SCIENTIFIC REPORTS

natureresearch

OPEN

Check for updates

Effects of biochar and crop straws on the bioavailability of cadmium in contaminated soil

Xuan Chen^{1,2}, Hong-Zhi He^{1,2,3}, Gui-Kui Chen^{1,2,3} & Hua-Shou Li^{1,2,3}

Numerous studies have been investigated the potential of biochar (BC) derived from various materials and crop straw (CS) to decrease the bioavailability of heavy metals in soil contaminated with cadmium (Cd), and thereby reduce their potential risk to human health and the ecological environment. However, little attention has been given to the comparison of heavy metal remediation efficiency using BC and CS such as peanut vine (PV) and rice straw (RS), especially in soil contaminated with Cd. Here, we explore if Cd bioavailability is affected in contaminated soil by BC and CS. Peanuts were grown in plastic pots, which contained BC or CS at 5% (dry weight, w/w) in controlled environment mesocosms. The bioavailability of Cd in contaminated soil was measured by Cd concentration in the plant and the concentrations of various forms of Cd in the soil. At the same plant age, growth with BC (compared with PV and RS) led to 13.56% and 8.28% lower rates of Cd content in the aboveground parts, 40.65% and 35.67% lower rates of Cd content in the seeds, yet 9.08% and 7.09% lower rates of Cd content in the roots, yet 35.80% and 28.48% lower rates of exchangeable Cd content in the soil. Moreover, BC amendment enhanced the biomass of peanut and physiological quality. Thus, BC had a greater impact on immobilizing Cd in the soil. The results imply that BC was more significantly (P < 0.05) remarkable in decreasing the Cd bioavailability and improving the biomass of peanut. BC has greater potential for enhancing soil quality and promoting peanut growth. In conclusion, this research demonstrates an understanding of employing BC as a promising inexpensive and eco-friendly amendment to remediate soil contaminated with Cd.

Nowadays, due to the expansion of industrial production and the rapid development of urbanization, the ever-increasing use of sewage irrigation, chemical fertilizers and pesticides in agricultural production result in the accumulation of heavy metals in the soil¹⁻³. Cd is a carcinogenic heavy metal^{4,5}, it has become one of the major sources of heavy metal pollution in soils and foods⁶⁻⁸. In the Toxic Substance and Disease Registry's (ATSDR) priority list of hazardous substances, it ranks seventh. And studies have reported the toxic effects of Cd on biological systems^{9,10}. In addition, Cd is extremely toxic to plants¹¹. Compared to other heavy metals, the toxicity of Cd is 2 to 20 times higher. Some studies have shown that Cd is the fourth most toxic metal to vascular plants^{12,13}. For plants, Cd is an unnecessary nutrient element, but plant roots can rapidly absorb it through nonspecific ion channels and via divalent metal transport $(DMT)^{14}$, and when the Cd content of leaves exceed $5-10 \mu g \cdot g^{-1}$, it becomes toxic¹⁵. Multiple studies have reported that excessive Cd can inhibit the normal metabolism of plants, which seriously affects the morphology and physiological processes of plants¹⁶. In general, Cd is absorbed by the roots, moved through via shoots and leaves, and then accumulated in grains and/or seeds^{17,18}. Physiologically, Cd could lead to excessive production of reactive oxygen species (ROS), resulting in oxidative damage to plants, which may change plant morphological characteristics and cause significant reduction in yield¹⁹. In addition, Cd could interfere with chloroplast metabolic function. Studies about cell ultrastructure of plants under Cd stress have shown that chloroplast has drastic changes, especially the decomposition of thylakoid membranes and an increase starch^{20,21}, which may be attributed to interference with chloroplast metabolism. Even worse, Cd could enter the human body system through food chain to affect human health²². When the accumulation of Cd reaches a certain amount, it would cause serious damage to the human kidney function and bones, and even lead to the

¹Key Laboratory of Tropical Agro-Environment, Ministry of Agriculture/South China Agricultural University, Guangzhou, 510642, PR China. ²Guangdong Engineering Research Center for Modern Eco-agriculture and Circular Agriculture/Key Laboratory of Agroecology and Rural Environment of Guangzhou, Regular Higher Education Institutions, Guangzhou, 510642, PR China. ³These authors contributed equally: Hong-Zhi He, Gui-Kui Chen and Hua-Shou Li. ^{Se}e-mail: lihuashou@scau.edu.cn occurrence of various cancers²³. At present, scholars have studied various methods to reduce the accumulation of Cd in crops.

Recently, studies have paid considerable attention to soil amendments such as BC and CS that immobilize contaminants whilst promoting plant growth, and these amendments have been widely applied for decreasing the bioavailability of heavy metals in soils^{23,24}. BC is a carbon-rich solid material made by pyrolysis of biomass under oxygen limited conditions²⁵. Typically, the physical and chemical properties of biochar what make it environmentally friendly²⁵, BC has a high porosity and large specific surface area, and strong ion exchange capacity, which offer a great potential for application to soils for carbon sequestration, greenhouse gas emission reduction, soil fertility improvement, and contaminated soil remediation²⁶⁻²⁸. BC is used as a soil conditioner to improve soil fertility as well as to adjust the soil pH, moisture, air, and temperature conditions²⁴. Previous researches have reported the biochemical effects of BC on heavy metal adsorption in soil and water in recent years^{29–31}. Cui et al.³² reported that wheat straw BC reduced the Cd bioavailability up to ~90% as compared to control (2.5 mg Cd/kg) and exhibited stable property based on the long-term incubation. Bashir et al.³³ showed that biochar significantly reduced the bioavailability of Cd and Cr by 85% and 63%, respectively. And when the application amount of biochar was 15 g·kg⁻¹, the extractable Cd in Cd-contaminated and Cr-Cd-contaminated soil could be reduced by 29% and 32%, respectively. Other studies presented that sugarcane straw derived BC could reduce the Zn, Cd, and Pb effectively in the pore water within acidified soil, thereby reducing their mobility and indicating that the main mechanism was the metal adsorption on BC³⁴. In a similar way, the application of CS as a fixative in soil contaminated with heavy metals may play an effective role for metal bioavailability. CS, as the main by-product of crops, not only has a positive effect on the soil structure, but also can loosen the soil, thus promoting the growth and development of crops³⁵. Moreover, many studies have demonstrated that straw returned to the field could produce dissolved organic carbon (DOC) which acted as an organic ligand to adsorb heavy metals in the soil³⁶. The organic ligand in soil solution could affect the formation of heavy metal complexes³⁷.

In this work, peanuts were used as experimental materials. Peanut (Arachis hypogaea) is one of the most important oilseed crops in the world and it has a strong absorption capacity for Cd^{38} . Some studies have shown that even on soils with relatively low total or available Cd concentrations ($<0.5 \text{ mg} \cdot \text{kg}^{-1}$), the Cd in seeds was higher than the maximum allowable concentration (MPC, 0.05 mg Cd/kg). In addition, the Cd concentrations at the top of plant always exceeded that in kernel or seeds. Furthermore, the testa in peanut kernel contained 50 times more Cd than the embryonic axis and cotyledons³⁹. Wang *et al.*⁴⁰ reported the Cd content in peanut kernel in Yantai ranged from 0.027 to 0.280 mg·kg⁻¹, with an average of 0.1048 mg·kg⁻¹, which surpassed hygienic quality standard of green peanut kernel issued by Ministry of Agriculture (HQSGPK). The rate beyond the HQSGPK reached 37.5% among all tested fields. So the development of peanut lines with food safety (Cd $\leq 0.2 \text{ mg/kg}$, GB 2762-2012); Cd $\leq 0.1 \text{ mg/kg}$ for the CODEX STAN 193-2015) would represent a major advance for the peanut industry. Our research goal is to develop an efficiency way for the immobilization of heavy metals in contaminated soil to assist the peanut safety.

Here, we examine how plants respond to Cd stress by measuring the biomass of peanut and physiological quality, and Cd content in different parts of peanut and exchangeable Cd content in soil. We performed a small scale experiment with peanut grown under BC addition and CS addition. The bioavailabilities of Cd in contaminated soil under BC addition and CS addition were analysed by laboratory simulation experiment. Results showed that BC and CS are good candidates for the remediation of soils contaminated with heavy metals. The efficiency of the BC and CS for the immobilization of heavy metals in contaminated soil should be carefully evaluated for each specific site prior to large-scale application. Therefore, the specific objectives of this paper were to, (1) determine the effect of BC and CS application on the bioavailability of Cd in contaminated soil, (2) compare the efficiency of BC and CS for remediation of soil contaminated with Cd, and (3) measure the effects of BC and CS application on plant yield and physiological response. This study aims to provide fundamental insights into the feasibility of using BC and CS to decrease the Cd bioavailability in contaminated paddy soil.

Materials and methods

Experiment materials. The topsoil (0–20 cm) was collected from an abandoned paddy field in Tianhe District, Guangzhou City, Guangdong Province, China, and it was classified as latosolic red soil according to the Chinese soil classification⁴¹. Soils were air-dried, crushed and sieved through the sieve of 2 mm meshes. The soil pH was 5.5, the organic matter content was 12.13 mg·kg⁻¹, and the total N, P and K concentrations of soil were 317.60 mg·kg⁻¹, 292.68 mg·kg⁻¹, 2236.61 mg·kg⁻¹, respectively, and the available N, P and K concentrations of soil were 27.13 mg·kg⁻¹, 1.14 mg·kg⁻¹, 64.14 mg·kg⁻¹, respectively. The total Cd content of the soils was below the detection limit of a graphite furnace (Z700P, Jena, Germany).

The peanut cultivar selected in this experiment was Yueyou No. 7 of Guangdong. The selected peanut shell BC was purchased from Sanli Company of Shangqiu, Henan Province, China. The total nitrogen and carbon content of BC were 7.20 g·kg⁻¹ and 521.31 g·kg⁻¹, respectively. PV and RS were collected from a farm in the Ecology Department of South China Agricultural University, their total nitrogen and carbon contents were 7.61 g·kg⁻¹, 501.77 g·kg⁻¹ and 6.36 g·kg⁻¹, 437.82 g·kg⁻¹, respectively. The total Cd contents of BC and CS were below the detection limit of graphite furnace atomic absorption spectrometry (Z700P, Jena, Germany). All three carbon-based materials were crushed and sieved through the sieve of 2-mm meshes and they were mixed well into the experimental soil.

Sampling and analysis. Soil and materials characterization. The physicochemical properties of soil were measured using the methods described by Bao⁴². Briefly, the soil pH was determined by a soil/water slurry ratio of 1:2.5 (w/v). The determination of soil organic matter was based on the method using potassium dichromate ($K_2Cr_2O_7$) and concentrated sulfuric acid (H_2SO_4) to oxidize the soil carbon before titrating with ferrous sulfate (FeSO₄). The total and available contents of phosphorus, potassium and nitrogen of the experimental soil were

determined according to Bao^{42,43}. The total Cd content of the soil was determined using a Z700P atomic absorption spectrometer (Z700P, Jena, Germany). The concentrations of various forms of Cd in soil were analyzed using a sequential extraction procedure developed by Tessier *et al.*⁴⁴.

The carbon contents of BC and CS were evaluated via C/N analyzer (TOC MULTI N/C 2000, Analytik Jena), and their nitrogen contents were determined according to Li *et al.*^{45,46}.

Plant analysis. Each plant sample was divided into four parts, including the shoot, shell, seed and root. The number of peanut pods and seeds in each treatment were counted for statistic analysis, and the dry matter weight was determined after the samples were oven-dried at 105 °C for 0.5 h and then oven-dried at 75 °C until their weight was constant.

Peanut leaves were collected and cut into pieces, then immersed in a mixture of ethanol and acetone (1:1; v/v) for measuring the concentrations of chlorophyll and proline by ultraviolet spectrophotometer. The concentrations of soluble protein, soluble sugars, and crude fat in peanut were measured using Commassie brilliant blue G-250 staining, anthrone colorimetry and Soxhlet extraction, respectively. All methods described above followed Modern Plant Physiology Experimental Guidelines⁴⁵. The Cd concentration in the plant samples was extracted using microwave digestion and determined using an atomic absorption spectrometer (Z700P, Jena, Germany).

Experiment design. The experiment was carried out on a farm (23°16′N, 113°37′E) of Ecology Department of South China Agricultural University in Tianhe District, Guangzhou City, Guangdong Province, China, under natural sunlight. To prepare a Cd-contaminated soil, 3L solution containing 48 mg of CdCl₂ in deionized water was sprayed onto 3 kg soil, then the soil was gradually diluted with 45 kg of clean soil until the final concentration of Cd^{2+} reached 1 mg kg⁻¹. Subsequently, a total of 4 kg of contaminated soil spiked with Cd was placed into a plastic pot (diameter 20 cm, height 16 cm). Peanut shell BC, PV, and RS were added into the pots at a rate of 5% (dry weight, w/w) and mixed thoroughly to ensure uniformity. This experiment consisted of 4 treatments and every treatment had 3 replications. The soil contaminated with Cd without any amendment was set as the control. To guarantee the regular growth of plants, each pot was supplemented with urea, $Ca(H_2PO_4)_2$, and KCl at a dose of 0.2 g·kg⁻¹, 0.4 g·kg⁻¹, and 0.3 g·kg⁻¹, and these pots were arranged in a randomized complete block design. The pots were irrigated with deionized water to 70% of the field water holding capacity, and allowed to equilibrate for 4 weeks. Peanut seeds of uniform size were sowed in a sandy bed. At the four-leaf stage, uniform seedlings were selected and transplanted into the plastic pots, 3 seedlings were kept in each pot. Regular watering (2 to 4 times a week) with deionized water was maintained to prevent drought stress to the plants. After 4 months (from March 8 to July 8, 2016) the mature peanut were harvested, the soil and the plant were sampled, ground and sieved for further analysis.

Statistical analysis. The data was analyzed via Microsoft Office Excel 2007 and Data Processing System (DPS 7.05), and the data of the study was presented as mean value and standard deviation. Significant differences were tested among treatments by one-way ANOVA and via post hoc least significant difference tests (LSD) for multiple comparisons at a 5% significance level. All statistical analyses were carried out using SPSS 17.0 and the data was graphed using Origin 8.0.

Results

Soil pH. BC and CS had strong effects on soil pH. Particularly in the BC treatment. Relative to the control, both BC and CS application could effectively increase the pH of soil (Fig. 1). The pH of soil was 24.00% higher at T_B . Meanwhile, CS increased pH by 17.54% (T_R) and 15.54% (T_P), respectively (P < 0.05), whereas there was non-significant difference between T_R and T_P treatments.

Infrared spectra of DOM in BC and CS. It can be seen from Table 1 and Fig. 2 that there were two obvious absorption peaks in the DOM of BC at 1400.80 cm⁻¹ and 3129.28 cm⁻¹. The absorption peak at 1400.80 cm⁻¹ showed that there were more CH_3 and CH_2 in the aliphatic group and fewer C = C and amido bonds on the aromatic group. A wide absorption band at 3129.28 cm⁻¹ indicated that the contents of phenols, –COOH, and alcohols were much higher.

From Figs. 3 and 4, it presented that in the DOM of PV, there were four obvious absorption peaks at 1069.43 cm^{-1} , 1400.74 cm^{-1} , 1645.79 cm^{-1} and 3135.39 cm^{-1} . The absorption peak at 1069.43 cm^{-1} indicated that there were more C-O on the phenols or the alcohols. Besides, there were more CH₃ and CH₂ in the aliphatic group at 1400.74 cm^{-1} and fewer amido bonds. Moreover, the absorption peak at 1645.79 cm^{-1} showed that the contents of aldehyde, ketone and aromatic group were much higher. In addition, the absorption peak was widened at 3135.39 cm^{-1} , and the contents of phenols and alcohols increased. Relative to PV, a total of six obvious absorption peaks appeared in the DOM of the decomposition products of PV. At 469.61 cm^{-1} and 536.89 cm^{-1} , the carbohydrate content was much higher. Also, the degree of absorption at 1000 cm^{-1} was relatively weak, which means that the C-O of phenols or alcohols was more converted to carbohydrates. Plus, the absorption peaks were widened at 3440.91 cm^{-1} and the two small absorption peaks at 3619.68 cm^{-1} and 3696.30 cm^{-1} indicated that an increase in the contents of -COOH, phenols and alcohols.

Furthermore, from Figs. 5 and 6, there were three obvious absorption peaks in the DOM of RS at 1054.71 cm^{-1} , 1400.63 cm^{-1} and 3259.00 cm^{-1} , and seven distinct absorption peaks in the DOM of the decomposition products of RS, including 470.14 cm^{-1} , 537.98 cm^{-1} , 1000 cm^{-1} , 1400.60 cm^{-1} , 3136.71 cm^{-1} , 3620.00 cm^{-1} , and 3696.17 cm^{-1} . The results were consistent with the DOMs of PV and the decomposition products of PV, respectively.

The contents of total Cd and various forms of Cd in soil. As expected, in both BC and CS, the total Cd contents in soil were all consistently higher than T_{CK} (Fig. 7) and the exchangeable Cd contents were lower than



Figure 1. Effect of biochar and crop straw application on soil pH. Treatments: T_{CK} : control, T_B : biochar addition, T_p : peanut straw addition, T_R : rice straw addition. Error bars indicate standard error of the means (n = 3). Different letters indicates significant difference among treatments (P < 0.05).

Absorption band position/cm ⁻¹	Absorption band assignment				
650-520	Stretching vibration of -OH (carbohydrates)				
870	Carbonate substance				
1020-970	Stretching vibration of C-O or stretching vibration of inorganic SiO (carbohydrates)				
1080-1020	Asymmetric stretching vibration of C-O (phenols or alcohols)				
1170-1150	Stretching vibrations of C-OH and C-O (aliphatic)				
1220-1210	Asymmetric stretching vibration of C-O or deformable vibration of N-H (hydroxyl)				
1250-1230	Stretching vibration of C-O or stretching vibration of SiO in organosilicon compounds (phenols)				
1460-1400	Symmetric deformable vibrations of $-CH_3$ and $-CH_2$, and asymmetric stretching vibration on hydroxyl group, or stretching vibration of C-OH (aliphatic)				
1555-1540	Deformable vibration of -N-H (secondary amide)				
1650-1600	Stretching vibration of $-C = O$, stretching vibration of $C = C$ on aromatic group or antisymmetric vibration of organic carboxylate COO- (aldehyde, ketone)				
1720-1690	Stretching vibration of -C = O, stretching vibration of C = O in hydroxyl group (hydrogen bond formed between molecules and within molecules)				
2870-2850	Symmetric stretching vibrations of -CH ₃ and -CH ₂				
2900	Stretching vibration of C-H (aliphatic)				
2930	Asymmetric stretching vibration of -CH ₂ (aliphatic)				
2950	Asymmetric stretching vibration of -CH ₃ (aliphatic)				
2060-3030	Stretching vibration of -C-H (aromatic nucleus)				
3500-3300	Stretching vibrations of -COOH and -OH or stretching vibration of N-H and hydrogen bond association				

 Table 1. Assignment of characteristic absorption bands in infrared spectra. According to Huang (2013), etc.

 T_{CK} (Fig. 8). In this study, growth under BC addition (compared with PV addition and RS addition) led to 18.97% and 29.90% higher rates of total Cd content in the soil, yet 35.80% and 28.48% lower rates of exchangeable Cd content in the soil (P < 0.05). Furthermore, BC and CS could effectively increase the contents of carbonate-bound Cd and organic-bound Cd in soil (Fig. 9). Particularly in the biochar treatment, the highest increase reached to 48.07% and 52.94%, respectively.

Cd accumulation in the tissues of peanut. The Cd concentrations in the tissues of peanut were given in Fig. 10. When compared at the same age, the concentrations of Cd in the different parts of the peanut followed the order: root > aboveground > shell > seed. When compared at the same tissue, the concentrations of Cd under different materials followed the order: root > aboveground > shell > seed. Compared to the control, with either T_B or T_R or T_P , the Cd concentrations in roots decreased by 13.99%, 7.89% and 5.40%, respectively (P < 0.05), and led to 20.30%, 13.70% and 7.80% lower rates of Cd concentrations in aboveground parts, yet 9.54%, 5.52% and 5.03% lower rates of Cd concentrations in shells, yet 28.77%, 5.84% and 5.30% lower rates of Cd concentrations in seeds. In addition, there was non-significant difference between T_R and T_P treatments.



Figure 2. Infrared spectra of DOM in biochar.



Figure 3. Infrared spectra of DOM in peanut vine.



Figure 4. Infrared spectra of DOM in the decomposition products of peanut vine.

Physiological parameters, biomass and yield of peanut. Both BC and CS had clear effects on many physiological parameters (Figs. 11 and 12). Particularly in the BC treatment. Relative to CS, BC had an approx. 95.61% higher chlorophyll, 95.65% higher proline, 81.25% higher soluble sugars, 71.32% higher soluble proline, and a 27.37% higher crude fat, as compared with the control. On the other hand, BC had 32.35% and 60.71% higher proline, yet 21.56% and 23.78% higher soluble sugars, yet 36.42% and 38.99% higher soluble proline, yet 9.20% and 17.13% higher crude fat compared with RS and PV. Nevertheless, all physiological parameters did not differ significantly between the two CS (P > 0.05).

Table 2 presents the biomass and yield of peanut with the applications of BC and CS. The applications of BC and CS significantly increased the biomass and yield of peanut. Particularly in the BC treatment, the highest decrease of biomass in aboveground parts and underground parts reached to 86.49%, and 75.47%, respective-ly(P < 0.05), and the highest decrease of yield in seeds per plant reached to 61.42% (P < 0.05).



Figure 5. Infrared spectra of DOM in rice straw.



Figure 6. Infrared spectra of DOM in the decomposition products of rice straw.



Figure 7. Effect of biochar and crop straw application on soil total Cd content. Treatments T_{CK} : control, T_B : biochar addition. T_P : peanut straw addition, T_R : rice straw addition. Error bars indicates standard error of the means (n = 3). Different letters indicate significant difference among treatments (P < 0.05).

Discussion

Biochar had a greater impact on immobilizing Cd in the soil. This work shows that the contents of total Cd and exchangeable Cd in the soil were affected by biochar and crop straw, with a higher total Cd and a lower exchangeable Cd in the biochar and crop straw treatments mainly due to the increase in pH. One of the mechanisms for the pH increase after the application of these two materials is probably due to the considerable amount of ash and base cations^{47–49}. On the one hand, for biochar, the basic cations existing on the feedstock



Figure 8. Effect of biochar and crop straw application on soil exchangeable Cd content. Treatments: T_{CK} : control, T_B : biochar addition, T_P : pearnut straw addition, T_R : rice straw addition. Error bars indicate strandard error of the means (n = 3). Different letteres indicate significant difference among treatments (P < 0.05).





biomass could be converted into oxides, hydroxides and carbonates produced in the pyrolysis process of biochar, which may contribute to the increase of soil pH^{50} . On the other hand, the high alkalinity of these two materials²⁵, and the subsequent release of base cations, especially Ca²⁺ and K⁺, and the replacement of soil exchangeable Al³⁺ and H⁺ by these cations on the soil's negatively charged sites could greatly increase soil pH^{51} . In addition, the decarboxylation of organic anions induced by these two materials, owing to increasing attack of organic anions by microbes, which may consume H⁺ from the soil solution^{52,53}.

The response of exchangeable Cd content to pH is important for understanding the effect of organic materials on the bioavailability of Cd. The increase of the pH resulted in the hydrolysis of heavy metal cations to form oxide precipitations, which decreased the content of exchangeable Cd in soil⁵⁴. In addition, many studies have shown that the application of organic materials (including crop straw and biochar) can significantly affect the adsorption and desorption behavior of heavy metal in soil^{55,56}. The most direct effect of biochar and crop straw was to bring a large amount of dissolved organic matter (DOM) into soil. DOM is the most active component of organic matter and has active groups. It is easy to chelate and complex with Cd as ligands, and then change the availability of Cd⁵⁷.

The negative charge on the surface of clay minerals, hydrated oxides and organic matter in soil increases with the increase of soil pH, which increases the adsorption of Cd^{2+} , which promotes the formation of $CdCO_3$ and $Cd(OH)_2$ precipitates⁵⁸, which may be the reason for the increase of the content of carbonate-bound Cd in soil⁵⁹, and the reason for the increasing the content of organic-bound Cd in the soil might be due to the abundant



Figure 10. Effect of biochar and crop straw addition on Cd accumulation in the tissues of peanut. Treatments: T_{CK} : control, T_B : biochar addition, T_P : peanut straw addition, T_R : rice straw addition. Error bars indicate strandard error of the means (n = 3). Different letters indicate significant difference among treatments (*P* < 0.05).





carboxyl groups in biochar and the decomposed products of crop straw contained more carboxyl groups, aldehyde groups, ketone groups, and aromatic substances, so the retention ability of ions was enhanced, and a significant colloidal characteristic was gradually exhibited⁶⁰. Furthermore, biochar and crop straw can reduce the toxicity of Cd to peanuts by increasing the concentration of total Cd in the soil. In our study, we found that these two materials can significantly increase the concentration of total Cd. The reason might be due to the numerous organic functional groups in the biochar, such as -C-OH, -C=O and COO–, which could complex heavy metal ions⁶¹. As can be seen from Fig. 3, the effects of the three carbon-based materials on the total Cd were consistent, the organic matter produced by the decomposition of crop straw also played the same role^{62,63}.

In summary, the inhibition effect of biochar and crop straw on Cd adsorption is mainly through chemisorption. The inhibition effect of biochar mainly relies on the ion exchange of metal cations on its surface⁶⁴ and the precipitation or complexation of metal ions on biochar with minerals or functional groups⁵⁴. The inhibition effect of crop straw mainly relies on the decomposition products produced by soil microbes, and there are plenty of functional groups on the surface of crop straw, which can complexation metal ions⁶⁵.

Biochar has a positive effect on reducing Cd content in peanut. The results of this study show that the addition of biochar and crop straw could effectively reduce the absorption of Cd in peanut. Some studies presented that the addition of biochar could inhibit the absorption of Cd in rapeseed⁶⁶ and other plants, such as



Figure 12. Effect of biochar and crop straw addition on crude fat in peanut. Treatments: T_{CK} : control, T_B : biochar addition, T_P : peanut straw addition, T_R : rice straw addition. Error bars indicates strandard error of the means (n = 3). Different letters indicate significant difference among treatments (P < 0.05).

	Biomass		Yield			
	Aboveground (g·plant ⁻¹)	Underground (g-	plant ⁻¹)	Number of effective pods per	Number of seeds	
Treatments		Roots	Seeds	Shells	plant	per plant
T _{CK}	$9.45 \pm 1.54c$	$1.61\pm0.29c$	$6.26\pm0.46c$	$3.75\pm0.34b$	$15.00 \pm 0.58c$	$19.00 \pm 1.15c$
T _B	$17.61 \pm 2.33a$	$4.05\pm0.09a$	$11.17 \pm 0.55a$	$5.17\pm0.32a$	$21.00\pm0.57a$	$30.67 \pm 0.58a$
T _p	14.00±1.38b	$2.16\pm0.09b$	$9.91 \pm 1.62 b$	$4.62 \pm 1.14 b$	$15.33\pm1.53b$	$21.00 \pm 1.53b$
T _R	$14.97 \pm 1.25b$	$2.37\pm0.24b$	$10.50 \pm 0.82b$	$4.85\pm0.77b$	$16.33\pm0.57b$	$22.00 \pm 1.15b$

Table 2. Effect of biochar and crop straw addition on the biomass and yield of peanut. Treatments: TCK: control, TB: biochar addition, TP: peanut straw addition, TR: rice straw addition. All values are presented as mean \pm standard error (n = 3), different letters in the same row indicate significant differences between treatments (P < 0.05).

.....

rice⁶⁷, wheat⁶⁸, spinach⁶⁹. In this research, the reason that biochar could reduce the Cd concentration in different parts of peanut might be that on the one hand, biochar could increase Cd concentration bound to the soil organic matter. On the other hand, it might be due to the reduction in pore water Cd concentration⁷⁰. For crop straw, studies have shown that the addition of crop straw not only increased the pH of soil⁷¹, but also improved the metal adsorption by ligands in organic matter^{63,72,73}, so as to reduce the absorption of Cd by plants. Ok *et al.*⁷⁴ found that the use of rapeseed residue amendment could decrease the easily accessible fraction of Cd by 5 to 14%, thus reducing the bioavailability of Cd. Xu *et al.*⁷² reported that after adding rice straw and wheat straw, the accumulation of Cd in maize shoots decreased by 69.5 and 66.9%, respectively. Zeng *et al.*⁶⁵. found that in the cassava–peanut intercropping system, the uptake of Cd by the two crops was significantly reduced after the addition of crop straw. In this work, the results indicate that the most easily accumulated Cd parts in peanuts were roots, followed by shoots and pulses(grains). The higher concentration of Cd in the roots suggested that one of the ways to inhibit the transfer of Cd from roots to shoots might be by locating toxic metals in the plant tissues⁷⁵.

Biochar and crop straw additions enhanced the biomass of peanut and physiological quality. The present study indicates that these two materials increased the physiological quality of peanuts compared to the control. In this study, the increase in chlorophyll contents might be due to the reversal of Cd-induced toxicities in the plants with organic materials applications^{68,76}. In addition, the increase in proline, soluble protein, soluble sugars, and crude fat with the additions of biochar and crop straw might be due to the increase in mineral nutrients and decrease in Cd concentration in the plants⁷⁷.

Further, the biochar and crop straw exerted a positive effect on enhancing the biomass and yield of peanut. On the one hand, the biomass and yield of peanut are well known to be improved by the additions of biochar and crop straw, and this improvement is attributed to the modification of the physicochemical and biological characteristics of the cultivated soil^{77,78}. On the other hand, the application of these two materials can reduce the bulk density and increase the total porosity of the soil, which could provide a good space for the root growth.

Conclusions and perspectives

This study showed biochar and crop straw had significant effects on decreasing of Cd availability in soil and Cd concentration in peanut pulses, especially biochar, it had a greater impact on immobilizing Cd. At the same peanut plant age, growth with biochar (compared with peanut vine and rice straw) led to 13.56% and 8.28% lower rates of Cd content in the aboveground parts, 40.65% and 35.67% lower rates of Cd content in the seeds, yet 9.08% and 7.09% lower rates of Cd content in the roots, and 35.80% and 28.48% lower rates of exchangeable Cd content in the soil. In the meantime, Cd uptake by the peanut markedly decreased, and the biomass and yield of peanut markedly increased. These findings suggested that biochar could be used as an ecological friendly amendment, and served for mitigates Cd pollution in the soil-grain system.

Received: 31 December 2019; Accepted: 6 May 2020;

Published online: 12 June 2020

References

- 1. Xuan, B., Wang, J., Duan, Z. B., Wang, K. & An, J. P. Review on contamination and remediation technology of heavy metal in agricultural soil. Advances in Environmental Protection. 7(1), 26–34 (2017).
- Tang, C. L., Sun, P. F., Yang, J. L., Huang, Y. P. & Wu, Y. H. Kinetics simulation of Cu and Cd removal and the microbial community adaptation in a periphytic biofilm reactor. *Bioresource Technology*. 276, 199–203 (2019).
- Sonali, D., Manju, S., Anubhuti, G., Vibha, R. & Debasis, C. Toxicity and detoxification of heavy metals during plant growth and metabolism. *Environmental Chemistry Letters*. 16(4), 1169–1192 (2018).
- Chen, L. et al. High cadmium adsorption on nanoscale zero-valent iron coated Eichhornia crassipes BC. Environmental Chemistry Letters. 17(1), 589–594 (2019).
- 5. Rohan, J. et al. Higher Cd adsorption on biogenic elemental selenium nanoparticles. Environmental Chemistry Letters. 14(3), 381–386 (2016).
- Sharma, S., Nagpal, A. K. & Kaur, I. Heavy metal contamination in soil, food crops and associated health risks for residents of Ropar wetland, Punjab, India and its environs. *Food Chemistry.* 255, 15–22 (2018).
- 7. Ronzan, M. *et al.* Cadmium and arsenic affect root development in *Oryza sativa* L. negatively interacting with auxin. *Environ Exp Bot.* **151**, 64–75 (2018).
- 8. Xu, L. *et al.* Adaption and restoration of anammox biomass to Cd (II) stress: Performance, extracellular polymeric substance and microbial community. *Bioresource Technology.* **290**, 121766 (2019).
- Adams, M. L., Zhao, F. J., McGrath, S. P., Nicholson, F. A. & Chambers, B. J. Predicting cadmium concentrations in wheat and barley grain using soil properties. J. Environ. Qual. 33(2), 532–541 (2004).
- Rahimzadeh, M. R., Kazemi, S. & Moghadamnia, A. A. Cadmium toxicity and treatment: an update Casp. J. Intern. Med. 8(3), 135–145 (2017).
- Nagajyoti, P. C., Lee, K. D. & Sreekanth, T. V. M. Heavy metals, occurrence and toxicity for plants: a review. *Environmental Chemistry Letters*. 8(3), 199–216 (2010).
- 12. Vassilev, A., Tsonev, T. & Yordanov, I. Physiological response of barley plants (Hordeum vulgare) to cadmium contamination in soil during ontogenesis. *Environ. Pollut.* **103**(2–3), 287–293 (1998).
- Jones, R., Lapp, T. & Wallace. D. Locating and Estimating Air Emissions from Sources of Cadmium and Cadmium Compounds. Office of Air and Radiation Report Prepared by Midwest Research Institute for the US Environmental Protection Agency. EPA-453/R-93-040 (1993).
- 14. Liu, J. G. *et al.* Correlations between cadmium and mineral nutrients in absorption and accumulation in various genotypes of rice under cadmium stress. *Chemosphere.* 52, 1467–1473 (2003).
- 15. White, P. J. & Brown, P. H. Plant nutrition for sustainable development and global health. Ann. Bot. 105, 1073-1080 (2010).
- Pietrini, F., Iannelli, M. A., Pasqualini, S. & Massacci, A. Interaction of cadmium with glutathione and photosynthesis in developing leaves and chloroplasts of Phragmites australis (Cav.) Trin. Ex Steudel. *Plant Physiol.* 133, 829–837 (2003).
- Li, Y., Pang, H., He, L., Wang, Q. & Sheng, X. Cd immobilization and reduced tissue Cd accumulation of rice (Oryza sativa wuyun-23) in the presence of heavy metal-resistant bacteria. *Ecotoxicol. Environ. Saf.* 138, 56–63 (2017).
- Ashraf, U. et al. Alterations in growth, oxidative damage, and metal uptake of five aromatic rice cultiv0ars under lead toxicity. Plant Physiol. Biochem. 115, 461–471 (2017).
- Khan, M. I. R., Nazir, F., Asgher, M., Per, T. S. & Khan, N. A. Selenium and sulfur influence ethylene formation and alleviate cadmium-induced oxidative stress by improving proline and glutathione production in wheat. J. Plant Physiol. 173, 9–18 (2015).
- 20. Pereira, L. S. *et al.* Cadmium induced changes in Solidago chilensis Meyen (Asteraceae) grown on organically fertilized soil with reference to mycorrhizae, metabolism, anatomy and ultrastructure. *Ecotoxicol. Environ. Saf.* **150**, 76–85 (2018).
- Daud, M. K., Quiling, H., Lei, M., Ali, B. & Zhu, S. J. Ultrastructural, metabolic and proteomic changes in leaves of upland cotton in response to cadmium stress. *Chemosphere* 120, 309–320 (2015).
- Kaplan, O., Ince, M. & Yaman, M. Sequential extraction of cadmium in different soil phases and plant parts from a fromer industrialized area. *Environmental Chemistry Letters*. 9(3), 397–404 (2011).
- Lin, L. J. et al. Effects of living hyperaccumulator plants and their straws on the growth and cadmium accumulation of cyphomandra betacea seedlings. Ecotoxicol. Environ. Saf. 155, 109–116 (2018).
- Kasak, K. et al. Biochar enhances plant growth and nutrient removal in horizontal subsurface flow constructed wetlands. Science of the Total Environment. 639, 67–74 (2018).
- Bashir, S., Zhu, J., Fu, Q. L. & Hu, H. Q. Cadmium mobility, uptake and anti-oxidative response of water spinach (Ipomoea Aquatic) under rice straw biochar, zeolite and rock phosphate as amendments. *Chemosphere*. 194, 579–587 (2018).
- Jeffery, S. *et al.* The way forward in biochar research: targeting trade-offs between the potential wins. *GCB Bioenergy.* 7, 1–13 (2015).
 Mahar, A., Wang, P., Li, R. H. & Zhang, Z. Q. Immobilization of lead and cadmium in contaminated soil using amendments: a review. *Pedosphere.* 25(4), 555–568 (2015).
- Mandal, S. et al. Designing advanced BC products for maximizing greenhouse gas mitigation potential. Environ. Sci. Technol. 46, 1367–1401 (2016).
- 29. Ahmad, M. et al. BC as a sorbent for contaminant management in soil and water: a review. Chemosphere. 99, 19-33 (2014b).
- Qi, F. J. et al. Pyrogenic carbon and its role in contaminant immobilization in soils. Crit. Rev. Environ. Sci. Technol. 47(10), 795–876 (2017).
- 31. Zhang, M. *et al.* BC reduces cadmium accumulation in rice grains in a tungsten mining area-field experiment: effects of BC type and dosage, rice variety, and pollution level. *Environmental Geochemistry and Health.* 41(1), 43–52 (2019).
- Cui, L., Noerpel, M. R., Scheckel, K. G. & Ippolito, J. A. Wheat straw biochar reduces environmental cadmium bioavailability. Environ. Int. 126, 69–75 (2019).
- Bashir, S. et al. Sugarcane bagasse-derived biochar reduces the cadmium and chromium bioavailability to mash bean and enhances the microbial activity in contaminated soil. Journal of Soil and Sediments. 18(3), 874–886 (2018).

- Puga, A. P., Melo, L. C. A., de Abreu, C. A., Coscione, A. R. & Paz-Ferreiro, J. Leaching and fractionation of heavy metals in mining soils amended with BC. Soil. Till. Res. 164, 25–33 (2016).
- 35. Bi, Y. Y. Study on straw resources evaluation and utilization in China. Beijing, Chinese Academy of Agricultural Science (2010).
- Kim, H. B. et al. Effect of dissolved organic carbon from sludge, rice straw and spent coffee ground BC on the mobility of arsenic in soil. Science of the Total Environment. 636, 1241–1248 (2018).
- 37. Xu, P. *et al.* The effect of biochar and crow straws on heavy metal bioavailability and plant accumulation in a Cd and Pb polluted soil. *Ecotoxicol Environ Saf.* **132**, 94–100 (2016).
- Christou, A., Theologides, C. P., Coasta, C., Kalavrouziotis, I. K. & Varnavas, S. P. Assessment of toxic heavy metals concentrations in soils and wild and cultivated plants species in Limni abandoned copper mining site, Cyprus. J. Geochem. Explor. 178, 16–22 (2017).
- Bell, M. J., Mclaughlin, M. J., Wright, G. C. & Cruickshank, A. Inter and intra-specific variation in accumulation of cadmium by peanut, soybean, and navybean. Australian Journal of Agricultural Research. 48(8), 1151–1160 (1997).
- Wang, Y. Y., Gao, B., Zhang, J. L. & Li, X. D. Effects of different sulfur application rates on physiological characteristics, yield and quality of peanut. Shandong Agricultural Sciences. 46(12), 67–71 (2014).
- 41. Gong, Z. T. Chinese soil taxonomic: theory approaches and application (ed. Chen, P. L.) 1-903 (Science Press, 1999).
- 42. Bao, S. D. Soil agro-chemistrical analysis (ed. Bao, S.) 1-495 (China Agriculture Press, 2000).
- Li, Z. Y., Zheng, L., Lu, L. H. & Li, L. Improvement in the H₂SO₄-H₂O₂ Digestion Method for Determining Plant Total Nitrogen. Chinese Agricultural Science Bulletin. 30(6), 159–162 (2014).
- Tessier, A., Campbell, P. G. & Bisson, M. Sequential extraction procedure for the speciation of particulate trace metals. *Anal. Chem.* 51, 844–851 (1979).
- 45. Science of Plant Physiology and Ecology, SIBS, CAS. Modern Plant Physiology Experimental Guidelines (ed. Tang, Z. C.) 1–415 (Science Press, 1999).
- 46. Huang, J. Y. Impact mechanism of straw returning on cadmium speciation and bioavailability of soils in tongling mining area. PhD diss. Hefei. Hefei University of Technology (2013).
- Li, S., Barreto, V., Li, R., Chen, G. & Hsieh, Y. P. Nitrogen retention of biochar derived from different feedstocks at variable pyrolysis temperatures. J. Anal Appl. Pyrolysis. 133, 136–146 (2018).
- Pocknee, S. & Sumner, M. E. Cation and notrogen contents of organic matter determine its soil liming potential. Soil Sci. Soc. Am. J. 61, 86–92 (1997).
- Noble, A. D., Zenneck, I. & Randall, P. J. Leaf litter ash alkalinity and neutralization of soil acidity. *Plant Soil.* 179, 293–302 (1996).
 Bashir, S. *et al.* Efficiency of C3 and C4 plant derived-biochar for Cd mobility, nutrient cycling and microbial biomass in https://doi.org/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10.1016/j.com/10016/j.com/1001
- contaminated soil. *Bulletin of Environmental Contamination and Toxicology.* 100(6), 834–838 (2018).
 51. Masud, M. M., Li, J. Y. & Xu, R. K. Use of alkaline slag and crop residue biochars to promote base saturation and reduce acidity of an acidic ultisol. *Pedosphere.* 24, 791–798 (2014).
- Wang, L. et al. Effect of crop residue biochar on soil acidity amelioration in strongly acidic tea garden soils. Soil Use Manag. 30, 119-128 (2014).
- Yan, F., Schubert, S. & Mengel, K. Soil pH increase due to biological decarboxylation of organic anions. Soil Biol & Biochem. 28, 17-24 (1996).
- Yuan, J. H., Xu, R. K., Qian, W. & Wang, R. H. Comparison of the ameliorating effects on an acidic ultisol between four crow straws and their biochars. *Journal of Soils and Sediments.* 11(5), 741–750 (2011).
- Wang, G. M. & Zhou, L. X. The dynamics of dissolved organic matter and associated water-soluble Cu in two Cu-contaminated soils amended with various organic matters. Acta Scientiae Circumstantiae. 23(4), 453–456 (2003).
- Zhu, L., Wu, J., Zhou, J. M., Chen, H. L. & Tang, D. M. Effect of dissolved organic matter on sorption-desorption behavior of copper in soil. *Journal of Agro-Environment Science*. 27(5), 1779–1785 (2008).
- Chen, T. B. & Chen, Z. J. Cadmium adsorption in soil influenced by dissolved organic matter derived from rice straw and sediment. *Chinese Journal Of Applied Ecology.* 13(2), 183–186 (2002).
- Liu, G. S., Xu, Z. J., Zhou, G. D. & Liu, W. P. Studies on the character and rule of cadmium release from red soils under the action of acid rain. *China Environmental Science*. 24(4), 419–423 (2004).
- 59. Zhou, H. J. *et al.* Effects of biochar on Cd forms in red soil and cinnamon soil. *Journal of Plant Nutrition and Fertilizers.* **25**(3), 433–442 (2019).
- Li, H. B. et al. Mechanisms of metal sorption by biochars: biochar characteristics and modifications. Chemosphere. 178, 466–478 (2017).
- 61. Qian, L. B. *et al.* Effective removal of heavy metal by biochar colloids under different pyrolysis temperatures. *Bioresour. Technol.* 206, 217–224 (2016).
- Gao, J. K., Lv, J. L., Wu, H. M., Dai, Y. C. & Nasir, M. Impacts of wheat straw addition on dissolved organic matter characteristics in cadmium-contaminated soils: insights from fluorescence spectroscopy and environmental implications. *Chemosphere.* 193, 1027–1035 (2017).
- 63. Mohamed, I. et al. Fractionation of copper and cadmium and their binding with soil organic matter in a contaminated soil amended with organic materials. J. Soil Sediment. 10(6), 973–982 (2010).
- 64. Li, H. B. et al. Mechanisms of metal sorption by biochars: Biochar characteristics and modifications. Chemosphere. **178**, 466–478 (2017).
- Zeng, L. P., Lin, X. K., Zhou, F., Qin, J. H. & Li, H. S. Biochar and crushed straw additions affect cadmium absorption in cassavapeanut intercropping system. *Ecotoxicol Environ Saf.* 167, 520–530 (2019).
- 66. Shaheen, S. M. & Rinklebe, J. Impact of emerging and low cost alternative amendments on the (im) mobilization and phytoavailability of Cd and Pb in a contaminated floodplain soil. *Ecol. Eng.* 74, 319–326 (2015).
- 67. Bian, R. *et al.* Biochar soil amendment as a solution to prevent Cd-tainted rice from China: results from a cross-site field experiment. *Ecol. Eng.* **58**, 378–383 (2013).
- 68. Yousaf, B. *et al.* Investigating the potential influence of biochar and traditional organic amendments on the bioavailability and transfer of Cd in the soil-plant system. *Environ. Earth Sci.* **75**, 1–10 (2016).
- 69. Younis, U. *et al.* Biochar enhances the cadmium tolerance in spinach (Spinacia oleracea) through modification of Cd uptake and physiological and biochemical attributes. *Environ. Sci. Pollut. Res.* 23, 21385–21394 (2016).
- 70. Lu, K. P. *et al.* Effect of bamboo and RS biochars on the mobility and redistribution of heavy metals (Cd, Cu, Pb and Zn) in contaminated soil. *J. Environ. Manag.* **186**, 285–292 (2017).
- Wang, S. *et al.* Speciation and phytoavailability of cadmium in soil treated with cadmium-contaminated RS. *Environ Sci Pollut Res.* 22, 2679–2686 (2015).
- 72. Xu, P. *et al.* The effect of biochar and crow straws on heavy metal bioavailability and plant accumulation in a Cd and Pb polluted soil. 2016. *Ecotoxicol Environ Saf.* **132**, 94–100 (2016).
- 73. Tang, W. L. *et al.* Inhibitory effects of rice residues amendment on Cd phytoavailability: a matter of Cd-organic matter interactions? *Chemosphere.* **186**, 227–234 (2017).
- 74. Ok, Y. S. *et al.* Effects of rapeseed residue on lead and cadmium availability and uptake by rice plants in heavy metal contaminated paddy soil. *Chemosphere.* **85**, 677–682 (2011).

- 75. Rizwan, M., Meunier, J. D., Hélène, M. & Keller, C. Effect of silicon on reducing cadmium toxicity in durum wheat (*Triticum turgidum* L. cv. Claudio W.) grown in a soil with aged contamination. *J. Hazard. Mater.* **209–210**, 326–334 (2012).
- Rizwan, R. *et al.* Exogenous proline and glycinebetaine mitigate cadmium stress in two genetically different spring wheat (Triticum aestivum L.) cultivars. *Braz. J. Bot.* 37(4), 399–406 (2014).
- Borchard, N., Siemens, J., Ladd, B., Möller, A. & Amelung, W. Application of biochars to sandy and silty soil failed to increase maize yield under common agricultural practice. Soil and Tillage Research. 144, 184–194 (2014).
- Jones, D. L., Rousk, J., Edwards-Jones, G., DeLuca, T. H. & Murphy, D. V. Biochar-mediated changes in soil quality and plant growth in a three-year field trial. Soil Biology and Biochemistry. 45, 113–124 (2012).

Acknowledgements

We thank Pror. Hailong Wang in Foshan University for his help in wiring of this paper. We gratefully acknowledge the National Key Research and Development Program of China (2017YFD0800903) and the Science & Technology Project of Guangdong, China (2015A020208012, 2019B030301007) for financial support of the research.

Author contributions

X.C. wrote the main manuscript text and prepared all figures and tables. H.Z.H. and G.K.C. oversaw the project and assisted with the writing of the manuscript. H.S.L. contributed to the conception of the study and assisted with Infrared Spectra analysis. All authors reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to H.-S.L.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2020