SCIENTIFIC REPORTS

natureresearch

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OPEN Catalyst-Free One-Pot Synthesis of Densely Substituted Pyrazole-**Pyrazines as Anti-Colorectal Cancer** Agents

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The first catalyst-free post-Ugi cascade methodology was developed for expeditious access to structurally diverse and complex pyrazole-pyrazines in one-pot. This novel cascade reaction features an intramolecular N2-arylation of pyrazoles with allenes at the C- β position of triple bond. Screening in the colorectal cancer cell lines HCT116 and SW620 validated the feasibility of the methodology for generating bioactive compounds. The lead compound 7h which is active against HCT116 and SW620 with IC₅₀ of 1.3 and 1.8 μ M, respectively, can be synthesized and purified in a gram process synthetic scale in 7 hours. The mechanical studies indicated that compound 7h can induce cell cycle arrest in the G2/M phase and inhibit proliferation and viability in human colon cancer cells. Overall, compound 7h is represented as a promising starting point for the development of new anti-colorectal cancer drugs.

The Ugi four-component reaction (U-4CR) as a typical multicomponent reaction (MCR) continues to attract considerable interest¹. The post-Ugi condensation is known as a versatile and highly efficient synthetic strategy for the preparation of heterocyclic compounds²⁻⁴. In addition, the Ugi MCR provides expeditious access to complex and drug like chemical scaffolds with high iterative efficiency from commercially available starting materials⁵⁻⁸.

In recent years, the indazole scaffold as an important core structure has been investigated extensively as antibacterial agent^{9,10}, HIV-1 integrase inhibitor (I)¹¹, γ -secretase modulator (GSM) (II)¹², Weel kinase inhibitor (III)^{13,14} and PARP-1/PARP-2 inhibitor (IV) for the treatment of cancer (Fig. 1)^{15,16}.

Although synthetic studies of indazole derivatives have been mainly devoted to the intramolecular N1-arylation with metal catalysts^{17–19}, the intramolecular N2-arylation was also feasible^{20,21}. More recently, privileged scaffolds represented by 2H-indazole were prepared via the U-4CR with Au@Al-SBA15¹⁷, silver¹⁹, or copper as a catalyst²²⁻²⁴. In view of the cost-effective and environmentally friendly methodologies developed with MCRs for the synthesis of heterocyclic compounds^{25–27}, further exploring sustainable reaction conditions for the synthesis of 2*H*-indazoles would be still desirable.

During the process by using the post-Ugi cascade reaction for the construction of heterocycles in drug discovery campaign^{28,29}, a new methodology in our group was developed to synthesize two types of the 2*H*-indazole scaffolds³⁰⁻³². The methodology features with mild reaction, simple operation procedure, a broad scope of starting materials, and the absence of a catalyst. The 2H-indazoles were screened in two cancer cell lines HCT116 and SW620. Compound 7h which was synthesized and purified in a gram scale in 7 hours, is only 10 times less active than the legendary cancer drug paclitaxel. Herein we disclosed detailed synthesis and biological testing.

Results and Discussion

Chemistry. Using an optimized solvent 2,2,2-trifluoroethanol (TFE) shown in Scheme 1, the reaction of 1H-indazole-3-carboxylic acid (1) with phenylpropiolaldehyde (2), aniline (3a), and phenyl isocyanide (4a) in TFE at room temperature afforded the Ugi product 5a in high yield as expected.

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HIV-1 integrase inhibitor, $IC_{50} = 0.626 \ \mu M$



Wee1 kinaise inhibitor, $IC_{50} = 5.2 \text{ nM}$ Clinical trial, phase 2

Figure 1. Bioactive structures of 2H-indazoles.



A β 42 inhibitor, IC₅₀ = 18 nM



PARP-1/PARP-2 inhibitor, $IC_{50} = 3.8$ and 2.1nM Clinical trial, phase 3



Scheme 1. Synthetic routes for compound 7a.

With a large amount of compound 5a in hand, we then explored conditions for the post-Ugi cyclization reaction. In order to find environmental-friendly conditions, we investigated the cyclization reaction by testing inorganic or organic base as a catalyst instead of a metal catalyst (Table 1). Strikingly the Ugi product 5a was converted to compound 7a with inorganic base K_2CO_3 (2.0 equiv.) in 63% yield. Compound 6a, the tautomeric precursor of compound 7a was also isolated with 25% yield and the structure of 6a was proved by NMR spectra (see supporting information). Further, the desired product 7a was isolated in 78% yield, and the side product 6a was reduced to 7% yield by elevating the microwave irradiation temperature from 80 °C to 90 °C. Enhanced temperature to 110 °C led to 89% yield for compound 7a. Interestingly, 1,8-Diazabicyclo[5.4.0]undec-7-ene (DBU) and diisopropanolamine (DIPA), organic bases, could still afford compound 7a (entries 4-7) with excellent yields (71-87%). With these exciting results, an investigation without a base or acid was processed. Unprecedentedly, the desired product 7a was obtained just by microwave heating in DMF for 10 min (entries 8–10). The microwave heating at 110 °C for 20 min in DMF could afford the desired product 7a in 95% yield. A variety of different solvents were then tested (entries 11-17). For protic solvents (TFE and EtOH), compound 6a could be isolated together with the product 7a (entries 11–13, 16–17) with lower yields. Notably, switching the solvent DMF to MeCN or tetrahydrofuran (THF), we failed to isolate either the product 7a or the intermediate 6a (entries 14, 15). Therefore, the optimized reaction condition is entry 10: the reaction was heated with microwave irradiation at 110 °C for 20 min in DMF in the absence of a catalyst.

entry	cat	solv	conditions	yield (%) 6a ^a	yield (%) 7a ^a
1	K ₂ CO ₃	DMF	MW, 80 °C, 20 min	25	63
2	K ₂ CO ₃	DMF	MW, 90 °C, 10 min	7	78
3	K ₂ CO ₃	DMF	MW, 110 °C, 10 min	_	89
4	DBU	DMF	MW, 90 °C, 10 min	_	76
5	DBU	DMF	MW, 110 °C, 10 min	_	87
6	DIPA	DMF	MW, 90 °C, 10 min	_	71
7	DIPA	DMF	MW, 110 °C, 10 min	_	83
8	_	DMF	MW, 90 °C, 10 min	7	77
9	-	DMF	MW, 110 °C, 10 min	_	85
10	-	DMF	MW, 110°C, 20 min	_	95
11	_	TFE	MW, 90 °C, 10 min	12	23
12	_	TFE	MW, 110°C, 10 min	30	40
13	_	TFE	MW, 110 °C, 20 min	32	51
14	_	MeCN	MW, 90°C, 10 min	_	_
15	_	THF	MW, 90 °C, 10 min	_	_
16	_	EtOH	MW, 90 °C, 10 min	_	12
17	_	EtOH	MW, 110 °C, 10 min	10	15

Table 1. Optimization of the reaction conditions for the synthesis of compound 7a. ^aYield (%) based onthe integral of the HPLC peaks detected at 254 nm. MW = microwave irradiation. Reaction conditions: 5a(0.1 mmol), additive (2.0 eq.) in DMF (1.0 mL) under microwave irradiation.

Encouraged by good yields for both the Ugi and cyclization reactions, we investigated the cyclization reaction with the unpurified Ugi product **5a**. In one pot, the cascade reaction produced the final product **7a** in 64% yield for two steps. We then investigated the scope of this one-pot procedure by varying starting materials (in Scheme 2). First, either electron withdrawing or donating phenyl amines and aromatic or aliphatic substituted isocyanides were well tolerated. Strikingly, compounds **7d** and **7h** with small cyclopropanamine were also obtained with good yields. Phenyl pyrazoles (**7j-m**, **7o** and **7s**) or methyl indazoles (**7n**, **7p-r**) didn't make a significant difference for the reaction yield. It is worth to emphasize that the Ugi adduct **5** did not require purification by column chromatography, with the crude product having no discernible impact on the overall yield. In all cases, the initial Ugi products **5a-s** were used directly in the next step without further purification after the removal of solvent removal. A variety of different starting materials were successfully employed under the optimized conditions for the construction of structurally diverse pyrazole-pyrazines **7a-s** with yields in the range of 55–68% shown in Scheme 2, indicating a good functional group tolerance. The structure of compound **7i** was unequivocally confirmed by X-ray crystallography, as shown in Scheme 2 as well.

Mechanistically, under microwave irradiation the tautomer 8 could be formed. Microwave irradiation would promote an isomerization at the triple bond to generate the intermediate allene 8', which in turn would react with the pyrazole to afford the unstable compound 6. It was surprising to find that the intramolecular N2-arylation occurred without a catalyst, which is obviously different from the previous reports^{18–24,30,31}. This two-step operation provided the facile access to a variety of complex fused pyrazole-pyrazines 7 in good yields, making it suitable for the construction of small compound libraries and scale-up synthesis.

To expand this catalyst-free cyclization for the synthesis of other scaffolds, we investigated if the triple bond would still react with a 2*H*-indazole when it was present in different Ugi inputs. We replaced aniline with 2-propynylamine **10** to provide a triple bond in the amino-Ugi input as depicted in Scheme 3. Similarly to the synthesis of compound 7, the U-4CR with dazole carboxylic acid **1**, 2-propynylamine **10**, aldehyde **9** and isocyanide **4** progressed smoothly (Scheme 3). The U-4CR was completed at room temperature overnight and evaporated to dryness to give the crude product **11**, which was then subjected to the optimal conditions with the microwave irradiation at 110 °C for 20 min in DMF without a catalyst to give compound **14**. With this condition, we proceeded to investigate the synthesis scope of this new series of pyrazine-pyrazines **14**. In all cases, initial Ugi products **11** were subjected to the cyclization reaction following the solvent removal. A variety of different starting



Scheme 2. Scope of the Ugi cascade reaction route leading to fused pyrazole-pyrazines 7a-s. Reaction conditions: 1 (1.0 mmol), 2 (1.0 mmol), 3 (1.0 mmol) and 4 (1.0 mmol) in TFE (2.0 mL) under air; After the removal of solvent, diluted by DMF (2.0 mL), under microwave irradiation at 110 °C, for 20 min. Overall yield of isolated product. See general procedure for synthesizing 7 in SI.

materials were successfully employed in one-pot for the construction of structurally diverse pyrazole-pyrazines **14a-j** with yields of 53–64%.

With a similar mechanism to the preparation of compound 7, the tautomerization of the Ugi adduct 11 provides intermediate 12 and allenamide 12' under the microwave irradiation. The nucleophic attach of the allene bond from the pyrazole nitrogen would sequentially afford the unstable intermediate 13. The double bond isomerization would then give the stable structure 14. Remarkably, two types of structurally distinct and complex final products 7 or 14 can be synthesized without a catalyst in one-pot, just by varying a triple bond in different Ugi inputs.



Scheme 3. Scope of the Ugi cascade reaction route leading to fused pyrazole-pyrazines 14a-j. Reaction conditions: 1 (1.0 mmol), 9 (1.0 mmol), 10 (1.0 mmol) and 4 (1.0 mmol) in TFE (2.0 mL) under air; After the removal of solvent, diluted by DMF (2.0 mL), under microwave irradiation at 110 °C, for 20 min. Overall yield of isolated product. See general procedure for synthesizing 14 in SI.

Biology and structure activity relationship (SAR). To evaluate the potential for developing a drug lead from compounds 7a-s and 14a-j, the MTT assay was used to measure cancer cell viability upon the drug treatment $(Table 2)^{33}$. Compounds 7a and 7c with an unsubstituted phenyl and compound 7i with 2-chloro-substituted phenyl at the R_1 position of 7A (Scheme 2) were completely inactive, while compounds 7b, 7f and 7g with either an electron donating (-OMe) or withdrawing group (-Br) at the 4-position of the phenyl group were potent with IC_{50} less than 5 μ M in both cancer cells, indicating the importance of substituents at the 4-position of the phenyl group. The critical role of a 4-substituted phenyl group at the R₁ position was further confirmed with derivatives 7j, 7k, 7l, 7m, 7o, and 7s, which possess a different pyrazolopyridone core structure. Compounds 7l and 7o with a 4-substituted phenyl group demonstrated a good activity; in contrast, other compounds including compound 7m with a 3-substituted phenyl group was not active at 20 µM. Surprisingly, four compounds (compounds 7n and 7p-7r) with a methyl substituted indazole at the R₃ position of 7A (Scheme 2) were all inactive against HCT116 and SW620 cell lines at 20 μ M. Furthermore, replacement of the non-substituted phenyl (R₁ of **7A**) group of compound 7a with a simple cyclopropyl group (7d) increased the activity significantly. Especially when the R_2 group (7A, Scheme 2) was switched from the phenylmethyl group (7a) to a longer phenylethyl group, compound (7h) is equally potent for both cell lines with IC_{50} of $1.3 \,\mu$ M and $1.8 \,\mu$ M against HCT116 and SW620 cell lines, respectively. Finally, all fused pyrazole-pyrazines 14a-j (Scheme 3) were not active at 20 µM.

As shown in Fig. 2A, the compound 7h efficiently inhibited colon cancer cell viability in a time- and dose-dependent manner. Moreover, soft agar assay *in vitro* was performed to evaluate the effect of the compound 7h in colony formation, and the results demonstrated that smaller and lesser colonies were formed in treated groups (1, 2 and 4 μ mol/L) compared with control groups in both glioblastoma cell lines (Fig. 2B,C). These results supported that the compound 7h dramatically inhibited cell viability and proliferation in human colon cells.

To explore the underlying mechanism responsible to compound **7h**-induced cell proliferation, cell cycle distribution was analyzed using flow cytometry in colon cancer cells treated with or without the inhibitor. Control

compound	НСТ116 (μМ)	SW620 (µM)
7a	>20	>20
7b	2.1 ± 0.2	1.4 ± 0.2
7c	>20	18.7±6.0
7d	1.2 ± 0.1	5.3 ± 1.2
7e	>20	>20
7 f	2.7 ± 0.7	3.8±0.6
7 g	1.8 ± 0.2	1.6 ± 0.3
7 h	1.3 ± 0.4	1.8 ± 0.3
7i	>20	>20
7j	>20	>20
7k	17.2 ± 5.1	14.5 ± 3.7
71	2.9 ± 0.8	1.4 ± 0.5
7 m	>20	>20
7n	>20	>20
70	3.4 ± 1.0	2.1 ± 0.9
7p	>20	>20
7 q	>20	>20
7 r	>20	>20
7 s	8.5 ± 1.0	11.3 ± 3.3
14a	>20	>20
14b	>20	>20
14c	>20	>20
14d	>20	>20
14e	>20	>20
14 f	>20	>20
14 g	>20	>20
14h	>20	>20
14i	>20	>20
14j	>20	>20
Paclitaxel	0.14 ± 0.05	-

 Table 2.
 Anticancer activities of compounds 7 and 14^a. ^aAll MTT assays were repeated three times by using six samples per assay.

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and treated cells were harvested and stained with PI and then measured by flow cytometry. As shown in Fig. 3, compound **7h** affected the cell cycle progression in a dose-dependent manner in HCT116 and SW620 cell lines. Representative histograms further showed that the percentage of G2/M-phase cells increased after exposing to different concentrations of compound **7h**, implying that this compound induced cell cycle arrest at the G2/M phase to inhibit cell proliferation.

Conclusions

In summary, two series of densely substituted and distinct fused pyrazole-pyrazines were synthesized by using an Ugi reaction following an intramolecular *N*2-arylation of pyrazoles with allenes at the C- β position in absence of a catalyst in one-pot. Under the mild reaction condition, simple operation procedure, commercial availability of starting materials, and good overall yield, the novel methodology will offer a quick access to diverse indazole and pyrazole analogues for biological testing in a cost-effective and environmental-friendly manner. The gram scale synthesis of compound 7 can be accomplished within in one day (see SI for details). To our best knowledge, this is the first report for post-Ugi cascade reaction to prepare pyrazole-pyrazine scaffolds without a catalyst in one-pot. This catalyst-free cascade methodology would be highly applicable to other MCRs such as Passerini³⁴, Petasis³⁵, Betti³⁶, Kabachnik-Fields³⁷, and GBBR³⁸.

The preliminary screening results of compounds 7 in the colon cancer cell lines HCT116 and SW620 are promising. Considering that compounds 7b and 7h are only 10 times less active than the most popular anticancer drug paclitaxel, 7b and 7h provide starting points for the further optimization in the drug discovery value chain for the treatment of colorectal cancer. Further efforts are ongoing for applicable design strategy of this cascade cyclization to synthesize new scaffolds and optimize 7h toward a clinical candidate.

Experimental Section

General information. ¹H and ¹³C NMR were recorded on a Bruker 400 spectrometer. ¹H NMR data are reported as follows: chemical shift in ppm (δ), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet), coupling constant (Hz), relative intensity. ¹³C NMR data are reported as follows: chemical shift in ppm (δ). HPLC-MS analyses were performed on a Shimadzu-2020 LC-MS instrument using the following conditions:



Figure 2. Anticancer activities of compound 7**h** against human colon cancer cells. (A) HCT116 and SW620 cells were treated with the different concentrations of compound 7**h** for 24, 48 and 72 h. Cell viability was measured with the MTT assay. Data are the mean \pm SD of three independent experiments, and each experiment was conducted in sextuplicate. (B, C) The soft agar assay was employed to detect colony formation *in vitro* after treating with the indicated concentrations of compound 7**h** for 14 days. The colonies were visualized with the images (B) and quantitated by histogram (C). All data were demonstrated as the mean \pm SD of three independent experiments. *P < 0.05; **P < 0.01; ***P < 0.001 versus vehicle.

Shim-pack VPODS C18 column (reverse phase, 150×2.0 mm); 80% acetonitrile and 20% water over 6.0 min; flow rate of 0.4 mL/min; UV photodiode array detection from 200 to 300 nm. The reaction was irradiated at the required ceiling temperature (the reaction temperature was monitored by an external surface sensor using the Biotage Initiator reactor) using maximum power for the stipulated time. Then it was cooled to 50 °C with gas jet cooling. The products were purified by Biotage IsoleraTM Spektra Systems and Hexane/EtOAc solvent systems. All reagents and solvents were obtained from commercial sources and used without further purification.

General procedure for compound 7. A solution of 3-phenylpropiolaldehyde (1.0 mmol) and amine (1.0 mmol) in CF_3CH_2OH (TFE) (2 mL) was stirred at room temperature for 10 min in a 10 mL microwave vial. Then, carboxylic acid (1.0 mmol) and isocyanide (1.0 mmol) were added separately. After completion of the reaction (monitored by TLC), the solvent was removed under a nitrogen stream (The Ugi product 5a and 5b were purified). The residue was dissolved in DMF (2.0 mL), sealed and heated in microwave at 110 °C for 20 min. After the microwave vial was cooled to room temperature, the solvent was removed under reduced pressure and then diluted with EtOAc (15 mL) and washed with brine. The organic layer was dried over Na_2SO_4 and concentrated. The residue was purified by silica gel column chromatography using a gradient of ethyl acetate/hexane (0–60%) to afford the relative pyrazoles products **7a-s**. (The crude Ugi compound **5a** was heated in microwave at 80 °C for 20 min to give products **6a** (22%) and **7a** (60%)).

$$\begin{split} &N-(1-(benzylamino)-1-oxo-4-phenylbut-3-yn-2-yl)-N-phenyl-1H-indazole-3-carboxamide (\textbf{5a}), 420 \text{ mg light} \\ &yellow solid, yield 87\%, HPLC purity >96\%, mp 117-119 ^{\circ}C. ^{1}H NMR (400 MHz, CDCl_3) & 8.27 (d, J = 8.3 Hz, 1 H), 7.89 (d, J = 8.8 Hz, 1 H), 7.61-7.40 (m, 5 H), 7.35 (d, J = 7.1 Hz, 4 H), 7.31-7.05 (m, 7 H), 6.77 (d, J = 7.2 Hz, 1 H), 6.04 (s, 1 H), 4.54 (s, 1 H), 4.13 (d, J = 5.5 Hz, 2 H). ^{13}C NMR (100 MHz, CDCl_3) & 160.63, 154.76, 148.85, 137.41, 136.32, 136.16, 129.34, 129.23, 128.69, 128.61, 128.50, 128.29, 127.94, 127.78, 126.89, 124.82, 124.44, 121.34, 120.98, 117.72, 82,57, 81.38, 58.45, 44.44. HRMS (ESI) m/z calcd for C_{31}H_{25}N_4O_2^+ (M + H)^+ 485.1972, found 485.1973. \end{split}$$

N-(1-(benzylamino)-1-oxo-4-phenylbut-3-yn-2-yl)-*N*-(4-methoxyphenyl)-1*H*-indazole-3-carboxamide (**5b**), 465 mg white solid, yield 90%, HPLC purity >94%, mp 118–120 °C. ¹H NMR (400 MHz, DMSO- d_6) δ 12.20 (s, 1 H), 8.41–7.83 (m, 1 H), 7.78 (d, *J* = 7.7 Hz, 2 H), 7.74 (d, *J* = 8.1 Hz, 1 H), 7.65–7.47 (m, 2 H), 7.45 (d, *J* = 7.7 Hz, 2 H), 7.31 (t, *J* = 7.4 Hz, 1 H), 7.24 (d, *J* = 7.3 Hz, 1 H), 7.22–7.06 (m, 3 H), 6.86 (t, *J* = 7.1 Hz, 1 H), 6.76 (d, *J* = 8.5 Hz, 2 H), 5.49 (s, 1 H), 4.54 (d, *J* = 5.8 Hz, 2 H), 3.65 (s, 3 H). ¹³C NMR (100 MHz, DMSO- d_6) δ 158.67, 132.54, 132.30, 130.92, 130.29, 129.59, 129.32, 129.15, 127.50, 127.33, 126.11, 125.71, 124.49, 119.15, 116.04, 115.11, 114.98, 113.97, 79.65, 55.41, 48.78. HRMS (ESI) m/z calcd for C₃₂H₂₇N₄O₃⁺ (M + H)⁺ 515.2078, found 515.2075.

N-benzyl-4-benzylidene-1-oxo-2-phenyl-1,2,3,4-tetrahydropyrazino[1,2-b]indazole-3-carboxamide (**6a**), 106 mg white solid, yield 22%, HPLC purity >98%, mp 120–122 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.21 (d,



Figure 3. Compound **7h** induces cell cycle arrest at G2/M phase. Cell cycle of HCT116 and SW620 was analyzed by flow cytometry following treatment with compound **7h** for 48 h. Graph is a representative of percentage mean of three independent experiments. Histograms represent the percentages of cell distribution in G0/G1-, S- and G2/M-phase.

$$\begin{split} J = 8.5 \, \text{Hz}, 1 \, \text{H}), 7.93 \, (\text{s}, 1 \, \text{H}), 7.83 \, (\text{d}, J = 8.8 \, \text{Hz}, 1 \, \text{H}), 7.48 \, (\text{dd}, J = 13.4, 7.1 \, \text{Hz}, 3 \, \text{H}), 7.44 - 7.41 \, (\text{m}, 4 \, \text{H}), 7.38 \, (\text{t}, J = 7.2 \, \text{Hz}, 3 \, \text{H}), 7.33 \, (\text{dd}, J = 9.3, 6.0 \, \text{Hz}, 2 \, \text{H}), 7.24 - 7.13 \, (\text{m}, 3 \, \text{H}), 6.92 \, (\text{d}, J = 7.0 \, \text{Hz}, 2 \, \text{H}), 5.83 \, (\text{s}, 1 \, \text{H}), 5.74 \, (\text{s}, 1 \, \text{H}), 4.31 \, (\text{m}, 2 \, \text{H}). ^{13} \text{C} \, \text{NMR} \, (100 \, \text{MHz}, \text{CDCl}_3) \, \delta \, 166.70, 156.95, 149.63, 139.98, 136.72, 133.07, 130.19, 129.54, 129.03, 128.72, 128.31, 127.80, 127.65, 127.43, 126.26, 125.70, 124.67, 123.29, 122.87, 121.70, 118.08, 62.75, 44.24. \, \text{HRMS} \, (\text{ESI}) \, \text{m/z} \, \text{calcd for } \text{C}_{31} \text{H}_{25} \text{N}_4 \text{O}_2^+ \, (\text{M} + \text{H})^+ \, 485.1972, \text{found} \, 485.1972. \end{split}$$

N,4-dibenzyl-1-oxo-2-phenyl-1,2-dihydropyrazino[1,2-b]indazole-3-carboxamide (7**a**), 310 mg white solid, yield 64%, HPLC purity >98%, mp 124–126°C. ¹H NMR (400 MHz, CDCl₃) δ 8.32 (d, *J* = 8.3 Hz, 1 H), 7.92 (d, *J* = 8.8 Hz, 1 H), 7.52 (s, 1 H), 7.47 (s, 3 H), 7.43–7.31 (m, 5 H), 7.25 (s, 2 H), 7.23 (s, 2 H), 7.20 (s, 2 H), 6.78 (d, *J* = 5.7 Hz, 2 H), 5.82 (s, 1 H), 4.60 (s, 2 H), 4.17 (d, *J* = 5.3 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.67, 154.79, 148.91, 137.49, 136.37, 136.16, 129.40, 129.28, 128.76, 128.51, 128.30, 127.99, 127.85, 126.95, 124.89, 124.50, 121.40, 121.04, 117.76, 44.11, 32.36. HRMS (ESI) m/z calcd for C₃₁H₂₅N₄O₂+ (M + H)+ 485.1972, found 485.1973.

N,4-dibenzyl-2-(4-methoxyphenyl)-1-oxo-1,2-dihydropyrazino[1,2-b]indazole-3-Carboxamide (**7b**), 330 mg white solid, yield 64%, HPLC purity >98%, mp 120–122 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.32 (d, *J* = 8.3 Hz, 1 H), 7.92 (d, *J* = 8.7 Hz, 1 H), 7.52 (t, *J* = 7.3 Hz, 1 H), 7.41–7.35 (m, 3 H), 7.31 (s, 1 H), 7.28 (s, 1 H), 7.27 (s, 1 H), 7.26–7.25 (m, 1 H), 7.21 (dd, *J* = 12.9, 5.6 Hz, 4 H), 6.95 (d, *J* = 8.9 Hz, 2 H), 6.81 (d, *J* = 6.6 Hz, 2 H), 5.85 (t, *J* = 5.7 Hz, 1 H), 4.59 (s, 2 H), 4.22 (d, *J* = 5.8 Hz, 2 H), 3.87 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.79, 160.01, 148.94, 137.62, 136.26, 129.76, 128.76, 128.48, 127.92, 126.93, 124.79, 121.14, 120.78, 117.71, 114.62, 55.50, 44.09, 32.37. HRMS (ESI) m/z calcd for C₃₂H₂₇N₄O₃⁺ (M + H)⁺ 515.2078, found 515.2077.

4-benzyl-1-oxo-N-phenethyl-2-phenyl-1,2-dihydropyrazino[1,2-b]indazole-3-Carboxamide (7c), 313 mg white solid, yield 63%, HPLC purity >98%, mp 122–125 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.31 (d, J = 8.2 Hz, 1 H), 7.91 (d, J = 8.7 Hz, 1 H), 7.51 (dd, J = 15.8, 7.4 Hz, 2 H), 7.46 (s, 2 H), 7.39 (d, J = 7.7 Hz, 3 H), 7.34 (d, J = 7.8 Hz, 2 H), 7.28 (d, J = 6.4 Hz, 2 H), 7.25–7.25 (m, 1 H), 7.24–7.18 (m, 3 H), 6.96 (d, J = 7.4 Hz, 2 H), 5.55 (s, 1 H), 4.53 (s, 2 H), 3.26 (dd, J = 13.0, 6.4 Hz, 2 H), 2.37 (t, J = 7.1 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.88, 154.77, 148.97, 137.67, 136.57, 129.29, 128.81, 128.67, 128.56, 128.42, 127.98, 126.90, 124.87, 121.07, 117.77, 40.84, 34.56, 32.40. HRMS (ESI) m/z calcd for C₃₂H₂₇N₄O₂+ (M + H)⁺ 499.2129, found 499.2129.

N,4-*dibenzyl*-2-*cyclopropyl*-1-*oxo*-1,2-*dihydropyrazino*[1,2-*b*]*indazole*-3-*Carboxamide* (**7d**), 305 mg white solid, yield 68%, HPLC purity >98%, mp 119–122 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.21 (d, *J* = 8.4 Hz, 1 H), 7.81 (d, *J* = 8.7 Hz, 1 H), 7.48 (dd, *J* = 11.2, 4.3 Hz, 1 H), 7.36–7.33 (m, 3 H), 7.31 (d, *J* = 8.1 Hz, 3 H), 7.24 (s, 2 H), 7.19 (d, *J* = 7.4 Hz, 3 H), 6.81 (s, 1 H), 4.62 (d, *J* = 5.8 Hz, 2 H), 4.28 (s, 2 H), 3.20–3.08 (m, 1 H), 1.02 (d, *J* = 6.9 Hz, 2 H), 0.94 (s, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 161.10, 148.73, 137.47, 136.76, 129.37, 128.95, 128.63, 128.53, 128.38, 128.13, 127.78, 126.82, 124.61, 121.26, 120.88, 117.74, 44.47, 32.28, 29.52, 8.87. HRMS (ESI) m/z calcd for C₂₈H₂₅N₄O₃⁺ (M + H)⁺ 449.1972, found 449.1977.

4-benzyl-N-(2,6-dimethylphenyl)-1-oxo-2-phenyl-1,2-dihydropyrazino[1,2-b]indazole-3-carboxamide (7e), 290 mg white solid, yield 58%, HPLC purity >98%, mp 125–127 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.30 (d, *J*=8.4 Hz, 1 H), 7.89 (d, *J*=8.7 Hz, 1 H), 7.55 (d, *J*=7.7 Hz, 2 H), 7.51 (d, *J*=7.9 Hz, 2 H), 7.48 (d, *J*=6.4 Hz, 1 H), 7.44 (d, *J*=7.5 Hz, 2 H), 7.40–7.34 (m, 1 H), 7.28 (d, *J*=7.3 Hz, 2 H), 7.20 (t, *J*=7.3 Hz, 1 H), 7.06 (dd, *J*=16.5, 9.0 Hz, 2 H), 6.94 (d, *J*=7.5 Hz, 2 H), 4.76 (s, 2 H), 1.71 (s, 6 H). ¹³C NMR (100 MHz, CDCl₃) δ 158.93, 155.15, 149.15, 136.81, 134.76, 132.11, 129.74, 129.60, 129.48, 128.68, 128.48, 128.05, 127.87, 126.94, 125.04, 121.40, 121.05, 117.92, 32.88, 18.21. HRMS (ESI) m/z calcd for C₃₂H₂₇N₄O₂⁺ (M + H)⁺ 499.2129, found 499.2126.

N,4-*dibenzyl*-2-(4-*bromophenyl*)-1-*oxo*-1,2-*dihydropyrazino*[1,2-*b*]*indazole*-3-*carboxamide* (**7f**), 365 mg white solid, yield 65%, HPLC purity >98%, mp 171–173 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.30 (d, *J* = 8.3 Hz, 1 H), 7.93 (d, *J* = 8.7 Hz, 1 H), 7.54 (dd, *J* = 11.6, 8.7 Hz, 3 H), 7.43–7.37 (m, 1 H), 7.35 (d, *J* = 7.1 Hz, 2 H), 7.28 (s, 4 H), 7.24 (t, *J* = 5.3 Hz, 4 H), 6.86–6.75 (m, 2 H), 5.87 (s, 1 H), 4.59 (s, 2 H), 4.23 (d, *J* = 5.9 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.46, 137.45, 132.60, 130.30, 128.85, 128.76, 128.43, 128.02, 127.06, 125.10, 121.05, 117.85, 44.14, 32.38. HRMS (ESI) m/z calcd for C₃₁H₂₄BrN₄O₂⁺ (M + H)⁺ 563.1077, found 563.1081 and 565.1061.

4-benzyl-2-(4-methoxyphenyl)-1-oxo-N-phenethyl-1,2-dihydropyrazino[1,2-b]indazole-3-carboxamide (7g), 300 mg white solid, yield 57%, HPLC purity >98%, mp 124–126 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.31 (d, J = 8.3 Hz, 1 H), 7.91 (d, J = 8.7 Hz, 1 H), 7.56–7.47 (m, 1 H), 7.41–7.35 (m, 3 H), 7.31–7.26 (m, 3 H), 7.25–7.19 (m, 5 H), 6.97 (dd, J = 7.6, 5.6 Hz, 4 H), 5.55 (t, J = 5.5 Hz, 1 H), 4.51 (s, 2 H), 3.84 (s, 3 H), 3.31 (dd, J = 13.1, 7.0 Hz, 2 H), 2.44 (t, J = 7.1 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.96, 159.97, 155.08, 148.92, 137.75, 137.63, 129.66, 128.82, 128.67, 128.50, 128.41, 127.93, 126.94, 124.77, 121.11, 117.70, 114.55, 55.58, 40.81, 34.62, 32.41. HRMS (ESI) m/z calcd for C₃₃H₂₉N₄O₃+ (M + H)+ 529.2234, found 529.2236.

4-benzyl-2-cyclopropyl-1-oxo-N-phenethyl-1,2-dihydropyrazino[1,2-b]indazole-3-carboxamide (7h), 290 mg white solid, yield 63%, HPLC purity >98%, mp 121–123 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, J = 8.4 Hz, 1 H), 7.82 (d, *J* = 8.7 Hz, 1 H), 7.53–7.44 (m, 1 H), 7.37–7.32 (m, 1 H), 7.29 (dd, *J* = 10.2, 3.5 Hz, 4 H), 7.23 (d, *J*=7.6 Hz, 3 H), 7.19 (t, *J*=7.5 Hz, 3 H), 6.42 (s, 1 H), 4.23 (s, 2 H), 3.78 (dd, *J*=12.7, 6.8 Hz, 2 H), 3.11–2.99 (m, 1 H), 2.91 (t, J = 6.9 Hz, 2 H), 1.01 (d, J = 6.8 Hz, 2 H), 0.88 (s, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 161.42, 156.16, 148.73, 138.02, 137.60, 129.50, 128.84, 128.63, 128.41, 127.76, 126.92, 126.86, 124.58, 121.21, 120.86, 120.69, 117.73, 41.13, 34.94, 32.29, 29.49, 8.84. HRMS (ESI) m/z calcd for C₂₉H₂₇N₄O₂⁺ (M+H)⁺ 463.2129, found 463.2131. Gram scale synthesis of compound 7h. A solution of 3-phenylpropiolaldehyde (5.0 mmol) and cyclopropanamine (5.0 mmol) in TFE (10 mL) was stirred at room temperature for 10 min. Then, 1H-indazole-3-carboxylic acid (5.0 mmol) and phenethyl isocyanide (5.0 mmol) were added separately. After completion of the reaction (monitored by TLC), the solvent was removed under reduced pressure. The residue was dissolved in DMF (10.0 mL), sealed and heated in microwave at 110 °C for 20 min. After the microwave vial was cooled to room temperature, the solvent was removed under reduced pressure and then diluted with EtOAc (60 mL). After washed with brine, the organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column chromatography using a gradient of ethyl acetate/hexane (0-60%) to afford compound 7 h (1.18 g)with 51% yield in 7 hours.

N,4-*dibenzyl*-2-(2-*chlorophenyl*)-1-*oxo*-1,2-*dihydropyrazino*[1,2-*b*]*indazole*-3-*carboxamide* (**7i**), 310 mg white solid, yield 60%, HPLC purity >98%, mp 166–168 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.31 (d, *J* = 8.4 Hz, 1 H), 7.91 (d, *J* = 8.7 Hz, 1 H), 7.51 (dd, *J* = 10.2, 5.3 Hz, 2 H), 7.45–7.36 (m, 4 H), 7.34 (d, *J* = 7.8 Hz, 2 H), 7.29–7.23 (m, 3 H), 7.21 (dd, *J* = 13.0, 5.5 Hz, 3 H), 6.84 (d, *J* = 7.0 Hz, 2 H), 6.16 (s, 1 H), 4.68 (d, *J* = 15.9 Hz, 1 H), 4.44 (d, *J* = 15.9 Hz, 1 H), 4.27 (dd, *J* = 14.5, 6.6 Hz, 1 H), 4.07 (dd, *J* = 14.5, 5.3 Hz, 1 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.24, 154.12, 148.90, 137.27, 136.18, 134.08, 133.02, 131.55, 131.03, 130.14, 128.77, 128.74, 128.29, 128.11, 128.01, 127.90, 127.81, 126.94, 125.02, 124.48, 121.55, 121.08, 120.71, 117.83, 43.99, 32.46. HRMS (ESI) m/z calcd for C₃₁H₂₄ClN₄O₂+ (M + H)+ 519.1582, found 519.1586.

7-benzyl-N-(2,6-dimethylphenyl)-4-oxo-2,5-diphenyl-4,5-dihydropyrazolo[1,5-a]pyrazine-6-carboxamide (7j), 325 mg white solid, yield 62%, HPLC purity >97%, mp 135–137 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J*=7.1 Hz, 2 H), 7.54 (d, *J*=7.6 Hz, 2 H), 7.48 (d, *J*=7.2 Hz, 2 H), 7.45–7.41 (m, 3 H), 7.41–7.34 (m, 3 H), 7.32 (s, 1 H), 7.28 (d, *J*=7.0 Hz, 2 H), 7.19 (t, *J*=7.3 Hz, 1 H), 7.07–7.00 (m, 1 H), 6.94 (d, *J*=7.5 Hz, 2 H), 4.59 (s, 2 H), 1.70 (s, 6 H). ¹³C NMR (100 MHz, CDCl₃) δ 159.17, 154.89, 153.62, 137.09, 136.93, 134.69, 134.15, 132.35, 131.94, 129.61, 129.35, 129.17, 128.95, 128.82, 128.52, 128.48, 127.70, 126.86, 126.24, 124.94, 122.32, 103.82, 32.81, 18.25. HRMS (ESI) m/z calcd for C₃₄H₂₉N₄O₂+ (M + H)⁺ 525.2285, found 525.2299.

7-benzyl-4-oxo-N-phenethyl-2,5-diphenyl-4,5-dihydropyrazolo[1,5-a]pyrazine-6-carbox amide (7k), 340 mg white solid, yield 65%, HPLC purity >98%, mp 129–131 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.88 (d, *J* = 7.2 Hz, 2 H), 7.45 (d, *J* = 7.2 Hz, 3 H), 7.44–7.40 (m, 4 H), 7.39 (s, 1 H), 7.35 (s, 1 H), 7.32–7.26 (m, 4 H), 7.21 (dd, *J* = 14.1, 7.1 Hz, 4 H), 6.98 (d, *J* = 7.0 Hz, 2 H), 5.75 (t, *J* = 5.6 Hz, 1 H), 4.36 (s, 2 H), 3.23 (dd, *J* = 13.4, 7.0 Hz, 2 H), 2.36 (t, *J* = 7.3 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 161.13, 154.49, 153.34, 137.88, 137.60, 136.91, 134.24, 132.03, 129.26, 129.07, 128.95, 128.83, 128.76, 128.53, 128.45, 128.44, 126.89, 126.78, 126.24, 124.82, 121.61, 103.73, 40.81, 34.58, 32.35. HRMS (ESI) m/z calcd for C₃₄H₂₉N₄O₂⁺ (M + H)⁺ 525.2285, found 525.2285.

7-benzyl-5-(4-chlorophenyl)-4-oxo-N-phenethyl-2-phenyl-4,5-dihydropyrazolo[1,5-a] pyrazine-6-carboxamide (7l), 330 mg white solid, yield 59%, HPLC purity >98%, mp 155–157 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.89 (d, J = 7.2 Hz, 2 H), 7.44 (dd, J = 10.2, 4.2 Hz, 3 H), 7.42–7.35 (m, 5 H), 7.33–7.26 (m, 4 H), 7.24 (s, 1 H), 7.21 (dd, J = 11.4, 5.2 Hz, 3 H), 6.99 (d, J = 6.9 Hz, 2 H), 5.60 (t, J = 5.3 Hz, 1 H), 4.36 (s, 2 H), 3.32 (dd, J = 12.9, 6.9 Hz, 2 H),

2.48 (t, $J\!=\!7.0$ Hz, 2 H). 13 C NMR (100 MHz, CDCl₃) δ 160.96, 154.34, 153.50, 137.69, 137.53, 135.27, 135.04, 134.11, 131.91, 129.72, 129.46, 128.90, 128.85, 128.78, 128.64, 128.37, 127.00, 126.90, 126.25, 124.47, 121.73, 103.99, 40.75, 34.52, 32.36. HRMS (ESI) m/z calcd for $C_{34}H_{28}ClN_4O_2^+$ (M + H)+ 559.1895, found 559.1895.

7-benzyl-5-(3-bromophenyl)-4-oxo-N-phenethyl-2-phenyl-4,5-dihydropyrazolo[1,5-a] pyrazine-6-carboxamide (7m), 343 mg white solid, yield 57%, HPLC purity >97%, mp 206–209 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.91 (d, J = 5.9 Hz, 2 H), 7.62–7.42 (m, 4 H), 7.42–7.33 (m, 2 H), 7.29 (t, J = 7.5 Hz, 2 H), 7.22 (t, J = 7.2 Hz, 3 H), 7.16 (d, J = 7.4 Hz, 2 H), 6.81 (d, J = 95.5 Hz, 3 H), 4.68 (s, 2 H), 2.19 (s, 6 H). ¹³C NMR (100 MHz, CDCl₃) δ 153.73, 137.23, 135.64, 134.49, 131.75, 130.35, 129.70, 129.16, 129.07, 128.89, 128.62, 127.32, 126.98, 126.30, 122.84, 103.65, 31.58, 18.73. HRMS (ESI) m/z calcd for C₃₄H₂₈BrN₄O₂⁺ (M + H)⁺ 603.1390, found 603.1395.

4-benzyl-9-methyl-1-oxo-N-phenethyl-2-phenyl-1,2-dihydropyrazino[1,2-b]indazole-3-carboxamide (7**n**), 280 mg white solid, yield 55%, HPLC purity >97%, mp 196–198 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (s, 1 H), 7.79 (d, *J* = 8.8 Hz, 1 H), 7.50–7.42 (m, 3 H), 7.38 (d, *J* = 7.2 Hz, 2 H), 7.36–7.30 (m, 3 H), 7.27 (d, *J* = 7.2 Hz, 2 H), 7.25–7.18 (m, 4 H), 6.95 (d, *J* = 6.9 Hz, 2 H), 5.63 (s, 1 H), 4.48 (s, 2 H), 3.24 (d, *J* = 6.4 Hz, 2 H), 2.49 (s, 3 H), 2.35 (t, *J* = 7.3 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.97, 154.87, 147.83, 137.76, 137.62, 136.64, 134.88, 130.76, 129.26, 129.13, 128.77, 128.63, 128.59, 128.55, 128.41, 128.07, 126.89, 126.79, 123.74, 121.80, 121.04, 119.38, 117.39, 40.83, 34.55, 32.37, 21.97. HRMS (ESI) m/z calcd for C₃₃H₂₉N₄O₂+ (M + H)⁺ 513.2285, found 513.2285.

7-benzyl-5-(3,4-dimethoxyphenyl)-4-oxo-N-phenethyl-2-phenyl-4,5-dihydropyrazolo[1,5-a] pyrazine-6-carboxamide (**7o**), 356 mg white solid, yield 61%, HPLC purity >98%, mp 187–190 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.90 (d, *J* = 7.3 Hz, 2 H), 7.44 (t, *J* = 6.9 Hz, 4 H), 7.38 (dd, *J* = 8.5, 4.8 Hz, 2 H), 7.28 (d, *J* = 7.2 Hz, 2 H), 7.26–7.15 (m, 4 H), 6.99 (d, *J* = 7.3 Hz, 2 H), 6.83 (dd, *J* = 12.3, 6.9 Hz, 3 H), 5.68 (d, *J* = 4.2 Hz, 1 H), 4.35 (s, 2 H), 3.89 (s, 3 H), 3.83 (s, 3 H), 3.32 (dd, *J* = 13.0, 6.7 Hz, 2 H), 2.47 (t, *J* = 7.0 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 161.26, 154.73, 153.27, 149.52, 149.35, 137.85, 137.62, 134.23, 132.04, 129.49, 128.94, 128.83, 128.78, 128.52, 128.38, 126.88, 126.79, 126.24, 125.16, 121.36, 120.62, 112.04, 110.94, 103.65, 56.18, 56.09, 40.76, 34.75, 32.33. HRMS (ESI) m/z calcd for C₃₆H₃₃N₄O₄+ (M + H)⁺ 585.2496, found 585.2504.

4-benzyl-2-(4-chlorophenyl)-9-methyl-1-oxo-N-phenethyl-1,2-dihydropyrazino[1,2-b] indazole-3-carboxamide (7**p**), 327 mg white solid, yield 60%, HPLC purity >98%, mp 199–202 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (s, 1 H), 7.80 (d, *J* = 8.8 Hz, 1 H), 7.41 (d, *J* = 8.6 Hz, 2 H), 7.35 (d, *J* = 8.7 Hz, 3 H), 7.28 (dd, *J* = 7.7, 3.3 Hz, 3 H), 7.24 (d, *J* = 8.7 Hz, 5 H), 6.96 (d, *J* = 6.8 Hz, 2 H), 5.56 (s, 1 H), 4.50 (s, 2 H), 3.32 (dd, *J* = 12.9, 6.9 Hz, 2 H), 2.50 (s, 3 H), 2.47 (t, *J* = 7.0 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 160.83, 154.73, 147.90, 137.61, 137.57, 135.15, 135.02, 130.91, 129.88, 129.45, 128.87, 128.75, 128.41, 128.36, 127.73, 127.02, 126.92, 123.60, 121.90, 121.19, 119.33, 117.46, 40.77, 34.49, 32.41, 21.99. 327 mg white solid, yield 60%. HRMS (ESI) m/z calcd for C₃₃H₂₈ClN₄O₂⁺ (M + H)⁺ 547.1895, found 547.1896.

4-benzyl-2-(3-chlorophenyl)-9-methyl-1-oxo-N-phenethyl-1,2-dihydropyrazino[1,2-b] indazole-3-carboxamide (7**q**), 344 mg white solid, yield 63%, HPLC purity >98%, mp 197–200 °C. ¹H NMR (400 MHz, CDCl₃) δ 11.36 (s, 1 H), 7.99 (s, 1 H), 7.78 (d, *J* = 8.8 Hz, 1 H), 7.40 (d, *J* = 7.5 Hz, 2 H), 7.29 (d, *J* = 8.6 Hz, 5 H), 7.26–7.16 (m, 4 H), 6.92 (dd, *J* = 87.9, 57.8 Hz, 4 H), 4.58 (s, 2 H), 4.13 (s, 2 H), 3.04 (s, 2 H), 2.24 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 162.33, 156.29, 147.60, 137.81, 137.07, 135.01, 134.92, 130.73, 130.41, 128.90, 128.69, 128.51, 127.13, 126.99, 126.82, 125.39, 123.80, 123.74, 122.55, 121.15, 119.00, 117.47, 52.31, 33.82, 31.39, 21.66. HRMS (ESI) m/z calcd for C₃₃H₂₈ClN₄O₂+ (M + H)+ 547.1895, found 547.1898.

4-benzyl-2-(4-fluorophenyl)-9-methyl-1-oxo-N-phenethyl-1,2-dihydropyrazino[1,2-b] indazole-3-carboxamide (7**r**), 313 mg white solid, yield 59%, HPLC purity >96%, mp 185–187 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.03 (s, 1 H), 7.78 (d, *J* = 8.8 Hz, 1 H), 7.35 (t, *J* = 7.4 Hz, 3 H), 7.28 (d, *J* = 7.1 Hz, 4 H), 7.25–7.15 (m, 4 H), 7.10 (t, *J* = 8.4 Hz, 2 H), 6.96 (d, *J* = 6.8 Hz, 2 H), 5.70 (d, *J* = 5.4 Hz, 1 H), 4.44 (s, 2 H), 3.30 (dd, *J* = 13.0, 6.8 Hz, 2 H), 2.49 (s, 3 H), 2.45 (t, *J* = 7.1 Hz, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 161.33, 160.87, 154.94, 147.83, 137.66, 137.51, 135.07, 132.35, 130.83, 130.53, 130.45, 128.83, 128.70, 128.48, 138.37, 127.99, 126.98, 126.88, 123.57, 121.78, 120.95, 119.26, 117.46, 116.33, 116.11, 40.81, 34.55, 32.42, 21.98. ¹⁹F NMR (377 MHz, CDCl₃) δ -111.28. HRMS (ESI) m/z calcd for C₃₃H₂₈FN₄O₂+ (M + H)+ 531.2191, found 531.2184.

N,7-dibenzyl-5-(4-bromophenyl)-4-oxo-2-phenyl-4,5-dihydropyrazolo[1,5-a]pyrazine-6-carboxamide (7s), 340 mg white solid, yield 58%, HPLC purity >97%, mp 205–208 °C. ¹H NMR (400 MHz, CDCl₃) δ 10.77 (s, 1 H), 7.89 (d, *J* = 7.3 Hz, 2 H), 7.45 (d, *J* = 7.2 Hz, 2 H), 7.39 (dd, *J* = 17.3, 10.1 Hz, 4 H), 7.29 (d, *J* = 6.2 Hz, 5 H), 7.24–7.19 (m, 3 H), 7.14 (d, *J* = 8.1 Hz, 2 H), 6.57 (s, 2 H), 5.03 (s, 2 H), 4.49 (s, 2 H). ¹³C NMR (100 MHz, CDCl₃) δ 162.84, 155.43, 153.20, 139.47, 137.23, 135.71, 134.21, 132.59, 131.95, 129.18, 129.11, 128.89, 128.86, 128.63, 128.58, 128.00, 126.94, 126.24, 123.13, 122.00, 119.97, 103.04, 54.05, 31.21. HRMS (ESI) m/z calcd for C₃₃H₂₆BrN₄O₂+ (M + H)⁺ 589.1234, found 589.1233.

General procedure for compound 14. The same procedure with compound 7. (The Ugi product **11a** was purified.)

N-(2-((4-methoxybenzyl)amino)-2-oxo-1-phenylethyl)-*N*-(prop-2-yn-1-yl)-1H-indazole-3-carboxamide, (**11a**), 420 mg white solid, yield 93%, HPLC purity >95%, mp 155–157 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.24 (s, 1 H), 7.35 (s, 4 H), 7.33 (d, *J* = 15.9 Hz, 3 H), 7.26 (s, 1 H), 7.23 (d, *J* = 8.1 Hz, 1 H), 7.19 (d, *J* = 8.6 Hz, 2 H), 6.77 (d, *J* = 8.5 Hz, 2 H), 4.51 (s, 2 H), 4.49 (s, 1 H), 3.73 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 158.99, 140.57, 129.66, 129.18, 128.91, 128.63, 127.20, 123.91, 122.74, 122.5, 114.00, 110.10, 80.16, 71.79, 55.26, 43.38. HRMS (ESI) m/z calcd for C₂₇H₂₅N₄O₃⁺ (M + H)⁺ 452.1848, found 452.1844.

N-(4-methoxybenzyl)-2-(4-methyl-1-oxopyrazino[1,2-b]indazol-2(1 H)-yl)-2-phenylacet amide (14a), 380 mg white solid, yield 62%, HPLC purity >98%, mp 195–197 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, *J* = 8.3 Hz, 1 H), 7.93 (d, *J* = 8.7 Hz, 1 H), 7.57–7.49 (m, 1 H), 7.44 (dd, *J* = 8.5, 3.9 Hz, 5 H), 7.40–7.32 (m, 1 H), 7.10 (d, *J* = 8.5 Hz, 2 H), 7.02 (s, 1 H), 6.91 (s, 1 H), 6.83 (d, *J* = 5.1 Hz, 1 H), 6.71 (d, *J* = 8.5 Hz, 2 H), 4.44 (qd, *J* = 14.5, 5.5 Hz, 2 H), 3.73 (s, 3 H), 2.56 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 167.68, 159.07, 155.61, 148.62, 134.64,

129.42, 129.29, 129.22, 129.10, 127.54, 124.19, 124.12, 121.03, 120.86, 119.28, 117.79, 117.39, 114.01, 59.13, 55.23, 43.65, 14.52. HRMS (ESI) m/z calcd for $C_{27}H_{25}N_4O_3^+$ (M + H)⁺ 453.1921, found 453.1917.

2-(4-fluorophenyl)-2-(4-methyl-1-oxopyrazino[1,2-b]indazol-2(1H)-yl)-N-phenethylacet amide (14b), 272 mg white solid, yield 60%, HPLC purity >98%, mp 199–202 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, J = 8.3 Hz, 1 H), 7.94 (d, J = 8.7 Hz, 1 H), 7.57–7.49 (m, 1 H), 7.41–7.30 (m, 3 H), 7.14–7.08 (m, 3 H), 7.06 (t, J = 6.8 Hz, 4 H), 7.00–6.92 (m, 2 H), 6.89 (s, 1 H), 3.70 (dq, J = 13.5, 7.0 Hz, 1 H), 3.57 (td, J = 13.0, 6.5 Hz, 1 H), 2.81 (t, J = 6.8 Hz, 2 H), 2.56 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 167.78, 164.19, 161.71, 155.52, 148.65, 138.18, 130.92, 130.84, 130.52, 130.49, 128.64, 128.48, 127.65, 126.43, 124.32, 124.04, 121.00, 120.66, 119.54, 117.53, 117.43, 116.43, 116.22, 58.34, 40.86, 35.49, 14.51. ¹⁹F NMR (377 MHz, CDCl₃) δ -111.72. HRMS (ESI) m/z calcd for C₂₇H₂₄FN₄O₂+ (M + H)⁺ 455.1878, found 455.1884.

 $\begin{array}{l} 2-(4-methyl-1-oxopyrazino[1,2-b]indazol-2(1 H)-yl)-N-phenethyl-2-(3,4,5-trimethoxyphenyl) \ acetamide (14c), 334 mg white solid, yield 58%, HPLC purity >98%, mp 201–204 °C. ¹H NMR (400 MHz, CDCl₃) <math display="inline">\delta$ 8.33 (d, *J* = 8.3 Hz, 1 H), 7.95 (d, *J* = 8.7 Hz, 1 H), 7.58–7.50 (m, 1 H), 7.45–7.38 (m, 1 H), 7.19–7.14 (m, 2 H), 7.11 (dd, *J* = 13.8, 7.1 Hz, 3 H), 6.90 (s, 1 H), 6.75 (s, 1 H), 6.68 (s, 2 H), 6.44 (d, *J* = 5.2 Hz, 1 H), 3.87 (s, 3 H), 3.81 (s, 6 H), 3.71–3.52 (m, 2 H), 2.84 (tq, *J* = 13.8, 7.1 Hz, 2 H), 2.59 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 167.80, 155.49, 153.87, 148.67, 138.85, 138.19, 129.54, 128.63, 128.57, 127.64, 126.57, 124.30, 124.17, 121.03, 120.82, 119.31, 117.47, 117.43, 106.43, 60.90, 59.39, 56.37, 41.11, 35.54, 14.60. HRMS (ESI) m/z calcd for C₃₀H₃₁N₄O₅⁺ (M + H)⁺ 527.2289, found 527.2291.

 $\label{eq:N-(4-methoxybenzyl)-2-(4-methoxyphenyl)-2-(4-methyl-1-oxopyrazino[1,2-b]indazol-2(1 H)-yl)acetamide} (14d), 265 mg white solid, yield 55%, HPLC purity >98%, mp 200–203 °C. ¹H NMR (400 MHz, CDCl₃) <math display="inline">\delta$ 8.13 (dd, J=17.1, 8.3 Hz, 1 H), 7.91 (d, J=8.7 Hz, 1 H), 7.50 (t, J=7.7 Hz, 1 H), 7.41 (d, J=8.7 Hz, 2 H), 7.30 (dd, J=14.7, 7.7 Hz, 1 H), 7.10 (s, 1 H), 7.07–6.99 (m, 3 H), 6.93 (dd, J=9.2, 5.9 Hz, 3 H), 6.61 (dd, J=15.3, 8.3 Hz, 2 H), 4.49–4.35 (m, 2 H), 3.83 (s, 3 H), 3.75–3.66 (m, 3 H), 2.55 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 168.03, 160.25, 158.90, 155.64, 148.59, 130.64, 129.28, 129.13, 127.47, 126.44, 124.13, 124.04, 120.94, 120.82, 119.16, 117.82, 117.33, 114.75, 113.85, 58.88, 55.39, 55.17, 43.57, 14.52. HRMS (ESI) m/z calcd for C₂₈H₂₇N₄O₄+ (M + H)+ 483.2027, found 483.2028.

 $\begin{array}{l} 2-(4-methyl-1-oxopyrazino[1,2-b]indazol-2(1 H)-yl)-N-phenethyl-2-(thiophen-2-yl)acetamide (14e), 278 mg \\ \text{white solid, yield 63\%, HPLC purity >98\%, mp 174-177 °C. <math display="inline">^1\text{H}$ NMR (400 MHz, CDCl₃) δ 8.28 (d, J = 8.3 Hz, 1 H), 7.94 (d, J = 8.6 Hz, 1 H), 7.53 (t, J = 7.6 Hz, 1 H), 7.39 (dd, J = 12.4, 5.7 Hz, 2 H), 7.25 (d, J = 3.3 Hz, 1 H), 7.09 (s, 1 H), 7.05 (d, J = 6.5 Hz, 5 H), 7.02 (s, 2 H), 6.81 (d, J = 5.1 Hz, 1 H), 3.61 (q, J = 6.7 Hz, 2 H), 2.89–2.73 (m, 2 H), 2.58 (s, 3 H). ^{13}C NMR (100 MHz, CDCl₃) δ 167.24, 155.13, 148.67, 138.14, 135.70, 129.64, 128.59, 128.41, 128.19, 127.64, 127.20, 126.33, 124.32, 124.07, 121.05, 120.78, 119.70, 117.48, 116.80, 54.56, 40.98, 35.37, 14.55. HRMS (ESI) m/z calcd for C₂₅H₂₃N₄O₂S⁺ (M + H)⁺ 443.1536, found 443.1532.

 $\begin{array}{l} 2\mbox{-}cyclohexyl-2\mbox{-}(4\mbox{-}methyl-1\mbox{-}oxpyrazino[1,2\mbox{-}b]indazol-2(1\mbox{H})\mbox{-}vl)\mbox{-}N\mbox{-}phenethylacetamide (14f), 282 mg white solid, yield 64\%, HPLC purity >98\%, mp 178\mbox{-}180 ^{\circ}C. ^{1}H NMR (400 MHz, CDCl_3) & 8.32 (d, J = 8.3 Hz, 1 H), 7.96 (d, J = 8.6 Hz, 1 H), 7.59\mbox{-}7.51 (m, 1 H), 7.45\mbox{-}7.37 (m, 1 H), 7.23 (s, 1 H), 7.01 (d, J = 4.0 Hz, 3 H), 7.00\mbox{-}6.93 (m, 2 H), 6.68 (s, 1 H), 5.25 (d, J = 10.8 Hz, 1 H), 3.54 (q, J = 6.7 Hz, 2 H), 2.76 (tq, J = 14.0, 7.0 Hz, 2 H), 2.67 (s, 3 H), 2.23 (q, J = 10.9 Hz, 1 H), 1.78 (t, J = 16.2 Hz, 2 H), 1.65 (d, J = 11.2 Hz, 2 H), 1.45\mbox{-}1.28 (m, 2 H), 1.25\mbox{-}1.03 (m, 3 H), 1.00\mbox{-}0.84 (m, 1 H). ^{13}C NMR (100 MHz, CDCl_3) & 168.80, 155.69, 148.62, 138.15, 128.53, 128.29, 127.64, 126.25, 124.31, 124.19, 121.02, 120.80, 119.73, 117.43, 116.12, 40.45, 37.64, 35.45, 29.95, 28.25, 26.06, 25.43, 25.31, 14.56. HRMS (ESI) m/z calcd for C_{27}H_{31}N_4O_2^+ (M + H)^+ 443.2442, found 443.2450. \end{array}$

 $\begin{array}{l} 2-(4\mbox{-}b]\mbox{ind}\ phen-2\mbox{-}yl)\mbox{-}2-(4\mbox{-}methyl\mbox{-}1\mbox{-}oxopyrazino\mbox{[}1\mbox{-}2\mbox{-}b\mbox{]}\mbox{-}n\mbox{-}phenethyl\mbox{-}acta mide} \\ (\mathbf{14g}), 307\mbox{ mg white solid, yield 59\%, HPLC purity >98\%, mp 193-196 °C. ^1H NMR (400\mbox{ MHz, CDCl}_3) & 8.22 \\ (d, J=8.0\mbox{ Hz}, 1\mbox{ H}), 8.03-7.89\mbox{ (m, 1 H}), 7.54\mbox{ (dd, } J=10.4, 4.8\mbox{ Hz}, 1\mbox{ H}), 7.40\mbox{ (dd, } J=9.8, 5.0\mbox{ Hz}, 1\mbox{ H}), 7.28\mbox{ (dd, } J=8.2, 3.8\mbox{ Hz}, 1\mbox{ H}), 7.16\mbox{ (s, 1 H}), 7.07\mbox{ (s, 1 H}), 7.03\mbox{ (d, } J=18.3\mbox{ Hz}, 3\mbox{ H}), 7.01\mbox{-}6.74\mbox{ (m, 4 H}), 3.61\mbox{ (qd, } J=13.4, 6.7\mbox{ Hz}, 2\mbox{ H}), 2.90\mbox{-}2.71\mbox{ (m, 2 H}), 2.60\mbox{ (d, } J=2.8\mbox{ Hz}, 3\mbox{ H}). ^{13}\mbox{ CNMR}\mbox{ (100\mbox{ MHz}, CDCl}_3) & 166.68, 155.05, 148.69, 137.95, 137.22, 131.94, 128.52, 128.37, 127.76, 126.36, 125.49, 124.55, 123.90, 121.08, 120.62, 120.18, 117.61, 116.31, 110.06, 53.79, 40.91, 35.28, 14.54.\mbox{ HRMS}\mbox{ (ESI)}\mbox{ m/z calcd for $C_{25}H_{22}BrN_4O_2S^+\mbox{ (M + H)}^+ 521.0641\mbox{ found} 521.0646. \end{array}$

 $\begin{array}{l} 2-(4,9-dimethyl-1-oxopyrazino[1,2-b]indazol-2(1 H)-yl)-N-phenethyl-2-phenylacetamide (14h), 238 mg \\ \text{white solid, yield 53\%, HPLC purity >98\%, mp 174–177 °C. ¹H NMR (400 MHz, CDCl₃) <math display="inline">\delta$ 8.06 (s, 1 H), 7.83 (d, J=8.8 Hz, 1 H), 7.44–7.38 (m, 3 H), 7.36 (d, J=9.6 Hz, 3 H), 7.21–7.11 (m, 3 H), 7.09 (d, J=7.4 Hz, 2 H), 6.88 (d, J=12.2 Hz, 1 H), 6.82 (s, 1 H), 6.51 (d, J=43.6 Hz, 1 H), 3.70 (dq, J=13.5, 6.9 Hz, 1 H), 3.57 (td, J=13.0, 6.7 Hz, 1 H), 2.83 (t, J=7.0 Hz, 2 H), 2.52 (s, 6 H). ¹³C NMR (100 MHz, CDCl₃) δ 167.92, 155.67, 147.50, 138.28, 134.66, 134.17, 130.31, 129.31, 129.10, 128.97, 128.70, 128.55, 126.49, 123.45, 121.40, 119.31, 117.25, 117.07, 59.23, 40.95, 35.56, 21.94, 14.50. HRMS (ESI) m/z calcd for C₂₈H₂₆N₄O₂+ (M + H)+ 451.2129, found 451.2126.

 $\begin{array}{l} 2-(9\text{-}bromo\text{-}4\text{-}methyl\text{-}1\text{-}oxopyrazino}[1,2\text{-}b]indazol\text{-}2(1\,H)\text{-}yl)\text{-}N\text{-}phenethyl\text{-}2-(p\text{-}tolyl)acet amide (14i), 316 mg white solid, yield 60\%, HPLC purity >98\%, mp 207-209 °C. ¹H NMR (400 MHz, CDCl₃) & 8.44 (d,$ *J*= 1.6 Hz, 1 H), 7.79 (d,*J*= 9.1 Hz, 1 H), 7.57 (dd,*J*= 9.1, 1.8 Hz, 1 H), 7.27 (s, 1 H), 7.25 (s, 1 H), 7.20 (d,*J*= 8.3 Hz, 2 H), 7.15 (d,*J*= 7.4 Hz, 2 H), 7.12 (d,*J*= 6.8 Hz, 1 H), 7.11-7.06 (m, 2 H), 6.89 (s, 1 H), 6.83 (s, 1 H), 6.53 (t,*J*= 5.6 Hz, 1 H), 3.69 (td,*J*= 13.5, 7.0 Hz, 1 H), 3.57 (td,*J*= 13.0, 6.7 Hz, 1 H), 2.83 (t,*J*= 7.0 Hz, 2 H), 2.52 (s, 3 H), 2.37 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) & 167.91, 155.29, 147.04, 139.37, 138.27, 131.26, 131.16, 130.09, 128.95, 128.70, 128.53, 126.46, 123.64, 123.17, 122.08, 119.11, 118.97, 118.35, 117.79, 59.30, 40.98, 35.56, 21.21, 14.43. HRMS (ESI) m/z calcd for C₂₈H₂₆BrN₄O₂+ (M + H)+ 529.1234, found 529.1234.

 $2-(4-methyl-1-oxopyrazino[1,2-b]indazol-2(1 H)-yl)-N-phenethyl-2-phenylacetamide (14j), 280 mg white solid, yield 64%, HPLC purity >98%, mp 189–191 °C. ¹H NMR (400 MHz, CDCl₃) <math>\delta$ 8.38–8.27 (m, 1 H), 7.94 (d, J = 8.6 Hz, 1 H), 7.53 (t, J = 7.7 Hz, 1 H), 7.39 (s, 4 H), 7.37 (d, J = 3.5 Hz, 2 H), 7.20–7.14 (m, 2 H), 7.11 (dd, J = 16.8, 4.0 Hz, 3 H), 6.88 (dd, J = 13.2, 3.3 Hz, 2 H), 6.47 (s, 1 H), 3.70 (dq, J = 13.5, 6.9 Hz, 1 H), 3.58 (dq, J = 13.3, 6.6 Hz, 1 H), 2.84 (t, J = 6.9 Hz, 2 H), 2.55 (s, 3 H). ¹³C NMR (100 MHz, CDCl₃) δ 167.84, 155.51, 148.63, 138.24,

134.53, 129.36, 129.18, 128.99, 128.71, 128.57, 127.60, 126.51, 124.25, 124.16, 121.03, 120.87, 119.28, 117.68, 117.42, 59.24, 40.98, 35.54, 14.55. HRMS (ESI) m/z calcd for $C_{27}H_{25}N_4O_2^+$ (M + H)⁺ 437.1972, found 437.1966.

Measurement of cell viability and proliferation. Human colon cancer cell lines HCT116 and SW620 were cultured under standard culture conditions at 37 °C and 5% CO₂ atmosphere in the specific medium (McCOY'S 5 A and DMEM) supplemented with 10% fetal bovine serum, 100 UI/mL penicillin and 100 mg/L streptomycin. The short-term effects of compounds 7 and 14 on cell growth were measured with the MTT assay. Briefly, HCT116 and SW620 cells were seeded into 96-well plates (3,000 cells/well) and incubated overnight at 37 °C, then treated with 0, 0.2, 0.4, 0.8, 1.6, 3.2, 6.4 and 12.8 µmol/L test compound for 24, 48 and 72 h. Next, 20 µL MTT solution (5 mg/mL) was added into each well and incubated for another 4 h followed by media removal and solubilization in 200 µL DMSO. The absorbance value was determined at 570 nm using a microplate reader (Bio-Tek, Winooski, VT, USA). The long-term effects of compounds 7 and 14 on cell growth were assessed with a clone formation assay. Cells were cultured at a density of 200 cells per well in 6-well plates for 36 h. Then, different concentrations of escin (0, 1, 2 and 4 µmol/L) were added. Cells were cultured for approximately 14 d until the cells grew visible colonies. The medium was discarded, and the colonies were stained with crystal violet (C0121, Beyotime) for 15 min at room temperature. After carefully washing with PBS, the stained colonies in each well were photographed and the number of colonies with more than 50 cells was counted manually.

Flow cytometry analysis. HCT116 and SW620 were exposed to different concentrations of compound 7h for 24 h, and then operated in flow cytometry analysis. Briefly, the harvested cells were fixed with 70% cold ethanol in 4 °C for 24 h. Then, the fixed cells were incubated in 200 μ L PBS solution containing 1 μ L propidium iodide (BD, CA, USA) and 1 μ L RNase (Sigma Aldrich, USA). Finally, cells were analyzed using BD AccuriTM C6 flow cytometry (BD Biosciences, USA). The FlowJo software was used to analyze the cell cycle arrest.

Received: 19 January 2020; Accepted: 14 May 2020; Published online: 09 June 2020

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Acknowledgements

The authors would like to thank the Science and Technology Research Program of Chongqing Municipal Education Commission (KJQN201801321, KJZD-M201801301 and KJQN201901305), the Natural Science Foundation Project of CQ CSTSC (cstc2018jszx-cyzd0110 and cstc2018jszx-cyzdX0023) and the Scientific Research Foundation of the Chongqing University of Arts and Sciences (2017RBX10). The authors would also like to thank Ms H.Z. L. for obtaining the LC/MS, HRMS and NMR data.

Author contributions

J.X. and H.T. conceived the details of the study, performed the chemicals synthesis and characterization. Y.Z. and D.T. performed and analyzed biological experiments. H.L., F.Z. and Z.C. conceived the study design, supervised the synthetic part and revised the manuscript. All the work was performed under the supervision of Z.X.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at https://doi.org/10.1038/s41598-020-66137-z.

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