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A novel anti *Candida albicans* drug screening system based on high-throughput microfluidic chips

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Due to the antibacterial resistance crisis, developing new antibacterials is of particular interest. In this study, we combined the antifungal drug amphotericin B with 50,520 different small molecule compounds obtained from the Chinese National Compound Library in an attempt to improve its efficacy against *Candida albicans* persister cells. To systematically study the antifungal effect of each compound, we utilized custom-designed high-throughput microfluidic chips. Our microfluidic chips contained microchannels ranging from 3 μm to 5 μm in width to allow *Candida albicans* cells to line up one-by-one to facilitate fluorescence-microscope viewing. After screening, we were left with 10 small molecule compounds that improved the antifungal effects of amphotericin B more than 30% against *Candida albicans* persister cells.

In recent decades, fungal infections have increased in severity. People are more at-risk for opportunistic fungal infections than ever, especially those who undergo organ transplants, prosthetic heart valve implants, or chemotherapy; those who use immunosuppressants or broad-spectrum antibacterials; those with HIV infections; premature infants; and the elderly^{1–5}. At present, there is a very small variety of effective antifungal agents from which to choose. As a result, chronic, secondary fungal infections often prove fatal long before the primary disease^{6,7}. The abuse of broad-spectrum antibacterials has led to the development of microbiological drug resistance, partially through the activity of persister cells. Persisters were first described by Joseph Bigger in 1944, in one of the first studies on the mechanism of penicillin's action⁸. Bigger discovered that although penicillin lysed the cell walls in a growing population of *Staphylococcus spp.*, a small number of cells persisted and survived. Lewis, K noted that these cells were not antibacterial-resistant mutants but dormant, non-dividing cells⁹. Michael D. LaFleur continued work on the topic and proposed the idea of *Candida albicans* persisters in 2006¹⁰. *Candida albicans* is the most common clinical fungal infection. It is both a component of normal human microflora and a dangerous opportunistic pathogen. Invasive candidiasis can grow deep in tissue and is generally chronic and persistent, especially in patients with human immunodeficiency. Systemic *Candida* infections have a surprisingly high mortality rate: 29–76%^{11–13}. Yamashita Ichiro *et al.* first reported the flocculation gene *FLO8* in the Yeast *Saccharomyces*¹⁴. Fang Cao suggested that *FLO8* may function downstream of the cAMP/PKA pathway, and together with *EFG1*, regulates the expression of hypha-specific genes and genes that are important for the virulence of *Candida albicans*¹⁵. In 2009, our research group screened out kinase GIN4 and transcription factor *FLO8*. In this work, we constructed a gene fragment using green fluorescence proteins (GFPs) to target *FLO8* in normal *Candida albicans* and induce more persisters. This multiplication of *FLO8* persisters allowed for easy observation by fluorescence microscopy, which enabled the related new drugs screening based on GFPs expression.

However, the current selection of anti *Candida albicans* drug is extremely limited: fluconazole, amphotericin B, caspofungin, and terbinafine make up the bulk of them. Moreover, these antifungals are toxic and exhibit unpleasant side effects¹⁶. It is imperative that new antibacterials are developed to address these limitations, including ones that are effective against persisters. K. Lewis *et al.* has reported that combining fluconazole with small

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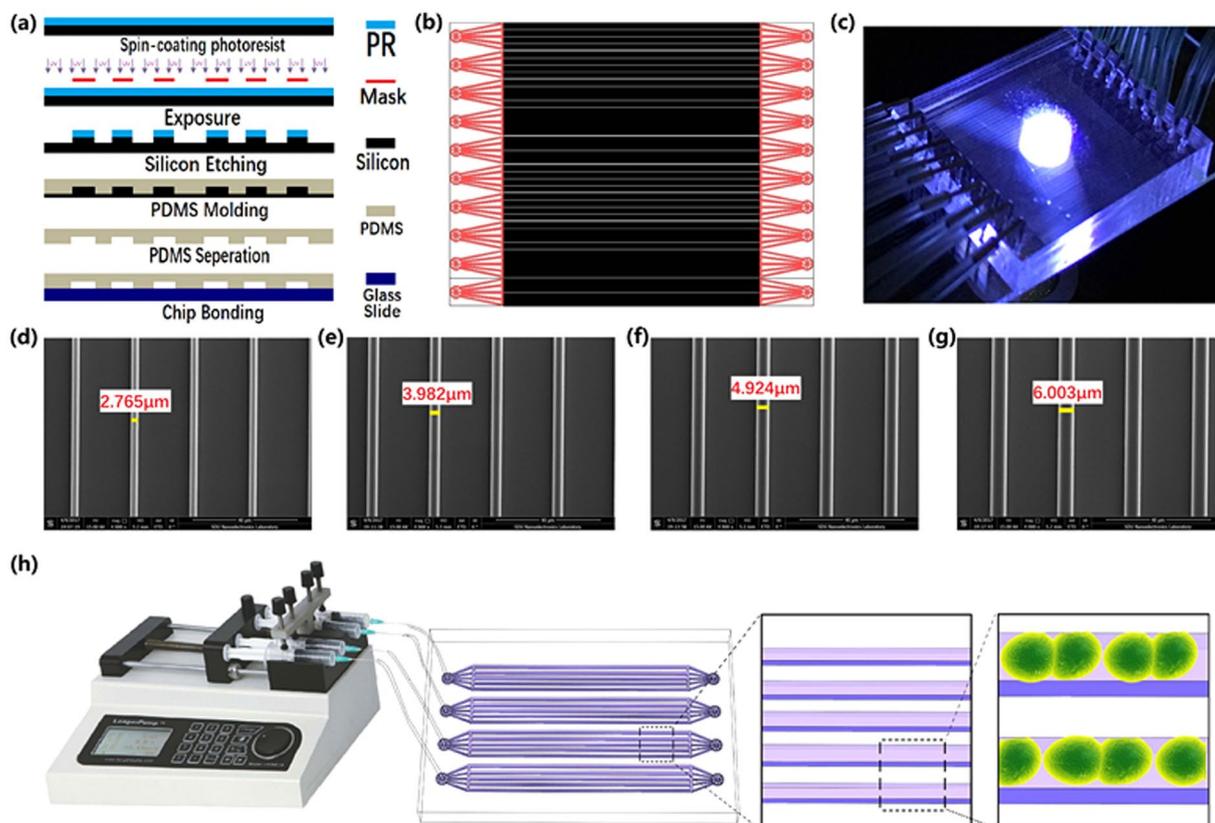


Figure 1. (a) The fabrication process of the drug-screening microfluidic chip; (b) The layout; (c) Photo of the microfluidic chip; (d) SEM microscope photo of the 3- μm microchannel; (e) The 4- μm microchannel; (f) The 5- μm microchannel; (g) The 6- μm microchannel; (h) System sketch. We thank Longer Precision Pump Co., Ltd. China for the authorization to use the pump photo.

molecule compounds can improve activity against *Candida albicans*¹⁷. Here, we report an approach to improve the efficacy of anti *Candida albicans* drug by combining amphotericin B with 50,520 different small molecule compounds obtained from the Chinese National Compound Library, and developed a new microfluidic chip for conducting *Candida albicans* antifungal screening. The miniaturization brought forth by microfluidics generally allows shorter time to results, integrates sample preparation, and makes fluid handling portable¹⁸. Microfluidic chips were reported to conduct flow cytometric analysis of fluorescently stained cells from different organisms¹⁹, rapid detection and identification of bacteria²⁰, fungal separation and PCR amplification²¹. Balaban *et al.* firstly used microfluidic-chip-like device to observe the persisters²².

Our microfluidic chips contain thousands of parallel microchannels through which many new drugs can be screened at the same time (Fig. 1). Microchannels are generally only a few micrometers wide, which allows a working volume of mere nanoliters by reducing reagent consumption to 1/100000 of traditional 384w-MTP^{23–27}. Using our microfluidic chips, we performed the screening of 50,520 new drugs, and finally found 10 small molecule compounds that enhance amphotericin B's anti-*Candida-albicans* effects against persisters by more than 30%^{28,29}.

Results

New anti candida albicans drugs development. Previously, our team located the *FLO8* gene, which is related to the activity of *Candida albicans* persisters¹⁶. To target *FLO8*, we constructed a gene fragment with GFP. We then transferred this gene fragment to *Candida albicans* 3153A to obtain more *Candida albicans* persisters, and to make them readily observable using fluorescent microscopy.

To find new drug candidates, we selected 50,520 small molecule compounds from the Chinese National Compound Library and systematically combined each of them with amphotericin B. The resulting concentration of each new amphotericin B combination was 3.5 $\mu\text{g}/\text{mL}$. Suspended cells were injected into 10 microfluidic chips at the same time, and it took 48 hours for the cells to grow into biofilms. 100 compounds could be screened in one chip, and 20 chips were processed each time. The images of cells were taken under fluorescence microscope at 24 hours after drugs were loaded. The drug efficacy was evaluated by counting the alive persister cells in the microchannels.

Microfluidic chips design and fabrication. The microchannels of microfluidic chip was designed based on the size of *Candida albicans*. The width and depth of microchannels are in the range of 3 μm –6 μm in order to let *Candida albicans* pass through and line up one by one. One chip has 20 units, each unit include twenty 3 μm -microchannels, twenty 4 μm -microchannels, twenty 5 μm -microchannels, and twenty

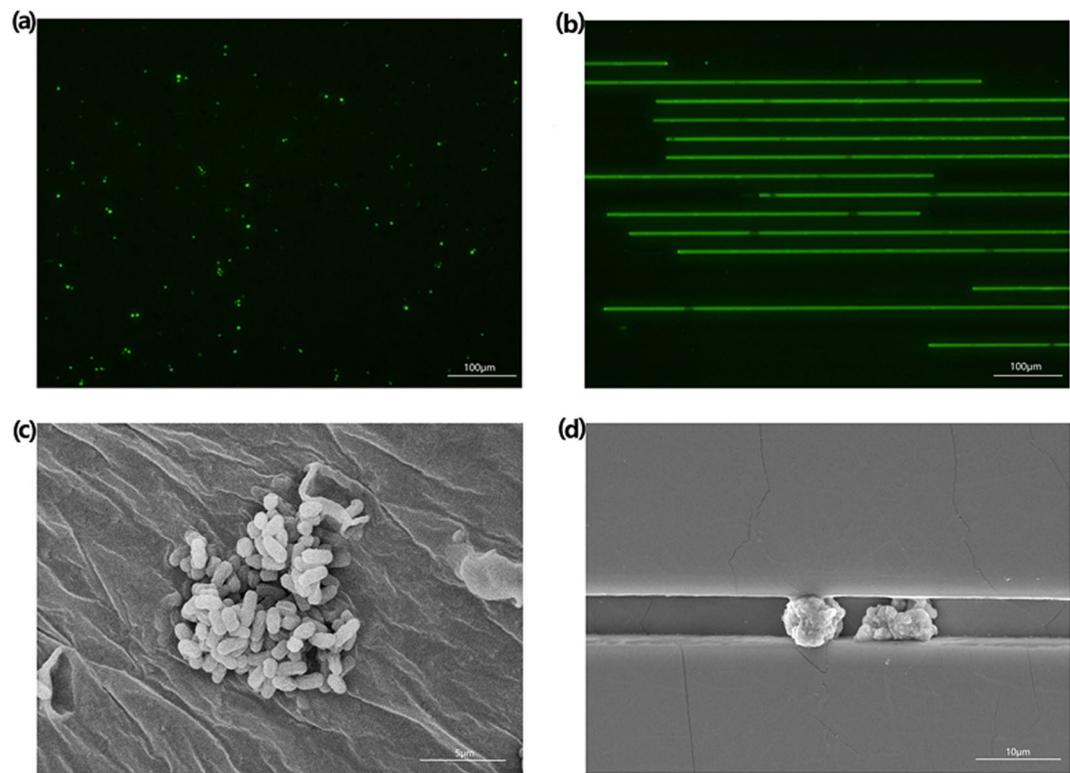


Figure 2. The comparison of *Candida albicans* in traditional culture and microfluidic chips. (a) Fluorescence microscope view of normal *Candida albicans*; (b) Fluorescence microscope view of *Candida albicans* in microchannels; (c) SEM view of normal *Candida albicans*; (d) SEM view of *Candida albicans* in microchannels.

6 μm -microchannels. Each unit has an inlet and an outlet. The layout is shown in Fig. 1(a), and the fabrication process schematic is illustrated in Fig. 1(b). The silicon wafer with microchannels then was used hard mode for PDMS chips. The SEM views of microchannels in different sizes are shown in Fig. 1(d–g). The PDMS chip and a pre-cleaned glass slide were treated with oxygen plasma, it was immediately brought into contact against the slide to form closed channels. The microscope picture of the whole chip is as shown in Fig. 1(c), and the microfluidic platform was setup as shown in Fig. 1(h).

Figure 2(a,c) show the fluorescent and SEM microscope views of cells cultured in Petri dish, and Fig. 2(b,d) are those of cells in microchannels. In our study we discovered that 4- μm and 5- μm microchannels were most suitable for the growth of *Candida albicans*, while 3- μm microchannels were too narrow to allow for *Candida albicans* cell passage. Due to the obstructions created by the 3- μm microchannels, it was difficult for the pump to load the culture solution. Conversely, the 6- μm microchannels were too wide for the majority of *Candida albicans* cells and created cell overlap, making persister visualization difficult. Cells grow up in piles in the culture Petri dish, while they line up one by one in order in the microchannels. It showed similar growth curves in microchannels compared to traditional 384-well plate.

New drug candidates found using high-throughput microfluidic chip screening. The *Candida albicans* suspension was loaded into a 1-ml injector, and an injection pump was used to inject the solution into microfluidic chips at a rate of 5 $\mu\text{m}/\text{min}$. 100 small molecules and amphotericin B mixtures were screened in each chip, and 20 microfluidic chips were conducted at the same time. Adhesive *Candida albicans* settled down and spreaded on the PDMS channel surface in 2 hours, and Fresh RPMI-1640 was provided every 6 hours up to 36 hours, at which point the microchannels had been completely filled by growing *Candida albicans* in line (Fig. 3a–c). Then 2 μL of a drug solution was loaded into each microfluidic chip inlet, and the microchannels were filled via vacuum suction at the outlet. Due to the adhesive characteristics of *Candida albicans* cells, the cells remained in the microchannels throughout the culture and drug-input process. After the cells were exposed to the drugs for 1 hour, fresh RPMI-1640 culture medium was loaded into the injection pump to flush out the drug. The microfluidic chip was then observed under fluorescent microscopy to check for persisters (Fig. 3d–f).

GFP constructed in *FLO8* is only active when cells are alive. However, in order to confirm *Candida albicans* cell death, LIVE-DEAD[®] staining was used to verify a compound's fungicidal effects. After incubation, the microchannels were observed under fluorescent microscopy (Fig. 3g–i). Green fluorescent indicated live cells, and red fluorescent indicated dead cells. This method made it easy to observe whether or not the drug compounds exhibited a fungicidal effect on *Candida albicans*.

To construct an evaluation equation, we considered the number of *Candida albicans* persisters per unit of microchannel length.

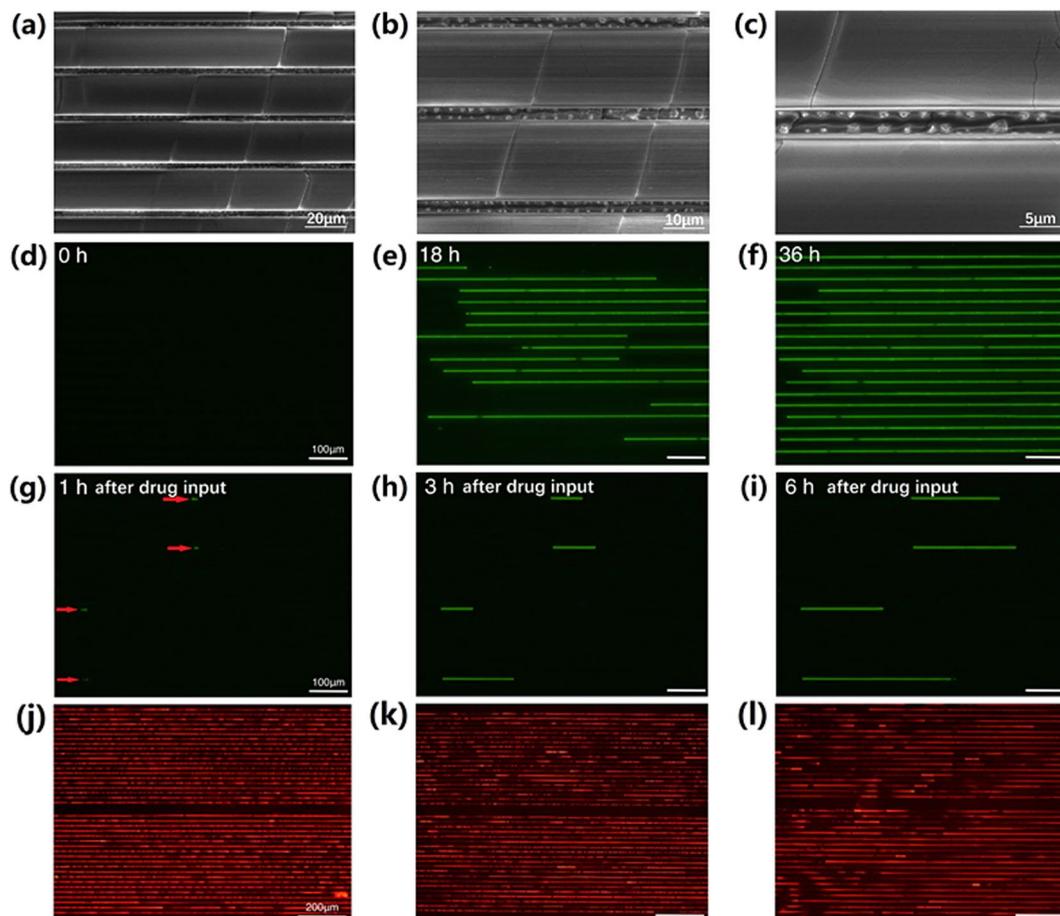


Figure 3. *Candida albicans* cultured in a microfluidic chip as seen under SEM at different magnifications (a–c) and fluorescent microscopy at (d) $t = 0$, (e) $t = 18$ h, (f) $t = 36$ h. One hour after new drug import into microchannels, *Candida albicans* persists in a microfluidic chip under fluorescent microscopy at (g) $t = 1$ h, (h) $t = 3$ h, and (i) $t = 6$ h (The four red arrows identify the persisters.) *Candida albicans* LIVE/DEAD[®] staining after drug exposure in a microfluidic chip at (j) $t = 1$ h, (k) $t = 3$ h, and (l) $t = 6$ h.

$$n = \sum \frac{(pl)i}{al} \times \frac{1}{L}$$

Here, n refers to the number of live *Candida albicans* persisters per unit length after exposed to drugs, pl is the total length of *Candida albicans* persisters under fluorescent microscope, and al represents the average length of a *Candida albicans* cell, which is usually $4.5 \mu\text{m}$. L is the total length of microchannels, and each microchannel is 2 cm long.

Fungicidal effects were evaluated based on the number of surviving *Candida albicans* persisters remaining in the microchannels. 20 replicates were done for each drug, and the average value of 20 tests was used to describe the final efficacy. We observed significant efficacy differences between amphotericin B by itself versus the amphotericin B and small molecule combinations (Fig. 4).

Using high-throughput screening, we discovered 10 small molecule compounds (out of 50,520) that increase amphotericin B's fungicidal effect on *Candida albicans* persisters by over 30% (Fig. 5). Our tentative hypothesis is that the 10 amphotericin B combinations displayed increased efficacy because of the presence of azolates and azolate derivatives. Compounds 2,4,5,6, and 9 contain benzothiazole structures (Fig. 5) while compounds 4,5, and 6 contain 3 azole structures. Compound 3 and 7 are imidazoles. Compound 9 and 10 are thiazoles. Several of the most potent clotrimazole potentiators contain a 1,3-benzothiazole scaffold (compounds 2,4,5,6 and 9)^{17,30}. The antifungal properties of benzothiazole compounds have been reported³¹. One example is 6-amino-2-n-pentylthiobenzothiazole, which has been found to inhibit *Candida* filamentation³². Unfortunately, some benzothiazoles exhibit cytotoxicity, such as 1,3-benzothiazole. However, such a liability may be irrelevant for topical applications. Alternatively, it may be possible to reduce the scaffold's cytotoxicity by synthesizing and testing chemical analogues¹⁷. Other azole potentiators have been found to function via diverse mechanisms, including inhibiting calcineurin³³, HSP90^{34,35}, and drug efflux pumps^{36,37}.

It is also possible that the amphotericin B combinations contain conjugated systems containing nitrogen and sulfur atoms which enhance the fungicidal effect. Two examples of π - π conjugated systems are $\text{CH}_2=\text{CH}-\text{CH}=\text{O}$

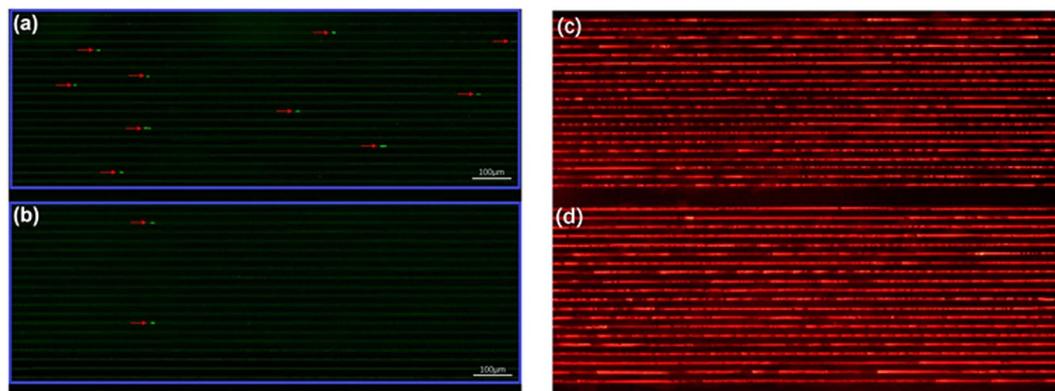


Figure 4. Surviving *Candida albicans* persists after treatment with (a) Normal amphotericin B and (b) Amphotericin B in combination with small molecule compounds. The red arrows point to the persisters. (c,d) Are the LIVE-DEAD[®] fluorescent images corresponding to (a,b).

Structure	ID Number	Mol. Weight	Mol. Formula	Percent increase in efficacy	
				Microfluidic chip*	384 well plate#
	F0288-0118	296.364	C18H20N2O2	35.99%	37%
	F0308-0828	408.513	C23H24N2O3S	35.11%	35%
	F0371-0041	446.493	C25H19FN2O3S	43.83%	44%
	F0665-0283	545.676	C28H27N5O3S2	32.45%	30%
	F0665-0476	412.528	C20H20N4O2S2	34.74%	34%
	F0665-0496	517.623	C26H23N5O3S2	38.25%	38%
	F0668-0385	370.419	C19H16F2N4S	33.33%	33%
	F0694-0141	351.445	C18H13N3OS2	40.66%	40%
	F0760-0864	506.167	C19H10Br2N2O3S	33.33%	31%
	F0760-2126	427.904	C21H18ClN3O3S	46.63%	47%

* It is the average value of 20 replicated tests in the microchannels.
It is the average value of 16 replicated tests on 384-well plate.

Figure 5. 10 small molecule compounds improve the fungicidal effect of amphotericin B by more than 30% against *Candida albicans* persister cells.

and $\text{CH}_2=\text{CH}-\text{C}\equiv\text{N}$. The presence of such conjugations may alter amphotericin B's three-dimensional structure. The altered structure may inhibit *Candida albicans*' drug efflux pump, thereby keeping amphotericin B in the cell.

Discussion

The development of new drugs is a lengthy process, and high-throughput microfluidic platform screening is an innovative method that can be utilized to advance drug research. It's a highly-sensitive method that provides rapid, accurate results. In addition, its setup is compact. As shown in Table 1, the highly integrated microchannels enabled microfluidic chips to process over 1000 kinds of drugs simultaneously, and 50520 drugs could be screened in one week. Traditional HTS tests the efficacy percentage by Alamar Blue, which based on oxidation-reduction reaction. It is tedious to calculate the absorbance at 570 nm and 600 nm wavelengths with complicated steps³⁸. The microfluidic chips ensure the passage of *Candida albicans* that pass through and line up one-by-one in the microchannels, which makes the drug evaluation more convenient and accurate because of the straightforward calculation and detection of small probability of residual bacteria.

Conclusion

Through our high-throughput microfluidic chips, we have erected a novel platform for *Candida albicans* antifungal drug screening. We were able to screen 50,520 potential drugs easily and accurately. Ten of these compounds improved the efficacy of the anti-*Candida albicans* drug amphotericin B, increasing it by more than 30%. This discovery may contribute to the clinical treatment of refractory, recurrent *Candida albicans* infections and may also create a new path for drug development.

	Microfluidic chips	Traditional HTS Methods (384w-MTP as example) ³⁹
Principle	Direct living cells counting	AlamarBlue Redox method, or 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) method ^{40,41} , Indirect method
Quantity	100 kinds of drug	20 kinds of drug
Operation time for 50520 drugs screening	1 week	125 days
Observation	Visually, intuitively	Indirectly
Reagent	Totally <1 μ L	>100 μ L/well

Table 1. Comparison of our custom-designed microfluidic chips and traditional HTS methods.

Materials and Methods

Cell culture and Regents. *Candida albicans* 3153A is wildtype strained. The *FLO8* plasmid with GFP was purchased in BioSune Biology Company, Shanghai, China. Stock cultures of *Candida albicans* strains were routinely cultured in YPD (1% yeast extract, 2% peptone, 2% glucose) solid medium containing 1.5% agar at 37°C for 24 to 48 h. *Candida albicans* Yeast inocula cells were prepared by transferring a single colony into YPD medium with overnight incubation at 37°C in an incubator shaker at about 100 rpm. Cells were harvested by centrifugation at 6,000 g for 3 min and washed twice in sterile phosphatebuffered saline (PBS [pH 7.2 to 7.4]). Then the cells were resuspended in RPMI 1640 medium (Gibco) with L-glutamine, buffered to pH 7.0 with 0.165 M morpholinepropanesulfonic acid (MOPS [Sigma-Aldrich]) and adjusted to the desired density of $2 \times 10^3 \sim 1 \times 10^4$ ⁴². CLSI M27-A3 microdilution methodology was used to test *in vitro* susceptibility to amphotericin B (Amresco)⁴³. LIVE-DEAD® Funga Light yeast viability kit was purchased from Invitrogen. The link for 50,520 different small molecule compounds is: <http://www.cncl.org.cn/>.

New anti candida albicans drugs preparation. The initial concentration of each small molecule compound solution (dissolved in DMSO) was 1000 μ g/mL and total of 1 μ L. In order to ensure the maximum concentration of dissolved <1% compounds and the DMSO final effect on *Candida albicans*, we added in the original compounds on the basis of 2 μ L DMSO and 7 μ L RPMI-1640 dilution of small molecules compound. We then mixed 10 μ L of each small-molecular-compound with 100 μ L of 2- μ g/mL amphotericin B, following the NCCLS M27-A2 microdilution method⁴⁴. The resulting concentration of each new amphotericin B combination was 3.5 μ g/mL. We use injection pump (Longer Precision Pump Co., Ltd. China) to inject new drugs.

PDMS microfluidic chips fabrication. The microfluidic chip was fabricated by multilayer soft lithography technique using polydimethylsiloxane (PDMS; Sylgard 184A and B). One photomask was first generated with microscal patterns designed by computer-aided design software L-Edit and transferred on Cr mask substrate. The positive photoresist AZ5214 was spun on a 6-inch silicon wafer, and got exposed under UV light with Cr mask on top. After the photoresist was development, the silicon wafer was heated at 120°C for 5 minutes to remove the moisture, and then Oxford 180 dry etcher was used to etch the silicon channels. After silicon etching, the photoresist was removed by acetone and Isopropanol. The silicon wafer with microchannels then was used hard mode.

To make a PDMS chip, a ratio of Sylgard A:B = 10:1 by weight was mixed thoroughly and poured onto the silicon mold in a Petri dish at a thickness of approximately 5 mm. After all of the bubbles in the PDMS were vacuumed out, the Petri dish containing the PDMS was cured at 80°C for 60 minutes. Then the PDMS layer was then peeled off the mold and perforated to produce inlets and outlets for cell solution and medicine loading. Both the PDMS chip and clean glass slide were treated with oxygen plasma at 50 W for 5 minutes, and then they were lined up and bonded permanently.

SEM sample preparation. We referred to the publication by W. Krzysciak to prepare the SEM sample preparation⁴⁵ PDMS microfluidic chip was separated from the glass substrate, and *Candida albicans* cells were kept in the microchannels because they were adhesive. The PDMS chip was washed three times with 0.1 M PBS, and treated with 1% osmium tetroxide for 1 h at room temperature. Then the PDMS was fixed with 2% glutaraldehyde for 2 h at room temperature. The PDMS was dehydrated using graded ethanol (10%, 30%, 50%, 70% and 100%) and putted in vacuum freeze-drying machine for 8 hours. Finally, the PDMS was mounted on microscope stubs and sputtered with a thin layer of gold, and then examined using a Leica S440 scanning electron microscope.

Comments. The photo of injection pump in Fig. 1h has been provided by Longer Precision Pump Co., Ltd. China, where we bought the injection pump. We got the company's authorization to use this photo.

References

- Guarner, J. & Brandt, M. E. Histopathologic diagnosis of fungal infections in the 21st century. *Clinical microbiology reviews* **24**, 247–280 (2011).
- Hameed, S. & Fatima, Z. Novel regulatory mechanisms of pathogenicity and virulence to combat MDR in *Candida albicans*. *International journal of microbiology* **2013**, 1–10 (2013).
- Naggie, S. & Perfect, J. R. Molds: hyalohyphomycosis, phaeohyphomycosis, and zygomycosis. *Clinics in chest medicine* **30**, 337–353 (2009).
- Silva, N. C., Nery, J. M. & Dias, A. L. Aspartic proteinases of *Candida* spp.: role in pathogenicity and antifungal resistance. *Mycoses* **57**, 1–11 (2014).
- Hutter, I., Mueller, E., Kristiansen, P. M., Kresak, S. & Tiefenauer, L. Polymer-based microfluidic device for measuring membrane protein activities. *Microfluidics and Nanofluidics* **14**, 421–429, <https://doi.org/10.1007/s10404-012-1061-0> (2013).

6. Perlin, D. S. Mechanisms of echinocandin antifungal drug resistance. *Annals of the new York Academy of Sciences* **1354**, 1–11 (2015).
7. Ruhnke, M. *et al.* Emergence of fluconazole-resistant strains of *Candida albicans* in patients with recurrent oropharyngeal candidosis and human immunodeficiency virus infection. *Journal of Clinical Microbiology* **32**, 2092–2098 (1994).
8. Bigger, J. Treatment of Staphyloeoceal Infections with Penicillin by Intermittent Sterilisation. *Lancet*, 497–500 (1944).
9. Lewis, K. Persister cells, dormancy and infectious disease. *Nature Reviews Microbiology* **5**, 48 (2007).
10. LaFleur, M. D., Kumamoto, C. A. & Lewis, K. *Candida albicans* biofilms produce antifungal-tolerant persister cells. *Antimicrobial agents and chemotherapy* **50**, 3839–3846 (2006).
11. Costa, E., Silva, S., Tavaría, F. & Pintado, M. Antimicrobial and antibiofilm activity of chitosan on the oral pathogen *Candida albicans*. *Pathogens* **3**, 908–919 (2014).
12. Garbino, J. *et al.* Prevention of severe *Candida* infections in nonneutropenic, high-risk, critically ill patients: a randomized, double-blind, placebo-controlled trial in patients treated by selective digestive decontamination. *Intensive care medicine* **28**, 1708–1717 (2002).
13. Pfaller, M., Diekema, D., Procop, G. & Rinaldi, M. Multicenter comparison of the VITEK 2 antifungal susceptibility test with the CLSI broth microdilution reference method for testing amphotericin B, flucytosine, and voriconazole against *Candida* spp. *Journal of clinical microbiology* **45**, 3522–3528 (2007).
14. Yamashita, I. & Fukui, S. Mating signals control expression of both starch fermentation genes and a novel flocculation gene *FLO8* in the yeast *Saccharomyces*. *Agricultural and biological chemistry* **47**, 2889–2896 (1983).
15. Cao, F. *et al.* The *FLO8* transcription factor is essential for hyphal development and virulence in *Candida albicans*. *Molecular biology of the cell* **17**, 295–307 (2006).
16. Ehrenkauser, G. M., Suresh, S., Solow-Cordero, D. & Singh, U. High-Throughput Screening of Entamoeba Identifies Compounds Which Target Both Life Cycle Stages and Which Are Effective Against Metronidazole Resistant Parasites. *Front Cell Infect Microbiol* **8**, 276, <https://doi.org/10.3389/fcimb.2018.00276> (2018).
17. LaFleur, M. D., Lucumi, E., Napper, A. D., Diamond, S. L. & Lewis, K. Novel high-throughput screen against *Candida albicans* identifies antifungal potentiators and agents effective against biofilms. *J Antimicrob Chemother* **66**, 820–826, <https://doi.org/10.1093/jac/dkq530> (2011).
18. Sista, R. *et al.* Development of a digital microfluidic platform for point of care testing. *Lab on a Chip* **8**, 2091–2104 (2008).
19. Palková, Z., Váňová, L., Valer, M. & Preckel, T. Single-cell analysis of yeast, mammalian cells, and fungal spores with a microfluidic pressure-driven chip-based system. *Cytometry Part A: The Journal of the International Society for Analytical Cytology* **59**, 246–253 (2004).
20. Boehm, D. A., Gottlieb, P. A. & Hua, S. Z. On-chip microfluidic biosensor for bacterial detection and identification. *Sensors and Actuators B: Chemical* **126**, 508–514 (2007).
21. Safaviéh, M. *et al.* Advances in *Candida* detection platforms for clinical and point-of-care applications. *Critical reviews in biotechnology* **37**, 441–458 (2017).
22. Balaban, N. Q., Merrin, J., Chait, R., Kowalik, L. & Leibler, S. Bacterial persistence as a phenotypic switch. *Science* **305**, 1622–1625 (2004).
23. Smith, A. M. *et al.* A high-throughput turbidometric assay for screening inhibitors of protein disulfide isomerase activity. *J Biomol Screen* **9**, 614–620, <https://doi.org/10.1177/1087057104265292> (2004).
24. Widder, M. *et al.* A modified 384-well-device for versatile use in 3D cancer cell (co-)cultivation and screening for investigations of tumor biology *in vitro*. *Eng Life Sci* **18**, 132–139, <https://doi.org/10.1002/elsc.201700008> (2018).
25. Mayr, L. M. & Bojanic, D. Novel trends in high-throughput screening. *Current Opinion in Pharmacology* **9**, 580–588, <https://doi.org/10.1016/j.coph.2009.08.004> (2009).
26. Fan, Y., Nguyen, D. T., Akay, Y., Xu, F. & Akay, M. Engineering a brain cancer chip for high-throughput drug screening. *Scientific reports* **6**, 25062 (2016).
27. Zhang, J. *et al.* Real-time immuno-polymerase chain reaction in a 384-well format: detection of vascular endothelial growth factor and epidermal growth factor-like domain 7. *Anal Biochem* **463**, 61–66, <https://doi.org/10.1016/j.ab.2014.06.024> (2014).
28. Sjogren, A. K. *et al.* A novel multi-parametric high content screening assay in ciPTEC-OAT1 to predict drug-induced nephrotoxicity during drug discovery. *Arch Toxicol* **92**, 3175–3190, <https://doi.org/10.1007/s00204-018-2284-y> (2018).
29. Vriend, J. *et al.* Screening of Drug-Transporter Interactions in a 3D Microfluidic Renal Proximal Tubule on a Chip. *AAPS J* **20**, 87, <https://doi.org/10.1208/s12248-018-0247-0> (2018).
30. Kashyap, S. J. *et al.* Thiazoles: having diverse biological activities. *Medicinal Chemistry Research* **21**, 2123–2132 (2012).
31. Bujđáková, H., Kuchta, T., Sidóová, E. & Gvozďáková, A. Anti-*Candida* activity of four antifungal benzothiazoles. *FEMS microbiology letters* **112**, 329–333 (1993).
32. Fabry, S. *et al.* Inhibition of germ tube formation, filamentation and ergosterol biosynthesis in *Candida albicans* treated with 6-amino-2-n-pentylthiobenzothiazole. *Folia microbiologica* **44**, 523–526 (1999).
33. Sanglard, D., Ischer, F., Marchetti, O., Entenza, J. & Bille, J. Calcineurin A of *Candida albicans*: involvement in antifungal tolerance, cell morphogenesis and virulence. *Molecular microbiology* **48**, 959–976 (2003).
34. Cowen, L. E. Hsp90 orchestrates stress response signaling governing fungal drug resistance. *PLoS pathogens* **5**, e1000471 (2009).
35. Dongari-Bagtzoglou, A., Kashleva, H., Dwivedi, P., Diaz, P. & Vasilakos, J. Characterization of mucosal *Candida albicans* biofilms. *PLoS one* **4**, e7967 (2009).
36. Lamping, E. *et al.* Characterization of three classes of membrane proteins involved in fungal azole resistance by functional hyperexpression in *Saccharomyces cerevisiae*. *Eukaryotic cell* **6**, 1150–1165 (2007).
37. Harriott, M. M., Lilly, E. A., Rodriguez, T. E., Fidel, P. L. & Noverr, M. C. *Candida albicans* forms biofilms on the vaginal mucosa. *Microbiology* **156**, 3635–3644, <https://doi.org/10.1099/mic.0.039354-0> (2010).
38. Punithavathy, P. M., Nalina, K. & Menon, T. Antifungal susceptibility testing of *Candida tropicalis* biofilms against fluconazole using calorimetric indicator resazurin. *Indian Journal of Pathology and Microbiology* **55**, 72–74, <https://doi.org/10.4103/0377-4929.94861> (2012).
39. Escher, B. I., Neale, P. A. & Villeneuve, D. L. The advantages of linear concentration-response curves for *in vitro* bioassays with environmental samples. *Environ Toxicol Chem* **37**, 2273–2280, <https://doi.org/10.1002/etc.4178> (2018).
40. Tang, X. L., Suo, H., Zheng, R. C. & Zheng, Y. G. An efficient colorimetric high-throughput screening method for synthetic activity of tyrosine phenol-lyase. *Anal Biochem*, <https://doi.org/10.1016/j.ab.2018.08.026> (2018).
41. Zhang, R. *et al.* Identification of novel inhibitors of histone acetyltransferase hMOF through high throughput screening. *Eur J Med Chem* **157**, 867–876, <https://doi.org/10.1016/j.ejmech.2018.08.026> (2018).
42. Sun, J., Liu, X., Jiang, G. & Qi, Q. Inhibition of nucleic acid biosynthesis makes little difference to formation of amphotericin B-tolerant persisters in *Candida albicans* biofilm. *Antimicrobial agents and chemotherapy* **59**, 1627–1633 (2015).
43. Nobile, C. J. & Mitchell, A. P. Regulation of Cell-Surface Genes and Biofilm Formation by the *C. albicans* Transcription Factor Bcr1p. *Current Biology* **15**, 1150–1155 (2005).
44. Silici, S. & Koc, A. Comparative study of *in vitro* methods to analyse the antifungal activity of propolis against yeasts isolated from patients with superficial mycoses. *Letters in applied microbiology* **43**, 318–324 (2006).
45. Krzysciak, W. *et al.* Effect of a *Lactobacillus Salivarius* Probiotic on a Double-Species *Streptococcus Mutans* and *Candida albicans* Caries Biofilm. *Nutrients* **9**, <https://doi.org/10.3390/nu9111242> (2017).

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Author Contributions

Lin Han designed the study, and Qingguo Qi brought up the idea; Le Qiang carried out most of the experiments and wrote this manuscript; Jing Guo was in charge of small molecules. Yingkuan Han participated in part of the results analysis; Jianfeng Jiang was in charge of Scanning Electron Microscope (SEM) sample preparation, and Xiaowen Su conducted SEM operation. Hong Liu provided valuable suggestions for the experiments.

Additional Information

Competing Interests: The authors declare no competing interests.

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