# SCIENTIFIC REPORTS

Received: 21 February 2018 Accepted: 15 August 2018 Published online: 28 August 2018

## **OPEN** Seaweed assemblages under a climate change scenario: Functional responses to temperature of eight intertidal seaweeds match recent abundance shifts

Cristina Piñeiro-Corbeira 1, Rodolfo Barreiro 1, Javier Cremades<sup>1</sup> & Francisco Arenas 2

Field evidence is essential to assess the consequences of climate change but a solid causal link often requires additional information obtained under controlled laboratory conditions. Additionally, the functional response to temperature may also help to discriminate species potentially more vulnerable to warming. Using a highly resolved temperature gradient, we examined the temperature dependence of photosynthesis and respiration in eight intertidal seaweeds that recently followed opposite abundance trends in NW Iberia. The temperature dependence of photosynthesis was consistently different between the macroalgae that increased and those that decreased their abundance in the last decade and a half, with photosynthesis twice more sensitive in the upward group. Unlike photosynthesis, the temperature dependence of respiration was unrelated to the abundance trend group, implying that the net metabolic scaling with temperature varied between the two groups of seaweeds. Overall, our results provide experimental support to the role of temperate as a likely driver of the changes in abundance recorded by field-monitoring studies. They also suggest that the temperature dependence of photosynthesis and respiration assessed in short-term experiments may serve as a biomarker of the potential vulnerability of some seaweed to the consequences of water warming.

Macroalgae are the dominant primary producers in coastal areas, providing essential ecosystem functions and services such as habitat and/or food for many marine organisms<sup>1</sup>. Thus, shifts in the composition of seaweed assemblages can have large consequences for the entire coastal community<sup>2,3</sup>. Temperature is widely acknowledged as a key determinant of the geographic range of marine benthic macroalgae and the large-scale biogeographic distribution pattern of many seaweeds has been explained by coupling species' thermal traits with local seawater temperatures<sup>1,4</sup>.

Seawater temperatures are changing worldwide and the global oceans warmed an average of 0.1 °C per decade over the last 40 years<sup>5</sup>. Although a global phenomenon, the warming rate varies among regions/latitudes<sup>6</sup>. For example, sea surface temperature (SST) near the coast along the Iberian Peninsula seems to have warmed somewhat faster than the global average<sup>7</sup>. Understandably, the consequences of ocean warming for marine ecosystems have received increasing attention in recent years. There is a growing evidence that increasing temperatures have direct physical consequences for marine organisms and that warming, jointly with other anthropogenic stressors, is already affecting marine communities<sup>3,8–10</sup>.

In the particular case of the seaweeds, range shifts and abundance changes have been documented for various regions around the world<sup>10-14</sup>. In the Northeast Atlantic, some northern cold-adapted canopy forming seaweeds decreased in abundance in recent years while other meridional species with an affinity for warmer temperatures increased in regions such as North Spain, French Brittany, or British Isles<sup>10,11,13,15,16</sup>. Further North, warming seems responsible for the increased kelp biomass and the shift to a shallower depth of the biomass peak

<sup>1</sup>BioCost Research Group, Facultad de Ciencias and Centro de Investigaciones Científicas Avanzadas (CICA), Universidad de A Coruña, 15071, A Coruña, Spain. <sup>2</sup>CIIMAR, Centro Interdisciplinar de Investigação Marinha e Ambiental, Terminal de Cruzeiros do Porto de Leixões, Av. General Norton de Matos s/n, 4450-208, Matosinhos, Portugal. Correspondence and requests for materials should be addressed to C.P.-C. (email: c.pcorbeira@udc.es)

recently reported for one kelp forest at the border of the Artic Ocean<sup>17</sup>. Elsewhere, a rapid climate-driven regime shift has been reported for an extensive portion of the Australian temperate reef communities, where mixed kelps-fucacean canopies were lost after a succession of extreme heat waves and were replaced by persistent seaweed turfs with consequences for fish and marine invertebrate assemblages<sup>14</sup>. Previously, large-scale destructions of kelp forests were observed along the Pacific coasts of America after extreme climatic events but, unlike Australia, these forests recovered as environmental conditions returned to normal<sup>18,19</sup>

Range shifts and/or abundance changes are essential evidences for a realistic assessment of the consequences of rising temperatures for macroalgae<sup>3</sup>. However, field-monitoring studies may still be insufficient for establishing causal relationships or to foresee the effects of an even warmer future. Our inference ability can be greatly improved by coupling field observations with analyses of the functional response of seaweeds to increasing temperature under controlled laboratory conditions<sup>20–23</sup>. In thermo-conformer organisms like seaweeds, ambient temperature influences biochemical reactions with direct consequences for key physiological rates such as photosynthesis and respiration<sup>24</sup>. Respiration generally increases with temperature without reaching a clear optimum, while photosynthesis typically rise up to a plateau at a maximal level and then rapidly declines near the upper critical temperature<sup>1,25</sup>. Temperature optima for photosynthesis tend to be in agreement with the distribution range and species from colder latitudes often reach peak values at lower temperatures than temperate or tropical ones<sup>4,25–28</sup>.

Another thermal trait that also varies among species and even among populations, is the temperature dependence of metabolic rates<sup>1</sup>. The latter is typically characterized with  $Q_{10}$ , the factor by which a rate increases for a 10 °C rise in temperature. However, parameterizing in 10 °C increments introduces a distortion and the Boltzmann-Arrhenius model derived from chemical kinetics is a better alternative to characterize the temperature dependence<sup>29,30</sup>. In this formulation, the temperature dependence is given by the slope of the Boltzmann-Arrhenius model, which with sign reversed is known as the activation energy  $E_a^{31}$ . Unlike  $Q_{10}$ ,  $E_a$  is temperature independent and avoids the distortion introduced by parameterizing in 10 °C increments<sup>29,30</sup>. Thermal dependence in seaweeds can be greatly affected by ambient temperature and metabolic rates often increase more in response to increasing temperature when the individuals grew in cooler environments<sup>32,33</sup>. As a result, a higher temperature dependence has been reported for individuals collected in winter compared to those sampled in summer<sup>34</sup>, or in populations from cooler locations along a latitudinal gradient<sup>35</sup>. Individuals from cooler environments also experienced faster drops in net photosynthesis when they were exposed to temperatures that surpassed their temperature optimum<sup>35</sup>.

Despite the growing number of reports on the impact of global change on the distribution and abundance of seaweeds, few studies have compared the thermal functional response of seaweeds that followed divergent abundance trends in recent years<sup>22,36</sup>. This study examines the physiological response to an experimental temperature gradient in eight intertidal seaweeds that changed their patterns of site occupancy differently in NW Iberia in the last decades: four that decreased their frequency and four that increased<sup>10</sup>. To provide further support for the role of temperature as a decisive driver of the observed shifts, we tested whether there were consistent differences in the functional responses to temperature shown by each set of seaweeds. The seaweeds targeted in this study are major members of the seaweed assemblage in NW Iberia<sup>37,38</sup>; a better knowledge of the actual drivers of their recent frequency shifts seems highly relevant to predict the future configuration of the intertidal ecosystem.

#### Material and Methods

**Sample collection.** *Mastocarpus stellatus* (Stackhouse) Guiry, *Chondrus crispus* Stackhouse, *Himanthalia elongata* (Linnaeus) S.F.Gray and *Fucus vesiculosus* Linnaeus were chosen as representatives of seaweeds with a downward trend in site occupancy. A previous survey showed that these species had 48% (*M. stellatus*) to 70% (*C. crispus, H. elongata, F. vesiculosus*) lower site occupancy in 2014 than in 1998/99<sup>10</sup>. On the other hand, *Corallina caespitosa* R.H.Walker, J.Brodie & L.M.Irvine, *Bifurcaria bifurcata* R. Ross, *Cystoseira tamariscifolia* (Hudson) Papenfuss and *Cystoseira baccata* (S.G.Gmelin) P.C.Silva were selected as representatives of seaweeds that followed an upward trend because they experienced gains of 11% (*C. tamariscifolia*) to 75% (*C. caespitosa*) in the frequency of occupied sites over the same period of time<sup>10</sup>.

Live fronds of the eight macroalgal species were collected from intertidal locations in NW Iberia in February 2016. The study included Fucales with apical growth (*H. elongata, F. vesiculosus, C. baccata, C. tamariscifolia, B. bifurcata*) and multiaxial red algae with pseudoparenchymatous tissues where the growth of new tissue is not restricted to a meristematic region (*M. stellatus, C. crispus, C. caespitosa*). To guarantee that the functional responses were consistently recorded in metabolically active tissue, samples consisted of whole fronds in the case of the pseudoparenchymatous red algae and the moderately sized *B. bifurcata*. By contrast, the samples of large-sized Fucales consisted of moderately sized young plants (*F. vesiculosus*), plants with young receptacles (*H. elongata*), or distal, young apical fronds (*C. tamariscifolia* and *C. baccata*) were used for convenience (frond size 15 cm long in all cases). As a result, whole fronds samples and young frond/tissue regions were evenly partitioned between the two trend sets compared in our experimental design. Samples were transported in a portable cool box to the laboratory where grazers and epiphytic organism were carefully removed. For each species, a subset of evenly sized specimens was acclimated to laboratory conditions before the experiment during two weeks with aerated sea water at around 13 °C; water was renewed every two days along the acclimation period to avoid nutrient depletion.

**Functional responses: estimation of productivity and respiration rates.** To examine the functional response of the target species to rising temperatures, we assessed the respiration and primary productivity rates over a thermal gradient under controlled laboratory conditions. Respiration and net primary productivity were determined throughout oxygen fluxes by continuously recording (every 30 seconds) the concentration of dissolved  $O_2$  with a HQ40D oxygen probe (Hach Lange<sup>®</sup>) within incubation chambers. Seaweed individuals were

incubated under temperature-controlled conditions in custom-made, sealed methacrylate incubation units (2 L) equipped with a submersible pump to homogenize the medium. Lighting was provided by a variable number of 30 W Biolux Osram® fluorescent lamps. The setup allowed the simultaneous and independent incubation of three samples, successively exposed for 20 min (30 min for dark conditions) to seven light intensities (0, 30,108, 237, 557, 971, 1515  $\mu$ mol photons  $\cdot$  m<sup>-2</sup>  $\cdot$  s<sup>-1</sup>). Light intensity ranged from light-limited conditions (dark incubation) to light-saturation and allowed us to calculate photosynthesis-irradiance (P-E) curves. Independent P-E curves were constructed at nine temperatures for each species. The experiment included water temperatures normally experienced by seaweeds in NW Iberian along the year (12 °C, 15 °C, 18 °C and 21 °C), but also extremes beyond the limits usually seen in the region (6 °C, 9 °Č, 24 °C, 27 °C and 30 °C)<sup>39,40</sup>. Samples were acclimated to incubation-temperature for 12h before each experiment. P-E curves for each combination of species and temperature were recorded following a strictly randomized order to avoid any bias. In addition, filtered sea water (5 µm) was used to prevent any spurious influence of phytoplankton on photosynthesis/respiration estimates. The effectiveness of the filtering procedure was repeatedly confirmed along the experiment by recording O<sub>2</sub> changes in control (no seaweed) incubation units. After incubation, samples were oven-dried at 45 °C for 48 h to estimate dry biomass. Dark respiration  $(R_d)$  and net productivity (NP) rates were estimated as the slope of the linear regression between  $O_2$  concentration and time; gross photosynthesis (GP) was calculated by adding the absolute value of  $R_d$ to estimates of NP at each light intensity. All rates were expressed as mg  $O_2 \cdot g^{-1} DW \cdot h^{-1}$ .

The maximum rate of gross photosynthesis at saturating light ( $GP_{max}$ ) and the photosynthetic efficiency at low irradiance ( $\alpha$ ) were determined by fitting the model of Eilers and Peeters<sup>41</sup>:

$$GP = 1/(E^2/\alpha \times E_{opt}^2) + (E/GP_{max}) - (2E/\alpha \times E_{opt}) + (1/\alpha)$$
(1)

where  $E_{opt}$  is the optimum light intensity. The maximum rate of net photosynthesis ( $NP_{max}$ ) was calculated as  $GP_{max} - R_d$ . Model fitting was done by non-linear regression with the help of the package Phytotools<sup>42</sup> for R<sup>43</sup>.

 $T_{opt}$  and rate at  $T_{opt}$  of  $NP_{max}$ ,  $GP_{max}$  and  $R_d$  were estimated by fitting the three parameter Gaussian model recommended by June, *et al.*<sup>44</sup>:

$$y = y_{opt} e^{(-0.5((T - T_{opt})/b)^2)}$$
(2)

where y is the rate  $(NP_{\text{max}}, GP_{\text{max}}, \text{or } R_d)$  estimated at each temperature T,  $y_{\text{opt}}$  is the value of the rate at the optimum temperature  $(T_{\text{opt}})$  and b is the steepness of the curve. The Gaussian model was fitted with the non-linear routine implemented in Statgraphics Centurion XVI (StatPoint Technologies, Inc.).

**Activation Energy from the Arrhenius equation.** The temperature dependence of photosynthesis and dark respiration was described with the Boltzmann-Arrhenius model:

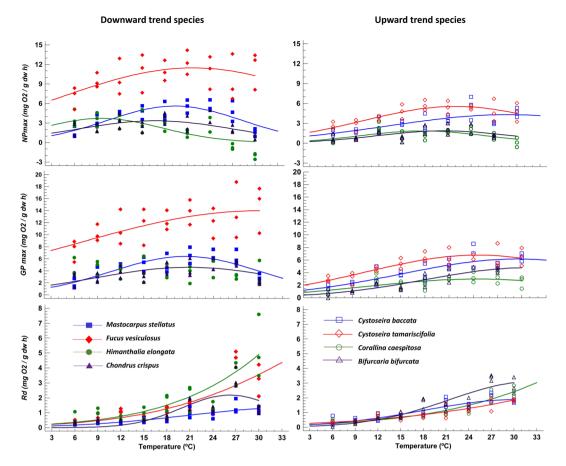
$$R = R_0 e^{-E_d/kT} \tag{3}$$

where  $R_0$  is a scaling coefficient and  $e^{-E_d/kT}$  is the Boltzmann factor that describes an exponential relationship between rate and temperature (*T* in kelvin). In this model, the temperature dependence of the trait is measured by the activation energy  $E_a$ , while *k* is the Boltzmann's constant<sup>31</sup>. Accordingly, we calculated the  $E_a$  of  $GP_{max}$ ,  $NP_{max}$  and  $R_d$  for each species. Since the Arrhenius model describes an exponential relationship while the temperature response function can be hump-shaped, we fitted the model only to the approximately exponential rise phase of our rate-temperature curves<sup>45</sup>. The rising component of each curve was delimited by iteratively removing rate measurements at the upper terminal temperatures until monotonicity was observed in the response. To do this, a quadratic function was initially fitted to the relationship between the log-transformed rate and 1/kT using ordinary least squares (OLS) regression. If the *p*-value of the model coefficient for the order 2 term was significant, the response was considered hump-shaped, the highest terminal temperature removed and the procedure repeated until the model coefficient for the order 2 term was non-significant. Then, a linear model was fitted to the data and we used R<sup>2</sup>-adjusted values to assess whether the linear model fitted better than the quadratic one; if necessary, the highest terminal temperature was removed until the R<sup>2</sup>-adjusted value was equal or larger for the linear than for the quadratic model. The whole procedure was accomplished with the help of Statgraphics Centurion XVI (StatPoint Technologies, Inc.).

**Data analysis.** For each physiological rate ( $GP_{max}$ ,  $NP_{max}$ ,  $R_d$  and  $\alpha$ ), we used conditional sums of squares as a global test to assess whether the activation energy differed among species<sup>46</sup>. Then, we calculated 95% confidence intervals corrected for multiple comparisons with the Bonferroni method to determine species or sets of species with significantly different activation energies. Bonferroni-adjusted 95% confidence intervals were likewise used to assess interspecific differences in  $T_{opt}$  and in rate estimates at  $T_{opt}$ .

#### Results

**Effects of temperature on photosynthesis and respiration.** Irrespective of temperature, both net and gross maximum photosynthesis were conspicuously higher in *F. vesiculosus* than in other seaweed species. Metabolic estimates in *F. vesiculosus* ranged  $5.1-14.2 \text{ mg O}_2 \cdot \text{g}^{-1}\text{DW} \cdot \text{h}^{-1}$  for  $NP_{\text{max}}$  and  $5.5-18.7 \text{ mg O}_2 \cdot \text{g}^{-1}\text{DW} \cdot \text{h}^{-1}$  for  $GP_{\text{max}}$  compared to  $2.6-7.0 \text{ mg O}_2 \cdot \text{g}^{-1}\text{DW} \cdot \text{h}^{-1}$  for  $NP_{\text{max}}$  and  $0.0-8.7 \text{ mg O}_2 \cdot \text{g}^{-1}\text{DW} \cdot \text{h}^{-1}$  for  $GP_{\text{max}}$  in other algae (Fig. 1). Other than *F. vesiculosus*, the productivity rates were very similar in downward and upward seaweeds. Regardless of the species, the response of  $NP_{\text{max}}$  and  $GP_{\text{max}}$  to temperature was consistently unimodal; photosynthesis increased gradually with temperature up to an optimum and decreased afterwards. Nonetheless and despite the consistently unimodal shape, there were differences between species. For example, the fall phase



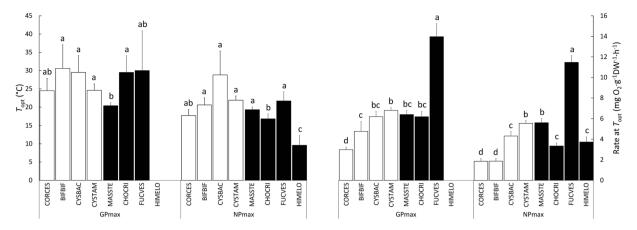
**Figure 1.** Temperature dependence of  $NP_{max}$  (top),  $GP_{max}$  (middle) and  $R_d$  (bottom) in seaweeds that showed divergent downward (left) and upward (right) abundance trends in NW Iberia along the last decade. Lines are non-linear fits of a three-parameter Gaussian model (see text for further detail). Due to poor fitting, no line is shown for  $GP_{max}$  in *H. elongata*.

Occurrence		<i>GP</i> <sub>max</sub>		R <sub>d</sub>		NP <sub>max</sub>		
trend	Species <sup>a</sup>	T <sub>opt</sub> Rate at T <sub>opt</sub>		T <sub>opt</sub>	Rate at T <sub>opt</sub>	T <sub>opt</sub>	Rate at T <sub>opt</sub>	
Upward	CORCES	24.5±3.39	3.0±0.25	ND	$2.4 \pm 0.22$	$17.7 \pm 1.62$	$1.9\pm0.27$	
Upward	BIFBIF	30.6±6.47	$4.8 \pm 0.94$	ND	$2.8 \pm 0.32$	$20.56 \pm 1.94$	$1.9\pm0.25$	
Upward	CYSBAC	$29.5 \pm 4.58$	6.2±0.51	ND	$1.8 \pm 0.06$	$28.8 \pm 6.54$	$4.3\pm0.44$	
Upward	CYSTAM	$24.6 \pm 1.73$	6.8±0.28	ND	$1.8\pm0.05$	$21.9 \pm 1.13$	$5.5\pm0.28$	
Downward	MASSTE	$20.4 \pm 0.80$	$6.4 \pm 0.40$	ND	$1.2 \pm 0.13$	$19.3 \pm 0.76$	$5.6\pm0.39$	
Downward	CHOCRI	$29.5\pm4.58$	6.2±0.51	$26.2 \pm 1.34$	$2.2 \pm 0.25$	$16.8 \pm 1.37$	3.4±0.33	
Downward	FUCVES	$30.0 \pm 10.89$	$14.0 \pm 1.26$	ND	3.2±0.61	$21.7 \pm 2.49$	$11.5\pm0.65$	
Downward	HIMELO	ND	ND	ND	$5.3\pm1.22$	9.6±2.67	$3.7\pm0.50$	

**Table 1.** Parameter estimates from fitted temperature-responses curves for  $GP_{max}$ ,  $NP_{max}$  and  $R_d$  of macroalgae that decreased ("downward") or increased ("upward") their frequency of occurrence in NW Iberia in recent times.  $T_{opt}$  (°C) is the temperature at which maximum rates occur; rates at  $T_{opt}$  were determined for  $GP_{max}$  and  $NP_{max}$  (mg  $O_2 \cdot g^{-1}DW \cdot h^{-1}$ ). For most species,  $T_{opt}$  could not be determined for  $R_d$  (ND = not determined); in those cases,  $R_d$  at  $T_{opt}$  is the rate recorded at 30 °C. Values are estimate  $\pm$  asymptotic SE except for  $R_d$  at  $T_{opt}$  where values a mean  $\pm$  SE. <sup>a</sup>CORCES = *Corallina caespitosa*; BIFBIF = *Bifurcaria bifurcata*; CYSBAC = *Cystoseira baccata*; CYSTAM = *Cystoseira tamariscifolia*; MASSTE = *Mastocarpus stellatus*; CHOCRI = *Chondrus crispus*; FUCVES = *Fucus vesiculosus*; HIMELO = *Himanthalia elongata*.

at temperatures beyond the optimum seemed more pronounced in three of the four downward seaweeds (not in *F. vesiculosus*) than in upward algae. No differences were found in the photosynthetic efficiency at low irradiance ( $\alpha$ ) between upward and downward trend species (results not shown).

Gaussian model estimates of  $T_{opt}$  differed significantly among species both for  $GP_{max}$  and for  $NP_{max}$  (Table 1 and Fig. 2). However, no consistent pattern could be detected between the upward and downward sets of species. Instead, many  $T_{opt}$  estimates for  $NP_{max}$  ranged between 17 °C and 22 °C and were statistically undistinguishable.



**Figure 2.** Comparison of  $T_{opt}$  and rate at  $T_{opt}$  for  $GP_{max}$  and  $NP_{max}$  estimated by non-linear regression for eight seaweeds that followed divergent upward (white bars) and downward (black bars) abundance trends in NW Iberia along the last decade. 95% confidence intervals calculated after adjusting the experiment wise error rate with the Bonferroni method. No bar shown for  $GP_{max}$  in *H. elongata* due to poor fitting. See Table 1 for species abbreviations.

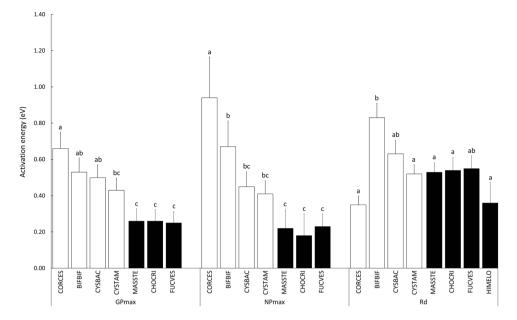
		<i>GP</i> <sub>max</sub>				R <sub>d</sub>			NP <sub>max</sub>				
Occurrence trend	Species <sup>a</sup>	Ea	R <sup>2</sup> -adj	P-value	Range (°C)/N	Ea	R <sup>2</sup> -adj	P-value	Range (°C)/N	Ea	R <sup>2</sup> -adj	P-value	Range (°C)/N
Upward	CORCES	$0.66 \pm 0.090$	80.3	< 0.0001	6-24/21	$0.35\pm0.048$	77.2	< 0.0001	6-24/21	$0.94 \pm 0.225$	65.9	0.0024	6-21/18
Upward	BIFBIF	$0.53 \pm 0.078$	65.9	< 0.0001	6-30/27	$0.83\pm0.080$	86.4	< 0.0001	9-27/21	$0.67 \pm 0.142$	65.5	0.0005	6-21/18
Upward	CYSBAC	$0.50\pm0.070$	74.0	< 0.0001	6-24/21	$0.63\pm0.076$	76.1	< 0.0001	6-27/24	$0.45 \pm 0.082$	62.2	< 0.0001	6-24/21
Upward	CYSTAM	$0.43 \pm 0.067$	71.8	< 0.0001	6-21/18	$0.52\pm0.050$	81.2	< 0.0001	6-30/27	$0.41 \pm 0.073$	66.8	< 0.0001	6-21/18
Downward	MASSTE	$0.26 \pm 0.068$	47.3	0.0016	9-24/18	$0.53 \pm 0.052$	81.0	< 0.0001	6-30/27	$0.22 \pm 0.069$	40.1	0.0048	9-24/18
Downward	CHOCRI	$0.26 \pm 0.063$	42.0	0.0004	6-27/24	$0.54 \pm 0.071$	75.3	< 0.0001	6-24/21	$0.18 \pm 0.104$	14.0	0.1044	6-24/21
Downward	FUCVES	$0.25 \pm 0.064$	49.4	0.0011	6-21/18	$0.55 \pm 0.070$	71.8	< 0.0001	6-30/27	$0.23 \pm 0.066$	43.7	0.0028	6-21/18
Downward	HIMELO	$-0.12 \pm 0.067$	9.0	0.0717	6-30/27	$0.36 \pm 0.113$	35.8	0.0053	6-24/21	$-0.30 \pm 0.121$	27.1	0.0223	6-30/27

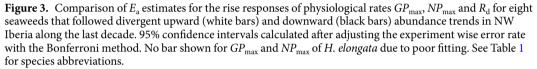
**Table 2.** Temperature dependence of  $GP_{max}$ ,  $NP_{max}$  and  $R_d$ . Values are the activation energy  $E_a$  ( $\pm$ SE; eV) calculated from the Boltzmann-Arrhenius model by OLS fit of the rise component of the rate-temperature curve. Range (°C) is the range of temperatures where the rate followed a monotonic rise while *N* is sample size. R<sup>2</sup>-adjusted is the percentage of the variability explained by the fitted model while *P*-value indicates the significance of the  $E_a$  estimate. <sup>a</sup>CORCES = *Corallina caespitosa*; BIFBIF = *Bifurcaria bifurcata*; CYSBAC = *Cystoseira baccata*; CYSTAM = *Cystoseira tamariscifolia*; MASSTE = *Mastocarpus stellatus*; CHOCRI = *Chondrus crispus*; FUCVES = *Fucus vesiculosus*; HIMELO = *Himanthalia elongata*.

Exceptions were the downward *H. elongata* (9.6 ± 2.67 °C) and the upward *C. baccata* (28.8 ± 6.54 °C).  $T_{opt}$  estimates for  $GP_{max}$  were slightly higher than for  $NP_{max}$ , ranging between 20 °C and 31 °C. No estimate could be obtained for *H. elongata*, but an examination of the original data reveals that the largest rates of  $GP_{max}$  also occurred at low temperatures around 6–9 °C. Like  $T_{opt}$ , Gaussian model estimates of  $NP_{max}$  and  $GP_{max}$  at  $T_{opt}$  also varied significantly among species. Again, the highest and the lowest estimates fell into algae from different trend sets: the downward *F. vesiculosus* (11.5 ± 0.65 mg O<sub>2</sub> · g<sup>-1</sup>DW · h<sup>-1</sup>  $NP_{max}$  and 14.0 ± 1.26 mg O<sub>2</sub> · g<sup>-1</sup>DW · h<sup>-1</sup>  $GP_{max}$ ) and the upward *C. caespitosa* (1.9 ± 0.27 mg O<sub>2</sub> · g<sup>-1</sup>DW · h<sup>-1</sup>  $NP_{max}$  and 3.0 ± 0.25 mg O<sub>2</sub> · g<sup>-1</sup>DW · h<sup>-1</sup>  $GP_{max}$ ). Still, no consistent pattern could be detected between abundance trend groups for any of these parameters.

Unlike photosynthesis, no  $T_{opt}$  could be observed for dark respiration ( $R_d$ ) because it increased monotonically with temperature in most algae (Fig. 1). The exception was the downward Irish moss *C. crispus* where  $R_d$ slightly decreased at the highest temperature, resulting in a  $T_{opt}$  estimate of  $26.2 \pm 1.34$  °C ( $R_d$  estimate at a  $T_{opt} = 2.2 \pm 0.25$  mg O<sub>2</sub> · g<sup>-1</sup>DW · h<sup>-1</sup>). In most cases,  $R_d$  changed gradually with temperature, although some downward seaweeds (*H. elongata* and *F. vesiculosus*) showed some evidence of an exponential increment of  $R_d$  at the highest experimental temperatures (>24 °C).

**Activation energy.** We were able to estimate activation energies for net and gross photosynthesis in seven of the eight seaweeds (Table 2). In the downward *Himanthalia elongata*, the range of experimental temperatures at which photosynthesis followed a monotonic increase was too small to allow a reliable estimate of  $E_a$ . Unlike  $T_{opt}$  our analyses of the temperature dependence of photosynthesis with the Boltzmann-Arrhenius model suggested consistent differences between upward and downward algae. The conditional sums of squares of  $E_a$  estimates





revealed highly significant interspecific differences in the temperature dependency of both  $GP_{max}$  (F = 4.4; P-value = 0.0004) and  $NP_{max}$  (F = 8.4; P-value < 0.0001). Upward species showed consistently higher activation energies than downward ones, indicating that productivity was more responsive to temperature increases below  $T_{opt}$  in the former.  $E_a$  ranged 0.43–0.66 eV for  $GP_{max}$  in upward algae compared to 0.25–0.26 eV in downward ones and Bonferroni-corrected pairwise comparisons indicated that the differences between both sets of species were statistically significant in most cases (Fig. 3). A similar pattern was observed for  $NP_{max}$ . Again, net photosynthesis was more responsive to temperature in upward than in downward algae, with activation energies ranging 0.40–0.94 eV in the former while only 0.18–0.22 eV in the latter. Bonferroni-corrected pairwise tests again revealed significant differences between upward and downward algae, although the slightly wider confidence intervals of some estimates lowered the statistical power of some comparisons compared to  $GP_{max}$ .

A very different picture emerged when we analysed the temperature dependence of dark respiration (Table 2). The conditional sums of squares test detected statistically significant differences among seaweeds (F = 2.7; P-value = 0.0124). Nonetheless, the Bonferroni-corrected pairwise comparisons indicated that this was mostly a consequence of the extreme temperature dependence shown by respiration in the upward *B. bifurcata* (0.83 ± 0.080 eV) (Fig. 3). Many algae, both upward and downward, had activation energies for  $R_d$  around 0.52–0.63 eV and while respiration still seemed less dependent on temperature in some species, these included both upward (*C. caespitosa*) and downward (*H. elongata*) examples.

#### Discussion

Growing experimental evidence indicates that losing species can diminish ecosystems functions such as productivity, biomass accumulation or nutrient uptake. However, the ecosystem consequences of species loss will probably be species-specific as functional performance varies among species<sup>47</sup>. In this context, finding simple proxies to anticipate a species' susceptibility to become extinct will improve our ability to predict the actual consequences of future reductions in species diversity. Our results found that, besides the varying metabolic rates and optimal temperatures recorded in the species examined, the temperature dependence of photosynthesis (as the activation energy  $E_a$  for *GPP*) was consistently larger in seaweeds that recently followed an upward abundance trend than in those that declined in abundance in recent years. These variation is in agreement with the hypothesis that temperature may have played a pivotal role in the shifts in abundance detected in these species. Furthermore, they also suggest that  $E_a$  might be a convenient tool to foresee the impact of ocean warming on seaweed populations.

The metabolic theory of ecology (MTE) predicts that  $E_a$  for heterotrophic metabolism should be close to 0.65 eV (Brown *et al.* 2004) while photosynthetic  $E_a$  should be smaller, estimated around 0.32 eV by Allen, *et al.*<sup>48</sup>. However, these values are intended to characterize processes at a global scale and have been derived from approximations based on the kinetics of the relevant rate-limiting biochemical reactions, e.g. the approximation of 0.32 eV for the temperature dependence of photosynthesis assumes that the rate of C<sub>3</sub> photosynthesis is limited by Rubisco carboxylation<sup>48</sup>. Consequently, the values predicted by MTE are found in interspecific studies<sup>30,31</sup> or when a sizeable number of intraspecific estimates are combined into a single mean value<sup>45</sup>. Meanwhile,  $E_a$  estimates for single traits obtained in individual intraspecific studies are usually very variable and typically range

between 0.2 and 1.2 eV, consistent with the variation observed for metabolic reactions<sup>30,45</sup>. Even the universality of a single value to characterize the temperature dependence of primary production at very large (ecosystem) scale has been questioned<sup>49</sup> and it has been proposed that  $E_a$  for primary production might be larger in aquatic than in terrestrial ecosystems<sup>50</sup>. In this context, the fact that our  $E_a$  estimates for photosynthesis do not match the theoretical value of 0.32 eV 0.44 for aquatic ecosystems according to<sup>50</sup> is just another example of the complexity of the temperature dependence of photosynthesis and is probably related to differences in ecology between species<sup>51</sup>. Unlike photosynthesis, the lack of consistent differences for respiration  $E_a$  among species with upward and downward abundance trends implies that respiration responded to temperature more uniformly across species than production. Moreover, the average respiration  $E_a$  in our experiment was  $0.54 \pm 0.05$  eV, below the 0.65 eV derived from theory but nearly identical to the median of 0.55 eV found in an comprehensive analysis of intraspecific responses to temperature<sup>45</sup>, indicating a better match with the predictions of MTE.

Temperature dependence is known to vary as a result of processes such as acclimation, acclimatization and adaptation<sup>29</sup>. In seaweeds, for example, sensitivity to increasing temperatures can be greater in winter than in summer, although this change appears to be more pronounced in polar than in temperate seaweeds<sup>1,34</sup>. Even within a single species, the temperature dependence of photosynthesis and respiration has been shown to grad-ually decrease from cooler to warmer latitudes<sup>35</sup>. In our case, because we used seaweeds collected at the same time of the year that were kept under uniform environmental conditions until the experiment, both acclimation and acclimatization can be safely disregarded as explanations for the differences in photosynthesis  $E_a$  detected between the two groups of species. Likewise, although our experimental design combined whole fronds for some species with young or distal apical fronds for others, the two types of sample were evenly represented in both trend groups, suggesting the sample type was not responsible for the consistent differences in  $E_a$  between trend groups. Instead, the variation in photosynthesis  $E_a$  possibly indicates that each group has a different potential to cope with the physiological consequences of water warming.

A comparison of the Boltzmann factor shows that the temperature dependence of respiration and photosynthesis was very similar in the upward trend group, predicting an average increase of twofold over the temperature range 283–293 K (10–20 °C) ( $e^{-E_d/293k}/e^{-E_d/283k} = 2.33$  for  $R_d$ , 2.11 for  $GP_{max}$  and 2.48 for  $NP_{max}$ ). In contrast, photosynthesis was much less sensitive to temperature ( $e^{-E_d/293k}/e^{-E_d/283k} = 1.28$  for  $GP_{max}$ , 1.17 for  $NP_{max}$ ) than respiration (2.01 for  $R_d$ ) in the downward trend, suggesting a higher risk to experience energy imbalances when the temperature increases over the physiological range. A comparable, but more extreme, imbalance between the temperature dependence of photosynthesis and respiration has been observed in Antarctic red algae<sup>52</sup>, suggesting that it might be characteristic of seaweeds adapted to low temperatures. In comparison, the temperature dependence of respiration is usually similar to or lower than that of photosynthesis in seaweeds from temperate and warm latitudes<sup>53–56</sup> but see<sup>23,57</sup>.

None of the other metabolic traits examined by us showed consistent differences among the two groups of species. Instead, functional responses were species-specific for physiological traits such as  $T_{opt}$ , maximum *GPP* and *NPP*, dark respiration at  $T_{opt}$ , or photosynthetic efficiency at low irradiance (alpha). Idiosyncratic or species-specific photosynthetic and respiration rates were expected since the seaweed examined here are considerably separated from a phylogenetic point of view. Metabolism in seaweeds is affected not only by the physical environment but also by biotic factors such as morphology, ontogeny, circadian and seasonal rhythms<sup>1</sup>. Studies on intraspecific and interspecific differences in temperature-photosynthesis response curves for many seaweed species indicate that both the amplitude and the position of the curve are usually correlated with the local temperature regime, reflecting local temperature adaptation<sup>25</sup>. For example, Antarctic seaweed populations show maximum net primary productivity rates at relatively cold temperatures, i.e. between 0 and 15 °C<sup>52</sup>, while tropical species of the genus *Asparagopsis* show maximum photosynthesis rates at 26 °C<sup>58</sup>. Intraspecific differences among populations revealing local adaption within species have been also recorded in seaweeds like *Chondrus crispus*<sup>33</sup> or the kelp *Ecklonia radiata*<sup>55</sup>.

Our proposal that the activation energy of photosynthesis and respiration may help to foresee the vulnerability to water warming does not necessarily imply that temperature alters the abundance of these seaweeds through an impact on the rates of photosynthesis and respiration. It has long been known that the geographic boundaries of seaweeds are set by limitations to growth (temperature inappropriate for population growth), but also by limitations to survival (temperature prevents the survival of the hardiest stage) or to reproduction (temperature prevents the maturation of some stage)<sup>25,59</sup>. Therefore, we suggest that the temperature dependence of photosynthesis and respiration might serve as a biomarker of the ability of some seaweeds to cope with water warming, even though temperature may actually have a stronger, more influential impact on life cycle traits other than respiration or production (e.g. reproduction).

Temperature is the main abiotic factor driving large scale distribution of seaweed species<sup>4,60</sup>. In the North Atlantic, large shifts in the distribution patterns of marine species have been experienced during glacial-interglacial cycles shaping the contemporary distribution and the genetic structure of seaweed populations<sup>61,62</sup>. In the last decades, quick shifts in species distribution<sup>3,10–13,15</sup> have been linked with rising temperatures in our oceans. Our experiment was aimed to characterize functional traits which could be relevant to assess seaweeds vulnerability to ocean warming. Previous, experimental studies have successfully used empirical lethal thresholds to define the current and future distribution of seaweeds across large spatial scales<sup>63,64</sup>. Here we found consistent differences in the temperature dependence of photosynthesis among species with opposite abundance trends, suggesting divergences in the metabolic scaling with temperature among these groups of seaweeds. Overall, our study supports the idea that temperature could be mechanistic cause of the shifts in abundance observed for these seaweeds in the last decades and provides a relatively simple experimental approach that could be further examined in other seaweeds undergoing similar climate-linked abundance changes.

#### References

- 1. Hurd, C. L., Harrison, P. J., Bischof, K. & Lobban, C. S. Seaweed Ecology and Physiology. (Cambridge University Press, 2014).
- 2. Harley, C. D. G. et al. Effects of climate change on global seaweed communities. J. Phycol. 48, 1064–1078 (2012).
- 3. Wernberg, T. et al. Seaweed communities in retreat from ocean warming. Curr. Biol. 21, 1828–1832 (2011).
- Lüning, K. Seaweeds: Their Environment, Biogeography and Ecophysiology. (Wiley, 1990).
  Intergovernmental Panel on Climate Change. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. ([Stocker, T. F., Qin, D., Plattner, G.-K., Tignor, M., Allen, S. K., Boschung, J., Nauels, A., Xia, Y., Bex, V. and Midgley, P. M. (eds)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 1535 pp, 2013).
- 6. Burrows, M. T. et al. The pace of shifting climate in marine and terrestrial ecosystems. Science 334, 652–655 (2011).
- 7. Barton, E. D., Field, D. B. & Roy, C. Canary current upwelling: More or less? Prog. Oceanogr. 116, 167-178 (2013).
- 8. Hoegh-Guldberg, O. & Bruno, J. F. The impact of climate change on the world's marine ecosystems. Science 328, 1523–1528 (2010).
- 9. Halpern, B. S. et al. A global map of human impact on marine ecosystems. Science 319, 948-952 (2008).
- Piñeiro-Corbeira, C., Barreiro, R. & Cremades, J. Decadal changes in the distribution of common intertidal seaweeds in Galicia (NW Iberia). Mar. Environ. Res. 113, 106–115 (2016).
- 11. Gallon, R. K. *et al.* Twenty years of observed and predicted changes in subtidal red seaweed assemblages along a biogeographical transition zone: Inferring potential causes from environmental data. *J. Biogeogr.* **41**, 2293–2306 (2014).
- Lima, F. P., Ribeiro, P., Queiroz, N., Hawkins, S. J. & Santos, A. M. Do distributional shifts of northern and southern species of algae match the warming pattern? *Global Change Biol.* 13, 2592–2604 (2007).
- Fernández, C. Current status and multidecadal biogeographical changes in rocky intertidal algal assemblages: The northern Spanish coast. Estuar. Coast. Shelf Sci. 171, 35–40 (2016).
- 14. Wernberg, T. et al. Climate-driven regime shift of a temperate marine ecosystem. Science 353, 169-172 (2016).
- Díez, I., Muguerza, N., Santolaria, A., Ganzedo, U. & Gorostiaga, J. M. Seaweed assemblage changes in the eastern Cantabrian Sea and their potential relationship to climate change. *Estuar. Coast. Shelf Sci.* 99, 108–120 (2012).
- Duarte, L. *et al.* Recent and historical range shifts of two canopy-forming seaweeds in North Spain and the link with trends in sea surface temperature. *Acta Oecol.* 51, 1–10 (2013).
- 17. Bartsch, I. *et al.* Changes in kelp forest biomass and depth distribution in Kongsfjorden, Svalbard, between 1996–1998 and 2012–2014 reflect Arctic warming. *Polar Biol.* **39**, 2021–2036 (2016).
- Dayton, P. K. & Tegner, M. J. Catastrophic storms, El Niño and patch stability in a southern california kelp community. Science 224, 283–285 (1984).
- Martinez, E. A., Cardenas, L. & Pinto, R. Recovery and genetic diversity of the intertidal kelp *Lessonia nigrescens* (Phaeophyceae) 20 years after El Nino 1982/831. *J. Phycol.* 39, 504–508 (2003).
- Colvard, N. B., Carrington, E. & Helmuth, B. Temperature-dependent photosynthesis in the intertidal alga Fucus gardneri and sensitivity to ongoing climate change. J. Exp. Mar. Biol. Ecol. 458, 6–12 (2014).
- Graiff, A., Liesner, D., Karsten, U. & Bartsch, I. Temperature tolerance of western Baltic Sea Fucus vesiculosus growth, photosynthesis and survival. J. Exp. Mar. Biol. Ecol. 471, 8–16 (2015).
- Hargrave, M. S., Foggo, A., Pessarrodona, A. & Smale, D. A. The effects of warming on the ecophysiology of two co-existing kelp species with contrasting distributions. *Oecologia*, 1–13 (2016).
- Biskup, S., Bertocci, I., Arenas, F. & Tuya, F. Functional responses of juvenile kelps, Laminaria ochroleuca and Saccorhiza polyschides, to increasing temperatures. Aquat. Bot. 113, 117–122 (2014).
- Ji, Y., Xu, Z., Zou, D. & Gao, K. Ecophysiological responses of marine macroalgae to climate change factors. J. Appl. Phycol. 28, 2953–2967 (2016).
- Eggert, A. In Seaweed Biology: Novel Insights into Ecophysiology, Ecology and Utilization (eds Christian Wiencke & Kai Bischof) 47–66 (Springer Berlin Heidelberg, 2012).
- Tom Dieck, I. Temperature tolerance and survival in darkness of kelp gametophytes (Laminariales, Phaeophyta): ecological and biogeographical implications. Mar. Ecol. Prog. Ser. 100, 253-264 (1993).
- Bolton, J. J. & Lüning, K. Optimal growth and maximal survival temperatures of Atlantic Laminaria species (Phaeophyta) in culture. Mar. Biol. 66, 89–94 (1982).
- Tom Dieck, I. & de Oliveira, E. C. The section Digitatae of the genus Laminaria (Phaeophyta) in the northern and southern Atlantic: crossing experiments and temperature responses. Mar. Biol. 115, 151–160 (1993).
- Brown, J. H. & Sibly, R. M. In *Metabolic Ecology: A Scaling Approach* (eds Sibly, R. M., James H. Brown, & Astrid Kodric-Brown) 21–33 (Wiley-Blackwell, 2012).
- Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M. & Charnov, E. L. Effects of size and temperature on metabolic rate. Science 293, 2248–2251 (2001).
- Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. & West, G. B. Toward a metabolic theory of ecology. *Ecology* 85, 1771–1789 (2004).
- 32. Davison, I. R., Greene, R. M. & Podolak, E. J. Temperature acclimation of respiration and photosynthesis in the brown alga *Laminaria saccharina. Mar. Biol.* 110, 449-454 (1991).
- Kübler, J. E. & Davison, I. R. Thermal acclimation of light-use characteristics of *Chondrus crispus* (Rhodophyta). *Eur. J. Phycol.* 30, 189–195 (1995).
- 34. Kremer, B. P. In The Biology of Seaweeds (eds Lobban, C. S. & Wynne, M. J.) 493-533 (University of California Press, 1981).
- Wernberg, T., de Bettignies, T., Joy, B. A. & Finnegan, P. M. Physiological responses of habitat-forming seaweeds to increasing temperatures. *Limnol. Oceanogr.* 61, 2180–2190 (2016).
- Martínez, B., Arenas, F., Trilla, A., Viejo, R. M. & Carreño, F. Combining physiological threshold knowledge to species distribution models is key to improving forecasts of the future niche for macroalgae. *Global Change Biol.*, 1422–1433 (2014).
- Boaventura, D., Ré, P., Cancela da Fonseca, L. & Hawkins, S. J. Intertidal rocky shore communities of the continental portuguese coast: Analysis of distribution patterns. *Mar. Ecol.* 23, 69–90 (2002).
- Cremades, J., Bárbara, I. & Veiga, A. J. Intertidal vegetation and its commercial potential on the shores of Galicia (NW Iberian Peninsula). *Thalassas: An International Journal of Marine Sciences* 20, 69–80 (2004).
- Alonso-Pérez, F. & Castro, C. G. Benthic oxygen and nutrient fluxes in a coastal upwelling system (Ria de Vigo, NW Iberian Peninsula): seasonal trends and regulating factors. *Mar. Ecol. Prog. Ser.* 511, 17–32 (2014).
- 40. Ospina-Alvarez, N. *et al.* Outside the paradigm of upwelling rias in NW Iberian Peninsula: Biogeochemical and phytoplankton patterns of a non-upwelling ria. *Estuar. Coast. Shelf Sci.* **138**, 1–13 (2014).
- 41. Eilers, P. H. C. & Peeters, J. C. H. A model for the relationship between light intensity and the rate of photosynthesis in phytoplankton. *Ecol. Model.* **42**, 199–215 (1988).
- 42. Phytotools: Phytoplankton Production Tools. R package. v. 1.0 (2015).
- R Core Team R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. www.R-project.org/ (2013).
- 44. June, T., Evans, J. R. & Farquhar, G. D. A simple new equation for the reversible temperature dependence of photosynthetic electron transport: a study on soybean leaf. *Funct. Plant Biol.* **31**, 275–283 (2004).

- Dell, A. I., Pawar, S. & Savage, V. M. Systematic variation in the temperature dependence of physiological and ecological traits. Proc. Natl. Acad. Sci. USA 108, 10591–10596 (2011).
- Sokal, R. R. & Rohlf, F. J. Biometry: the Principles and Practice of Statistics in Biological Research. 3rd edn, (W.H. Freeman and Co., 1995).
- 47. Mineur, F. et al. European seaweeds under pressure: Consequences for communities and ecosystem functioning. J. Sea Res. 98, 91–108 (2015).
- 48. Allen, A. P., Gillooly, J. F. & Brown, J. H. Linking the global carbon cycle to individual metabolism. Funct. Ecol. 19, 202-213 (2005).

 Anderson-Teixeira, K. J. & Vitousek, P. M. In Metabolic Ecology: A Scaling Approach (eds Sibly, R. M., James H. Brown, & Astrid Kodric-Brown) 99–111 (Wiley-Blackwell, 2012).

- Yvon-Durocher, G., Allen, A. P., Montoya, J. M., Trimmer, M. & Woodward, G. The temperature dependence of the carbon cycle in aquatic ecosystems. Adv. Ecol. Res. 43, 267–313 (2010).
- 51. Clarke, A. & Fraser, K. P. P. Why does metabolism scale with temperature? Funct. Ecol. 18, 243-251 (2004).
- 52. Eggert, A. & Wiencke, C. Adaptation and acclimation of growth and photosynthesis of five Antarctic red algae to low temperatures. *Polar Biol.* 23, 609–618 (2000).
- 53. Phooprong, S., Ogawa, H. & Hayashizaki, K. Photosynthetic and respiratory responses of *Gracilaria salicornia* (C. Ag.) Dawson (Gracilariales, Rhodophyta) from Thailand and Japan. J. Appl. Phycol. **19**, 795–801 (2007).
- Lapointe, B. E., Tenore, K. R. & Dawes, C. J. Interactions between light and temperature on the physiological ecology of *Gracilaria tikvahiae* (Gigartinales: Rhodophyta). *Mar. Biol.* 80, 161–170 (1984).
- 55. Staehr, P. A. & Wernberg, T. Physiological responses of *Ecklonia radiata* (Laminariales) to a latitudinal gradient in ocean temperature. J. Phycol. 45, 91-99 (2009).
- Zou, D. & Gao, K. Thermal acclimation of respiration and photosynthesis in the marine macroalga *Gracilaria lemaneiformis* (Gracilariales, Rhodophyta). J. Phycol. 49, 61–68 (2013).
- 57. Zou, D., Liu, S., Du, H. & Xu, J. Growth and photosynthesis in seedlings of *Hizikia fusiformis* (Harvey) Okamura (Sargassaceae, Phaeophyta) cultured at two different temperatures. *J. Appl. Phycol.* 24, 1321–1327 (2012).
- Zanolla, M. et al. Photosynthetic plasticity of the genus Asparagopsis (Bonnemaisoniales, Rhodophyta) in response to temperature: implications for invasiveness. Biol. Invasions 17, 1341–1353 (2015).
- Breeman, A. M. Relative importance of temperature and other factors in determining geographic boundaries of seaweeds: Experimental and phenological evidence. *Helgoländer Meeresun.* 42, 199–241 (1988).
- 60. Hoek, C. V. D. The distribution of benthic marine algae in relation to the temperature regulation of their life histories. *Biol. J. Linn. Soc.* 18, 81-144 (1982).
- Neiva, J., Assis, J., Fernandes, F., Pearson, G. A. & Serrão, E. A. Species distribution models and mitochondrial DNA phylogeography suggest an extensive biogeographical shift in the high-intertidal seaweed *Pelvetia canaliculata*. J. Biogeogr. 41, 1137–1148 (2014).
- Neiva, J., Pearson, G. A., Valero, M. & Serrão, E. A. Drifting fronds and drifting alleles: range dynamics, local dispersal and habitat isolation shape the population structure of the estuarine seaweed *Fucus ceranoides*. J. Biogeogr. 39, 1167–1178 (2012).
- Martínez, B. et al. Physical factors driving intertidal macroalgae distribution: physiological stress of a dominant fucoid at its southern limit. Oecologia 170, 341–353 (2012).
- 64. Martínez, B., Viejo, R. M., Carreño, F. & Aranda, S. C. Habitat distribution models for intertidal seaweeds: responses to climatic and non-climatic drivers. *J. Biogeogr.* 39, 1877–1890 (2012).

#### Acknowledgements

Authors thanks to Belén Carro and Anna Saborido for their help during the field work. F.A. received funding from the project INSEAFOOD within the Structured Program of R&D&I INNOVMAR - Innovation and Sustainability in the Management and Exploitation of Marine Resources (reference NORTE-01-0145-FEDER-000035) and funded by the Northern Regional Operational Programme (NORTE2020) through the European Regional Development Fund (ERDF). C.P.C., R.B. and J.C. acknowledges funding from Xunta de Galicia (Axudas para a consolidación e estruturación de unidades de investigación competitivas do SUG, grant GPC2015/025).

#### **Author Contributions**

F.A. and C.P.C. conceived the experiment. C.P.C. collected the data. C.P.C., R.B., F.A. analyzed data. C.P.C., R.B. wrote the manuscript and F.A. and J.C. participated in writing and editing the manuscript.

### **Additional Information**

Competing Interests: The authors declare no competing interests.

**Publisher's note:** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2018