern. All experiments resulted in the consumption of the ingesta offered, but smaller gastropods invested more time. This effect had also been observed for lizards¹²² and should be further investigated for Mollusca. Thus, we can conclude that in studies of feeding forces of gastropods with a similar radular type, teeth, and radular motion, adaptations to ingesta might only been detected by studying feeding efficiency and time invested. We hope that in the future more biomechanical, physiological and functional morphological studies will approach the topic of trophic specialization in molluscs via feeding force experiments on species possessing distinct radular morphologies and feeding on more types of ingesta.

Data availability

The datasets generated and analysed during the current study (the force measurements) are available from the corresponding author on reasonable request; all other datasets are included in this published article (and its Supplementary Information files).

Received: 20 January 2021; Accepted: 4 May 2021 Published online: 18 May 2021

References

- 1. Kiltie, R. Bite force as a basis for niche differentiation between rain forest peccaries (*Tayassu tajacu* and *T. pecari*). *Biotropica* 14(3), 188–195. https://doi.org/10.2307/2388025 (1982).
- Anderson, P. S. L. Making a point: Shared mechanics underlying the diversity of biological puncture. J. Exp. Biol. 221, jeb187294. https://doi.org/10.1242/jeb.187294 (2018).
- 3. Lappin, A. K. & Jones, M. H. E. Reliable quantification of bite-force performance requires use of appropriate biting substrate and standardization of bite out-lever. J. Exp. Biol. 217, 4303–4312. https://doi.org/10.1242/jeb.106385 (2014).
- Aguirre, L. F., Herrel, A., van Damme, R. & Matthysen, E. Ecomorphological analysis of trophic niche partitioning in a tropical savannah bat community. Proc. R. Soc. B 269(1497), 1271–1278. https://doi.org/10.1098/rspb.2002.2011 (2002).
- Wroe, S., McHenry, C. & Thomason, J. Bite club: comparative bite force in big biting mammals and the prediction of predatory behaviour in fossil taxa. Proc. R. Soc. B 272(1563), 619–625. https://doi.org/10.1098/rspb.2004.2986 (2005).
- Christiansen, P. Evolutionary implications of bite mechanics and feeding ecology in bears. J. Zool. Lond. 272(4), 423–443. https:// doi.org/10.1111/j.1469-7998.2006.00286.x (2007).
- Christiansen, P. & Wroe, S. Bite forces and evolutionary adaptations to feeding ecology in carnivores. *Ecology* 88(2), 347–358. https://doi.org/10.1890/0012-9658(2007)88[347:bfaeat]2.0.co;2 (2007).
- Herrel, A., De Smet, A., Aguirre, L. F. & Aerts, P. Morphological and mechanical determinants of bite force in bats: Do muscles matter?. J. Exp. Biol. 211, 86–91. https://doi.org/10.1242/jeb.012211 (2008).
- Herrel, A. & O'Reilly, J.C., Richmond, A.M., Evolution of bite performance in turtles. J. Evol. Biol. 15(6), 1083–1094. https:// doi.org/10.1046/j.1420-9101.2002.00459.x (2002).
- Herrel, A. & O'Reilly, J. C. Ontogenetic scaling of bite force in lizards and turtles. *Physiol. Biochem. Zool.* 79(1), 31–42. https://doi.org/10.1086/498193 (2006).
- 11. Vervust, B., Brecko, J. & Herrel, A. Temperature effects on snapping performance in the common snapper *Chelydra serpentine* (Reptilia, Testudines). J. Exp. Zool. A **315**(1), 41–47. https://doi.org/10.1002/jez.650 (2011).
- Kaliontzopoulou, A., Adams, D. C., Van der Meijden, A., Perera, A. & Carretero, M. A. Relationships between head morphology, bite performance and ecology in two species of *Podarcis* wall lizards. *Evol. Ecol.* 26, 825–845. https://doi.org/10.1007/s10682-011-9538-y (2012).
- Huber, D. R., Eason, T. G., Hueter, R. E. & Motta, P. J. Analysis of the bite force and mechanical design of the feeding mechanism of the durophagous horn shark *Heterodontus francisci. J. Exp. Biol.* 208, 3553–3571. https://doi.org/10.1242/jeb.01816 (2005).

- Huber, D. R., Weggelaar, C. L. & Motta, P. J. Scaling of bite force in the blacktip shark *Carcharhinus limbatus*. Zoology 109(2), 109–119. https://doi.org/10.1016/j.zool.2005.12.002 (2006).
- Huber, D. R., Dean, M. N. & Summers, A. P. Hard prey, soft jaws and the ontogeny of feeding mechanics in the spotted ratfish Hydrolagus colliei. J. R. Soc. Interface 5(25), 941–952. https://doi.org/10.1098/rsif.2007.1325 (2008).
- Van der Meij, M. A. A. & Bout, R. G. Scaling of jaw muscle size and maximal bite force in finches. J. Exp. Biol. 207, 2745–2753. https://doi.org/10.1242/jeb.01091 (2004).
- Herrel, A., Podos, J., Huber, S. K. & Hendry, A. P. Bite performance and morphology in a population of Darwin's finches: Implications for the evolution of beak shape. *Funct. Ecol.* 19(1), 43–48. https://doi.org/10.1111/j.0269-8463.2005.00923.x (2005).
- Wheater, C. P. & Evans, M. E. G. The mandibular forces and pressures of some predacious Coleoptera. J. Insect. Physiol. 35(11), 815–820. https://doi.org/10.1016/0022-1910(89)90096-6 (1989).
- Taylor, G. M. Maximum force production: Why are crabs so strong?. Proc. R. Soc. B 267(1451), 1475–1480. https://doi.org/10. 1098/rspb.2000.1167 (2000).
- Claussen, D. L., Gerald, G. W., Kotcher, J. E. & Miskell, C. A. Pinching forces in crayfish and fiddler crabs, and comparisons with the closing forces of other animals. J. Comp. Physiol. B 178(3), 333–342. https://doi.org/10.1007/s00360-007-0226-8 (2008).
- Van der Meijden, A., Herrel, A. & Summers, A. Comparison of chela size and pincer force in scorpions; getting a first grip. J. Zool. 280(4), 319–325. https://doi.org/10.1111/j.1469-7998.2009.00628.x (2010).
- Van der Meijden, A., Langer, F., Boistel, R., Vagovic, P. & Heethoff, M. Functional morphology and bite performance of raptorial chelicerae of camel spiders (Solifugae). J. Exp. Biol. 215, 3411–3418. https://doi.org/10.1242/jeb.072926 (2012).
- Goyens, J., Dirckx, J., Dierick, M., Van Hoorebeke, L. & Aerts, P. Biomechanical determinants of bite force dimorphism in Cyclommatus metallifer stag beetles. J. Exp. Biol. 217, 1065–1071. https://doi.org/10.1242/jeb.091744 (2014).
- Weihmann, T., Reinhardt, L., Weissing, K., Siebert, T. & Wipfler, B. Fast and powerful: Biomechanics and bite forces of the mandibles in the American cockroach *Periplaneta Americana*. *PLoS ONE* 10(11), e0141226. https://doi.org/10.1371/journal. pone.0141226 (2015).
- 25. Chapman, A. D. *Numbers of Living Species in Australia and the World*. 2nd edition. (Australian Biodiversity Information Services, Toowoomba, Australia, 2009).
- 26. Bouchet, P. et al. Classification and nomenclator of gastropod families. Malacologia 47(1-2), 1-368 (2005).
- 27. Wanninger, A. & Wollesen, T. The evolution of molluscs. Biol. Rev. 94, 102-115. https://doi.org/10.1111/brv.12439 (2019).
- Padilla, D. K. Form and function of radular teeth of herbivorous molluscs: Focus on the future. Am. Malacol. Bull. 18(1), 163–168 (2003).
- 29. Ankel, W. E. Erwerb und Aufnahme der Nahrung bei den Gastropoden. Verh. Dtsch. Zool. Ges. Zool. Anz. 11, 223-295 (1938).
- 30. Eigenbrodt, H. Untersuchungen über die Funktion der Radula einiger Schnecken. Z. Morphol. Oekol. Tiere 37, 735–791 (1941).
- Crampton, D. Functional anatomy of the buccal apparatus of Onchidoris bilamellata (Mollusca: Opisthobranchia). Trans. Zool. Soc. Lond. 34, 45–86. https://doi.org/10.1111/j.1096-3642.1977.tb00372.x (1977).
- Wägele, H. Rasterelektronenmikroskopische Untersuchungen an Radulae einiger Nordseeschnecken (Gastropoda: Prosobranchia) mit Anmerkungen zur Funktionsmorphologie. Drosera 83(1), 68–78 (1983).
- Hawkins, S. J. et al. A comparison of feeding mechanisms in microphagous, herbivorous, intertidal, prosobranchs in relation to resource partitioning. J. Molluscan Stud. 55(2), 151–165. https://doi.org/10.1093/mollus/55.2.151 (1989).
- Scheel, C., Gorb, S. N., Glaubrecht, M. & Krings, W. Not just scratching the surface: Distinct radular motion patterns in Mollusca. *Biol. Open* 9, bio.055699. https://doi.org/10.1242/bio.055699 (2020).
- 35. Solem, A. Patterns of radular tooth structure in carnivorous land snails. Veliger 17, 81-88 (1974).
- 36. Walsby, J. R. Feeding and the radula in the marine pulmonate limpet, Trimusculus reticulatus. Veliger 18, 139-145 (1975).
- 37. Jensen, K. R. A review of sacoglossan diets, with comparative notes on radular and buccal anatomy. *Malacol. Rev.* 13, 55-77 (1980).
- 38. Jensen, K. R. Observations on feeding methods in some Florida ascoglossans. J. Molluscan Stud. 47(2), 190–199 (1981).
- Jensen, K. R. Factor affecting feeding selectivity in herbivorous Ascoglossa (Mollusca: Opisthobranchia). J. Exp. Mar. Biol. Ecol. 66(2), 135–148. https://doi.org/10.1016/0022-0981(83)90035-7 (1983).
- Jensen, K. R. Morphological adaptations and plasticity of radular teeth of the Sacoglossa (= Ascoglossa) (Mollusca: Opisthobranchia) in relation to their food plants. *Biol. J. Linn. Soc.* 48(2), 135–155. https://doi.org/10.1111/j.1095-8312.1993.tb00883.x (1993).
- Jensen, K. R. Evolution of the Sacoglossa (Mollusca, Opisthobranchia) and the ecological associations with their food plants. *Evol. Ecol.* 11, 301–335. https://doi.org/10.1023/A:1018468420368 (1997).
- Steneck, R. S. & Watling, L. Feeding capabilities and limitation of herbivorous molluscs: A functional group approach. *Mar. Biol.* 68, 299–319. https://doi.org/10.1007/BF00409596 (1982).
- 43. Burch, J. B. & Jeong, K. H. The radula teeth of selected Planorbidae. Malacol. Rev. 17(1-2), 67-84 (1984).
- Kesler, D. H., Jokinen, E. H. & Munns, W. R. Jr. Trophic preferences and feeding morphology of two pulmonate snail species from a small New England pond, USa. *Can. J. Zool.* 64(11), 2570–2575. https://doi.org/10.1139/z86-377 (1986).
- Black, R., Lymbery, A. & Hill, A. Form and function: Size of radular teeth and inorganic content of faeces in a guild of grazing molluscs at Rottnest Island, Western Australia. J. Exp. Mar. Biol. Ecol. 121(1), 23–35. https://doi.org/10.1016/0022-0981(88) 90021-4 (1988).
- Nishi, M. & Kohn, A. J. Radular teeth of Indo-Pacific molluscivorous species of *Conus*: A comparative analysis. J. Molluscan Stud. 65, 483–497. https://doi.org/10.1093/mollus/65.4.483 (1999).
- Duda, T. F., Kohn, A. J. & Palumbi, S. R. Origins of diverse feeding ecologies within *Conus*, a genus of venomous marine gastropods. *Biol. J. Linn. Soc.* 73, 391–409. https://doi.org/10.1006/bijl.2001.054 (2001).
- Rintelen, T. V., Wilson, A. B., Meyer, A. & Glaubrecht, M. Escalation and trophic specialization drive adaptive radiation of freshwater gastropods in ancient lakes on Sulawesi, Indonesia. Proc. R. Soc. B 271(1557), 2541–2549. https://doi.org/10.1098/ rspb.2004.2842 (2004).
- 49. Ramesh, R. & Ravichandran, S. Feeding biology with reference to algal preference and scanning electron microscopy studies on the radula of *Turbo brunneus*. *Trends Appl. Sci. Res.* **3**(2), 189–195. https://doi.org/10.3923/tasr.2008.189.195 (2008).
- Krings, W., Brütt, J.-O., Gorb, S. N. & Glaubrecht, M. Tightening it up: Diversity of the chitin anchorage of radular teeth in paludomid freshwater-gastropods. *Malacologia* 63(1), 77–94. https://doi.org/10.4002/040.063.0108 (2020).
- Bleakney, J. S. Indirect evidence of a morphological response in the radula of *Placida dentritica* (Alder and Hancock, 1843) (Opisthobranchia, Ascoglossa/Sacoglossa) to different algae prey. *Veliger* 33(1), 111–115 (1990).
- 52. Padilla, D. K., Dilger, E. K. & Dittmann, D. E. Phenotypic plasticity of feeding structures in species of *Littorina*. Am. Zool. 40(6), 1161–1161 (2000).
- Padilla, D. K., Dittman, D. E., Franz, J. & Sladek, R. Radula production rates in two species of *Lacuna* Turton (Gastropoda: Littorinidae). *J. Molluscan Stud.* 62(3), 275–280. https://doi.org/10.1093/mollus/62.3.275 (1996).
- 54. Padilla, D. K. Inducible phenotypic plasticity of the radula in *Lacuna* (Gastropoda: Littorinidae). *Veliger* 41(2), 201–204 (1998).
- 55. Padilla, D. K. Food and environmental cues trigger an inducible offense. *Evol. Ecol. Res.* **3**(1), 15–25 (2001).
- Reid, D. G. & Mak, Y. M. Indirect evidence for ecophenotypic plasticity in radular dentition of *Littoraria* species (Gastropoda: Littorinidae). J. Molluscan Stud. 65(3), 355–370. https://doi.org/10.1093/mollus/65.3.355 (1999).

- Trussell, G. C. Phenotypic clines, plasticity, and morphological trade-offs in an intertidal snail. *Evolution* 54(1), 151–166. https:// doi.org/10.1111/j.0014-3820.2000.tb00016.x (2000).
- Ito, A., Ilano, A. S. & Nakao, S. Seasonal and tidal height variations in body weight and radular length in Nodilittorina radiata (Eydoux & Souleyet, 1852). J. Molluscan Stud. 68(3), 197–203. https://doi.org/10.1093/mollus/68.3.197 (2002).
- Andrade, S. C. S. & Solferini, V. N. The influence of size on the radula of *Littoraria angulifera* (Gastropoda: Littorinidae). *Malacologica* 49(1), 1–5. https://doi.org/10.4002/1543-8120-49.1.1 (2006).
- Grünbaum, D. & Padilla, D. K. An integrated modeling approach to assessing linkages between environment, organism, and phenotypic plasticity. *Integr. Comp. Biol.* 54(2), 323–335. https://doi.org/10.1093/icb/icu045 (2014).
- Molis, M., Scrosati, R. A., El-Belely, E. F., Lesniowski, T. J. & Wahl, M. Wave-induced changes in seaweed toughness entail plastic modifications in snail traits maintaining consumption efficacy. J. Ecol. 103(4), 851–859. https://doi.org/10.1111/1365-2745.12386 (2015).
- Breure, A. S. H. & Gittenberger, E. The rock-scraping radula, a striking case of convergence (Mollusca). Neth. J. Zool. 32(3), 307–312. https://doi.org/10.1163/002829681X00347 (1981).
- Weaver, J. C. et al. Analysis of an ultra hard magnetic biomineral in chiton radular teeth. Mater. Today 13(1-2), 42-52. https://doi.org/10.1016/S1369-7021(10)70016-X (2010).
- Lu, D. & Barber, A. H. Optimized nanoscale composite behaviour in limpet teeth. J. R. Soc. Interface 9(71), 1318–1324. https:// doi.org/10.1098/rsif.2011.0688 (2012).
- Grunenfelder, L. K. et al. Biomineralization: Stress and damage mitigation from oriented nanostructures within the radular teeth of Cryptochiton stelleri. Adv. Funct. Mater. 24(39), 6085. https://doi.org/10.1002/adfm.201470255 (2014).
- Barber, A. H., Lu, D. & Pugno, N. M. Extreme strength observed in limpet teeth. J. R. Soc. Interface 12(105), 20141326. https:// doi.org/10.1098/rsif.2014.1326 (2015).
- Ukmar-Godec, T. *et al.* Materials nanoarchitecturing via cation-mediated protein assembly: Making limpet teeth without mineral. *Adv. Mater.* 29(27), 1701171. https://doi.org/10.1002/adma.201701171 (2017).
- Ukmar-Godec, T., Kapun, G., Zaslansky, P. & Faivre, D. The giant keyhole limpet radular teeth: A naturally-grown harvest machine. J. Struct. Biol. 192(3), 392–402. https://doi.org/10.1016/j.jsb.2015.09.021 (2015).
- Krings, W., Kovalev, A., Glaubrecht, M. & Gorb, S. N. Differences in the Young modulus and hardness reflect different functions of teeth within the taenioglossan radula of gastropods. *Zoology* 137, 125713. https://doi.org/10.1016/j.zool.2019.125713 (2019).
- Krings, W., Marcé-Nogué, N., Karabacak, H., Glaubrecht, M. & Gorb, S. N. Finite element analysis of individual taenioglossan radula teeth. Acta Biomater. 115, 317–332. https://doi.org/10.1016/j.actbio.2020.08.034 (2020).
- Krings, W. Trophic specialization of paludomid gastropods from 'ancient' Lake Tanganyika reflected by radular tooth morphologies and material properties. Dissertation, Universität Hamburg. https://ediss.sub.uni-hamburg.de/handle/ediss/8654 (2020).
- Krings, W., Kovalev, A. & Gorb, S. N. Influence of water content on mechanical behaviour of gastropod taenioglossan radulae. Proc. R. Soc. B. 288, 20203173 https://doi.org/10.1098/rspb.2020.3173 (2021).
- Krings, W., Neiber, M. T., Kovalev, A., Gorb, S. N. & Glaubrecht, M. Trophic specialisation reflected by radular tooth material properties in an 'ancient' Lake Tanganyikan gastropod species flock. *BMC Ecol. Evol.* 21, 35. https://doi.org/10.1186/s12862-021-01754-4 (2021).
- 74. Hayes, T. The influence of diet on local distributions of Cypraea. Pac. Sci. 37(1), 27-36 (1983).
- 75. Padilla, D. K. The importance of form: Differences in competitive ability, resistance to consumers and environmental stress in an assemblage of coralline algae. *J. Exp. Mar. Biol. Ecol.* **79**(2), 105–127. https://doi.org/10.1016/0022-0981(84)90213-2 (1984).
- Blinn, W., Truitt, R. E. & Pickart, A. Feeding ecology and radular morphology of the freshwater limpet *Ferrissia fragilis. J. N. Am. Benthol. Soc.* 8(3), 237–242. https://doi.org/10.2307/1467327 (1989).
- Franz, C. J. Feeding patterns of *Fissurella* species on Isla de Margarita, Venezuela: Use of radulae and food passage rates. *J. Molluscan Stud.* 56(1), 25–35. https://doi.org/10.1093/mollus/56.1.25 (1990).
- Thompson, R. C., Johnson, L. E. & Hawkins, S. J. A method for spatial and temporal assessment of gastropod grazing intensity in the field: The use of radula scrapes on wax surfaces. J. Exp. Mar. Biol. Ecol. 218(1), 63–76. https://doi.org/10.1016/S0022-0981(97)00068-3 (1997).
- Iken, K. Feeding ecology of the Antarctic herbivorous gastropod Laevilacunaria antarctica Martens. J. Exp. Mar. Biol. Ecol. 236(1), 133–148. https://doi.org/10.1016/S0022-0981(98)00199-3 (1999).
- Forrest, R. E., Chapman, M. G. & Underwood, A. J. Quantification of radular marks as a method for estimating grazing of intertidal gastropods on rocky shores. J. Exp. Mar. Biol. Ecol. 258(2), 155–171. https://doi.org/10.1016/S0022-0981(01)00212-X (2001).
- Dimitriadis, V. K. Structure and function of the digestive system in Stylommatophora. In *The Biology of Terrestrial Molluscs* (ed. Barker, G. M.) 237–258 (CABI Publishing, 2001).
- Speiser, B. Food and feeding behaviour. In *The Biology of Terrestrial Molluscs* (ed. Barker, G. M.) 259–288 (CABI Publishing, 2001).
- Sutton, G. P. et al. Neural control exploits changing mechanical advantage and context dependence to generate different feeding responses in *Aplysia. Biol. Cybern.* 91(5), 333–345. https://doi.org/10.1007/s00422-004-0517-z (2004).
- Mangan, E. V. et al. A biologically inspired gripping device. Ind. Rob. 32(1), 49-54. https://doi.org/10.1108/014399105105732 91 (2005).
- Padilla, D. K. Structural resistance of algae to herbivores. A biomechanical approach. Mar. Biol. 90, 103–109. https://doi.org/10. 1007/BF00428220 (1985).
- Padilla, D. K. Algal structural defenses: Form and calcification in resistance to tropical limpets. *Ecology* 70(4), 835–842. https:// doi.org/10.2307/1941352 (1989).
- Krings, W. et al. In slow motion: Radula motion pattern and forces exerted to the substrate in the land snail Cornu aspersum (Mollusca, Gastropoda) during feeding. R. Soc. Open Sci. 6(7), 190222. https://doi.org/10.1098/rsos.190222 (2019).
- Mackenstedt, U. & Märkel, K. Radula structure and function. In *The Biology of Terrestrial Molluscs* (ed. Barker, G. M.) 213–236 (CABI Publishing, 2001).
- Neustadter, D. M., Drushel, R. F. & Chiel, H. J. Kinematics of the buccal mass during swallowing based on magnetic resonance imaging in intact, behaving *Aplysia californica. J. Exp. Biol.* 205(7), 939–958 (2002).
- Neustadter, D. M., Drushel, R. F., Crago, P. E., Adams, B. W. & Chiel, H. J. A kinematic model of swallowing in *Aplysia californica* based on radula/odontophore kinematics and in vivo magnetic resonance images. J. Exp. Biol. 205(20), 3177–3206 (2002).
- 91. Neustadter, D. M., Drushel, R. F., Crago, P. E. & Chiel, H. J. A 3-dimensional model of the odontophore of *Aplysia* throughout a swallowing cycle. *Soc. Neurosci. Abstr.* 27(943), 7 (2001).
- Kehl, C. E. et al. Soft-surface grasping: Radular opening in Alysia californica. J. Exp. Biol. 222, jeb191254. https://doi.org/10. 1242/jeb.191254 (2019).
- Montroni, D. et al. Structural characterization of the buccal mass of Ariolimax californicus (Gastropoda; Stylommatophora). PLoS ONE 14(8), e0212249. https://doi.org/10.1371/journal.pone.0212249 (2019).
- Holznagel, W. A nondestructive method for cleaning gastropod radulae from frozen, alcohol-fixed, or dried material. Am. Malacol. Bull. 14(2), 181–183 (1998).
- Wakeling, J. M., Kemp, K. M. & Johnston, I. A. The biomechanics of fast-starts during ontogeny in the common carp (*Cyprinus carpio*). J. Exp. Biol. 202, 3057–3067 (1999).

- 96. Hutchinson, J. R. & Garcia, M. Tyrannosaurus was not a fast runner. Nature 415, 1018–1021. https://doi.org/10.1038/4151018a (2002).
- 97. Alexander, R. M. The maximum forces exerted by animals. J. Exp. Biol. 115, 231-238 (1985).
- Heethoff, M. & Norton, R. A. A new use for synchrotron X-ray microtomography: Three-dimensional biomechanical modeling of chelicerate mouthparts and calculation of theoretical bite forces. *Invertebr. Biol.* 128(4), 332–339. https://doi.org/10.1111/j. 1744-7410.2009.00183.x (2009).
- Erickson, G. M., Lappin, A. K. & Viet, K. A. The ontogeny of bite-force performance in American alligator (Alligator mississippiensis). J. Zool. 260(3), 317–327. https://doi.org/10.1017/S0952836903003819 (2003).
- 100. Sustaita, D. & Hertel, F. *In vivo* bite and grip forces, morphology and prey killing behavior of North American accipiters (Accipitridae) and falcons (Falconidae). *J. Exp. Biol.* **213**, 2617–2628. https://doi.org/10.1242/jeb.041731 (2010).
- Sakamoto, M., Lloyd, G. T. & Benton, M. J. Phylogenetically structured variance in felid bite force: The role of phylogeny in the evolution of biting performance. J. Evol. Biol. 23(3), 463–478. https://doi.org/10.1111/j.1420-9101.2009.01922.x (2010).
- Sakamoto, M., Ruťa, M. & Venditti, C. Extreme and rapid bursts of functional adaptions shape bite force in amniotes. Proc. R. Soc. B 286(1894), 20181932. https://doi.org/10.1098/rspb.2018.1932 (2019).
- 103. Vogel, S. Comparative Biomechanics. Life's Physical World 2nd edn, 320 (Princeton University Press, 2013).
- 104. Alexander, D. E. Nature's Machines 189 (Academic Press, 2017).
- Becerra, F., Echeverria, A. I., Casinos, A. & Vassallo, A. I. Another one bites the dust: Bite force and ecology in three caviomorph rodents (Rodentia, Hystricognathi). J. Exp. Zool. 321(4), 220–232. https://doi.org/10.1002/jez.1853 (2014).
- Pfaller, J. B., Gignac, P. M. & Erickson, G. M. Ontogenetic changes in jaw-muscle architecture facilitate durophagy in the turtle Sternotherus minor. J. Exp. Biol. 214, 1655–1667. https://doi.org/10.1242/jeb.048090 (2010).
- Chapman, R. F. Mechanics of food handling by chewing insects. In *Regulatory Mechanisms in Insect Feeding* (eds Chapman, R. F. & de Boer, G.) 3–31 (Springer, 1995).
- 108. Chiel, H. J. Aplysia feeding biomechanics. Scholarpedia 2(9), 4165 (2007).
- 109. Marshall, C. D. *et al.* The ontogenetic scaling of bite force and head size in loggerhead sea turtles (*Caretta caretta*): Implications for durophagy in neretic, benthic habitats. *J. Exp. Biol.* **215**, 4166–4174. https://doi.org/10.1242/jeb.074385 (2012).
- Van der Meij, M. A. A. & Bout, R. G. The relationship between shape of the skull and bite force in finches. J. Exp. Biol. 211, 1668–1680. https://doi.org/10.1242/jeb.015289 (2008).
- Forbes-Harper, J. L. et al. Diet and bite force in red foxes: Ontogenetic and sex difference in an invasive carnivore. J. Zool. 303(1), 54–63. https://doi.org/10.1111/jzo.12463 (2017).
- 112. Ahn, A. N. & Full, R. J. A motor and a brake: Two leg extensor muscles acting at the same joint manage energy differently in a running insect. J. Exp. Biol. 205, 379–389 (2002).
- Guschlbauer, C., Scharstein, H. & Buschges, A. The extensor tibiae muscle of the stick insect: Bio-mechanical properties of an insect walking leg muscle. J. Exp. Biol. 210, 1092–1108. https://doi.org/10.1242/jeb.02729 (2007).
- Siebert, T., Weihmann, T., Rode, C. & Blickhan, R. *Cupiennius salei*: biomechanical properties of the tibia-metatarsus joint and its flexing muscles. J. Comp. Physiol. B 180, 199–209. https://doi.org/10.1007/s00360-009-0401-1 (2010).
- Krings, W. & Gorb, S. N. Substrate roughness induced wear pattern in gastropod radulae. *Biotribology* 26, 100164. https://doi. org/10.1016/j.biotri.2021.100164 (2021).
- Reingold, S. C. & Gelperin, A. Feeding motor programme in *Limax*. II. Modulation by sensory inputs in intact animals and isolated central nervous systems. *J. Exp. Biol.* 85, 1–20 (1980).
- Hurwitz, I. & Susswein, A. J. Adaptation of feeding sequences in *Aplysia oculifera* to changes in the load and width of food. J. Exp. Biol. 166, 215–235 (1992).
- Kupfermann, I. Feeding in *Aplysia*: A simple system for the study of motivation. *Behav. Biol.* 10(1), 1–26. https://doi.org/10.1016/S0091-6773(74)91644-7 (1974).
- Susswein, A. J. & Bennett, M. V. L. Plasticity of feeding behavior in the opisthobranch mollusc Navanax. J. Neurobiol. 10(6), 521–534. https://doi.org/10.1002/neu.480100603 (1979).
- Susswein, A. J., Achituv, Y., Cappell, M. C. & Bennett, M. V. L. Pharyngeal movements during feeding sequences in *Navanax inermis*: A cinematographic analysis. J. Comp. Physiol. A 155, 209–218. https://doi.org/10.1007/BF00612638 (1984).
- Pennings, S. C. Size-related shifts in herbivory: Specialization in the sea hare *Aplysia californica* Cooper. J. Exp. Mar. Biol. Ecol 142(1–2), 43–61. https://doi.org/10.1016/0022-0981(90)90136-Z (1990).
- 122. Verwaijen, D., Van Damme, R. & Herrel, A. Relationships between head size, bite force, prey handling efficiency and diet in two sympatric lacertid lizards. *Funct. Ecol.* 16(6), 842–850. https://doi.org/10.1046/j.1365-2435.2002.00696.x (2002).

Acknowledgements

We would like to thank Renate Walter from the Zoological Institute of the Universität Hamburg for support on the SEM, Thomas M. Kaiser from the CeNak of the Universität Hamburg for discussing results. We are highly grateful for the helpful comments of the four anonymous reviewers and of the editorial board member.

Author contributions

W.K. and S.G. initiated the project, discussed the data analysed, performed statistical analyses. W.K. took the photos. C.N. performed experiments, analysed data, and contributed to earlier versions of the manuscript. S.G. provided experimental design and contributed to the biomechanical aspects of the manuscript. M.T.N. performed species identification. A.K. contributed to data analysis. All authors contributed to and approved the final manuscript for publication.

Funding

Open Access funding enabled and organized by Projekt DEAL.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1038/s41598-021-89892-z.

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