scientific reports

Check for updates

OPEN A primary study of breeding system of Ziziphus jujuba var. spinosa

Feng Wang^{1,2,3}, Xiaohan Sun^{1,2,3}, Jibin Dong^{1,2,3}, Rong Cui^{1,2,3}, Xiao Liu^{1,2,3}, Xiangxiang Li^{1,2,3}, Hui Wang^{1,2,3}, Tongli He^{1,2,3}, Peiming Zheng^{1,2,3} & Renging Wang^{1,2,3}

Ziziphus jujuba var. spinosa has been used as a windbreak and for soil conservation and water retention. Previous studies focused on pharmacological effects and extraction of chemical components in this species, and very few explored the breeding system. The present study combined the analysis of floral morphology, behavior of flower visitors, and artificial pollination to reveal reproductive characteristics of the species. Its flowers are characterized by dichogamy, herkogamy, and stamen movement, which are evolutionary adaptations to its breeding system. There were more than 40 species of visiting insects, mainly Hymenoptera and Diptera, and the characteristics of dichogamous and herkogamous flower adapted to the visiting insects. The breeding system is outcrossing, partially self-compatible, and demand for pollinators. The fruit setting rate after natural pollination was 2%. Geitonogamy and xenogamy did not significantly increase the fruit setting rate, indicating that the low fruit setting rate was not due to pollen limitation by likely caused by resource limitation or fruit consumption. The fruit setting rate of zero in emasculated and in naturally and hand self-pollinated individuals suggested the absence of apomixis and spontaneous self-pollination. The above results can be utilized in studies on evolution and cultivation of Z. jujuba var. spinosa.

Breeding system in plants is an essential part of their life cycle and an important factor in their evolution¹. About 80% of angiosperms contain bisexual flowers, including hermaphroditism, monoecy, andromonoecy, gynomonoecy, and trimonoecy². Among those, hermaphrodites account for 70% of angiosperms and are generally considered to be ancestral types of angiosperms³. Understanding the reproductive characteristics of hermaphrodites will thus help us to interpret plant evolution. Studies have linked interference between sexual functions to hermaphroditism, and the theory of gender interference helps to explain the evolutionary adaptation of floral characteristics⁴. The movement of stamens and pistils provides additional evidence for the adaptive significance and evolutionary ecology of flowers^{5,6}. Different pollination methods were used to deal with a series of plants in order to realize the discussion of their reproductive ecological characteristics in previous studies⁷. Some hermaphroditic flowering plants have a temporal separation of male and female functions, which is traditionally considered an effective mechanism to prevent self-pollination in flowers 8.

The evolution of floral characteristics is influenced by pollinator choice and co-evolution with pollinators, and is the main driving force for angiosperm speciation⁹⁻¹¹. Attracting more pollinators to participate in pollen transfer and thereby improve the success rate of pollination is an important driving force of flower evolution⁸. The evolution of floral characteristics is not only related to pollinators, but also influenced by environmental factors, genetic linkage of different genes, and metabolism¹²⁻¹⁴. Flower color affects the behavior of pollinators and improves pollination efficiency¹⁵. Flowering phenology may be an important factor in maintaining the diversity of plant communities¹³, and it is also the result of selection pressure, which can reflect the suitability of plants¹⁶.

Although self-pollination may be an important component of the population mating system, previous research showed that insects play a role in promoting cross-pollination¹⁷. The destruction of habitats by human activities and climate change has dramatically reduced biodiversity¹⁸ and disrupted the plant-pollinator relationship^{19,20}. The functional diversity of pollination networks helps to maintain the diversity of plant communities²¹. About 87.5% of flowering plants rely on animals to deliver pollen²². Globally, the value of pollinators for crop production in 2005 was estimated at 153 billion euros per year²³. Governments, media, and scientists all over the world have raised concerns about the reduction in pollinator numbers and the loss of pollination services²⁴.

In the natural state, plants may self-pollinate, cross-pollinate with the help of insects and other external factors, or both. Different pollination methods have different effects on fruit and offspring quality²⁵. Many studies on reproductive ecology of plants implemented artificial control of pollination. Generally, self-pollination may

¹Institute of Ecology and Biodiversity, School of Life Sciences, Shandong University, 72 Binhai Road, Qingdao 266237, China. ²Shandong Provincial Engineering and Technology Research Center for Vegetation Ecology, Shandong University, Qingdao, China. ³Qingdao Forest Ecology Research Station of National Forestry and Grassland Administration, Shandong University, Qingdao, China. Remail: zhengpm@sdu.edu.cn



Figure 1. Single flower opening stages. T1-bud opening stage (bud dehiscence, showing no internal structure of flower), T2- initial opening stage (sepals initially open, showing the internal structure of flowers), T3-sepal flattening stage (sepals spreading horizontally, petals still enveloping stamens), T4- petal and stamen separation stage (petals open, separate from stamens), T5- petal flattening stage (petals horizontally distributed between sepals), T6- stamen flattening stage (the stamens bend to the horizontal direction, when the stigma is already bifurcated), T7-stamen drooping stage (stamen bending downward), T8- wilting stage.

result in less seed setting and seed yield²⁶ and the seed setting is significantly improved by cross-pollination^{7,27}. Artificial pollination ensures that plants receive enough pollen, which is then used to study pollen restrictions^{28,29}. For example, female and male reproductive success of female, male, and hermaphrodite individuals in the subdioecious shrub *Eurya japonica* (Theaceae) was studied using artificial pollination^{30,31}. Thus, artificial pollination is suitable for studies on reproductive ecology.

As a soil and water conservation species, *Ziziphus jujuba* var. *spinosa* acts as a windbreak, protects from windblown sand, and reduces soil erosion. The previous studies on this species were mainly focused on its pharmacological effects, such as extraction of chemical components^{32–35}, and on the effects of water, nutrition components, and arbuscular mycorrhizal fungi on its physiology and ecology^{36,37}. *Z. jujuba* var. *spinosa* is an excellent species of soil and water conservation in China. Its performance of forest and slope protection has obvious advantages in improving the ecological environment. It is also a new generation of wild fruit resources with great development potential, and has high economic and medical value. It is of great ecological significance to improve the germination rate and seedling survival rate of wild *Z. jujuba* var. *spinosa* seeds for the conservation and restoration of vegetation and the reduction of soil erosion. There is few researches focusing on the breeding system of *Z. jujuba* var. *spinosa*. Some researchers studied the pollen of *Z. jujuba*, and found that there were significant differences in pollen germination rate in different stages. The results of in vitro culture were more accurate^{69,70}.

The aim of the present study was to reveal the characteristics of pollination biological and reproductive system of *Z. jujuba* var. *spinosa* by examining floral morphology and conducting insect trapping and artificial pollination. The following specific topics were discussed: (1) the flower morphology adapting to its breeding system in evolution; (2) the relationship between the flowers and flower-visiting insects (species, quantity and behavior); (3) breeding system of *Z. jujuba* var. *spinosa* (self-crossing or cross-breeding). The results presented herein would provide information that can be used to study the evolution and cultivation of *Z. jujuba* var. *spinosa*.

Result

Floral features. Eight stages were identified from bud to senescence stage of a single *Z. jujuba* var. *spinosa* flower (Fig. 1): T1-bud opening stage (bud dehiscence, no internal structure of the flower), T2-initial opening stage (sepals initially open, internal structure of the flower visible), T3-sepal flattening stage (sepals spreading horizontally, petals still enveloping stamens), T4-petal and stamen separation stage (petals opened, separated from stamens), T5-petal flattening stage (petals horizontally distributed between sepals), T6-stamen flattening stage (the stamens bend to the horizontal direction, when the stigma is already bifurcated), T7 -stamen drooping stage (stamen bending downward), T8-wilting stage. T1–T5 stages occurred on the first day of flowering; T6 stage was reached the second day of flowering; and T7 stage followed one day later. The flowers are hermaphroditic. The flowers are hermaphroditic. Stamens and pistils were elongated in shape during the flowering (Fig. 2A,B),

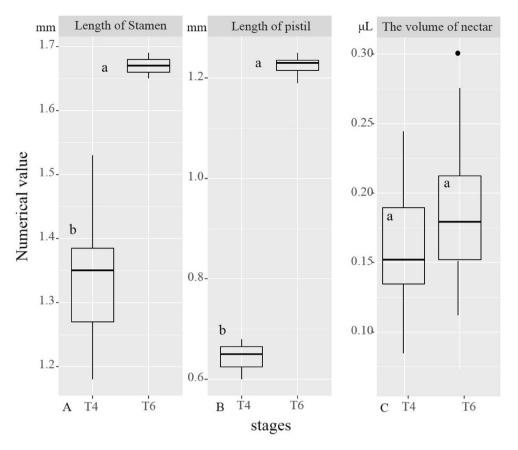


Figure 2. The length of stamen (**A**), length of pistil (**B**) and volume of nectar (**C**) at T4 and T6 periods were plotted. The lines in the box represent the median value of the data (n = 30). Different letters indicate a significant difference based on a generalised linear mixed model (GLMM). A shows that the length of stamens in T4 period is 1.34 ± 0.11 , and the length of stamens in T6 period is 1.67 ± 0.01 . *P*=0.002<0.05. There were significant differences between the two groups. B shows that the length of pistil in the T4 period is 0.64 ± 0.03 , and the length of pistil in the T6 period is 1.22 ± 0.02 . *P*=1.28e-14<0.001. There are very significant differences between the two groups. C shows that the nectar volume in the T4 period is 0.14 ± 0.04 , and the nectar volume in the T6 period is 0.17 ± 0.06 . *P*=0.178>0.05. There were no clear significant differences between the two groups.

Items of observation			Results of observation
Flower organs wilting order			Petal→Stamen
	Color changes		Aqua (No obvious changes)
Petal development	State changes		$Wrapping \rightarrow Separating \rightarrow Flattening \rightarrow Drooping$
	Color changes		Canary → Brown
Stamens development	Filament changes		Short \rightarrow Long Curve \rightarrow Upright \rightarrow Curve
	Distance from anthers to stigma		Short→Long
	Mode of anther dehiscence		Longitudinal dehiscence
	Number		4,5,6
Pistil development	Stigma	Color	Aqua→Brown
		Shape	$Cone \rightarrow Bifurcate \rightarrow Long$
	Ovary	Color	$Yellow \rightarrow Ivory \rightarrow Cyan(Fertilization) \rightarrow Red(Withered)$

Table 1. The functional floral morphology of Z. jujuba var. spinosa.

.....

and the stigma was bifurcated. Stamens moved slowly, from straight position to horizontal, and finally bending downward (Table 1); the color of petals and the pistil changed during the flowering period (Table 1).

Nectar volume. Nectar was secreted on the first and second day of flowering, and nectar secretion was measured in the T4 and T6 stages (Fig. 2C). The nectar volume in T4 and T6 was 0.14 ± 0.04 and 0.17 ± 0.06 , respectively. There was no significant difference in nectar production between T4 and T6 stages (P>0.05).

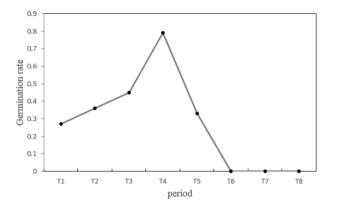


Figure 3. Pollen germination rate of Z. jujuba var. spinosa at different flowering stages.

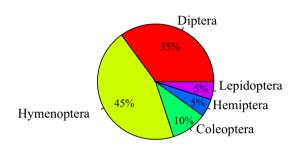


Figure 4. species of flower visiting insects.

Pollen germination rate. The number of pollen grains in the petal flattening stage (T6) and later was very limited and difficult to collect. Therefore, the pollen for pollen germination rate experiment was collected from flowers between the bud opening stage (T1) and the petal flattening stage (T5) (Figs. 1 and 3). The pollen germination rate gradually increased from the bud opening stage, peaked at the petal and stamen separation stage (T4), and declined thereafter. Overall, pollen activity was the highest at the petal and stamen separation stage (T4).

Flower visitors and their visiting behavior. About 40 species of insects, belonging to five orders, visited the flowers. The largest order was Hymenoptera, accounting for 45% of total insect visitors, followed by Diptera (35%), Coleoptera (10%), and Hemiptera (5%) and Lepidoptera (5%) (Fig. 4).

According to our investigation, Apidae and Syrphidae are the main flower visiting insect families. Moreover, the operating time of the visiting insects on the flower differed between insect families. The visitation time of Bibionidae and Chalcididae was significantly longer than that of the other families (P<0.01; Fig. 5). Differences were also observed in the visitation time among Bibionidae individuals (Fig. 5).

Figure 6 shows the daily visitation frequency of the insects from 8:00 to 17:00. The visitors were much more active at 11:00 and 14:00, and much less dynamic in the mornings (8:00–9:00), at dusk (16:00–17:00), and at noon (12:00–13:00).

Pollen-ovule ratio (P/O) and Outcrossing index (OCI). Ziziphus jujuba var. spinosa has two ovules³⁸. The average amount of pollen in a single flower was 7800 ± 300 grains (n = 30). The pollen-ovule ratio (P/O) was 3900 ± 150 , indicating that the breeding system is xenogamy³⁹. The OCI was 4 and calculated by summation of the following values: (1) flower diameter was 5.13 ± 0.22 mm (n = 30), and assigned a value of 2; (2) the stamen was mature before the pistil, and therefore its value was 1; (3) the pistil and stamen were spatially separated, hence the value of 1⁴⁰. According to Dafni standard, the breeding system is mainly outcrossing, partially self-compatible, and demands pollinators.

Artificial pollination experiment. Figure 7 shows the fruit setting rate of various artificial pollination treatments. The fruit setting rate of emasculated, hand self-pollinated, and naturally self-pollinated flowers was 0% (Fig. 7). The fruit setting rate of geitonogamy and xenogamy was about 4%, which was slightly higher than that of natural pollination (2%); the difference was not significant.

The upper and lower hinges of the box indicate 75 and 25th percentiles of the data, respectively. E, Emasculated; G, Geitonogamy; HS, Hand self-pollination; N, Natural pollination; NS, Natural self-pollination; X, Xenogamy. There was no significant difference between G, N and X.

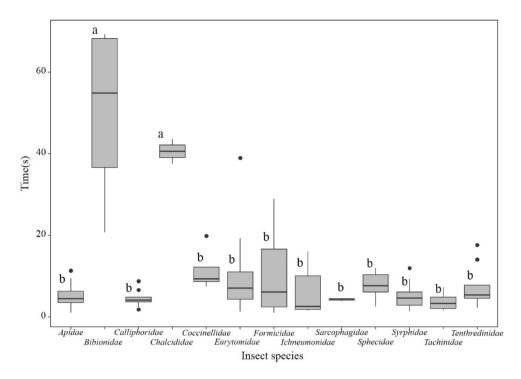


Figure 5. Observed visiting time of different families. The lines in the box represent the median value of the data. Different letters indicate significant differences in the results of multiple comparisons in which family-wise errors were adjusted using Tukey's method. P<0.01.

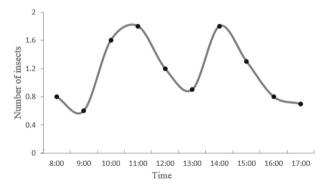


Figure 6. Average frequency of insect visiting flowers (between 8:00-17:00).

Discussion

Adaptive evolution of floral morphology. Self-pollination in hermaphrodites has advantages, but inbreeding also has a detrimental effect. During evolution, plants tend to cross-pollinate and evolve toward avoiding self-pollination⁴¹, such as herkogamy, dichogamy, and hermaphroditism⁵. The obvious way to promote outcrossing is by preventing self-pollination. A decline in self-pollination has been observed in a large number of plants, and there is evidence that the process of self-pollination is genetically controlled⁴². Studies have shown that dichogamy is a common but neglected feature of outcrossing in angiosperms. Dichogamy can reduce self-interference and thus reduce selfing⁴³. The observations of the flowering period in *Z. jujuba* var. *spinosa* revealed the characteristics of herkogamy and protandry (Figs. 1 and 2), both of which are mechanisms that tend to avoid self-pollination.

Four types of stamen movement has been reported in previous studies, and they include stimulated, simultaneous and slow, quick and explosive, and cascade^{44,45}. In this study, we found that stamen movement in *Z. jujuba* var. *spinosa*. belongs to simultaneous and slow type. Stamen movement of some plants can promote selfpollination, increase the probability of pollination, and facilitate reproduction^{46,47}. However, the slow movement of stamens does not promote self-pollination, but supports cross-pollination. Although herkogamy can reduce the possibility of selfing, the success of pollination is reduced because of the difference between pollinator contact sites on the anthers and stigmas, resulting in wasted resources⁴⁸.

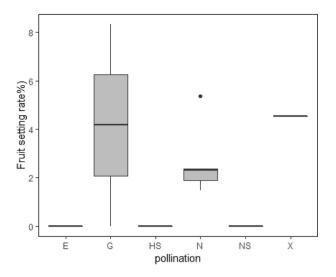


Figure 7. Fruit setting rate of artificial pollination experiment.

The pollen germination rate confirms the protandrous nature of the *Z. jujuba* var. *spinosa* reproductive system (Fig. 3). The stamens are scattered from the bud opening stage (T1) to the petal flattening stage (T5). At this time, the pistils are immature, and the stigmas are not receptive (Fig. 1). Similar pistil and stamen characteristics were reported for some species in Labiatae⁴⁹. This mechanism can effectively avoid self-pollination and increase the rate of outcrossing^{48,50}.

The nectar volumes in T4 (petal and stamen separation stage) and T6 (stamen flattening stage) were similar (Fig. 2C), indicating that pollinator attraction by nectar is always imparted during stamen (T4) and stigma maturation (T6). This further favors the outcrossing.

Characteristics of flower visiting insects. The sexual system of plants is fundamentally related to the pollination biology of plants⁵¹. Pollinators play important roles in plant reproduction⁵². Biological pollination is considered a key factor in biodiversity of natural populations⁵³, and plays an indispensable role in evolution. The productivity of crops can be improved by increasing species diversity and quantity of local wild flowers around farmlands⁵⁴.

Hymenoptera and Diptera are the dominant visitors of *Z. jujuba* var. *spinosa* (Fig. 4). These orders include important flower visitors and pollinators of many plant species and in various habitats⁵⁵. Although the pollination efficiency of non-bee insects is not as high as that of bees, they visit flowers more often and play an important role in global crop production⁵⁶. In fact, the species and number of pollinators are related to the pollination background and weather conditions—more kinds of insects are active on sunny days, and the opposite is true on cloudy days^{57–59}.

Additionally, the frequency of insect flower visitations is related to flowers morphology and environmental factors, such as temperature or light intensity. In the morning and afternoon, the frequency of insect visits was increased (Fig. 6). The decrease in flower visitation frequency at noon may be due to high temperature and strong solar radiation⁶⁰, which is not conducive to vigorous activities of the insects. At this time, some insects hide under the leaves to avoid strong solar radiation. The flower visiting behavior of insects and the characteristics of flower adapt to each other⁶¹, which is helpful for *Z. jujuba* var. *spinosa* to avoid inbreeding and increase the probability of outcrossing.

Breeding characteristics of *Z. jujuba* var. *spinosa*. The study of plant reproductive biology helps us to understand the reproductive strategies in different species, which will ensure successful reproduction as well as facilitate population survival and community evolution⁶². The breeding system of *Z. jujuba* var. *spinosa* was analyzed and determined through estimating the pollen-to-ovule ratio (P/O) and the outcrossing index (OCI), and by artificial pollination. The pollen ovule ratio (P/O) of a single flower was 3900 ± 150 , which belonged to specific outcrossing³⁹. The OCI value of 4 was indicative of the primarily outcrossing breeding system, and partial self-cross compatibility and need for pollinators⁴⁰. The two indexes thus indicate outcrossing as the breeding system in *Z. jujuba* var. *spinosa*.

The breeding system of this species was further investigated through artificial pollination. The fruit setting rate after natural pollination was only 2%, which may have been caused by certain limiting factors. Namely, geitonogamy and xenogamy did not significantly increase the fruit setting rate, indicating that pollen number was not a limiting factor. Thus, the low fruit setting might be due to resource limitation or fruit feeding⁶³. In this experiment, the fruit setting rate of Emasculated is 0, which indicates that there is no apomixis. The fruit set rate of Natural-self-pollination and Hand-self-pollination are all 0, indicating that there is no self-pollination. Certain species cannot self-pollinate even if they are inbred⁶⁴. Inbreeding decline affects the evolution of mating system and the persistence of small mutation load in a population⁶⁵. Declining tendency for inbreeding and

preference for outcrossing is present in many species^{66,67}. The floral characteristics of *Z. jujuba* var. *spinosa* reflect the adaptation to its breeding system. Dichogamy can ensure that self-pollination is avoided, but it can't prevent Geitonogamy. Because of the decrease in fitness caused by excessive far crossing, part of self compatibility (i.e. Geitonogamy) becomes the most suitable form of mating⁶⁸. It can improve genetic diversity and promote plant evolution while ensuring the success of reproduction.

Material and methods

Study species and site. *Ziziphus jujuba* var. *spinosa* is a hermaphroditic shrub or small tree in the Rhamnaceae. It is used as an excellent soil and water conservation species in China, and it is an important nectar and entomophilous plant.

This study was carried out on Qiangu Mountain, Qingdao City, Shandong Province, China (lat.36°29'53.33"N, long.120°43'44.31"E). The annual average temperature in the region is 12.1 °C, the maximum temperature is 38.6 °C, and the minimum temperature is – 18.6 °C. The annual average precipitation is 708.9 mm, 65% of which occur in summer.

The study was conducted at three sites, each with more than 10 individuals of *Z. jujuba* var. *spinosa*. Additional dominant trees in each site included *Pinus densiflora* Siebold & Zucc., *Robinia pseudoacacia* L., *Quercus acutissima* Carruth., and *Vitex negundo* L. var. *heterophylla* (Franch.) Rehder.

Floral morphology. Three individuals of *Z. jujuba* var. *spinosa* were randomly selected at each site, and three inflorescences were randomly chosen in each individual (about 40 flowers develop on each branch) to investigate the flowering process and floral morphology (Fig. 1). The number of flowers, flower shape, size, and color at different flowering stages were recorded (Table 1).

Nectar volume. Flowers were bagged with paper pollination bags to exclude pollinators before evaluation. According to the single flower opening stages of *Z. jujuba* var. *spinosa*, there was less nectar from T1 stage to T3 stage (Fig. 1). On the first day, the attraction of flowers to visiting insects mainly began in T4 stage, and on the second day, it mainly began in T6 stage. Nectar volume per flower was evaluated at petal and stamen separation stage (T4) or stamen flattening stage (T6) after anthesis using a 1 μ L microcapillary tube.

Pollen germination rate. Pollen germination rate was determined by in vitro culture method. Six flowers were collected at each flower developmental stage and fresh pollen was shaken onto a slide coated with a thin layer of MS medium. The total number of pollen was \geq 600. The slides were placed in a petri dish with two layers of wet filter paper to keep the culture environment moist. The slides were cultured in a dark incubator at 26 °C, and pollen germination rate was examined after 24 h under an optical microscope (CX31RTSF; Olympus, Tokyo, Japan). The germination rate was calculated using the following formula:

$$R_{PG}(\%) = \frac{n_l}{N_t} * 100$$

where R_{PG} is pollen germination rate (%), n_l is the number of pollen grains with tube length longer than pollen diameter, N_t is the number of pollen observed.

Flower visitors and their visiting behavior. From June 4 to June 11, 2019, the flower visiting insects were investigated hourly on marked inflorescences or individuals from 7:00–18:00 after the plants entered the full bloom period. The species and their behavior were observed and recorded. Flower visitation frequency was measured by stopwatch and camera, and the following three aspects were recorded: (1) the time needed for insects to operate flowers (such as drilling into the corolla) and to ingest nectar; (2) the time needed for insects to fly from one flower to the next; and (3) the pattern of insect movement among flowers.

Pollen-ovule ratio (P/O) and outcrossing index (OCI). Ten mature but indehiscent anthers were collected from three individuals (30 anthers in total) and placed in three centrifuge tubes, each containing 10 mL 0.1% hydrochloric acid solution. Pollen grains in each tube were incubated in a water bath with shaking for 4 h at 55 °C. The suspension was removed from the bath and immediately transferred onto a slide, and the number of grains was recorded. P/O was determined as the ratio of pollen grain number per flower (the number of grains per anther multiplied by the number of anthers) to the number of ovules.

The Dafni's OCI⁴⁰ is calculated at the species level based on floral morphological characteristics and used to predict the mating system. The value of OCI is obtained by adding the scores of each judgment condition. The OCI of *Z. jujuba* var. *spinosa* was calculated based on inflorescence diameter, flower size, and flowering behavior according to Dafni.

Artificial pollination experiment. From May to July, 2018, five mother trees were randomly selected at every site. Two flower branches 20–40 cm long were randomly selected for each mother tree, and flower buds on each branch were marked and counted. To prevent natural insect pollination, the selected branches were covered with nylon bags. The following treatments were conducted:

Control 1: Natural pollination, no emasculation, no bagging (N); Control 2: Natural self-pollination, no emasculation, bagging (NS); Treated 1: Geitonogamy, emasculation, and bagging (G); Treated 2: Hand self-pollination, no emasculation, bagging (HS);

Treated 3: Xenogamy, emasculation, and bagging (x);

Treated 4: Emasculation, bagging (to test whether self-pollination occurred at budding stage) (E).

During the experiment, three male parents were selected for pollen selection, and the suitable flower stage was chosen for pollination. Pollen was collected with a brush and transferred onto the stigma several times in succession.

Data availability

All data generated or analysed during this study are included in this published article (and its Supplementary Information files).

Received: 20 May 2020; Accepted: 26 April 2021 Published online: 14 May 2021

References

- 1. East, E. M. The role of reproduction in evolution. Am. Nat. https://doi.org/10.1086/279670 (1918).
- 2. Proctor, M., Yeo, P. F. & Lack, A. A Natural History of Pollination. (1996).
- Spigler, R. B. & Ashman, T.-L. Gynodioecy to dioecy: are we there yet?. Ann. Bot. 109, 531–543. https://doi.org/10.1093/aob/ mcr170%JAnnalsofBotany (2011).
- 4. Barrett, S. Sexual interference of the floral kind. Heredity 88, 154-159 (2002).
- 5. Li, Q.-J. et al. Flexible style that encourages outcrossing. Nature 410, 432-432. https://doi.org/10.1038/35068635 (2001).
- Sun, S., Gao, J. Y., Liao, W. J., Li, Q. J. & Zhang, D. Y. Adaptive significance of flexistyly in *Alpinia blepharocalyx* (Zingiberaceae): a hand-pollination experiment. *Ann. Bot.* 99, 661–666 (2007).
- 7. Kumar, B. D., Deepika, D. S. & Raju, A. S. Reproductive ecology of the semi-evergreen tree *Vitex negundo* (Lamiaceae). *Phytol. Balcanica* 23, 39–53 (2017).
- 8. Faegri, K. & Van Der Pijl, L. Principles of Pollination Ecology (Elsevier, Amsterdam, 2013).
- 9. Darwin, C. The Effects of Cross and Self Fertilisation in the Vegetable Kingdom (D. Appleton, Boston, 1877).
- 10. Baker, H. G. in Cold Spring Harbor Symposia on Quantitative Biology. 177-191 (Cold Spring Harbor Laboratory Press).
- 11. Heithaus, E. R., Opler, P. A. & Baker, H. G. Bat activity and pollination of *Bauhinia pauletia*: plant-pollinator coevolution. *Ecology* 55, 412–419 (1974).
- Armbruster, W. S. Can indirect selection and genetic context contribute to trait diversification? A transition-probability study of blossom-colour evolution in two genera. J. Evolut. Biol. 15, 468–486 (2002).
- Bradshaw, H. Jr. & Schemske, D. W. J. N. Allele substitution at a flower colour locus produces a pollinator shift in monkeyflowers. Nature 426, 176 (2003).
- Gómez, J. M. & Zamora, R. Ecological factors that promote the evolution of generalization in pollination systems, in *Plant-pollinator interactions: from specialization to generalization*. 145–166 (2006).
- 15. Barrett, S. C. & Harder, L. D. Ecology and evolution of plant mating. *Trends Ecol. Evolut.* 11, 73–79 (1996).
- 16. Elzinga, J. A. et al. Time after time: flowering phenology and biotic interactions. Trends Ecol. Evol. 22, 432-439 (2007).
- Huang, S.-Q., Xiong, Y.-Z. & Barrett, S. C. H. Experimental evidence of insect pollination in Juncaceae, a primarily wind-pollinated family. Int. J. Plant Sci. 174, 1219–1228. https://doi.org/10.1086/673247 (2013).
- 18. Sánchez-Bayo, F. & Wyckhuys, K. A. Worldwide decline of the entomofauna: a review of its drivers. Biol. Conserv. 232, 8-27 (2019).
- Memmott, J., Craze, P. G., Waser, N. M. & Price, M. V. Global warming and the disruption of plant–pollinator interactions. *Ecol. Lett.* 10, 710–717. https://doi.org/10.1111/j.1461-0248.2007.01061.x (2007).
- Winfree, R., Griswold, T. & Kremen, C. Effect of human disturbance on bee communities in a forested ecosystem. *Conserv. Biol.* 21, 213–223. https://doi.org/10.1111/j.1523-1739.2006.00574.x (2007).
- 21. Biesmeijer, J. C. *et al.* Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands. *Science* **313**, 351–354 (2006).
- Ollerton, J., Winfree, R. & Tarrant, S. How many flowering plants are pollinated by animals?. Oikos 120, 321–326. https://doi.org/ 10.1111/j.1600-0706.2010.18644.x (2011).
- Gallai, N., Salles, J.-M., Settele, J. & Vaissière, B. E. Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. *Econ.* 68, 810–821. https://doi.org/10.1016/j.ecolecon.2008.06.014 (2009).
- 24. Mayer, C. et al. Pollination ecology in the 21st century: key questions for future research. J. Pollinat. Ecol. 3, 8-23 (2011).
- Chavez, D. J. & Lyrene, P. M. Effects of self-pollination and cross-pollination of Vaccinium darrowii (Ericaceae) and other low-chill blueberries. Hortsci. Publ. Am. Soc. Hortic. Sci. 44, 1538–1541 (2009).
- Negussie, A., Achten, W. M. J., Verboven, H. A. F., Hermy, M. & Muys, B. Floral display and effects of natural and artificial pollination on fruiting and seed yield of the tropical biofuel crop *Jatropha curcas L. Global Change Biol. Bioenergy* 6, 210–218 (2014).
- Okubo, S., Yamada, M., Yamaura, T. & Akita, T. Effects of the pistil size and self-incompatibility on fruit production in *Curculigo latifolia* (Liliaceae). J. Jpn. Soc. Hortic. Sci. 79, 354–359 (2010).
- Benjamin, F. E. & Winfree, R. Lack of pollinators limits fruit production in commercial blueberry (*Vaccinium corymbosum*). Environ. Entomol. 43, 1574–1583 (2014).
- 29. Bennett, J. *et al.* A review of European studies on pollination networks and pollen limitation, and a case study designed to fill in a gap. *AoB PLANTS* https://doi.org/10.1093/aobpla/ply068 (2018).
- Wang, H., Matsushita, M., Tomaru, N., Nakagawa, M. & Arroyo, J. Differences in female reproductive success between female and hermaphrodite individuals in the subdioecious shrub *Eurya japonica* (Theaceae). *Plant Biol.* 17, 194–200 (2015).
- 31. Wang, H., Matsushita, M., Tomaru, N. & Nakagawa, M. High male fertility in males of a subdioecious shrub in hand-pollinated crosses. *AoB PLANTS* **8**, plw067 (2016).
- Shou, C., Wang, J., Zheng, X. & Guo, D. Inhibitory effect of jujuboside A on penicillin sodium induced hyperactivity in rat hippocampal CA1 area in vitro. Acta Pharmacol. Sin. 22, 986–990 (2001).
- Zhang, M. *et al.* Inhibitory effect of jujuboside A on glutamate-mediated excitatory signal pathway in hippocampus. *Planta Med.* 69, 692–695 (2003).
- 34. Yue, Y. *et al.* Wild jujube polysaccharides protect against experimental inflammatory bowel disease by enabling enhanced intestinal barrier function. *Food Funct.* **6**, 2568–2577 (2015).
- Han, D. et al. Jujuboside A protects H9C2 cells from isoproterenol-induced injury via activating PI3K/Akt/mTOR signaling pathway. Evidence-Based Complementary Alternative Medicine 2016 (2016).
- Lu, J., Liu, M., Mao, Y. & Shen, L. Effects of vesicular-arbuscular mycorrhizae on the drought resistance of wild jujube (*Zizyphs spinosus* Hu) seedlings. Front. Agric. China 1, 468–471 (2007).

- 37. Zhang, S. *et al.* Threshold effects of photosynthetic efficiency parameters of wild jujube in response to soil moisture variation on shell beach ridges, Shandong, China. *Plant Biosyst.* **148**, 140–149 (2014).
- 38. Wang, Q. Y. The developments of embryo and endosperm of Zizyphus jujuba mill. J. Integr. Plant Biol. 25, 32-37 (1983).
- Cruden, R. W. Pollen-ovule ratios: a conservative indicator of breeding systems in flowering plants. Evolution 31, 32–46. https:// doi.org/10.1111/j.1558-5646.1977.tb00979.x (1977).
- 40. Dafni, A. Pollination ecology: A practical approach. (1992).
- Barrett, S. C. H. The evolution of mating strategies in flowering plants. Trends Plant Sci. 3, 335–341. https://doi.org/10.1016/s1360-1385(98)01299-0 (1998).
- Carr, D. E. & Dudash, M. R. Recent approaches into the genetic basis of inbreeding depression in plants. *Philos. Trans. R. Soc. Lond. Ser. B: Biol. Sci.* 358, 1071–1084 (2003).
- Lloyd, D. G. & Webb, C. The avoidance of interference between the presentation of pollen and stigmas in angiosperms I. Dichogamy. NZ J. Bot. 24, 135–162 (1986).
- 44. Ren, M. Stamen movements in hermaphroditic flowers: diversity and adaptive significance. J. Plant Ecol. (Chin. Vers.) 34, 867–875 (2010).
- Xiao, C.-L. *et al.* Sequential stamen maturation and movement in a protandrous herb: mechanisms increasing pollination efficiency and reducing sexual interference. *AoB PLANTS* 9, plx019. https://doi.org/10.1093/aobpla/plx019 (2017).
- Nagy, E. S., Strong, L. & Galloway, L. F. Contribution of delayed autonomous selfing to reproductive success in mountain laurel, Kalmia latifolia (Ericaceae). Am. Midl. Nat. 142, 39–47 (1999).
- 47. Liu, K.-W. et al. Pollination: self-fertilization strategy in an orchid. Nature 441, 945 (2006).
- Ye, Z.-M., Jin, X.-F., Yang, J., Wang, Q.-F. & Yang, C.-F. Accurate position exchange of stamen and stigma by movement in opposite direction resolves the herkogamy dilemma in a protandrous plant, *Ajuga decumbens* (Labiatae). *AoB PLANTS* https://doi.org/10. 1093/aobpla/plz052 (2019).
- 49. Brantjes, N. & De Vos, O. The explosive release of pollen in flowers of Hyptis (Lamiaceae). New Phytol. 87, 425-430 (1981).
- 50. Guerrina, M., Casazza, G., Conti, E., Macrì, C. & Minuto, L. Reproductive biology of an Alpic paleo-endemic in a changing climate. J. Plant. Res. 129, 477–485 (2016).
- 51. Bawa, K. S. & Beach, J. H. Evolution of sexual systems in flowering plants. Ann. Mo. Bot. Garden 68, 254-274 (1981).
- Dietzsch, A. C., Stanley, D. A. & Stout, J. C. Relative abundance of an invasive alien plant affects native pollination processes. Oecologia 167, 469–479 (2011).
- Ollerton, J. The evolution of pollinator-plant relationships within the arthropods. Evolution and phylogeny of the arthropoda. Entomology Society of Aragon, Zaragoza, 741–758 (1999).
- 54. Blaauw, B. R. & Isaacs, R. Flower plantings increase wild bee abundance and the pollination services provided to a pollinationdependent crop. J. Appl. Ecol. 51, 890–898 (2014).
- Inouye, D. W., Larson, B. M., Ssymank, A. & Kevan, P. G. Flies and flowers III: ecology of foraging and pollination. J. Pollinat. Ecol. 16, 115–133 (2015).
- 56. Rader, R., Bartomeus, I., Garibaldi, L. A., Garratt, M. P. D. & Woyciechowski, M. Non-bee insects are important contributors to global crop pollination. *Proc. Natl. Acad. Sci. U.S.A.* 113, 146–151 (2016).
- 57. Corbet, S. A. Pollination and the weather. Isr. J. Plant Sci. 39, 13-30 (1990)
- Tuell, J. K. & Isaacs, R. Weather during bloom affects pollination and yield of highbush blueberry. J. Econ. Entomol. 103, 557–562 (2010).
- Ellis, C. R., Feltham, H., Park, K., Hanley, N. & Goulson, D. Seasonal complementary in pollinators of soft-fruit crops. *Basic Appl. Ecol.* 19, 45–55 (2017).
- Wang, W., Liu, Y., Chen, F.-D. & Dai, H.-G. Behavior and activity rhythm of flower-visiting insects on *Chrysanthemum morifolium* in Nanjing suburb. *Shengtaixue Zazhi* 27, 1167–1172 (2008).
- Waser, N. M., Chittka, L., Price, M. V., Williams, N. M. & Ollerton, J. Generalization in pollination systems, and why it matters. Ecology 77, 1043–1060 (1996).
- Navarro-Pérez, M., López, J., Rodríguez-Riaño, T. & Ortega-Olivencia, A. Reproductive system of two Mediterranean Scrophularia species with large, showy flowers. *Bot. Lett.* 166, 467–477 (2019).
- 63. Elle, E. Floral adaptations and biotic and abiotic selection pressures. *Plant adaptation: Molecular genetics and ecology. National Research Council of Canada, Ottawa, Ontario,* 111–118 (2004).
- Redmond, C. M. & Stout, J. C. Breeding system and pollination ecology of a potentially invasive alien *Clematis vitalba* L. Ireland. *J. Plant Ecol.* 11, 56–63. https://doi.org/10.1093/jpe/rtw137%JJournalofPlantEcology (2018).
- Byers, D. & Waller, D. Do plant populations purge their genetic load? Effects of population size and mating history on inbreeding depression. Ann. Rev. Ecol. Syst. 30, 479–513 (1999).
- Lande, R. & Schemske, D. W. The evolution of self-fertilization and inbreeding depression in plants I. Genetic models. *Evolution* 39, 24–40 (1985).
- 67. Zhang, C. et al. The genetic basis of inbreeding depression in potato. Nat. Genet. 51, 374-378 (2019).
- 68. Jones, C. E. & Little, R. J. Handbook of Experimental Pollination Biology (Scientific and Academic Editions, New York, 1983).
- Liu, P. et al. Study on the biological basis of pollination in Chinese Jujube (Zizyphus jujuba) and Wild Jujube (Z. spinosa). Journal of Fruit Science 21(3), 224–228 (2004).
- Sun, Y., Wu, C., Wang, D. & Wang, Z. Comparative Study on Floral Organ Structure, Pollen Morphology and Viability of Ziziphus acdiojujuba. Chin. Agric. Sci. Bull. 32(4), 87–91 (2016).

Acknowledgements

This work was financially supported by the Special Foundation for National Science and Technology Basic Resources Investigation of China (2019FY202300), Research Foundation of Qingdao Forest Ecosystem, and Fundamental Research Funds of Shandong University.

Author contributions

P. Z., H. W., and R. W. designed the experiments. F. W., X. S., R. C., X. L., X. L. and J. D. performed the experiments. T. H. identified insects. F. W. wrote the main manuscript text. All authors revised and approved the manuscript.

Competing interests

The authors declare no competing interests. We declare that we obtained the permission to collect Ziziphus jujuba var. spinosa in present study from Shandong University and Qingdao Landscape and Forestry Bureau. Meanwhile, the experimental research and field studies on *Z. jujuba* var. *spinosa* (either cultivated or wild) in present study, including the collection of plant material, were complied with ecology experiment guidelines of Shandong University.

Additional information

Correspondence and requests for materials should be addressed to P.Z.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2021