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OPEN Plasma FABP4 is associated with liver disease recovery during treatment-induced clearance of chronic HCV infection

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Direct-acting antivirals (DAAs) have dramatically improved the management of chronic hepatitis C (CHC). In this study, we investigated the effects of hepatitis C virus clearance on markers of systemic inflammation measured in plasma samples from CHC patients before, during and after DAA therapy. We identified a plasma soluble protein profile associated with CHC. Successful DAA therapy rapidly normalised the plasma inflammatory milieu, with the notable exception of soluble (s)CD163, a marker of macrophage activation, which remained elevated after viral clearance and segregated patients with high and low levels of cirrhosis. Patients who received DAA in combination with Ribavirin maintained elevated levels of CXCL10, consistent with an immune-stimulatory role of Ribavirin. As anticipated, DAA-treated patients experienced durable improvement in liver fibrosis measurements. Interestingly, pre-treatment levels of fatty acid-binding protein 4 (FABP4) were inversely associated with reduction of APRI and FIB-4 scores during treatment. Together, these results support the notion of a rapid restoration of many aspects of the inflammatory state in CHC patients in response to DAA therapy. Furthermore, the associations with sCD163 and FABP4 warrant further investigation into the role of macrophages in residual liver disease and fibrosis resolution after viral clearance.

Over 70 million individuals are infected with Hepatitis C virus (HCV) worldwide¹. During the last decade, direct acting antivirals (DAA) have become available for treatment of chronic hepatitis C (CHC) and have dramatically improved clinical outcomes. DAA combinations have proven to be highly efficient, allowing for sustained virologic response (SVR) rates approaching 100%², with a standard treatment course of 12 weeks that can even be shortened to 8 weeks in some instances³. These treatments have sparked hopes of eradicating HCV and the WHO has set the goal of a 90% reduction in new cases of chronic infection by 2030⁴. Nevertheless, even after successful elimination of HCV, risks of residual liver disease and development of hepatocellular carcinoma (HCC) remain⁵, It is therefore important to understand the dynamics of inflammation and fibrosis resolution during and after successful DAA treatment.

Although HCV infection localises to the liver, chronic hepatic inflammation causes systemic changes in blood cytokine and chemokine levels^{6,7}. Numerous studies have investigated whether plasma levels of such soluble markers could predict clinical outcome of interferon (IFN) α -based therapy^{8,9}. In the present study, we aimed to characterise how viral clearance affects plasmatic levels of cytokines and inflammatory markers in a cohort of CHC patients successfully treated with DAA. We hypothesised that plasma levels of some of these proteins may be associated with the evolution of liver disease and we therefore investigated associations with clinical measures of liver status such as liver stiffness measurement (LSM), Aspartate transaminase-to-platelet ratio Index (APRI)¹⁰ and Fibrosis index based on 4 factors (FIB-4)¹¹ scores.

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	HD	AC	DAA
	n=20	n=12	n=28
M/F	50%/50%	92%/8%	50%/50%
Age (years)	57.00 [7.75]	60.00 [12.25]	60.50 [11.25]
$BMI(kg/m^2)$	ND	30.77 [4.38]	25.94 [5.21]
Diabetes	ND	4 (33%)	4 (14%)
Alcoholic	ND	12 (100%)	11 (39%)
Cirrhosis	ND	12 (100%)	25 (89%)
Child Pugh A/B/C		6/5/1	18/7/0
Viral load (10 ⁶ /mL)		-	1.06 [3.18]
Genotype 1/2/3/4		-	19/2/6/1
Prior treatment		-	18 (64%)
Albumin (g/L)	ND	33.00 [8.75]	34.50 [8.25]
Bilirubin (µmol/L)	ND	18.50 [24.50]	14.00 [13.25]
ALT (µkat/L)	ND	0.54 [0.29]	1.27 [0.97]
AST (µkat/L)	ND	0.82 [0.36]	1.37 [1.04]
PT (INR)	ND	1.20 [0.23]	1.15 [0.22]
Thrombocytes (10 ⁹ /L)	ND	140.00 [70.00]	127.50 [83.50]
LSM (kPa)	ND	24.35 [19.25]	21.70 [16.25]
APRI score	ND	0.90 [0.95]	1.92 [1.99]
FIB-4 index	ND	4.43 [3.25]	6.22 [5.35]

Table 1. Cohort baseline characteristics. Values are count (and percentage) for categorical variables and median[IQR] for continuous variables. ND: Not determined.

Results

Plasma cytokine alterations associated with CHC and evolution during DAA therapy. To investigate the impact of HCV clearance on the immune system, 28 CHC patients were sampled before, during and after successful DAA treatment. In addition, blood samples from 20 healthy donors (HD), and 12 patients suffering from alcohol-induced cirrhosis (AC) were included as non-HCV infected comparison groups (Table 1). Plasma concentrations of 25 soluble factors, all known to be related to the immune response or inflammation, were measured in each plasma sample (Supplementary Dataset). Out of the 25 soluble factors measured, four were significantly different (p < 0.05) between HD and CHC patients who were going to initiate IFN-free DAA therapy: CCL5, CXCL10, sCD14 and sCD163 (Fig. 1a). CCL5 was detected at lower levels (p = 0.0095), whereas CXCL10, sCD14 and sCD163 were higher in the CHC patients compared to HD (p < 0.0001, p = 0.0095 and p < 0.0001, respectively). Interestingly, there were no significant differences in CCL5 and CXCL10 levels between CHC patients and AC patients (p = 0.4764 and 0.0964), but sCD14 and sCD163 were significantly higher in CHC patients compared to AC (p = 0.0346 and 0.0489, respectively; Fig. 1a).

Upon start of DAA therapy, differences with HD were quickly reduced (Fig. 1b), in contrast to the pattern observed in a previously treated cohort of CHC patients who received pegylated-IFN therapy and successfully achieved SVR (Supplementary Fig. S1). At follow-up (FU) after completion of DAA therapy, both CCL5 and sCD14 were restored to levels comparable to HD, whereas CXCL10 and sCD163, although significantly decreased compared to BL, remained significantly elevated in comparison to HD (p = 0.0138 and p < 0.0001, respectively; Fig. 1c).

Effect of Ribavirin on plasma cytokine profiles. In our cohort, half of the patients who underwent DAA therapy received RBV as part of their treatment regimen (Supplementary Fig. 2). We therefore stratified CHC patients according to the use of RBV to assess possible effects on plasma cytokine levels (Fig. 2). There were no other significant differences before treatment between these two subgroups regarding age, BMI, liver health indicators (ALT, AST, albumin, bilirubin, PT-INR, thrombocytes, LSM) or any of the measured soluble markers. The patients that received RBV-free therapy quickly normalised all soluble factors in their plasma except for sCD163, which remained elevated throughout treatment. In contrast, patients that received RBV in combination with DAA maintained elevated levels of both CXCL10 and sCD163 (Fig. 2a,b). Six months after the end of therapy, sCD163 remained significantly elevated in both subgroups (p = 0.0158 and p = 0.0004), whereas CXCL10 was significantly elevated only in the group that received RBV (p = 0.0012, Fig. 2c).

Levels of sCD163 distinguish patients with different degrees of liver cirrhosis throughout DAA therapy. To assess whether the stage of liver disease was associated with the plasmatic protein profile, we stratified cirrhotic patients in the DAA group according to Child Pugh class. There were 18 patients classified as Child Pugh A and 7 patients classified as Child Pugh B. Although differences with HD seemed more pronounced in the Child B group (Fig. 3a), those differences were also reduced upon start of therapy, with the exception of sCD163, which remained elevated and significantly different between the 2 subgroups throughout treatment (Fig. 3b). Even 6 months after successful therapy, sCD163 remained elevated in Child Pugh A patients (p = 0.0103) and even higher in Child Pugh B patients (p < 0.0001, Fig. 3c). We also investigated possible associations between



Figure 1. Baseline differences and evolution of plasma cytokines between healthy controls and patients. (a) Comparison between healthy donors (HD), alcoholic cirrhotic patients (AC) and chronic hepatitis C (CHC) patients before they initiated DAA treatment (DAA). (b) Evolution of plasma cytokine levels (median \pm IQR) during DAA therapy. Dotted lines and shaded area represent the median and 95% confidence interval of the median for healthy donors. Stars represent significant changes when compared with baseline levels in CHC patients *p < 0.05; **p < 0.001; ****p < 0.0001. (c) Comparison between HD and CHC 6 months after the end of DAA treatment (FU).

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levels of sCD163 at the FU time point, and various indicators of liver health and found significant correlations with AST levels ($\rho = 0.44$, p = 0.0198), Albumin ($\rho = -0.48$, p = 0.0101) and prothrombin time (PT-INR, $\rho = 0.41$, p = 0.0301, Fig. 3d). Thus, elevated sCD163 levels seem associated with liver disease and remain elevated in patients with advanced cirrhosis despite successful clearance of HCV with DAA therapy.

IFN-free DAA combinations improve fibrosis indicators. Liver stiffness measurements were available from time points before and after therapy in 18 of the 28 DAA-treated patients (two non-cirrhotic, 14 Child Pugh A and two Child Pugh B). Mixed effects models of paired LSMs demonstrated a significant decrease in stiffness values (p < 0.0001), between baseline and follow-up measurements in the DAA group. The decrease was significant in the Child Pugh A subgroup of patients (p = 0.0003), and there was a similar trend in Child Pugh B patients (Fig. 4a). This pattern indicated decreased liver fibrosis and inflammation. Since DAA therapy efficiently eliminates the virus and quickly normalises most aspects of the cytokine milieu in plasma, it is likely that this treatment reduces liver inflammation and a decrease in LSM may thus be insufficient to conclude that there is a decrease in fibrosis. We therefore calculated APRI and FIB-4 scores for all treated patients. Both APRI and FIB-4 decreased significantly by EOT (p < 0.0068 and p = 0.0038, respectively), and this effect was maintained at follow-up (p = 0.0061 and p = 0.0010, respectively) (Fig. 4b,c). This pattern reached significance in the Child Pugh A patient subgroup with a similar trend in Child Pugh B patients (Fig. 4b,c). These observations are consistent with the notion that DAA therapy reduces liver fibrosis, including in patients with advanced cirrhosis. In support of these results, six out of seven patients classified as Child Pugh B before the start of therapy were re-classified as Child Pugh A at follow-up, further indicating an improvement of liver status.



Figure 2. Effect of Ribavirin on cytokine levels. (a) Heatmap of median fold change of soluble marker levels in plasma compared to healthy donors for all measured cytokines in CHC patients who received a DAA regimen with (+RBV) or without (-RBV) Ribavirin. (b) Evolution of markers concentration (median \pm IQR) in CHC patients over the course of treatment. Dotted lines and shaded area represent the median and 95% confidence interval of the median for healthy donors. (c) Comparison between healthy donors (HD) and CHC patients who received a DAA regimen with (+RBV) or without (-RBV) Ribavirin. BL: baseline; W4: week 4; EOT: end of treatment; FU: follow-up.

Baseline levels of soluble FABP4 are associated with improvement of fibrosis indicators in response to DAA therapy. To investigate whether soluble plasma markers might be associated with fibrosis reduction, we performed Spearman correlation analyses between baseline levels of the 25 cytokines and the reduction in LSM, APRI and FIB-4 scores observed at FU (Fig. 5). IL-18 levels at baseline correlated positively with LSM reduction ($\rho = 0.54$, p = 0.028), and FABP4 levels at baseline correlated negatively with reductions in both APRI ($\rho = -0.53$, p = 0.0035) and FIB-4 ($\rho = -0.65$, p = 0.0002). To investigate whether those correlations may be driven primarily by single components of the APRI and FIB-4 scores, we analysed correlated significantly with platelet count increase and decrease in liver enzymes. Although FABP4 correlated significantly with platelet count increase ($\rho = -0.48$, p = 0.0097) and AST decrease ($\rho = -0.45$, p = 0.0153; Supplementary Fig. S3), those correlations were weaker than the correlation with both composite scores, suggesting that FABP4 levels are associated with the reduction in liver disease.

Discussion

Residual liver disease and risk of developing HCC are concerns in patients that have cleared their HCV infection, and it is therefore important to understand the dynamics of systemic and hepatic inflammation during and after successful DAA treatment. Here, we identify differences in the plasma cytokine milieu of HD and CHC patients, and show that DAA therapy rapidly reduces those differences with the exception of sCD163. Levels of sCD163 also distinguish patients with different levels of cirrhosis. We also observe that patients who receive RBV seem to maintain elevated plasma levels of CXCL10, consistent with an immune-stimulatory role of RBV. Finally, we find a significant association between baseline levels of FABP4 and subsequent reduction in APRI and FIB-4 measurements during DAA treatment, suggesting a link between FABP4 plasma levels and liver disease reduction.

Our data set identified clear differences in plasma inflammatory markers between CHC patients and HD. CCL5 was reduced in CHC patients whereas CXCL10, sCD14 and sCD163 were elevated. CXCL10 is associated with viral load and HCV triggers its production via NF- κ B activation¹². Soluble CD14 is a marker of monocyte activation¹³, whereas sCD163 is a marker of macrophage activation¹⁴, and all these three markers are associated with liver fibrosis in untreated CHC^{15–17}. However, elevated levels of sCD14 and sCD163 are not specific to HCV infection as previously observed in AC^{18–20}. Interestingly, in our cohort, although both sCD14 and sCD163 were elevated in AC, they were significantly higher in CHC patients, despite a lower proportion of Child Pugh B and C



Figure 3. Influence of cirrhosis grade on cytokine levels over the course of therapy. (**a**) Heatmap of median fold change of soluble marker levels in plasma compared to healthy donors for all measured cytokines in CHC patients with cirrhosis classified as Child Pugh A (Child A) or Child Pugh B (Child B). (**b**) Evolution of protein concentration (median \pm IQR) in CHC patients over the course of treatment. Dotted lines and shaded area represent the median and 95% confidence interval of the median for healthy donors. Crosses and stars indicate significant difference between Child Pugh A and B groups overall and for each timepoint, respectively. ^{††}p < 0.01; *p < 0.05; **p < 0.01. (**c**) Comparison between healthy donors (HD) and CHC patients with cirrhosis. BL: baseline; W4: week 4; EOT: end of treatment; FU: follow-up. (**d**) Spearman correlations between follow-up plasma concentration of sCD163 and different indicators of liver health. Dotted blue lines indicate 95% confidence intervals.

patients. Thus, the higher plasma levels of sCD14 and sCD163 in CHC compared to AC might reflect a combination of anti-viral immune activation and liver disease.

In contrast to IFN-containing regimens, which induced broad changes in the cytokine milieu, DAA therapy quickly reduced the differences in plasma protein levels observed with HD. This pattern likely reflects the very different mechanisms of action of the two treatments. DAA drugs specifically interfere with viral replication, whereas IFN is immune activating and modulates the expression of hundreds of IFN-regulated genes, which themselves can influence cytokine secretion via a complex network of interactions²¹. These findings are in accordance with recent work by Burchill *et al.*, showing a rapid decrease in the expression of genes associated with inflammation in PBMCs of HCV infected patients undergoing DAA therapy²². Certain features of T-cell and NK cell phenotype and function are also restored following DAA therapy²³⁻²⁶, supporting the idea of a general immune system normalisation after HCV clearance. However, recent studies also indicate that other immune cell traits, such as the appearance of regulatory T cells²⁷, NK cell receptor repertoire diversity²⁸ and the state of the MAIT cell compartment²⁹, are still altered after DAA therapy. Thus, the impact on the cellular immune system might in part persist after successful viral clearance. Hengst *et al.* observed a partial but incomplete restoration



Figure 4. Improvement in fibrosis indicator values over the course of DAA therapy. Evolution of LSM (**a**), APRI score (**b**) and FIB-4 index (**c**) during the course of DAA therapy. Stars indicate statistical difference between time-points computed with repeated measurement ANOVA using mixed effect models. *p < 0.05; **p < 0.01; ****p < 0.001; ****p < 0.0001.



Figure 5. Correlations between baseline cytokine levels and decrease in fibrosis indicators. Spearman correlations between baseline plasma concentration and the improvement in fibrosis indicator values over the course of DAA treatment were computed for all cytokines. Correlations that were statistically significant (p < 0.05) are shown here. Dotted blue lines indicate 95% confidence intervals.

of the cytokine milieu during DAA therapy and, notably, CXCL10 remained elevated throughout treatment³⁰. Interestingly, all patients in that cohort received RBV in combination with DAA, and our observations indicate that patients who receive RBV maintain higher levels of CXCL10. Studies have also shown that RBV has immuno-modulatory effects^{31,32} and can induce ISGs *in vitro*^{33,34}. In our study, only sCD163 remained significantly elevated in patients that received DAA regimen without RBV.

Levels of sCD163 were significantly decreased by DAA treatment, as described by others^{35,36}. However, levels remained significantly elevated in comparison to those seen in HD. This is in accordance with work by Mascia *et al.*,



Figure 6. Longitudinal evolution of circulating soluble marker levels and liver fibrosis following elimination of HCV in patients undergoing DAA therapy. Liver diagrams were adapted from Servier medical art (https://smart.servier.com/) under a Creative Commons Attribution 3.0 Unported Licence (https://creativecommons.org/licenses/by/3.0/legalcode).

showing sustained high levels of sCD163 in DAA-treated CHC patients 12 weeks after EOT³⁷. Additionally, we observed significant correlations between levels of sCD163 at FU and multiple indicators of liver health (AST, Albumin and PT-INR). Taken together, these findings by us and others suggest that persistent macrophage activation could play a role in residual liver disease despite a general dampening of inflammation (Fig. 6). Of note, the last timepoint evaluated in our study was 6 months after EOT, thus further studies would be required to assess whether elevated sCD163 reflects an irreversible consequence of CHC or a slower recovery of certain immune processes.

It is also important to understand the response of patients with more severe cirrhosis (Child Pugh B) to DAA combinations in comparison to patients with milder liver damage (Child Pugh A). Our data indicate that IFN-free DAA therapy normalises the cytokine profile, with the exception of sCD163, in both groups, and thus suggest that advanced liver damage is not a barrier to a normalised plasma cytokine milieu. Interestingly, six out of the seven Child Pugh B patients from our cohort were re-classified as Child Pugh A at the EOT, in line with improved liver function. We also observed a clear improvement in LSMs after treatment, as observed by others^{38–40}. However, this measurement does not allow clear discrimination between reduction in liver inflammation, improvement of fibrosis, or a combination of the two. The predictive power of liver elasticity measurements for fibrosis can also be reduced following therapeutic eradication of HCV⁴¹. To complement the LSM data, we therefore investigated the evolution of APRI and FIB-4 scores during therapy and showed a clear reduction in both of these indicators of liver fibrosis. Nevertheless, further studies comprising liver biopsies will be required to determine the extent of liver regeneration after successful DAA therapy.

We also investigated associations between plasma proteins and improvement of liver fibrosis and found that baseline levels of FABP4 inversely correlated with the improvement in both APRI and FIB-4 observed after treatment. FABP4 is expressed in adipocytes and macrophages, and high serum concentrations of FABP4 are associated with inflammation and risk for metabolic and vascular diseases⁴². Recently, it was shown that increased FABP4 levels in the blood correlate with poor prognosis in cirrhosis⁴³. In the same study, it was also shown that FABP4 gene expression was increased in cirrhotic livers and that liver macrophages seemed to be responsible for that increase. Interestingly, sCD163 is also released by macrophages and hepatic macrophages are known to play a critical role in liver inflammation, fibrosis and resolution of inflammation⁴⁴. Investigating the mechanisms leading to sCD163 and FABP4 release during CHC may thus help better understand the mechanisms of fibrosis resolution during DAA therapy. Of note, CHC cohorts are often very heterogeneous groups and further research is needed to fully understand the influence of multiple factors (such as HCV genotype, fatty liver content, obesity, diabetes, etc.) on residual inflammation and liver disease.

In conclusion, these results support the notion of a rapid restoration of the inflammatory state in CHC in response to DAA therapy, but also warrant further investigations of the role of macrophage activation in liver disease reduction after clearance of HCV infection.

Methods

Patients. To investigate the impact of HCV clearance on the immune system, CHC patients above 18 years of age scheduled to receive DAA therapy were sampled before, during and after treatment at the Department of Infectious Diseases and the Department of Gastroenterology and Hepatology at Karolinska University Hospital. Blood samples were collected from 28 CHC patients who received DAA therapy and successfully achieved SVR, and 13 CHC patients who received pegylated-IFN therapy and successfully achieved SVR. Exclusion criteria included concurrent Hepatitis B virus or HIV co-infections. In addition, blood samples from 20 HD, and 12 patients suffering from AC were included as non-HCV infected comparison groups. Of note, a proportion (64%)

of patients in the DAA treated group previously underwent IFN α -based therapy without achieving SVR. The HD group was matched in gender and age to the DAA group. The study was conducted in accordance with the declaration of Helsinki, approved by the regional ethics committee (Regional Ethics Review Board in Stockholm, approval number 2012/63-31/1) and all participants gave informed consent.

Liver stiffness measurement. Liver stiffness measurement was performed using transient elastography (FibroScan[®]). Paired LSMs before and after therapy were obtained for 18 of the 28 DAA-treated patients (2 non-cirrhotic, 14 Child Pugh A and 2 Child Pugh B liver cirrhosis patients). Measurements with an IQR >30 or a success rate <50% were excluded from the analysis.

Sample collection. Venous blood was collected in heparin-coated tubes and spun 10 min at 680 g at room temperature to separate plasma, which was stored at -80 °C immediately after separation. CHC patients receiving DAA therapy had blood collected at four time-points: before initiation of therapy, four weeks after initiation of therapy (W4), at the end of therapy (EOT, either 12 or 24 weeks after initiation of therapy) and at a follow-up time point collected approximatively six months after the EOT. For the IFN-treated group, 13 baseline samples, 11 W12 and 6 EOT samples (either 24 or 48 weeks after initiation of therapy) were included in the analysis.

Luminex assay and ELISA. Plasma samples were analysed using a custom 24-plex magnetic Luminex assay (R&D systems). All samples were diluted 1:2 and assays were performed according to manufacturer's protocol. Samples were acquired using the Bio-Plex Magpix multiplex reader (Bio-Rad) and analysed using the Bio-Plex Manager software. Soluble CD14 (sCD14) concentration in plasma samples was measured using the human CD14 Quantikine ELISA kit from R&D systems (DC140) according to manufacturer's protocol.

Statistical analysis. Statistical analyses were performed using Prism 8 (GraphPad). Mann-Whitney tests were used for comparisons between 2 groups and Kruskal-Wallis test with post-hoc Dunn's correction were used for comparing more than 2 groups. Repeated measure one-way analysis of variance (ANOVA) using mixed effect models and Holm-Sidak multiple testing correction were used to analyse changes over time. Association between variables were analysed using Spearman's rank correlation.

Data availability

All data generated or analysed during this study are included in this published article (and its Supplementary Information files).

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References

- Blach, S. et al. Global prevalence and genotype distribution of hepatitis C virus infection in 2015: a modelling study. Lancet Gastroenterology Hepatology 2, 161–176 (2017).
- 2. Burstow, N. J. et al. Hepatitis C treatment: where are we now? Int. J. Gen. Med. 10, 39-52 (2017).
- European Association for the Study of the Liver. Electronic address: easloffice@easloffice.euEuropean Association for the Study of the Liver. EASL Recommendations on Treatment of Hepatitis C 2018. J. Hepatol., https://doi.org/10.1016/j.jhep.2018.03.026 (2018).
- World Health Organization. Global Hepatitis Programme. *Global Health Sector Strategy on Viral Hepatitis*, 2016–2021 (2016).
 Waziry, R. *et al.* Hepatocellular carcinoma risk following direct-acting antiviral HCV therapy: A systematic review, meta-analyses,
- and meta-regression. J. Hepatol. 67, 1204–1212 (2017).
 6. Mascia, C. et al. Active HCV infection is associated with increased circulating levels of interferon-gamma (IFN-γ)-inducible protein-10 (IP-10), soluble CD163 and inflammatory monocytes regardless of liver fibrosis and HIV coinfection. *Clin. Res. Hepatol. Gastroenterol.*, https://doi.org/10.1016/j.clinre.2017.04.007 (2017).
- 7. Baskic, D. et al. Cytokine profile in chronic hepatitis C: An observation. Cytokine 96, 185-188 (2017).
- 8. Neesgaard, B., Ruhwald, M. & Weis, N. Inducible protein-10 as a predictive marker of antiviral hepatitis C treatment: A systematic review. *World J. Hepatol.* 9, 677–688 (2017).
- 9. Yoneda, S. *et al.* Association of serum cytokine levels with treatment response to pegylated interferon and ribavirin therapy in genotype 1 chronic hepatitis C patients. *J. Infect. Dis.* **203**, 1087–1095 (2011).
- 10. Snyder, N. *et al.* APRI: an easy and validated predictor of hepatic fibrosis in chronic hepatitis C. J. Clin. Gastroenterol. **40**, 535–542 (2006).
- 11. Vallet-Pichard, A. *et al.* FIB-4: an inexpensive and accurate marker of fibrosis in HCV infection. comparison with liver biopsy and fibrotest. *Hepatology* **46**, 32–36 (2007).
- Brownell, J. et al. Direct, interferon-independent activation of the CXCL10 promoter by NF-κB and interferon regulatory factor 3 during hepatitis C virus infection. J. Virology 88, 1582–1590 (2014).
- Shive, C. L., Jiang, W., Anthony, D. D. & Lederman, M. M. Soluble CD14 is a nonspecific marker of monocyte activation. AIDS 29, 1263–1265 (2015).
- 14. Møller, H. J. Soluble CD163. Scand. J. Clin. Lab. Invest. 72, 1-13 (2012).
- 15. Kazankov, K. *et al.* Soluble CD163, a macrophage activation marker, is independently associated with fibrosis in patients with chronic viral hepatitis B and C. *Hepatology* **60**, 521–530 (2014).
- Sandler, N. G. *et al.* Host response to translocated microbial products predicts outcomes of patients with HBV or HCV infection. *Gastroenterology* 141, 1220–30–1230.e1–3 (2011).
- Zeremski, M., Dimova, R., Astemborski, J., Thomas, D. L. & Talal, A. H. CXCL9 and CXCL10 Chemokines as Predictors of Liver Fibrosis in a Cohort of Primarily African-American Injection Drug Users With Chronic Hepatitis C. J. Infect. Dis. 204, 832–836 (2011).
- Grønbaek, H. *et al.* Soluble CD163, a marker of Kupffer cell activation, is related to portal hypertension in patients with liver cirrhosis. *Aliment. Pharmacol. Ther.* 36, 173–180 (2012).
- Sandahl, T. D. et al. Hepatic macrophage activation and the LPS pathway in patients with alcoholic hepatitis: a prospective cohort study. Am. J. Gastroenterol. 109, 1749–1756 (2014).
- Sandahl, T. D. *et al.* The macrophage activation marker sCD163 combined with markers of the Enhanced Liver Fibrosis (ELF) score
 predicts clinically significant portal hypertension in patients with cirrhosis. *Aliment. Pharmacol. Ther.* 43, 1222–1231 (2016).

- Schneider, W. M., Chevillotte, M. D. & Rice, C. M. Interferon-stimulated genes: a complex web of host defenses. Annu. Rev. Immunol. 32, 513–545 (2014).
- 22. Burchill, M. A. *et al.* Rapid reversal of innate immune dysregulation in blood of patients and livers of humanized mice with HCV following DAA therapy. *PLoS One* **12**, e0186213–17 (2017).
- Burchill, M. A., Golden-Mason, L., Wind-Rotolo, M. & Rosen, H. R. Memory re-differentiation and reduced lymphocyte activation in chronic HCV-infected patients receiving direct-acting antivirals. J. Viral Hepat. 22, 983–991 (2015).
- 24. Martin, B. et al. Restoration of HCV-specific CD8+ T cell function by interferon-free therapy. J. Hepatol. 61, 538-543 (2014).
- Spaan, M. et al. Immunological Analysis During Interferon-Free Therapy for Chronic Hepatitis C Virus Infection Reveals Modulation of the Natural Killer Cell Compartment. J. Infect. Dis. 213, 216–223 (2016).
- Serti, E. et al. Successful Interferon-Free Therapy of Chronic Hepatitis C Virus Infection Normalizes Natural Killer Cell Function. Gastroenterology 149, 190–200.e2 (2015).
- Langhans, B. et al. Increased peripheral CD4+ regulatory T cells persist after successful direct-acting antiviral treatment of chronic hepatitis C. J. Hepatol. 66, 888–896 (2017).
- Strunz, B. *et al.* Chronic hepatitis C virus infection irreversibly impacts human natural killer cell repertoire diversity. *Nat. Commun.* 9, 2275 (2018).
- 29. Hengst, J. et al. Nonreversible MAIT cell-dysfunction in chronic hepatitis C virus infection despite successful interferon-free therapy. Eur. J. Immunol. 46, 2204–2210 (2016).
- Hengst, J. et al. Direct-Acting Antiviral-Induced Hepatitis C Virus Clearance Does Not Completely Restore the Altered Cytokine and Chemokine Milieu in Patients With Chronic Hepatitis C. J. Infect. Dis. 214, 1965–1974 (2016).
- 31. Graci, J. D. & Cameron, C. E. Mechanisms of action of ribavirin against distinct viruses. Rev. Med. Virol. 16, 37-48 (2006).
- 32. Chung, R. T. *et al.* Mechanisms of action of interferon and ribavirin in chronic hepatitis C: Summary of a workshop. In **47**, 306–320 (2008).
- Thomas, E. et al. Ribavirin potentiates interferon action by augmenting interferon-stimulated gene induction in hepatitis C virus cell culture models. Hepatology 53, 32–41 (2011).
- 34. Wang, Y. *et al.* Ribavirin Contributes to Hepatitis C Virus Suppression by Augmenting pDC Activation and Type 1 IFN Production. *PLoS One* **10**, e0135232 (2015).
- 35. Lund Laursen, T. *et al.* Rapid and persistent decline in soluble CD163 with successful direct-acting antiviral therapy and associations with chronic hepatitis C histology. *Scand. J. Gastroenterol.* **61**, 1–8 (2018).
- Laursen, T. L. et al. Time-dependent improvement of liver inflammation, fibrosis and metabolic liver function after successful directacting antiviral therapy of chronic hepatitis C. J. Viral. Hepat. 155, 411–421 e41–8 (2019).
- Mascia, C. et al. Changes in inflammatory biomarkers in HCV-infected patients undergoing direct acting antiviral-containing regimens with or without interferon. PLoS One 12, e0179400 (2017).
- Bernuth, S. et al. Early changes in dynamic biomarkers of liver fibrosis in hepatitis C virus-infected patients treated with sofosbuvir. Digestive Liver Dis. 48, 291–297 (2016).
- Knop, V. et al. Regression of fibrosis and portal hypertension in HCV-associated cirrhosis and sustained virologic response after interferon-free antiviral therapy. J. Viral Hepat. 23, 994–1002 (2016).
- 40. Bruno, G. *et al.* Rapid improvement in liver fibrosis in HCV-infected patients with or without HIV infection and DAA-induced SVR: A 'turning-off' effect of liver inflammation? *J. Viral Hepat.* **24**, 174–175 (2017).
- D'Ambrosio, R. et al. The diagnostic accuracy of Fibroscan for cirrhosis is influenced by liver morphometry in HCV patients with a sustained virological response. J. Hepatol. 59, 251–256 (2013).
- 42. Kralisch, S. & Fasshauer, M. Adipocyte fatty acid binding protein: a novel adipokine involved in the pathogenesis of metabolic and vascular disease? *Diabetologia* 56, 10–21 (2013).
- 43. Graupera, I. *et al.* Adipocyte Fatty-Acid Binding Protein is Overexpressed in Cirrhosis and Correlates with Clinical Outcomes. *Sci. Rep.* **7**, 1829 (2017).
- 44. Krenkel, O. & Tacke, F. Liver macrophages in tissue homeostasis and disease. Nat. Rev. Immunology 17, 306-321 (2017).

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Author contributions

J.B.G. collected samples, performed experiments, analysed data and co-wrote the manuscript. D.F.G.M. participated in study design, collected samples and performed experiments. BS collected samples. T.C., S.A. and K.F. participated in patient recruitment. N.K.B., S.A. and K.F. participated in study design and data analysis. JKS coordinated the study, participated in study design, data analysis, and co-wrote the manuscript. All authors read and approved the final manuscript.

Competing interests

J.B.G., D.F.G.M., B.S., T.C., N.K.B., K.F. and J.K.S. report no competing interests. S.A. has served as a speaker and a consultant for AbbVie, Gilead, BMS. and MSD, and has received research funding from AbbVie and Gilead for an investigator-initiated study, not related to this study.

Additional information

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