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Estimation of dietary ¹⁴C dose coefficient using ¹³C-labelled compound administration analysis

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Carbon-14 released from nuclear facilities has been assessed to contribute significantly to the radiation dose that people are exposed to through the food chain. However, the current dose coefficient for members of public, which is the ratio of the 50-year committed effective dose to ingested 1 Bq ¹⁴C, recommended by the International Commission on Radiological Protection (ICRP) is not based on experimental human metabolic data for ¹⁴C in nutrients and diet. Therefore, to validate the coefficient, we administered ¹³C-labelled nutrients consisting of four amino acids, three fatty acids, and one monosaccharide to volunteers as substitutes for ¹⁴C labelled nutrients and measured the ¹³C concentration in various excreta samples. Although metabolic models were constructed from the excretion data, a significant fraction of administered ¹³C was not recovered from some nutrients. The dose coefficients of ¹⁴C in uniformly labelled Japanese diet, which were estimated under several assumptions about the unrecoverable fraction, varied from (6.2 ± 0.9) × 10⁻¹¹ to (8.9 ± 4.4) × 10⁻¹⁰ Sv Bq⁻¹ and were approximately comparable to the current value of 5.8 × 10⁻¹⁰ Sv Bq⁻¹ recommended by the ICRP. Further studies are necessary to elucidate the metabolism of ¹⁴C in various nutrients in the unrecoverable fraction.

Carbon-14 is released from places such as nuclear fuel reprocessing facilities and nuclear power plants¹. It is a major radionuclide contributing to the radiation dose to people around the nuclear fuel reprocessing facilities¹⁻³. The dose coefficient of ¹⁴C, the ratio of 50-year committed effective dose to ingested 1 Bq ¹⁴C, is an important parameter for the dose evaluation. The International Commission on Radiological Protection (ICRP), which provides recommendations and guidance on radiological protection, has developed models describing the biokinetics of radionuclides for the calculation of doses from data of intakes or bioassay samples. For estimating the dose coefficient of ¹⁴C in diet of members of public, the ICRP has utilized a generic one-compartment model for organic carbon metabolism⁴, whereas a multi-compartment model was used in a recent publication for unspecified ¹⁴C compounds ingested by workers⁵. The one-compartment model for members of public assumes that ¹⁴C is uniformly distributed throughout all body organs and tissues following entry into the systemic circulation. The organic-carbon compartment of the model has a half-life of 40 days, which is derived from the mass balance of carbon in the reference man of 70 kg body weight with body carbon weight of 16 kg and carbon intake and excretion of 300 g carbon per day each⁶. The dose coefficient of organic ¹⁴C in the diet (5.8×10^{-10} Sv Bq⁻¹) was defined using that metabolic model⁷, which is not based on actual experimental data obtained from ¹⁴C in foods or component compounds of major nutrients. Therefore, experimental data of ¹⁴C in those compounds are needed to validate the generic one-compartment model of the ICRP or revise it in future publications for members of public following publications for workers⁸.

In rat tissue and organs, some studies have examined the biokinetics of ¹⁴C incorporated into foods or component molecule of major nutrient, e.g., single oral administration of $[^{14}C(U)]$ -glucose, $[^{14}C(U)]$ -glycine, $[^{14}C(U)]$ -leucine, $[^{14}C(U)]$ -lysine, $[1^{-14}C]$ -oleic acid, $[1^{-14}C]$ -palmitic acid, and $[2^{-14}C]$ -thymidine⁹; and continuous ingestion of ¹⁴C-wheat¹⁰, $[^{14}C(U)]$ -glycine, $[^{14}C(U)]$ -leucine, and ¹⁴C-rapesed¹¹. In sheep, Crout *et al.* reported the biokinetics of ¹⁴C after oral administration of ¹⁴C-glucose¹². However, most of the human data has been restricted to the ¹⁴C in drug molecules in biomedical studies¹³ and diagnostic use such as ¹⁴C-glycocholic acid and ¹⁴C-xylose breath tests¹⁴, and only few investigations have reported the biokinetics of ¹⁴C in component

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molecule of major nutrients. Malmendier *et al.*¹⁵ examined the metabolism of ¹⁴C in intravenously administered $[1^{-14}C]$ -glucose, $[1^{-14}C]$ -glycerol, and $[1^{-14}C]$ -palmitate for 24 h in breath in normal and hyperlipemic patients. Simmons *et al.*¹⁶ reported the ¹⁴C excretion in urine and breath until 3 and 7 days, respectively, after intravenous $[1^{-14}C]$ -alanine administration for human volunteers. However, both studies monitored the breath for a short time of up to 7 days after the administration. Berlin and Tolbert¹⁷ reported the breath excretion of ¹⁴C following intravenous injection of glycine-[2-¹⁴C] decreased in four successive phases with half-lives of 0.12, 0.97, 6.1, and 71.5 days. Stenström *et al.*¹⁸ orally administered ¹⁴C-triolein and found two phase excretions in the breath with half-lives of 2 and several hundred days. Gunnarsson *et al.*¹⁹ also reported that the excretion of ¹⁴C in breath after oral ¹⁴C-triolein administration decreased in three phases with half-lives of 0.04, 2, and 150 days. Although the data are limited, several studies have reported fractions of excreta with longer half-lives than that of the ICRP generic one-compartment model as described above.

Ingested carbon is excreted via several physiological routes including in the breath, urine, faeces, hair, nails, epithelial cells of the skin and mucous membrane, mucus, and other secretions. The ICRP has classified the routes into four excretion pathways, breath, urine, faeces, and others (other*, hereafter) and estimated the distribution ratio of carbon via the pathways in their reference man⁶. However, the previous studies on carbon excretion in humans described above examined only the respiration and urinary pathways. Regarding the other* pathway, Stenström *et al.* reported the specific activity of ¹⁴C in hair from personnel at a research laboratory and pointed out the possibility of using hair as a bioassay sample²⁰. To the best of our knowledge, no data has been reported for the human body on the metabolism of ¹⁴C incorporated into compounds covering a broad range of nutrients and food, considering every four excretion routes of the ICRP.

We previously reported the biokinetics of carbon in the human body estimated using data of the ${}^{13}C/{}^{12}C$ ratio in breath, urine, faeces, and hair, representing carbon excretion through the breath, urine, faeces, and other* pathways, respectively, after a single oral administration of ${}^{13}C$ -labelled glucose to three healthy male volunteers²¹. The ${}^{13}C/{}^{12}C$ ratio in breath, urine, and hair samples was measured for up to ~112 days after administration. The excretion of ${}^{13}C$ in the evaluated pathways decreased in two exponential phases with fast and slow rates, and the ${}^{13}C/{}^{12}C$ ratio of the other* pathway was higher than that of the breath and urine pathways toward the end of the experiment. Although this finding suggests that the carbon pool of the other* pathway had a longer half-life than that of the breath and urine pathways, we could not get the statistically significant ${}^{13}C$ half-life of the slow decreasing fraction in the other* pathway.

In this paper, we report the results of experiments administering ¹³C-labelled four amino acids and three fatty acids to volunteers. First, ¹³C-labelled leucine and palmitic acid were orally administered once to the volunteers and the excretion through the four routes of interest were measured for up to ~160 day after the administration. Then, the contribution of carbon pools of each route to the cumulative burden of ¹⁴C for 50 years was calculated assuming the same metabolism of ¹³C and ¹⁴C. Since the urine and faeces routes did not have significant contributions, we focused on only the breath and other* routes in experiments on the remaining nutrient components. In addition, once daily dosing administered four successive times, instead of the previously used single administration, was adopted for a more accurate measurement of ¹³C excretion in the final stage of the experiments. An experiment on ¹³C-labeled glucose administered four times was also conducted and, finally, the dose coefficient of ¹⁴C in the Japanese diet was estimated using integrated biokinetic models of each nutrient component.

Results and Discussion

Design of experiments. We selected the following amino acids as representative of proteins: leucine from branched chain amino acids, glutamic acid from major amino acids, glycine from collagen components, and phenylalanine from aromatic amino acids. Palmitic, oleic, and linoleic acids were selected as representative saturated, mono-unsaturated, and poly-unsaturated fatty acids, respectively while carbohydrates were represented by glucose. All carbon atoms of the compounds used were labelled with ¹³C (Cambridge Isotopes, Inc., Tewksbury). The compounds were administered to healthy volunteers and samples of various excreta were collected from them and analysed for ¹³C. All the ¹³C/¹²C ratio data presented in this report are shown as increments from the background values measured for the samples before administration, and normalized to the same dose per body weight (1 g ¹³C/70 kg body weight) prior to the following analysis. When the ¹³C/¹²C ratio under the detection limits, <95% confidence limit of the background, was observed, the data of the detection day and after it were not used for the subsequent numerical analysis. The detection limit mostly depended on the amount of ¹³C administered per body weight and individual difference.

Selection of major excretion routes of ¹³**C in amino acid and fatty acid.** We orally administered ¹³C-labelled leucine or palmitic acid to three male volunteers who were sedentary workers and then collected their breath, urine, faeces, and hair samples at different times depending on the compounds and excreta (Fig. 1). As shown in Figure 1, the ¹³C/¹²C ratio of the breath and hair samples increased after administration then decreased in two exponential phases. The peak time of breath samples from the leucine-administered group (0.11 ± 0.02 days, mean \pm standard deviation) was earlier than that of the palmitic acid-administered group (0.30 ± 0.03 days, P < 0.01), suggesting that the compounds had different absorption rates from the alimentary tract (Fig. S1 in the supplementary information). The peak time of the ¹³C/¹²C ratio in faeces samples after administration of labelled leucine and palmitic acid varied from 0.75 to 1.9 days and from 0.75 to 3.75 days among subjects, respectively. The excretion rate of administered ¹³C into the faeces is considered to depend on subject-specific values such as residence time of faeces in the colon, which was independent of the administered compounds.

For each compound, we developed a metabolic model comprising four carbon excretion pathways: breath, urine, faeces, and other* (Fig. 2). Since the ¹³C/¹²C ratios of each excreta sample showed a dual exponential decrease after peaking, two compartments representing fast and slow carbon metabolism were set for each of the four excretion pathways. However, every compartment in those models does not represent any tissue and organ.



Figure 1. Normalized ratio of ¹³C/¹²C in excreta (breath, urine, faeces, hair) after oral administration of ¹³C-leucine and ¹³C-palmitic acid. Circles, diamonds, and squares represent volunteer 1, 2, and 3, respectively. Lines were fitted using model developed in this study.



Figure 2. Structure of model developed to analyse ¹³C excreted through each excretion pathway after ingestion of ¹³C labelled compounds. R_r is ratio of total recoverable carbon through four excretion pathways to administered ¹³C. d_m , d_{mn} , distribution factors of ¹³C to each compartment. C_m , C_{mn} , ¹³C mass in the carbon pool (g). k_{mn} , excretion rate constant (day⁻¹).

In addition, one upstream compartment was used for each pathway to fit the prediction curve of the model to the increase phase observed in the breath and faeces samples immediately after administration (Fig. 1 and Fig. S1). The rate of ¹³C excretion for each pathway (E_m , g d⁻¹) is given by the following equations:

$$E_m = \sum_n k_{mn} \frac{d_m d_{mn} k_m}{k_m - k_{mn}} (e^{-k_{mn}t} - e^{-k_m t})$$
(1)

where, *m* is the excretion pathway symbol of *B*, *U*, *F*, and *O* corresponding to the excretion pathways of breath, urine, faeces, and other*, respectively; *n* is the symbol of *f* and *s* corresponding to the compartments with fast and slow metabolism, respectively; k_m and k_{mn} are the excretion rate constants from compartment C_m and C_{mn} ,

respectively; d_m is the distribution ratio of ¹³C into compartment C_m to the administered ¹³C; d_{mn} is the distribution ratio of ¹³C into compartment C_{mn} to ¹³C from compartment C_m ; R_r is the recovery ratio of administered ¹³C. The sum of distribution ratios of compartments with fast and slow metabolism was fixed to unity (i.e. $d_{Bf} + d_{Bs} =$ 1). Since the sum of the distribution ratios of each pathway ($d_B + d_U + d_F + d_O$) was not forced to unity, to prevent underestimation of these parameters, R_r was over unity for some cases.

The parameter values were estimated using the data of each volunteer using a least square method (Table 1). Although some parameters were not statistically significant (p > 0.05, most parameters for the urine, faeces, and other* pathway in the palmitic acid model, and in the faeces pathway in the leucine model), we included those parameters in the following analysis.

We calculated the 50-year cumulative body burdens of ¹³C from orally ingested 1 g ¹³C in leucine and palmitic acid using the above models. Then, the variation of the burden in response to a \pm 10% change of each parameter was calculated using the mixed data from all subjects for each compound group. The results are shown in Fig. 3 for the top 10 parameters causing larger variation and most were in the breath and other* pathways, whereas only two parameters causing smaller variations in the faeces pathway were included. All the statistically insignificant parameters in the regression analysis were not sensitive to the burden. The contribution of urine and faeces pathways to the burden from orally ingested 1 g of ¹³C in leucine were (0.37 ± 0.25)% and (0.7 ± 1.5)%, respectively, and those for palmitic acid were (0.038 ± 0.030)% and (0.034 ± 0.070)%, respectively. Therefore, we concluded that the urine and faeces pathway could be omitted from the following experiment on carbon dynamics in our study. For glucose, we previously reported the negligible contribution of the urine and faeces pathways to the cumulative burden²¹.

Metabolisms of ¹³C administered as various ¹³C -labelled compounds. We also administered the remaining target nutrient compounds,¹³C-labelled glutamic acid, glycine, phenylalanine, oleic acid, linoleic acid, and glucose to the volunteers who were healthy university students, three males and females each, and then we collected only breath and hair samples. All volunteers were administered each compound once daily for 4 successive days.

Figure 4 shows the ${}^{13}C/{}^{12}C$ ratio of the breath and hair samples after the first administration of each ${}^{13}C$ -labelled compound. The ${}^{13}C/{}^{12}C$ ratios of hair samples decreased faster in the groups of volunteers administered oleic acid, linoleic acid, and glucose than in the other groups and reached the detection limit before the end of experiments. Consequently, we did not analyse parameters relating the slow carbon compartment in the other* pathway for some subjects. In contrast, the ${}^{13}C/{}^{12}C$ ratios of the groups of volunteers administered glycine and phenylalanine were kept relatively high throughout the experimental period, and were higher than those in the breath samples of the same volunteers toward the end of the experiment. These results reflected the higher utilization of carbon in amino acids for protein synthesis such as in hair growth than carbon in fatty acids and monosaccharides. The results of leucine and palmitic acid-administered group also showed a similar tendency (Fig. 1). However, amino acids in samples from volunteers in the glutamic acid-administered group decreased faster than other amino acids did. This observation is consistent with the reported high use of dietary glutamic acid in the small intestine²².

The metabolic model was also developed for each compound. The values of parameters of the breath and other* pathways were estimated and are summarized in Table 2. The parameters for subjects with only fast C compartment in the other* pathway were included in the summarization. Since the difference of results between sexes was not clear, the average values for individual data of both sexes were also shown in Table 2 and discussed below. The estimated distribution ratio of ¹³C for the breath pathway (d_B) for leucine, glutamic acid, glycine, and phenylalanine were (68 ± 5)%, (84 ± 8)%, (80 ± 9)%, and (64 ± 7)%, respectively. Berlin and Tolbert¹⁷ reported an 86% recovery through the breath pathway after intravenous administration of glycine-[2-¹⁴C], which was consistent with the value of glycine in our experiment. Compared to glutamic acid and glycine, smaller proportions of leucine and phenylalanine were excreted through the breath pathway while a considerable portion was excreted through the other* pathway (d_O , (30 ± 12)% and (35 ± 8)%, respectively). Leucine stimulates protein synthesis via the mammalian target of rapamycin complex 1 (mTORC1) mediating signalling pathway²³ and, therefore, it might activate this pathway and consequently reduced its oxidation. However, such an effect has not been reported for phenylalanine. The distribution ratio of ¹³C of glucose in the breath pathway (d_B , 90 ± 15 %) was similar to the value reported in our previous paper ($86 \pm 10\%$)²¹.

The mean slow excretion rate constants (k_{Bs}) of the breath pathway varied from 0.061 to 0.21 day⁻¹ among the examined amino acids. These values were 5–20 times larger than the slow transfer coefficient of 0.0099 day⁻¹ from the systemic tissue compartment to the carbon dioxide compartment in the generic model for workers recommended by the ICRP-134⁵. However, the slow excretion constants (k_{Os}) of the other* pathway for the examined amino acids were from 0.0059 to 0.012 day⁻¹ and comparable to the value of the slow compartment in the generic model for workers. Our result provides evidence to supports the existence of a carbon pool with a slow metabolic rate.

Estimation of dose coefficient for members of public from ingested dietary ¹⁴**C**. As shown in Table 2B,C, the recovery ratios (R_r) of the administered ¹³C in oleic and linoleic acids were markedly small at 0.62 and 0.68, respectively, while most of the administered ¹³C in the other examined compounds including palmitic acid were recovered in our experiment (Tables 1 and 2). Takeda *et al.*⁹ reported that the ¹⁴C concentration in adipose tissue in rats 100 days after administration of ¹⁴C-labelled oleic acid was three times higher than that of palmitic acid. Although palmitic acid is a main constituent of adipose tissue fatty acids in humans²⁴, in this study, we assumed that the unrecoverable fraction of ¹³C in both unsaturated fatty acids and all nutrient compounds examined was stored in adipose tissue.

We further presumed that the carbon in adipose tissue had a mean residence time of 1.6 years, as estimated by Arner *et al.*²⁵ for adipocyte lipids by analysing ¹⁴C originating from nuclear weapon tests. Since the ICRP does not provide the tissue weighting factor for adipose tissue, we also set two assumptions for this, 0 and 0.12 used

	Volunteer											
	No. 1			No. 2			No. 3			All		
Parameter	Value	S.E.	P-value	Value	S.E.	P-value	Value	S.E.	P-value	Mean	S.D.	
(A) Estimated p	parameter	s of develop	ed model fo	r ¹³ C in leu	icine.							
d_B	0.62	0.02	< 0.01	0.62	0.02	< 0.01	0.76	0.01	< 0.01	0.68	0.05	
d_{Bf}	0.50	0.02	< 0.01	0.45	0.02	< 0.01	0.52	0.02	< 0.01	0.48	0.02	
d_U	0.012	0.002	< 0.01	0.011	0.003	< 0.01	0.011	0.003	< 0.01	0.011	0.001	
d _{Uf}	0.46	0.12	< 0.01	0.53	0.12	< 0.01	0.35	0.13	< 0.01	0.46	0.09	
d _o	0.24	0.01	< 0.01	0.36	0.02	< 0.01	0.22	0.01	< 0.01	0.30	0.12	
d _{Of}	0.40	0.02	< 0.01	0.34	0.03	< 0.01	0.38	0.03	< 0.01	0.40	0.03	
d_F	0.014	0.431	0.97	0.0083	0.0028	< 0.01	0.0047	0.0004	< 0.01	0.0092	0.0049	
d_{Ff}	0.28	4.0e+01	0.99	0.13	1.2e+02	>0.99	0.51	2.8e+00	0.86	0.30	0.17	
$k_B(\mathbf{d}^{-1})$	5.2	0.7	< 0.01	6.9	1.5	< 0.01	4.2	0.6	< 0.01	5.6	0.5	
$k_{Bf}(\mathbf{d}^{-1})$	20	5	< 0.01	16	5	< 0.01	17	4	< 0.01	18	3	
k_{Bs} (d ⁻¹)	0.076	0.009	< 0.01	0.064	0.007	< 0.01	0.043	0.004	< 0.01	0.061	0.015	
$k_{Uf}(\mathbf{d}^{-1})$	0.72	0.17	< 0.01	0.96	0.30	< 0.01	0.70	0.19	< 0.01	0.82	0.18	
k_{Us} (d ⁻¹)	0.024	0.012	0.05	0.042	0.016	< 0.01	0.021	0.014	0.15	0.030	0.010	
$k_{Of}(\mathbf{d}^{-1})$	0.29	0.03	< 0.01	0.35	0.05	< 0.01	0.28	0.04	< 0.01	0.32	0.07	
k_{Os} (d ⁻¹)	0.013	0.001	< 0.01	0.012	0.001	< 0.01	0.014	0.001	< 0.01	0.012	0.002	
$k_F(\mathbf{d}^{-1})$	0.43	1.1e+03	>0.99	2.9	1.1e+04	>0.99	1.7	9.3e+01	0.99	1.7	1.2	
$k_{Ff}(\mathbf{d}^{-1})$	0.43	1.1e+03	>0.99	2.8	1.1e+04	>0.99	1.5	0.0	< 0.01	1.7	1.2	
$k_{Fs}(d^{-1})$	0.0099	4.7e-01	0.98	0.10	0.02	< 0.01	0.17	9.6e+01	0.99	0.087	0.072	
R _r	0.89			1.2			0.91			1.0	0.2	
	Voluntee	ers										
Parameter	No. 4			No. 5			No. 6			All		
	Value	S.E.	P-value	Value	S.E.	P-value	Value	S.E.	P-value	Mean	S.D .	
(B) Estimated p	parameter	s of develop	ed model for	r ¹³ C in pal	mitic acid							
d_B	0.62	0.08	< 0.01	0.92	0.07	< 0.01	0.77	0.07	< 0.01	0.80	0.21	
d_{Bf}	0.73	0.30	0.02	0.53	0.15	< 0.01	0.46	0.06	< 0.01	0.59	0.14	
d_U	0.0062	0.0017	< 0.01	0.0088	0.0050	0.08	0.0080	0.0014	< 0.01	0.0077	0.0013	
d_{Uf}	0.52	0.85	0.05	0.55	0.22	0.01	0.40	0.12	< 0.01	0.49	0.08	
d_{O}	0.017	0.021	0.04	0.040	0.071	0.57	0.051	6.3	0.99	0.036	0.017	
d _{Of}	0.21	0.63	0.07	0.66	0.61	0.28	0.23	2.8e+01	0.99	0.37	0.26	
d_F	0.057	0.420				x					0.018	
d_{Ff}		0.450	0.09	0.030	0.012	0.01	0.024	0.006	< 0.01	0.037	0.010	
$k_{-}(d^{-1})$	0.94	0.430	0.09	0.030 0.75	0.012 1.3e+07	0.01 >0.99	0.024 0.84	0.006 1.2e+01	<0.01 0.94	0.037 0.84	0.10	
n _B (u)	0.94 2.8	0.430 0.49 5.6e+01	0.09 0.06 0.96	0.030 0.75 2.7	0.012 1.3e+07 2.0e+01	0.01 >0.99 0.89	0.024 0.84 4.2	0.006 1.2e+01 2.3	<0.01 0.94 0.07	0.037 0.84 3.1	0.10	
$k_{Bf}(\mathbf{d}^{-1})$	0.94 2.8 2.8	0.430 0.49 5.6e+01 5.6e+01	0.09 0.06 0.96 0.96	0.030 0.75 2.7 2.7	0.012 1.3e+07 2.0e+01 2.0e+01	0.01 >0.99 0.89 0.89	0.024 0.84 4.2 1.8	0.006 1.2e+01 2.3 0.8	<0.01 0.94 0.07 0.02	0.037 0.84 3.1 2.4	0.10 0.93 0.5	
$\frac{k_B(d^{-1})}{k_{Bs}(d^{-1})}$	0.94 2.8 2.8 0.15	0.49 5.6e+01 5.6e+01 0.04	0.09 0.06 0.96 0.96 <0.01	0.030 0.75 2.7 2.7 0.076	0.012 1.3e+07 2.0e+01 2.0e+01 0.024	0.01 >0.99 0.89 0.89 <0.01	0.024 0.84 4.2 1.8 0.081	0.006 1.2e+01 2.3 0.8 0.012	<0.01 0.94 0.07 0.02 <0.01	0.037 0.84 3.1 2.4 0.10	0.10 0.93 0.5 0.04	
$\frac{k_{B}(d^{-1})}{k_{Bf}(d^{-1})}$ $\frac{k_{Bf}(d^{-1})}{k_{Uf}(d^{-1})}$	0.94 2.8 2.8 0.15 2.8	0.49 5.6e+01 5.6e+01 0.04 5.7e+01	0.09 0.06 0.96 0.96 <0.01 0.96	0.030 0.75 2.7 2.7 0.076 1.8	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8	0.01 >0.99 0.89 0.89 <0.01 0.83	0.024 0.84 4.2 1.8 0.081 1.4	0.006 1.2e+01 2.3 0.8 0.012 0.8	<0.01 0.94 0.07 0.02 <0.01 0.09	0.037 0.84 3.1 2.4 0.10 2.1	0.10 0.93 0.5 0.04 0.7	
$ \begin{array}{c} k_{B}(\mathbf{d}^{-1}) \\ k_{Bs}(\mathbf{d}^{-1}) \\ k_{Uf}(\mathbf{d}^{-1}) \\ k_{Us}(\mathbf{d}^{-1}) \end{array} $	0.94 2.8 2.8 0.15 2.8 0.22	0.430 0.49 5.6e+01 5.6e+01 0.04 5.7e+01 0.09	0.09 0.06 0.96 0.96 <0.01 0.96 0.02 [‡]	0.030 0.75 2.7 2.7 0.076 1.8 0.12	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8 0.12	0.01 >0.99 0.89 <0.89 <0.01 0.83 0.29	0.024 0.84 4.2 1.8 0.081 1.4 0.14	0.006 1.2e+01 2.3 0.8 0.012 0.8 0.03	<0.01 0.94 0.07 0.02 <0.01 0.09 <0.01	0.037 0.84 3.1 2.4 0.10 2.1 0.16	0.10 0.93 0.5 0.04 0.7 0.05	
$ \begin{array}{c} k_{B}(\mathbf{d}^{-1}) \\ k_{Bf}(\mathbf{d}^{-1}) \\ k_{Uf}(\mathbf{d}^{-1}) \\ k_{Us}(\mathbf{d}^{-1}) \\ k_{Of}(\mathbf{d}^{-1}) \end{array} $	0.94 2.8 2.8 0.15 2.8 0.22 0.60	0.430 0.49 5.6e+01 5.6e+01 0.04 5.7e+01 0.09 1.3e+00	0.09 0.06 0.96 0.96 <0.01 0.96 0.02 [‡] 0.64	0.030 0.75 2.7 2.7 0.076 1.8 0.12 0.71	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8 0.12 0.65	0.01 >0.99 0.89 0.89 <0.01 0.83 0.29 0.27	0.024 0.84 4.2 1.8 0.081 1.4 0.14 0.23	0.006 1.2e+01 2.3 0.8 0.012 0.8 0.03 0.03 0.31	<0.01 0.94 0.07 0.02 <0.01 0.09 <0.01 0.45	0.037 0.84 3.1 2.4 0.10 2.1 0.16 0.52	0.10 0.93 0.5 0.04 0.7 0.05 0.26	
$\begin{array}{c} k_{B}(\mathbf{d}^{-1}) \\ k_{Bf}(\mathbf{d}^{-1}) \\ k_{Bs}(\mathbf{d}^{-1}) \\ k_{Uf}(\mathbf{d}^{-1}) \\ k_{Us}(\mathbf{d}^{-1}) \\ k_{Of}(\mathbf{d}^{-1}) \\ k_{Os}(\mathbf{d}^{-1}) \end{array}$	0.94 2.8 2.8 0.15 2.8 0.22 0.60 0.051	0.430 0.49 5.6e+01 5.6e+01 0.04 5.7e+01 0.09 1.3e+00 0.040	$\begin{array}{c} 0.09 \\ \hline 0.06 \\ \hline 0.96 \\ \hline 0.96 \\ \hline 0.96 \\ \hline 0.02^{\ddagger} \\ \hline 0.64 \\ \hline 0.21 \\ \hline \end{array}$	0.030 0.75 2.7 2.7 0.076 1.8 0.12 0.71 0.045	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8 0.12 0.65 1.1e-01	0.01 >0.99 0.89 <0.01 0.83 0.29 0.27 0.68	0.024 0.84 4.2 1.8 0.081 1.4 0.14 0.23 0.0044	0.006 1.2e+01 2.3 0.8 0.012 0.8 0.03 0.31 7.5e-01	<0.01 0.94 0.07 0.02 <0.01 0.09 <0.01 0.45 0.99	0.037 0.84 3.1 2.4 0.10 2.1 0.16 0.52 0.033	0.10 0.93 0.5 0.04 0.7 0.05 0.26 0.025	
$\begin{array}{c} k_{B}(\mathbf{d}^{-1}) \\ k_{Bf}(\mathbf{d}^{-1}) \\ k_{Bs}(\mathbf{d}^{-1}) \\ k_{Uf}(\mathbf{d}^{-1}) \\ k_{Us}(\mathbf{d}^{-1}) \\ k_{Of}(\mathbf{d}^{-1}) \\ k_{Os}(\mathbf{d}^{-1}) \\ k_{F}(\mathbf{d}^{-1}) \end{array}$	0.94 2.8 2.8 0.15 2.8 0.22 0.60 0.051 2.7	0.430 0.49 5.6e+01 5.6e+01 0.04 5.7e+01 0.09 1.3e+00 0.040 2.0	$\begin{array}{c} 0.09 \\ 0.06 \\ 0.96 \\ 0.96 \\ < 0.01 \\ 0.96 \\ 0.02^{\sharp} \\ 0.64 \\ 0.21 \\ 0.18 \end{array}$	0.030 0.75 2.7 2.7 0.076 1.8 0.12 0.71 0.045 0.45	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8 0.12 0.65 1.1e-01 1.3e+03	0.01 >0.99 0.89 <0.01 0.83 0.29 0.27 0.68 >0.99	0.024 0.84 4.2 1.8 0.081 1.4 0.14 0.23 0.0044 1.9	0.006 1.2e+01 2.3 0.8 0.012 0.8 0.03 0.31 7.5e-01 6.5e+02	<0.01 0.94 0.07 0.02 <0.01 0.09 <0.01 0.45 0.99 >0.99	0.037 0.84 3.1 2.4 0.10 2.1 0.16 0.52 0.033 1.7	0.10 0.93 0.5 0.04 0.7 0.05 0.26 0.025 1.1	
$\begin{array}{c} k_B(\mathbf{d}^{-1}) \\ k_{Bf}(\mathbf{d}^{-1}) \\ k_{Bs}(\mathbf{d}^{-1}) \\ k_{Uf}(\mathbf{d}^{-1}) \\ k_{Us}(\mathbf{d}^{-1}) \\ k_{Of}(\mathbf{d}^{-1}) \\ k_{Cs}(\mathbf{d}^{-1}) \\ k_F(\mathbf{d}^{-1}) \\ k_{Ff}(\mathbf{d}^{-1}) \end{array}$	0.94 2.8 2.8 0.15 2.8 0.22 0.60 0.051 2.7 8.2	0.430 0.49 5.6e+01 5.6e+01 0.04 5.7e+01 0.09 1.3e+00 0.040 2.0 1.7e+02	$\begin{array}{c} 0.09 \\ 0.06 \\ 0.96 \\ 0.96 \\ < 0.01 \\ 0.96 \\ 0.02^{\$} \\ 0.64 \\ 0.21 \\ 0.18 \\ 0.96 \\ \end{array}$	0.030 0.75 2.7 2.7 0.076 1.8 0.12 0.71 0.045 0.45	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8 0.12 0.65 1.1e-01 1.3e+03 1.5e+03	0.01 >0.99 0.89 <0.01 0.83 0.29 0.27 0.68 >0.99 >0.99	0.024 0.84 4.2 1.8 0.081 1.4 0.14 0.23 0.0044 1.9 1.9	0.006 1.2e+01 2.3 0.8 0.012 0.8 0.03 0.31 7.5e-01 6.5e+02 6.5e+02	<0.01 0.94 0.07 0.02 <0.01 0.09 <0.01 0.45 0.99 >0.99 >0.99	0.037 0.84 3.1 2.4 0.10 2.1 0.16 0.52 0.033 1.7 3.5	0.10 0.93 0.5 0.04 0.7 0.05 0.26 0.025 1.1 4.1	
$\begin{array}{c} k_B(\mathbf{d}^{-1}) \\ k_{Bf}(\mathbf{d}^{-1}) \\ k_{Bs}(\mathbf{d}^{-1}) \\ k_{Uf}(\mathbf{d}^{-1}) \\ k_{Us}(\mathbf{d}^{-1}) \\ k_{Of}(\mathbf{d}^{-1}) \\ k_{Os}(\mathbf{d}^{-1}) \\ k_F(\mathbf{d}^{-1}) \\ k_{Ff}(\mathbf{d}^{-1}) \\ k_{Fs}(\mathbf{d}^{-1}) \end{array}$	0.94 2.8 2.8 0.15 2.8 0.15 2.8 0.22 0.60 0.051 2.7 8.2 0.15	0.430 0.49 5.6e+01 5.6e+01 0.04 5.7e+01 0.09 1.3e+00 0.040 2.0 1.7e+02 0.04	$\begin{array}{c} 0.09 \\ \hline 0.06 \\ \hline 0.96 \\ \hline 0.96 \\ \hline 0.96 \\ \hline 0.02^{\ddagger} \\ \hline 0.64 \\ \hline 0.21 \\ \hline 0.18 \\ \hline 0.96 \\ \hline < 0.01 \\ \end{array}$	0.030 0.75 2.7 2.7 0.076 1.8 0.12 0.71 0.045 0.45 0.45	0.012 1.3e+07 2.0e+01 2.0e+01 0.024 8.8 0.12 0.65 1.1e-01 1.3e+03 1.5e+03 4.1e+03	0.01 >0.99 0.89 <0.01 0.83 0.29 0.27 0.68 >0.99 >0.99 >0.99	0.024 0.84 4.2 1.8 0.081 1.4 0.23 0.0044 1.9 1.9 1.9	0.006 1.2e+01 2.3 0.8 0.012 0.8 0.03 0.31 7.5e-01 6.5e+02 6.5e+02 0.1	<0.01 0.94 0.07 0.02 <0.01 0.09 <0.01 0.45 0.99 >0.99 >0.99 <0.01	0.037 0.84 3.1 2.4 0.10 2.1 0.16 0.52 0.033 1.7 3.5 0.31	0.10 0.93 0.5 0.04 0.7 0.05 0.26 0.025 1.1 4.1 0.15	

Table 1. Parameters are explained in Fig. 2. S.E., Standard error. S.D., Standard deviation. *Rr*, recovery ratio (i.e. dB + dU + dF + dO).

for the remaining organs in the ICRP-103 guidelines²⁶. The dose coefficients obtained with each assumption are called non conservative estimation (ER) and conservative estimation (EC) hereafter. The recovered fraction was assumed to be distributed uniformly in the whole-body carbon with a tissue weighting factor of unity. In considering the possible existence of additional carbon compartment, which was not detected in our study but have a longer half-life than that in the present slow carbon compartments, we adopted the EC as a moderate estimation. In addition to these estimations, we calculated the most conservative estimation (EX) adopting the tissue weighting factor of unity for unrecovered carbon. We solely adopted the value for the maximal estimation.

Figure 5A shows the 50-year committed effective dose from 1 Bq of the ¹⁴C in labelled compounds after single ingestion calculated using each model developed. The ER values of the amino acids were generally higher than those of the fatty acids, whereas the EC and EX values were the opposite. Glucose showed similar ER values



Figure 3. Sensitivity analysis of model parameters. A, leucine and B, palmitic acid. Changes in 50-year cumulative burden versus $\pm 10\%$ changes in parameter values were calculated. The top 10 parameters are shown in decreasing order. Solid and open bars, -10% and +10% change in parameters, respectively.

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to those of the unsaturated fatty acids, whereas the EC and EX values are similar to those of amino acids. The assumption about the unrecoverable fraction played an extremely important role in our estimation of the dose because of the assumed long half-life of unrecovered carbon. The relative contribution of each pathway including the unrecoverable fraction to the EC is shown in Fig. 5B. It is noteworthy that parameters of the urine and faeces pathway in the first experiment were used for the other compounds (details are shown below), and the contribution of these pathways was very minor (<0.8%). As shown in Fig. 5B, the unrecoverable fraction contributed significantly to the committed dose when we adopted a tissue weighting factor of 0.12 to adipose tissue.

We estimated the dose coefficient for members of public based on ingestion of a Japanese diet²⁷, which was uniformly labelled by ¹⁴C and had the nutritional composition shown in Table 3. The nutrition in the diet was classified into groups which was represented by each single compound examined in our study. The proportion of carbon ingested as the nutrition groups to total carbon ingestion rate was obtained by using literature data^{28,29} (details are shown below). The total daily ingestion of 300 g carbon per day of ICRP reference man was divided by the proportion of carbon ingestion rate each and inputted to each single compound model. The dose coefficient was obtained by summing up the 50-year committed effective dose from each representative compound comprising the diet with a total ingestion of 1 Bq ¹⁴C.

The ER of the Japanese diet was estimated to be $(6.2 \pm 0.9) \times 10^{-11}$ Sv Bq⁻¹ (Fig. 5C), which was nine and two times smaller than the current dose coefficient of 5.8×10^{-10} Sv Bq⁻¹ for members of public as recommended by the ICRP-72⁷ and 1.6×10^{-10} Sv Bq⁻¹ for workers adopting the generic multi-compartment model by ICRP-134⁵, respectively. The EC and EX were $(5.6 \pm 2.6) \times 10^{-10}$ and $(8.9 \pm 4.4) \times 10^{-10}$ Sv Bq⁻¹, respectively (Fig. 5C). These conservative values were not significantly different from the current dose coefficient for members of public⁷.

As described before, there is no data about metabolism of carbon in a broad range of nutrients administered orally, this work firstly provides the experimental base of dose coefficient for members of public. Since the unrecoverable fraction was very important for the estimation of the dose coefficient in our research, further studies are necessary to clarify the metabolism of ¹⁴C in various nutrients.

Materials and Methods

Administration of ¹³C-labelled compounds. In the first experiment to select the major excretion routes for carbon in amino and fatty acids, an aliquot of uniformly ¹³C-labelled leucine and palmitic acid (Cambridge Isotopes, Inc., Tewksbury) was administered to healthy adult Japanese male sedentary workers aged 41 \pm 9 and 40 \pm 5 years old (n =3 each), respectively. The dose was 1.0 g¹³C per person. We followed the administration procedure described in a previous paper²¹. Briefly, the ¹³C-labelled compounds were orally ingested by each volunteer at 12:15 immediately before eating lunch on experimental day 0. An additional 0.1 g of ¹³C in the compounds was similarly administered on day 112 after the initial administration to mark that day in the hair samples (see below). To avoid background fluctuations in the ¹³C/¹²C ratio by over-eating of ¹³C-rich foods, nutritionally balanced foods were provided to the volunteers throughout the experimental period from day -7 to 112, although they were able to eat other foods under certain circumstances.

In the second experiment using various ¹³C-labelled compounds, uniformly ¹³C-labelled glutamic acid, glycine, phenylalanine, oleic acid, linoleic acid, and glucose (Cambridge Isotopes, Inc., Tewksbury) were administered to healthy adult Japanese male and female volunteers (n = 3, each) aged 25 ± 7 years once a day at 12:05



Figure 4. Ratio of ${}^{13}C/{}^{12}C$ in excreta (breath and hair) after oral administration of ${}^{13}C$ -labelled compounds.

on 4 successive days. The doses of those compounds were 8.9, 7.1, 11.9, 11.0, 16.4, and $35.6 \text{ mg} {}^{13}\text{C} \text{ kg}^{-1} \text{ day}^{-1}$, respectively. Additional doses of ${}^{13}\text{C}$ -labelled compounds were administered on day 160 for marking the ${}^{13}\text{C}/{}^{12}\text{C}$ ratio of hair samples and the volunteers freely ate meals without calorific restrictions.

Sample collection and determination of ${}^{13}C/{}^{12}C$ ratio. In the first experiment, breath, urine, faeces, and hair samples were periodically collected and the ${}^{13}C/{}^{12}C$ ratio was analysed following the procedure described in a previous paper²¹. Briefly, breath and urine samples were collected daily when the volunteers rose in the morning from day -7 to 112. Faecal samples were collected once before day 0 and at every defecation from day 0 to 14. On day 0, additional breath and urine samples were collected frequently after administration. Five hair samples from each subject were collected on day 119 and used as representative samples of the other* pathway. Then, 10 cm of each hair starting from the root was cut into 1-mm sections and the ${}^{13}C/{}^{12}C$ ratio determination of each showed two peaks in the ${}^{13}C/{}^{12}C$ ratio corresponding to the first and second administration of ${}^{13}C$ -compounds. According to the section number of the two peaks, the experimental date for each section between the two peaks was determined under an assumption of uniform hair growth. In the second experiment, breath samples were collected until day 160 and hair samples were obtained by brushing on day 167.

The ${}^{13}C/{}^{12}C$ ratio was measured using a mass spectrometer (Delta V Advantage; Thermo Electron, Waltham, MA, USA) coupled with an elemental analyser (FlashEA 1112NC; Thermo Electron) and a gas chromatograph (MAT GC; Thermo Electron) for combusting organic samples and purifying CO₂. Breath samples were collected by the volunteers themselves, and some samples showed clearly inconceivable results such as similar ${}^{13}C/{}^{12}C$ ratios over considerable periods of breath sample collection, which may have been caused by collecting the samples on a single day instead of once daily sampling on sequential days. The whole data of such individuals were discarded, which decreased the number of samples from three in Table 2. Some volunteers also dropped out of the sampling procedure for various reasons.

Constructing multi-compartment metabolic model by regression analysis. The ¹³C distribution ratios and excretion rate constants of all compartments in the four excretion pathways for leucine and palmitic acid were determined using the least squares fitting method with the statistical analysis software R version 2.15.2., while only parameters for the breath and other* pathway were determined for the other six examined

	Glutamic acid									Glycine								
	Male			Female			All		Male			Female			All			
Parameter	Mean	S.D.	n	Mean	S.D.	n	Mean	S.D.	n	Mean	S.D.	n	Mean	S.D.	n	Mean	S.D.	n
(A) Estimated pa	rameters o	of develope	d mo	del for ¹³ C	in glutamic	acid	and glycin	ie.									_	
d_B	0.83	0.11	3	0.85	0.05	3	0.84	0.08	6	0.74	0.01	2	0.84	0.10	3	0.80	0.09	5
d _{Bf}	0.75	0.05	3	0.72	0.01	3	0.73	0.03	6	0.37	0.05	2	0.36	0.03	3	0.36	0.03	5
d _o	0.048	0.009	3	0.029	0.005	3	0.038	0.012	6	0.18	0.02	3	0.18	0.02	3	0.18	0.02	6
d _{Of}	0.41	0.07	3	0.59	0.16	3	0.50	0.15	6	0.47	0.02	3	0.52	0.01	3	0.50	0.03	6
$k_B (\mathrm{d}^{-1})$	42	16	3	56	32	3	49	24	6	37	14	2	25	9	3	30	12	5
$k_{Bf}(\mathbf{d}^{-1})$	11	0	3	15	5	3	13	4	6	9.7	3.5	2	11	4	3	11	3	5
$k_{Bs} (d^{-1})$	0.27	0.05	3	0.15	0.06	3	0.21	0.08	6	0.16	0.00	2	0.18	0.02	3	0.17	0.02	5
$k_{Of}(\mathbf{d}^{-1})$	0.21	0.01	3	0.17	0.02	3	0.19	0.03	6	0.11	0.02	3	0.098	0.003	3	0.10	0.01	6
$k_{Os}(d^{-1})$	0.0051	0.0014	3	0.016	0.011	3	0.010	0.009	6	0.0080	0.0006	3	0.0091	0.0030	3	0.0086	0.0021	6
R _r	0.90	0.12	3	0.91	0.05	3	0.90	0.08	6	0.93	0.02	3	1.0	0.1	3	1.0	0.1	6
Parameter	Phenylal	Phenylalanine								Oleic aci	d							
	Male		Female			All				Male		Female			All			
	Mean	S.D.	n	Mean	S.D.	n	Mean	S.D .	n	Mean	S.D .	n	Mean	S.D .	n	Mean	S.D.	n
(B) Estimated pa	rameters o	of develope	d mo	del for ¹³ C	in phenylal	anine	e and oleic	acid.										
d_B	0.59	0.06	3	0.69	0.01	3	0.64	0.07	6	0.62	0.08	3	0.55	0.07	3	0.59	0.08	6
d_{Bf}	0.37	0.04	3	0.41	0.04	3	0.39	0.05	6	0.53	0.24	3	0.66	0.05	3	0.59	0.17	6
d _o	0.26	—	1	0.38	0.06	3	0.35	0.08	4	0.0067	—	1	0.012	0.006	2	0.010	0.005	3
d _{Of}	0.35	—	1	0.30	0.03	3	0.31	0.04	4	-	—	0	0.37	_	1	0.37	-	1
$k_B(\mathbf{d}^{-1})$	52	22	3	49	18	3	51	18	6	7.0	2.9	3	15	11	3	11	8	6
$k_{Bf}(d^{-1})$	9.1	0.24	3	8.3	4.2	3	8.7	2.7	6	4.6	1.8	3	1.9	0.5	3	3.2	1.9	6
$k_{Bs} (d^{-1})$	0.070	0.012	3	0.073	0.043	3	0.071	0.028	6	0.12	0.04	3	0.097	0.011	3	0.11	0.03	6
$k_{Of}(d^{-1})$	0.14	-	1	0.13	0.03	3	0.13	0.02	4	0.095	—	1	0.17	0.07	2	0.14	0.06	3
$k_{Os}(\mathbf{d}^{-1})$	0.0061	—	1	0.0058	0.0013	3	0.0059	0.0011	4	-	—	0	0.0074	—	1	0.0074	-	1
R _r	0.90	—	1	1.1	0.1	3	1.0	0.1	4	0.61	—	1	0.62	0.10	2	0.62	0.07	3
	Linoleic acid								Glucose									
Parameter	Male		Female			All			Male			Female			All			
	Mean	S.D .	n	Mean	S.D.	n	Mean	S.D .	n	Mean	S.D.	n	Mean	S.D .	n	Mean	S.D .	n
(C) Estimated pa	rameters o	of develope	d mo	del for ¹³ C	in linoleic a	icid a	nd glucose	e										
d_B	0.61	0.15	3	0.63	0.18	2	0.62	0.13	5	0.81	0.15	3	0.99	0.08	3	0.90	0.15	6
d_{Bf}	0.48	0.17	3	0.43	0.23	2	0.46	0.17	5	0.71	0.14	3	0.85	0.00	3	0.78	0.11	6
d _o	0.012	0.006	3	0.013	0.005	3	0.013	0.005	6	0.014	0.001	3	0.016	0.001	3	0.015	0.004	6
d _{Of}	0.51	—	1	0.30	—	1	0.41	0.15	2	0.59	0.15	3	0.68	0.06	3	0.63	0.11	6
$k_{B}(d^{-1})$	21	10	3	16	4	2	19	8	5	43	14	3	69	7	3	56	17	6
$k_{Bf}(d^{-1})$	3.4	1.9	3	6.7	8.2	2	4.7	4.7	5	5.7	3.8	3	2.6	0.5	3	4.1	3.0	6
$k_{Bs} (d^{-1})$	0.095	0.039	3	0.085	0.034	2	0.091	0.033	5	0.094	0.055	3	0.064	0.014	3	0.079	0.049	6
$k_{Of}(d^{-1})$	0.097	0.023	3	0.14	0.05	3	0.12	0.04	6	0.14	0.01	3	0.30	0.10	3	0.22	0.11	6
$k_{Os}(d^{-1})$	0.0079	_	1	0.0056	_	1	0.0068	0.0016	2	0.0056	0.0037	3	0.022	0.014	3	0.014	0.013	6
R _r	0.67	0.15	2	0.69	0.17	3	0.68	0.14	5	0.83	0.15	3	1.0	0.1	3	0.93	0.15	6

Table 2. Parameters are explained in Fig. 2. S.D., standard deviation.

compounds. The detailed procedures used were described in a previous published paper²¹. When urine and hair samples had no increase phase, the values of k_U and k_O were fixed to those of k_B obtained from the same volunteers in the regression analysis.

Estimation of dose coefficient for members of public from dietary ¹⁴**C**. The 50-year cumulative body burden of ¹⁴C after a single ingestion of 1 Bq of ¹⁴C in each compound (B_C Bq d) was calculated using the metabolic model and its parameters described above, under the assumption of the same metabolic behaviour of ¹³C and ¹⁴C. The parameters of the urine and faeces pathway for leucine and palmitic acid were used for the other amino acids and fatty acids. For glucose, the parameters of the urine and faeces pathways reported in our previous paper²¹ were adopted. The total cumulative burden was derived as a sum of burdens from the recoverable and unrecoverable fractions with designated proportions R_r and $1-R_r$, respectively. For the recoverable fraction, the burden was calculated using following equation:



Figure 5. Dose estimation for ingested ¹⁴C. (**A**) Committed effective dose from 1 Bq of ¹⁴C in each compound. Error bars show standard deviation among subjects (n = 3-6). (**B**) contribution of ¹³C via each excretion pathway and unrecoverable carbon to the conservative committed dose (EC). (**C**) Dose coefficient for members of public from dietary ¹⁴C according to nutrition survey in Japan. Error bars show standard deviation.

	Nutrition											
					Lipids			Sum				
	Proteins				SFA MU		PU		Carbohydrates			
Carbon ingestion* (g-dry d ⁻¹)	71.8				17.6	22.6	20.8	254.0	386.8			
	BCAA	Glycine	AAA	Others								
Proportion of amino acid group in protein ingestion rate** (%)	18.2	4.3	12.7	64.8								
Carbon concentration [†] (%)	54	32	60	53	75	77	77	44				
Carbon ingestion rate (g d ⁻¹)	7.1	1.0	5.5	24.7	13.2	17.4	16.0	111.8	196.6			
Proportion to total carbon ingestion rate	0.036	0.0050	0.028	0.13	0.067	0.089	0.082	0.57	1.0			
Represented model	Leucine	Glycine	Phenyl-alanine	Glutamic acid	Palmitic acid	Oleic acid	Linoleic acid	Glucose				

Table 3. Nutritional composition in Japanese diet and corresponding models used to calculate dose coefficient. *, National Health and Nutritional Survey by the Ministry of Health, Labour and Welfare, Japan²⁷; **, proportion of each amino acid group intake to protein intake²⁸; ^{†29}. SFA, saturated fatty acid; MU, mono-unsaturated fatty acid; PU, poly-unsaturated fatty acid and others. BCAA, branched chain amino acid (leucine, isoleucine, and valine); AAA, aromatic amino acid (phenylalanine, histidine, and tryptophan).

$$B_C \approx \sum_m \frac{d_m}{k_m} + \sum_m \sum_n \frac{d_m d_{mn}}{k_{mn}}$$
(2)

where, every d_m was divided by R_r when R_r was larger than unity.

The 50-year committed effective dose (Sv) from ingestion of 1 Bq ¹⁴C was calculated from the 50-year cumulative burden by using the ratio of absorbed dose to ¹⁴C radioactivity, radiation weighting factor of beta-ray from ¹⁴C (1.0) and tissue weighting factors. As previously described, for the unrecoverable fraction, we used a mean residence time of 1.6 years (excretion rate constant of 0.0017 d⁻¹)²⁵ and tissue weighting factors of 0, 0.12, and unity for the estimation of ER, EC, and EX, respectively.

The Japanese typical nutrition intake rates were cited from the annual nationwide National Health and Nutrition Survey conducted by the Ministry of Health, Labour and Welfare, Japan²⁷. In addition, we used the proportion of each amino acid as reported by Iwaya²⁸ to obtain the ingestion rate of each amino acid group including branched chain amino acid (BCAA), major amino acid, glycine, and aromatic amino acid (AAA) groups (Table 3). We assumed that the metabolism of each amino acid group was the same as that of the represented

compound examined in our study as shown in Table 3. The lipids and carbohydrates were also represented by each single compound. Then, the ingestion rate of carbon in each nutrition group was calculated according to the weight percentage of carbon in the representative compound (Table 3)²⁹, followed by obtaining the proportion of carbon ingestion rate of each group to the total carbon ingestion rate. Then, the input to each compound model was calculated from the total carbon ingestion rate of 300 g carbon per day of ICRP reference man⁶ and the proportion of carbon ingestion rate. Finally, the dose coefficient for members of public from dietary ¹⁴C was calculated by summing up the committed effective doses of ¹⁴C and weighting proportion of ¹⁴C in each compound up to 1 Bq ¹⁴C in the diet.

Ethical considerations. The study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in the prior approval of the Institute for Environmental Sciences Review Board for Human Subject Experiments. Written informed consent was obtained from each volunteer prior to participation.

Data availability

The datasets generated and analysed during the current study are available from the corresponding author on a reasonable request.

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Author contributions

T.M. conceived the idea and designed the study; H.T. participated in the study design; T.M., K.M. and T.Y. acquired the data; T.M. and T.T. analysed the data; H.H., Ya. T. and Yu. T. participated in the study coordination; T.M. and S.H. interpreted the data and wrote the text; H.T., T.T., H.H., Ya. T. and Yu. T., commented on the manuscript writing; All authors critically reviewed the data and the manuscript; SH supervised the study.

Competing interests

The authors declare no competing interests.

Additional information

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