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OPEN Update analysis on the association between Methionine synthase rs1805087 A/G variant and risk of prostate cancer

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Previous studies have investigated the association of the rs1805087 A/G variant of Methionine synthase gene with the susceptibility to prostate cancer (PCa). Nevertheless, the conclusions remain divergent. We performed a systemic analysis with odds ratios (ORs) and 95% confidence intervals (95% CIs) to assess Methionine synthase rs1805087 A/G variant and PCa risk. Furthermore, we utilized in silico analysis to investigate the relationship between Methionine synthase expression and the overall survival (OS) time. Totally, 10,666 PCa patients and 40,750 controls were included. We observed that Methionine synthase rs1805087 A/G variant is associated with an elevated risk of PCa (G-allele vs. A-allele: OR = 1.06, 95% CI = 1.01–1.11, P = 0.013; heterozygous model: OR = 1.08, 95% CI=1.02-1.14, P=0.009; dominant model: OR=1.08, 95% CI=1.02-1.14, P=0.007). During stratified analysis, similar results were obtained in Asian populations, hospital-based, high quality studies and that with large sample size. Moreover, in silico analysis indicated the *Methionine synthase* expression is down-regulated in both young and old PCa subjects (P < 0.05). Compared with the normal subjects, the down-regulated expression of Methionine synthase was found in PCa cases with Gleason score 6 to 9. Our study showed that Methionine synthase rs1805087 A/G variant may be associated with susceptibility of PCa, especially in Asian populations, hospital-based studies and that with high quality and large sample size. Furthermore, Methionine synthase rs1805087 A/G variant may be related to the prognosis of PCa.

Abbreviations

CIs	Confidence intervals
GWAS	Genome-wide association study
HWE	Hardy-Weinberg equilibrium of controls
IHS	Immunohistochemical staining
MAF	Minor allele frequencies
HB	Hospital-based
MLH1	Mutl homolog—1
NA	Not applicable
OS	Overall survival
PB	Population-based
NOS	Newcastle-Ottawa Scale
ORs	Odds ratios

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PBS	Phosphate buffer saline
PCR-RFLP	Polymerase chain reaction-restriction fragment length polymorphism
PCa	Prostate cancer
PMS2	Postmeiotic segregation increased—2)
RT-PCR	Real-time PCR
TCGA	The cancer genome Atlas
vs	Versus

Prostate cancer (PCa) remains the most commonly occurring non-cutaneous carcinoma. It can be considered as the leading cause of cancer-related deaths among men in Western countries¹. Although the incidence and mortality rate of PCa in Asian populations is lower than that in Western countries², they have been increasing remarkably in recent years³. In China, the overall incidence is also increasing. PCa has become the most common solid tumor in urban male individuals. Therefore this highlights the need of strategies to prevent advanced PCa^{4,5}. Up to now, various factors, such as age, hormone exposure and gene mutations, have been proved to be correlated with the development of PCa^{6,7}. Particularly, growing evidence has indicated that genetic factors may play a crucial role in the etiology of PCa. For instance, the susceptibility of PCa would be increased around two to fivefold in patients suffering from Lynch syndrome. This disease is caused by gene mutations including *PMS2* (*Postmeiotic segregation increased*—2) and *MLH1* (*Mutl homolog*—1)^{8–10}.

Previous studies have shown that folate metabolizing genes play a central role in carcinogenesis. They are involved in the process of DNA repair and methylation^{11,12}. *Methionine synthase* is located on chromosome 1 (1q43) and with 34 exons. It encodes a core enzyme in folate pathway with 1,265 amino acids (molecular weight 140.5 kDa)¹³. *Methionine synthase* rs1805087 A/G variant is the most common mutation. It can lead to an aspartic acid to glycine transition at position 919 of the polypeptide chain¹⁴⁻¹⁶. Previous researches have indicated that this variant is involved in DNA methylation and elevation of homocysteine levels, therefore regulating the enzymatic activity of *Methionine synthase*¹⁷. The association of *Methionine synthase* rs1805087 A/G variant with susceptibility to PCa was investigated by several studies; however, their conclusions were divergent. In 2009, a systemic analysis evaluated the association between *Methionine synthase* rs1805087 A/G variant and susceptibility of PCa; nevertheless, they indicated no significant effects of this mutation on PCa risk¹⁸. From then on, another research assessed the association between this polymorphism and PCa susceptibility in Han Chinese population. They revealed that *Methionine synthase* rs1805087 A/G variant and PCa susceptibility in larger sample size using multiple analyses to acquire convincing conclusion¹⁸⁻³².

Methods

Identification of relevant literature. We conducted a comprehensive literature search according to EMbase, PubMed, Google Scholar, Web of Science, and Chinese SinoMed databases. The keywords are as follows: (MTR OR METH OR methionine synthase) AND (variant OR single nucleotide polymorphism) AND (prostate cancer OR tumor) (last search updated on May 01, 2020). In addition, the reference lists of reviews or supplementary material of source documents were also retrieved for further research.

Inclusion criteria and exclusion criteria. Studies were enrolled in our analysis according to the following criteria: (a) evaluating the relationship between PCa and *Methionine synthase* rs1805087 A/G polymorphism; (b) containing adequate information for all genotypes; and (c) case–control studies. Furthermore, studies should be removed if: (a) no control population was included; (b) research focuses on other diseases rather than cancer; (c) repeated previous publications.

Data extraction and quality assessment. Relevant data were independently screened by two of the authors according to the selection criteria. The quality assessment of the included studies was investigated by Newcastle–Ottawa Scale (NOS). The NOS score ranges from 0 to 9 stars. A research can be considered as high-quality if it obtained seven or more stars. The following items were extracted: the first author's name, publication year, race, source of control, score of NOS, genotype frequency, age range, Hardy–Weinberg Equilibrium (HWE) of cases and controls, sample size, and method. G-allele is a minor allele (mutated gene) for *Methionine synthase* rs1805087 A/G variant. On the other hand, the D-allele is a wild type and considered to be a low-risk allele. Five genetic models were selected in the current study: allelic comparison (G-allele *vs.* A-allele), homozygote model (GG *vs.* AA), heterozygote comparison (GA vs. AA), dominant contrast (GG+GA *vs.* AA), and recessive model (GG *vs.* GA+AA).

Statistical analysis. The strength of relationship between *Methionine synthase* rs1805087 A/G variant and risk of PCa was assessed by odds ratios (ORs) and 95% confidence intervals (CIs). We adopt *Z*-test to measure statistical significance of ORs. Assumption of heterogeneity was calculated by Chi-square-based *Q*-test. Fixed-effect model (Mantel–Haenszel method) was used when *P* value of *Q*-test was more than 0.05; otherwise, the random-effect model was used^{33,34}. Subgroup analyses were conducted by race, sample size of case, quality assessment, and source of controls. *P* value for HWE was evaluated by web-based program (https://ihg2. helmholtz-muenchen.de/cgibin/hw/hwa1.pl) ³⁵. *P* value > 0.05 revealed an HWE balance. Sensitivity analysis was adopted to explore the effect of single study on the OR by sequential exclusion of individual study³⁶. *I*² value < 50% indicates no statistical heterogeneity among studies. We used the Begg's funnel plot and Egger's test

to evaluate the publication bias^{37,38}. P>0.05 indicates no statistical significance. All the statistical process was conducted using software STATA v11.0 (Stata Corporation, College Station, TX).

Study population and immunohistochemical staining (IHS). Totally, 200 pathologically confirmed PCa patients were enrolled from our centers. Distribution of characteristics of these patients has been listed in our previous study⁴². The written informed consent was required from each participant. The above study protocol has been approved by ethics committee of the Affiliated Changzhou No.2 People's Hospital of Nanjing Medical University and Affiliated Hospital of Jiangnan University. IHS was used to test the *Methionine synthase* expression in PCa cases enrolled in our centers. We incubated the paraffin section of PRAD in hydrogen per-oxide (1%), and then washed it in phosphate buffer saline (PBS). Goat serum was utilized to block binding of non-specific proteins. These sections were incubated with anti *Methionine synthase* antibody at 1:200. Immuno-reactive site was brown using diaminobenzidine. All methods in the present study were conducted in accordance with the relevant guidelines and regulations.

In silico analysis of *Methionine synthase* **expression.** We employed the online database to explore the expression of *Methionine synthase* in PCa and control counterparts (https://gemini.cancer-pku.cn/)³⁹. We also used The Cancer Genome Atlas (TCGA) samples to investigate the expression in PCa based on patients' age, Gleason score, and nodal metastasis status. This database contains expression profiles of 52 PCa subjects and 496 controls (https://ualcan.path.uab.edu/analysis.html). We evaluated the effects of *Methionine synthase* rs1805087 A/G variant by SNAP tool (https://rostlab.org/services/snap/). String online server was also employed to assess the network of Methionine synthase interaction (https://string-db.org/)⁴⁷.

Ethics approval and consent to participate. The present study was approved by ethics committee of the Affiliated Changzhou No.2 People's Hospital of Nanjing Medical University and ethics committee of Affiliated Hospital of Jiangnan University.

Results

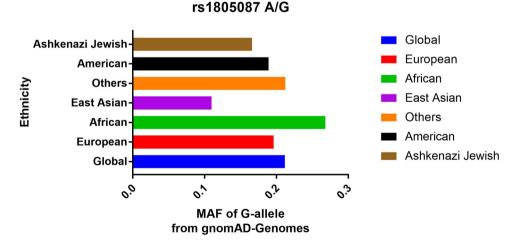
Characteristics of relevant studies. A total of 15 case–control studies were included in our study. The PRIZMA statement has been presented in the Supplementary material Table 1. Totally, 10,666 PCa patients and 40,750 controls were included in the current analysis (Table 1). Subgroup analyses were based on the following criteria: (a) race: 10 studies were conducted in Europeans, 3 studies were based on Asian populations, only one study was on Africans and South Americans; (b) source of controls: there were 6 HB studies and 9 PB studies in the present analysis; (c) quality assessment: 12 studies were of high quality and 3 studies were of low quality. (d) Sample size of case: 9 were large sample size studies and the rest were small sample size studies. Furthermore, we investigated the minor allele frequencies (MAF) in worldwide populations according to genome database (https://www.ncbi.nlm.nih.gov/snp). In Global, G=0.2118; Europeans, G=0.1961; Americans, G=0.1890; Africans, G=0.2679; East Asians, G=0.1098; Ashkenazi Jewish, G=0.1660; and Others, G=0.2123 (Fig. 1).

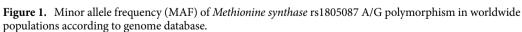
Systemic analysis. In the overall analysis, we observed that *Methionine synthase* rs1805087 A/G variant was associated with increased risk of PCa (Table 2) under three genetic models. For GG+GA vs. AA comparison: OR = 1.08, 95% CI = 1.02–1.14, P value for heterogeneity = 0.183, P = 0.007, $I^2 = 24.5$ (Fig. 2A); for G-allele vs. A-allele contrast: OR = 1.06, 95% CI = 1.01–1.11, $P_{heterogeneity} = 0.122$, P = 0.013, $I^2 = 30.9$; and for GA vs. AA model: OR = 1.08, 95% CI = 1.02–1.14, P value for heterogeneity = 0.353, P = 0.009, $I^2 = 8.9$. In stratification analysis by race, similar positive results were revealed in Asian descendants. During GG vs. AA model: OR = 1.93, 95% CI = 1.14 - 3.26, $P_{heterogeneity} = 0.390$, P = 0.014, $I^2 = 0$; for GG vs. GA + AA model: OR = 1.72, 95% CI = 1.02 - 2.89, 0.000 $P_{\text{heterogeneity}} = 0.767$, P = 0.041, $I^2 = 0$. In stratified analysis based on quality assessment, a positive relationship was revealed between Methionine synthase rs1805087 A/G variant and PCa risk in high quality studies (allelic contrast: OR = 1.05, 95% CI = 1.01–1.11, $P_{\text{heterogeneity}} = 0.417$, P = 0.029, $I^2 = 2.8$; heterozygous model: OR = 1.07, 95% CI = 1.01–1.13, $P_{\text{heterogeneity}} = 0.696$, P = 0.020, $I^2 = 0$; dominant comparison: OR = 1.07, 95% CI = 1.01–1.13, $P_{\text{heterogeneity}} = 0.576, P = 0.018, I^2 = 0$), but not in low quality studies (allelic contrast: OR = 1.27, 95% CI = 0.81-1.98, 1.25, 1.2 $P_{\text{heterogeneity}} = 0.019, P = 0.295, I^2 = 74.8$; heterozygous model: OR = 1.31, 95% CI = 0.80-2.16, $P_{\text{heterogeneity}} = 0.046$, P = 0.281, $I^2 = 67.5$; dominant comparison: OR = 1.35, 95% CI = 0.79-2.32, $P_{heterogeneity} = 0.019$, P = 0.270, $I^2 = 74.8$) (Fig. 2B). Furthermore, in subgroup analysis based on source of control and sample size, similar findings were obtained between Methionine synthase rs1805087 A/G variant and PCa susceptibility in hospital-based studies (allelic contrast: OR = 1.15, 95% CI = 1.02 - 1.29, $P_{heterogeneity} = 0.137$, P = 0.018, $I^2 = 40.3$; heterozygous model: OR = 1.15, 95% CI = 1.01 - 1.32, $P_{heterogeneity} = 0.245$, P = 0.041, $I^2 = 25.3$; dominant comparison: OR = 1.16, 95% CI = 1.02 - 1.32, $P_{heterogeneity} = 0.133$, P = 0.023, $I^2 = 40.9$, Fig. 3A) and studies with larger sample size (allelic contrast) and studies with larger sample size (allelic contrast) and studies with larger sample size (allelic contrast). trast: OR = 1.05, 95% CI = 1.00–1.11, $P_{\text{heterogeneity}} = 0.226$, P = 0.039, $I^2 = 27.8$; GA vs. AA model: OR = 1.07, 95% CI = 1.00 - 1.13, P value for heterogeneity = 0.309, P = 0.036, I² = 16.2; GG + GA vs. AA model: OR = 1.07, 95% CI = 1.01–1.13, *P* value for heterogeneity = 0.277, *P* = 0.029, I^2 = 20.8, Fig. 3B).

In silico analysis and IHS for *Methionine synthase* expression. We employed in silico tool to investigate the expression of *Methionine synthase* among 496 PCa patients and 52 controls. The overall result showed that the methionine synthase expression is down-regulated in both young and old PCa subjects (P < 0.05, Fig. 4A). In addition, we assessed whether the level of *Methionine synthase* expression affected the OS time of patients with PCa. As shown in Fig. 4B, PCa patients with high expression of *Methionine synthase* may have a shorter OS time in the first 5 years. Nevertheless, with the passage of time, this positive relationship would not be

First							Age range (years)	Case				Contro	ol			Sample	
author	Year	Race	Case	Control	Source	NOS	Case	Control	GG	GA	AA	HWE	GG	GA	AA	HWE	size	Method
Ebra- himi	2017	Asian	100	100	НВ	6	NA	NA	13	53	34	0.276	6	37	57	0.999	<1,000	PCR– RFLP
Qu	2016	Asian	1817	2026	HB	8	66.7 ± 7.2	66.9 ± 6.8	20	316	1,481	0.496	15	319	1692	0.993	>1,000	RT-PCR
López- Cortés	2013	SA	104	110	РВ	8	NA	NA	3	9	92	< 0.001	1	4	105	0.001	<1,000	PCR– RFLP
Jackson	2013	African	199	205	HB	7	67.8 ± 7.8	61.7 ± 10.7	20	82	97	0.664	24	82	99	0.274	<1,000	Taqman
Weiner	2012	Euro- pean	370	285	РВ	6	69±8	59±17	15	134	221	0.339	16	96	173	0.580	<1,000	RT-PCR
Cai	2010	Asian	217	220	НВ	6	72.4±12.2	72.8±12.3	5	27	185	0.003	3	29	188	0.139	<1,000	PCR– RFLP
Collin	2010	Euro- pean	49	261	РВ	8	62.6±5.1	NA	1	16	32	0.534	10	77	174	0.686	<1,000	GWAS
Mura- bito	2007	Euro- pean	172	231	РВ	7	66.0(43- 85)	76.0(83– 95)	7	55	110	0.970	9	69	153	0.728	<1,000	GWAS
Stevens	2009	Euro- pean	1,094	1,105	РВ	7	50-69	30-108	42	351	701	0.814	53	324	728	0.032	>1,000	Taqman
Yeager	2007	Euro- pean	1,162	1,112	РВ	7	NA	NA	48	376	738	0.990	38	340	734	0.858	>1,000	GWAS
ProtecT	2008	Euro- pean	1,600	2076	РВ	7	NA	NA	52	515	1,033	0.207	84	637	1,355	0.402	>1,000	Taqman
Eeles	2008	Euro- pean	1,850	1886	РВ	7	NA	NA	84	590	1,176	0.364	71	547	1,268	0.079	>1,000	GWAS
Amund- adottir	2006	Euro- pean	1619	30,779	РВ	7	mean 73.0	mean 52.0	60	466	1,093	0.242	1,044	9,160	20,575	0.532	>1,000	GWAS
Marchal	2008	Euro- pean	181	204	НВ	7	70.7±7.29	70.3±7.82	9	54	118	0.391	11	55	138	0.088	<1,000	Taqman
Kimura	2000	Euro- pean	132	150	НВ	9	65.6±6.0	62.0±11.4	4	41	87	0.753	4	44	102	0.773	<1,000	PCR– RFLP

Table 1. Study characteristics of *Methionine synthase* rs1805087 A/G variant included in this analysis. *GWAS* Genome-wide association study, *HWE* Hardy–Weinberg equilibrium of case and controls, *HB* Hospital-based, *NA* Not applicable, *PB* Population-based, *PCR–RFLP* polymerase chain reaction-restriction fragment length polymorphism, *RT-PCR* real-time PCR, *SA* South America.





populations according to genome database.

statistically significant (P > 0.05). TGCA database⁴⁰ showed evidence that the expression of *Methionine synthase* in PCa cases with Gleason score 6 to 9 was lower than that in normal subjects (P < 0.05, Fig. 5A). In addition, the *Methionine synthase* expression was both down-regulated in PCa patients with and without lymph node metastasis (P < 0.05, Fig. 5B). In order to assess whether the *Methionine synthase* rs1805087 A/G variant could impact the protein expression, we used SNAP tool to predict the mutation of *Methionine synthase* (Fig. 6A). The mutation score is 3, which indicate that *Methionine synthase* rs1805087 A/G variant is deleterious and effect (Fig. 6B).

				G-allele vs. A-allele								GG vs.	AA			GG+GA vs. AA				GG vs. GA+AA			
Variables	No	Cases/controls	OR (95% CI)	Р	Pheter	I ²	OR (95% CI)	Р	Pheter	<i>I</i> ²	OR (95% CI)	Р	Pheter	I ²	OR (95% CI)	Р	Pheter	I ²	OR (95% CI)	Р	P _{heter}	<i>I</i> ²
Total	15	10,666/40,750	1.06 (1.01–1.	.11)	0.013	0.122	30.9	1.08 (1.02- 1.14)	0.009	0.353	8.9	1.07 (0.93- 1.22)	0.346	0.376	6.8	1.08 (1.02- 1.14)	0.007	0.183	24.5	1.04 (0.91- 1.19)	0.589	0.573	0
Race																							
European	10	8,229/38,089	1.04 (0.99-1	.10)	0.106	0.758	0	1.06 (1.00- 1.13)	0.052	0.771	0	1.02 (0.89- 1.18)	0.749	0.673	0	1.06 (1.00- 1.12)	0.060	0.781	0	1.00 (0.87- 1.16)	0.659	0.974	0
Asian	3	2,134/2,346	1.33 (0.94–1.	.88)	0.109	0.050	66.6	1.31 (0.84– 2.05)	0.233	0.044	68.0	1.93 (1.14- 3.26)	0.014	0.390	0	1.38 (0.87- 2.20)	0.174	0.024	73.2	1.72 (1.02- 2.89)	0.041	0.767	0
South America	1	104/110	2.77 (1.05-7.	.29)	0.039	-	-	2.57 (0.77- 8.62)	0.127	-	-	3.42 (0.35– 33.49)	0.290	-	-	2.74 (0.93- 8.07)	0.067	-	-	3.24 (0.33- 31.63)	0.312	-	-
African	1	199/205	0.95 (0.71-1.	.28)	0.746	-	-	1.02 (0.67- 1.55)	0.923	-	-	0.85 (0.44- 1.64)	0.629	-	-	0.98 (0.66- 1.45)	0.928	-	-	0.84 (0.45- 1.58)	0.593	-	-
Source																							
РВ	9	8,020/37,845	1.04 (0.99–1.	.10)	0.095	0.287	17.5	1.06 (1.00- 1.13)	0.053	0.474	0	1.03 (0.89- 1.19)	0.688	0.467	0	1.06 (1.00- 1.12)	0.057	0.389	5.6	1.01 (0.87- 1.17)	0.901	0.461	0
НВ	6	2,646/2,905	1.15 (1.02-1.29)	0.018	0.137	40.3		1.15 (1.01- 1.32)	0.041	0.245	25.3	1.31 (0.92- 1.87)	0.138	0.303	17.2	1.16 (1.02- 1.32)	0.023	0.133	40.9	1.22 (0.86- 1.73)	0.273	0.568	0
Sample size																							
Large	6	9,142/38,984	1.05 (1.00-1.11)	0.039	0.226	27.8		1.07 (1.00- 1.13)	0.036	0.309	16.2	1.06 (0.92- 1.23)	0.424	0.258	23.4	1.07 (1.01- 1.13)	0.029	0.277	20.8	1.04 (0.90- 1.21)	0.588	0.258	23.4
Small	9	1524/1766	1.11 (0.97–1.26)	0.115	0.121	38.1		1.16 (0.99– 1.37)	0.064	0.390	5.4	1.09 (0.78- 1.52)	0.604	0.386	5.9	1.15 (0.99– 1.34)	0.069	0.178	30.1	1.02 (0.74- 1.41)	0.909	0.662	0
Quality					-																		
High	12	9,979/40,145	1.05 (1.01-1.11)	0.029	0.417	2.8		1.07 (1.01- 1.13)	0.020	0.696	0	1.05 (0.91- 1.21)	0.479	0674	0	1.07 (1.01- 1.13)	0.018	0.576	0	1.03 (0.90- 1.18)	0.678	0.676	0
Low	3	687/605	1.27 (0.81-1.98)	0.295	0.019	74.8		1.31 (0.80- 2.16)	0.281	0.046	67.5	1.56 (0.54– 4.49)	0.411	0.047	67.3	1.35 (0.79– 2.32)	0.270	0.019	74.8	1.16 (0.68- 1.97)	0.588	0.142	48.7

Table 2. Stratification analysis of *Methionine synthase* rs1805087 A/G polymorphism on the susceptibility to PCa. *No.* Number of case–control studies, $P_{heter} P$ value for heterogeneity. *HB* Hospital-based, *PB* population-based.

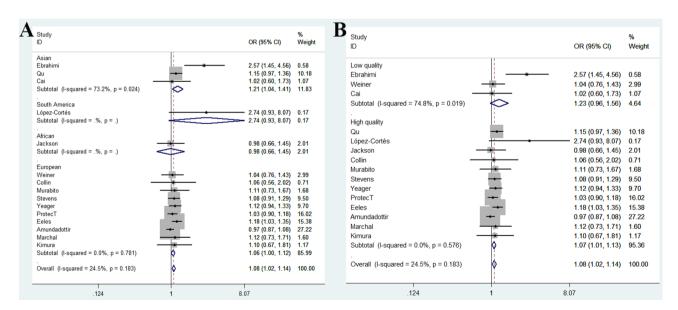


Figure 2. Forest plot shows odds ratio for the association between *Methionine synthase* rs1805087 A/G variant and PCa risk in subgroup analysis by race (\mathbf{A}) and quality assessment (\mathbf{B}) (GG+GA vs. AA, fixed-effects).

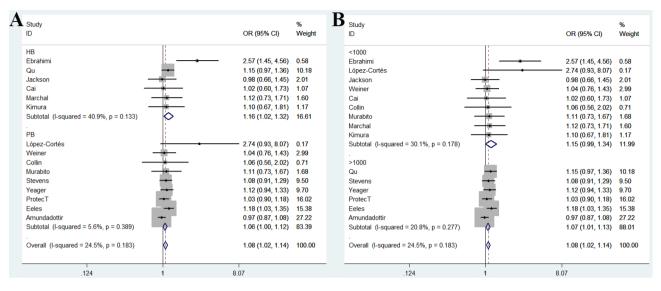


Figure 3. Forest plot of *Methionine synthase* rs1805087 A/G polymorphism in stratified analyses by source of control (**A**) and sample size (**B**).

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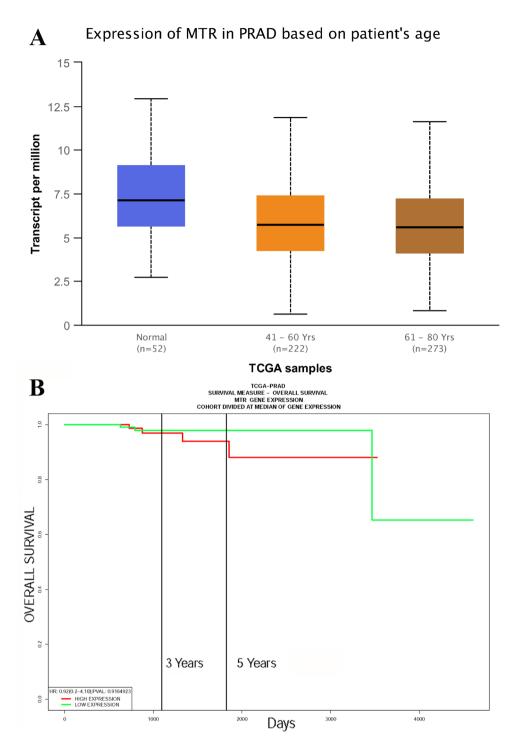
As described in Fig. 7A, at least 30 proteins are predicted to participate in the protein–protein interaction with Methionine synthase in *Homo sapiens*. The top 10 proteins are: MTRR: Methionine synthase reductase; MTHFR: Methylenetetrahydrofolate reductase; MMADHC: Methylmalonic aciduria and homocystinuria type D protein, mitochondrial; CBSL: Cystathionine beta-synthase-like protein; MMACHC: Methylmalonic aciduria and homocystinuria type C protein; CTH: Cystathionine gamma-lyase; AHCY: Adenosylhomocysteinase; SHMT1: Serine hydroxymethyltransferase; MAT1A: S-adenosylmethionine synthase isoform type-1; MTHFD1: Methyl-enetetrahydrofolate dehydrogenase (Fig. 7B). In order to further investigate the *Methionine synthase* expression in PCa tissues, we adopted IHS to assess its expression among PCa participants in our hospitals. Compared with the earlier stage, the down-regulated expression of *Methionine synthase* was found in more advanced PCa participants (stage > T3b versus = <T2c, P < 0.05, Fig. 8).

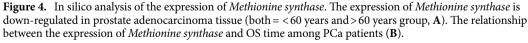
Sensitivity analysis and publication bias. The sensitivity analysis was conducted to investigate the effect of individual study on the OR. As described in Fig. 9A, no single study would significantly impact the overall OR. Both the Begg's funnel plot (Fig. 9B) and Egger's tests (Fig. 9C) were performed to assess the publication bias. No evidence of publication bias was revealed among any of the genetic models. For G-allele vs. A-allele: t=2.01, P=0.066; GA vs. AA: t=2.03, P=0.064; GG vs. AA: t=1.95, P=0.073; GG+GA vs. AA: t=2.06, P=0.060; and GG vs. GA + AA model: t=1.92, P=0.077.

Discussion

Identification of genetic mutations that related to susceptibility of carcinoma is useful to predict cancer risk and reveal the pathogenesis of various tumors^{41,42}. PCa remains one of the most commonly occurring carcinomas among men in Western countries. It has been clarified that several risk factors, such as family history, hormone exposure, lifestyle, may be associated with susceptibility of PCa⁴³⁻⁴⁶. Previous articles showed that *Methionine synthase* may be involved in the development of PCa²⁹. However, the assolation between *Methionine synthase* rs1805087 A/G variant and PCa risk remains ambiguous. For instance, a case–control study based on Iran descendants found that *Methionine synthase* rs1805087 A/G variant could influence stability and activity of its expression²². This polymorphism might be related to PCa risk in male individuals. However; another researchers did not indicate similar conclusions²⁶. A meta-analysis conducted 10 years ago revealed no significant associations between *Methionine synthase* rs1805087 A/G variant and PCa risk¹⁸. From then on, more and more researchers ascertained their results in different populations^{19,20,22,23,25,26}. Hence, in our systemic analysis, all eligible studies based on the inclusion criteria were summerized to ascertain a precise conclusion on the association between *Methionine synthase* rs1805087 A/G variant and PCa risk. Totally, 10,666 PCa patients and 40,750 controls were included to investigate the *Methionine synthase* polymorphism.

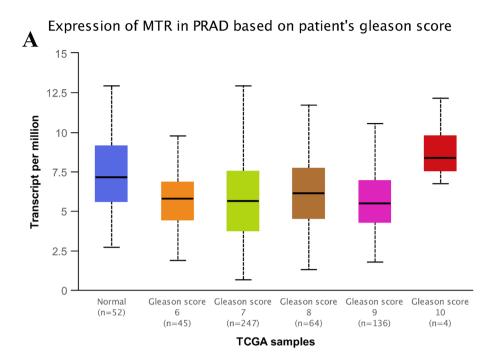
In our analysis, 15 case–control studies were included. The overall results revealed a positive association between *Methionine synthase* rs1805087 A/G variant and susceptibility of PCa under three genetic models. The conclusions acquired from our study were: person carrying the *Methionine synthase* G-allele may have an increased PCa risk. In stratification analysis by race, we observed similar positive results in Asian populations. *Methionine synthase* rs1805087 A/G variant might elevate PCa susceptibility as seen in high quality studies, hospital-based studies, and that with large sample size. Furthermore, we used online gene expression mini-database





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to investigate *Methionine synthase* expression in PCa and control counterparts. We found that *Methionine synthase* expression is down-regulated in both young and old PCa subjects. Compared with the normal subjects, the down-regulated expression of *Methionine synthase* was found in PCa cases with Gleason score 6 to 9, which was consistent with results of our IHS analysis. We also evaluated whether the *Methionine synthase* expression level influence the PCa patients' OS time. PCa patients with high *Methionine synthase* expression may have a shorter



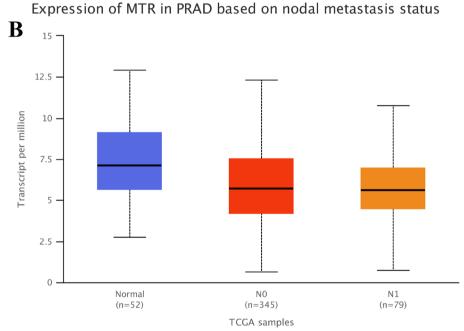
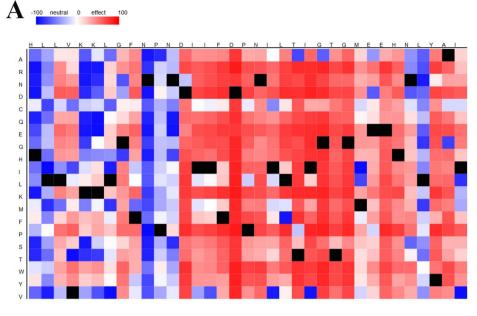


Figure 5. The expression of *Methionine synthase* in PCa based on patients' Gleason score (**A**) and nodal metastasis status (**B**).

OS time in the first 5 years. However, with the passage of time, this positive relationship would not be statistically significant. In addition, we used SNAP tool to evaluate the variation of *Methionine synthase*. The mutation score is 3, which indicate that *Methionine synthase* rs1805087 A/G variant is deleterious and effect.

Although considerable resources have been generated to evaluate association between *Methionine synthase* rs1805087 A/G variant and PCa susceptibility, there are some limitations that should be considered. To start with, only three Asian studies involving *Methionine synthase* rs1805087 A/G variant and PCa risk were retrieved, which indicates that the total participants count for Asian population remains relatively low for more comprehensive analysis. Second, all included case-control studies were retrospective, which may cause selection bias during the process of analysis. Third, lacking of some original data such as family history, lifestyle, and smoking



B

Wildtype Amino Acid♦	Position♦	Variant Amino Acid♦	Predicted Effect	Score	Expected Accuracy
D	919	A	neutral	-57	78%
D	919	R	neutral	-85	93%
D	919	N	neutral	-78	87%
D	919	D	neutral	-85	93%
D	919	С	effect	1	53%
D	919	Q	neutral	-89	93%
D	919	E	neutral	-86	93%
D	919	G	effect	3	53%
D	919	н	neutral	-70	82%
D	919	1	neutral	-45	72%
D	919	Ĺ	neutral	-52	78%
D	919	К	neutral	-78	87%
D	919	М	neutral	-17	57%
D	919	F	effect	74	85%
D	919	Р	neutral	-13	57%
D	919	S	neutral	-76	87%
D	919	т	neutral	-74	87%
D	919	W	effect	82	91%
D	919	Y	effect	54	75%
D	919	V	neutral	-49	72%

Figure 6. Evaluation of *Methionine synthase* rs1805087 A/G variant by SNAP tool (**A**). The mutation score is 3, which indicate that *Methionine synthase* rs1805087 A/G variant is deleterious and effect (**B**).

exposure, may limit the efficacy to further calculate adjusted OR. Fourth, studies with only English or Chinese language were included; therefore, some articles written in other languages may be missing, which may cause some bias in risk estimation. In spite of the limitations, there are several advantages in our analysis. First, HWE is very significant while evaluating genetic variations. *P* value for HWE is more than 0.05 in most of the included studies, which shows that the conclusion from the included studies is very stable. Second, studies with larger sample size particularly improved the statistical efficiency. Third, we indicated no publication bias while evaluating *Methionine synthase* rs1805087 A/G variant, so the conclusions of the present analysis are more convincing. As described in Fig. 7A, at least 30 proteins are predicted to participate in the protein–protein interaction with *Methionine synthase* in *Homo sapiens*. Nevertheless, there is not enough research on their further mechanism in PCa. Further researches are warranted to explore the interactions in more details.

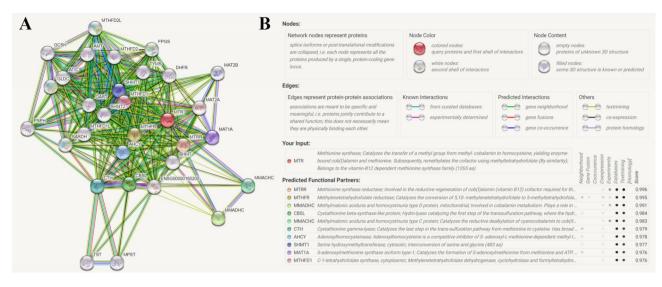
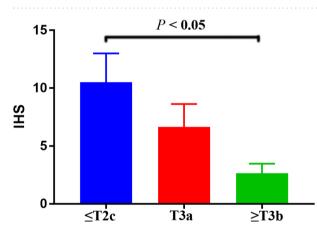
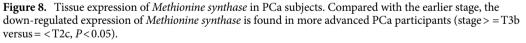


Figure 7. Methionine synthase crosstalk with other protein evaluated by String server. At least 30 proteins are predicted to participate in the protein–protein interaction with Methionine synthase in *Homo sapiens* (**A**). The top 10 proteins are: *MTRR* Methionine synthase reductase, *MTHFR* Methylenetetrahydrofolate reductase, *MMADHC* Methylmalonic aciduria and homocystinuria type D protein, mitochondrial, *CBSL* Cystathionine beta-synthase-like protein, *MMACHC* Methylmalonic aciduria and homocysteinase, *SHMT1* Serine hydroxymethyltransferase, *MAT1A* S-adenosylmethionine synthase isoform type-1, *MTHFD1* Methylenetetrahydrofolate dehydrogenase (**B**).





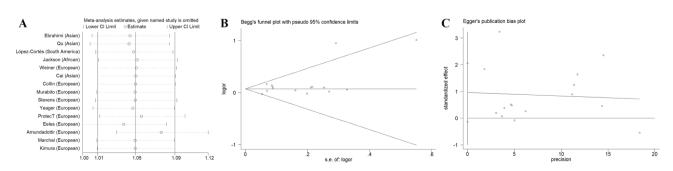


Figure 9. Publication bias analysis of *Methionine synthase* rs1805087 A/G variant and PCa risk. Sensitivity results were assessed by removing every single study in turn (**A**). No evidence of publication bias can be revealed through both Begg's funnel plot (**B**) and Egger's test(**C**).

Conclusions

Taken together, our study indicated that *Methionine synthase* rs1805087 A/G variant is associated with PCa susceptibility, especially in Asian descendants, hospital-based studies, high quality studies and that with large sample size. Moreover, *Methionine synthase* rs1805087 A/G variant may be related to the prognosis of PCa. Further studies containing more information such as lifestyle and smoking exposure, are warranted to assess this association in details.

Availability of data and materials

All the data analyzed in the present study is included in the manuscript.

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References

- 1. Bray, F. et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 68, 394–424 (2018).
- 2. Dy, G. W., Gore, J. L., Forouzanfar, M. H., Naghavi, M. & Fitzmaurice, C. Global burden of urologic cancers, 1990–2013. *Eur. Urol.* 71, 437–446 (2017).
- 3. Ito, K. Prostate cancer in Asian men. Nat. Rev. Urol. 11, 197-212 (2014).
- 4. Yang, L., Yuan, Y., Sun, T., Li, H. & Wang, N. Population-based cancer incidence analysis in Beijing, 2008–2012. Chin. J. Cancer Res. 27, 13–21 (2015).
- Chen, W., Zheng, R., Zeng, H. & Zhang, S. The updated incidences and mortalities of major cancers in China, 2011. Chin. J. Cancer 34, 502–507 (2015).
- 6. Stewart, R. W., Lizama, S., Peairs, K., Sateia, H. F. & Choi, Y. Screening for prostate cancer. Semin. Oncol. 44, 47-56 (2017).
- 7. Brawley, O. W., Ankerst, D. P. & Thompson, I. M. Screening for prostate cancer. CA Cancer J. Clin. 59, 264–273 (2009).
- 8. Engel, C. et al. Risks of less common cancers in proven mutation carriers with lynch syndrome. J. Clin. Oncol. 30, 4409-4415 (2012).
- 9. Broeke, S. W. et al. Cancer Risks for PMS2-Associated Lynch Syndrome. J. Clin. Oncol. 36, 2961–2968 (2018).
- 10. Raymond, V. M. et al. Elevated risk of prostate cancer among men with Lynch syndrome. J. Clin. Oncol. 31, 1713–1718 (2013).
- 11. Ly, A., Hoyt, L., Crowell, J. & Kim, Y. I. Folate and DNA methylation. Antioxid Redox Signal. 17, 302–326 (2012).
- 12. Duthie, S. J. Folate & cancer: how DNA damage, repair and methylation impact on colon carcinogenesis. J. Inherit Metab. Dis. 34, 101–109 (2011).
- Chen, L. H. et al. Human methionine synthase. cDNA cloning, gene localization, and expression. J. Biol. Chem. 272, 3628–3634 (1997).
- Leclerc, D. et al. Cloning and mapping of a cDNA for methionine synthase reductase, a flavoprotein defective in patients with homocystinuria. Proc. Natl. Acad. Sci. USA 95, 3059-3064 (1998).
- Matthews, R. G., Sheppard, C. & Goulding, C. Methylenetetrahydrofolate reductase and methionine synthase: biochemistry and molecular biology. *Eur. J. Pediatr.* 157, S54-59 (1998).
- 16. Leclerc, D. *et al.* Molecular cloning, expression and physical mapping of the human methionine synthase reductase gene. *Gene* 240, 75–88 (1999).
- 17. Sharp, L. & Little, J. Polymorphisms in genes involved in folate metabolism and colorectal neoplasia: A HuGE review. Am. J. Epidemiol. 159, 423-443 (2004).
- Collin, S. M. et al. Association of folate-pathway gene polymorphisms with the risk of prostate cancer: A population-based nested case-control study, systematic review, and meta-analysis. Cancer Epidemiol. Biomark. Prev. 18, 2528–2539 (2009).
- 19. Qu, Y. Y. *et al.* Functional variants of the 5-methyltetrahydrofolate-homocysteine methyltransferase gene significantly increase susceptibility to prostate cancer: Results from an ethnic Han Chinese population. *Sci. Rep.* **6**, 36264 (2016).
- Jackson, M. D. et al. Complex interaction between serum folate levels and genetic polymorphisms in folate pathway genes: biomarkers of prostate cancer aggressiveness. *Genes Nutr.* 8, 199–207 (2013).
- Murabito, J. M. et al. A genome-wide association study of breast and prostate cancer in the NHLBI's Framingham Heart Study. BMC Med. Genet. 8, S6 (2007).
- Ebrahimi, A., Hosseinzadeh, Colagar, A. & Karimian, M. Association of human methionine synthase-A2756G transition with prostate cancer: A case-control study and in silico analysis. Acta Med. Iran 55, 297–303 (2017).
- López-Cortés, A. et al. Genetic polymorphisms in MTHFR (C677T, A1298C), MTR (A2756G) and MTRR (A66G) genes associated with pathological characteristics of prostate cancer in the Ecuadorian population. Am. J. Med. Sci. 346, 447–454 (2013).
- 24. Eeles, R. A. et al. Multiple newly identified loci associated with prostate cancer susceptibility. Nat. Genet. 40, 316-321 (2008).
- Weiner, A. S. et al. Role of polymorphic variants of MTR gene A2756G and SHMT1 gene C1420T in the development of prostatic cancer in residents of the Western Siberian Region of Russia. Bull. Exp. Biol. Med. 152, 466–469 (2012).
- 26. Cai, D. *et al.* Association of polymorphisms in folate metabolic genes and prostate cancer risk: a case-control study in a Chinese population. *J. Genet.* **89**, 263–267 (2010).
- 27. Yeager, M. *et al.* Genome-wide association study of prostate cancer identifies a second risk locus at 8q24. *Nat. Genet.* **39**, 645–649 (2007).
- Stevens, V. L. *et al.* No association of single nucleotide polymorphisms in one-carbon metabolism genes with prostate cancer risk. *Cancer Epidemiol. Biomark. Prev.* 17, 3612–3614 (2008).
- Marchal, C. et al. Association between polymorphisms of folate-metabolizing enzymes and risk of prostate cancer. Eur. J. Surg. Oncol. 34, 805-810 (2008).
- Amundadottir, L. T. et al. A common variant associated with prostate cancer in European and African populations. Nat. Genet. 38, 652–658 (2006).
- Kimura, F. et al. Methyl group metabolism gene polymorphisms and susceptibility to prostatic carcinoma. Prostate 45, 225–231 (2000).
- Collin, S. M. et al. Associations of folate, vitamin B12, homocysteine, and folate-pathway polymorphisms with prostate-specific antigen velocity in men with localized prostate cancer. Cancer Epidemiol Biomark. Prev. 19, 2833–2838 (2010).
- 33. Mantel, N. & Haenszel, W. Statistical aspects of the analysis of data from retrospective studies of disease. J. Natl. Cancer Inst. 22, 719–748 (1959).
- 34. DerSimonian, R. & Laird, N. Meta-analysis in clinical trials. Control Clin. Trials 7, 177-188 (1986).
- 35. Zamora-Ros, R. *et al.* Dietary intakes and food sources of phenolic acids in the European Prospective Investigation into Cancer and Nutrition (EPIC) study. *Br. J. Nutr.* **110**, 1500–1511 (2013).

- Tobias, A. & Campbell, M. J. Modelling influenza epidemics in the relation between black smoke and total mortality. A sensitivity analysis. J. Epidemiol. Commun. Health 53, 583–584 (1999).
- Egger, M., Smith, G., Schneider, M. & Minder, C. Bias in meta-analysis detected by a simple, graphical test. BMJ 315, 629–634 (1997).
- Begg, C. B. & Mazumdar, M. Operating characteristics of a rank correlation test for publication bias. *Biometrics* 50, 1088–1101 (1994).
- Tang, Z., Li, C., Zhang, K., Yang, M. & Hu, X. GE-mini: a mobile APP for large-scale gene expression visualization. *Bioinformatics* 33, 941–943 (2017).
- 40. Chandrashekar, D. S. *et al.* UALCAN: A portal for facilitating tumor subgroup gene expression and survival analyses. *Neoplasia* 19, 649–658 (2017).
- Zhang, L. F. et al. VEGF gene rs3025039C/T and rs833052C/A variants are associated with bladder cancer risk in Asian descendants. J. Cell Biochem. 120, 10402–10412 (2019).
- 42. Shao, H. B. *et al.* Human methionine synthase A2756G polymorphism increases susceptibility to prostate cancer. *Aging (Albany NY)* **10**, 1776–1788 (2018).
- Tseng, M., Breslow, R. A., Graubard, B. I. & Ziegler, R. G. Dairy, calcium, and vitamin D intakes and prostate cancer risk in the National Health and nutrition examination epidemiologic follow-up Study cohort. Am. J. Clin. Nutr. 81, 1147–1154 (2005).
- Kesse, E. et al. Dairy products, calcium and phosphorus intake, and the risk of prostate cancer: results of the French prospective SU.VI.MAX (Supplémentation en Vitamines et Minéraux Antioxydants) study. Br. J. Nutr. 95, 539–545 (2006).
- 45. Mitrou, P. N. et al. A prospective study of dietary calcium, dairy products and prostate cancer risk (Finland). Int. J. Cancer 120, 2466-2473 (2007).
- Xu, B. et al. New insights into the association between AXIN2 148 C/T, 1365 C/T, and rs4791171 A/G variants and cancer risk. Cancer Cell Int. 19, 119 (2019).
- Szklarczyk, D. et al. STRING v11: protein–protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 47, D607-613 (2019).

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Author contributions

L.S. and L.Z. contributed to the study design, Z.Z., Y.Y.M. and K.X. searched the databases, H.W., Y.F.S. and W.Y. collected the full-text papers and screened the data, K.X. and W.Y. wrote the manuscript. L.S. and W.Z. were involved in the preparation of figures and tables. Y.F.S. and Y.Y.M. revised the manuscript, Y.Y.M. and L.Z. provided financial support. All authors approved the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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