SCIENTIFIC REPORTS

Received: 27 June 2017 Accepted: 6 February 2018 Published online: 12 March 2018

OPEN Genome-wide association analysis identifies new candidate risk loci for familial intracranial aneurysm in the French-Canadian population

Sirui Zhou^{1,2}, Ziv Gan-Or^{1,3}, Amirthagowri Ambalavanan^{1,3}, Dongbing Lai⁴, Pingxing Xie¹, Cynthia V. Bourassa¹, Stephanie Strong^{1,3}, Jay P. Ross^{1,3}, Alexandre Dionne-Laporte¹, Dan Spiegelman¹, Nicolas Dupré⁵, Tatiana M Foroud⁴, Lan Xiong^{2,6}, Patrick A. Dion^{1,7} & Guy A. Rouleau^{1,7}

Intracranial Aneurysm (IA) is a common disease with a worldwide prevalence of 1–3%. In the French-Canadian (FC) population, where there is an important founder effect, the incidence of IA is higher and is frequently seen in families. In this study, we genotyped a cohort of 257 mostly familial FC IA patients and 1,992 FC controls using the Illumina NeuroX SNP-chip. The most strongly associated loci were tested in 34 Inuit IA families and in 32 FC IA patients and 106 FC controls that had been exome sequenced (WES). After imputation, one locus at 3p14.2 (*FHIT*, rs1554600, $p = 4.66 \times 10^{-9}$) reached a genome-wide significant level of association and a subsequent validation in Nunavik Inuit cohort further confirmed the significance of the *FHIT* variant association (rs780365, FBAT-O, p = 0.002839). Additionally, among the other promising loci ($p < 5 \times 10^{-6}$), the one at 3q13.2 (rs78125721, $p = 4.77 \times 10^{-7}$), which encompasses CCDC80, also showed an increased mutation burden in the WES data (CCDC80, SKAT-O, p = 0.0005). In this study, we identified two new potential IA loci in the FC population: FHIT, which is significantly associated with hypertensive IA, and CCDC80, which has potential genetic and functional relevance to IA pathogenesis, providing evidence on the additional risk loci for familial IA. We also replicated the previous IA GWAS risk locus 18g11.2, and suggested a potential locus at 8p23.1 that warrants further study.

Intracranial Aneurysm (IA) has a prevalence of 1–3% in the general population^{1,2}. The rupture of an IA can lead to subarachnoid hemorrhages (SAH), which has devastating consequences. Environmental and genetic factors, such as hypertension and smoking³, family history and ethnicity all contribute to the risk of IA. Because of the complexity of IA, genome-wide association studies (GWAS) have become the predominant strategy used to look for genetic factors associated with IA. These studies used several large cohorts with IA patients, mainly of Finnish, Japanese or European descent. Several risk loci were discovered in these GWA studies: 8q11.23 (SOX17), 9p21.3-23.1 (CDKN2A-CDKN2BAS)^{4,5} and 2q33.1 from the European and Japanese cohorts; 18q11.2 (RBBP8), 13q13.1 (STARD13-KL) and 10q24.32⁶ from the Finnish and Japanese cohorts; 1q23.1, 3p25.2, 7p21.2, 9q31.3⁶ and 4q31.22 (EDNRA)⁷ from two Japanese cohorts, and 7p21.1 (HDAC9)⁸ from a cohort with European ancestry. A more recent Finnish IA study revealed additional GWAS risk loci, including 2q23.3, 5q31.3 and 6q24.2, represented by low-frequency SNPs⁹. Multiple GWAS signals suggest that the genetic etiology of IA may be complex and population specific. The risk loci found in each GWA study was estimated to only account for 4.1-6.1% of the heritability in the respective cohort⁹.

¹Montreal Neurological Institute and Hospital, McGill University, Montréal, QC, Canada. ²Department of Medicine, Faculty of Medicine, Université de Montréal, Montréal, QC, Canada. ³Department of Human Genetics, McGill University, Montréal, QC, Canada. ⁴Department of Medical and Molecular Genetics, Indiana University School of Medicine, Indianapolis, IN, USA. ⁵Faculty of Medicine, Université Laval, Quebec, QC, Canada. ⁶Centre de recherche, Institut universitaire en santé mentale de Montréal, Montréal, QC, Canada. ⁷Department of Neurology and Neurosurgery, McGill University, Montréal, QC, Canada. Correspondence and requests for materials should be addressed to G.A.R. (email: guy.rouleau@mcgill.ca)



Figure 1. Genome-wide assocation analysis in the FC IA discovery cohort. Imputed using HRC panel, 7,614,484 variants passed QC are included in making the manhattan plot. X-axis shows the physical position along the genome. Y-axis shows the $-\log 10$ (p-value) for association. Red line indicates the level of genome-wide significant association ($p = 5 \times 10^{-8}$); blue line indicates the level of suggestive association ($p = 5 \times 10^{-6}$). Green dots indicate *FHIT* SNPs.

It has been reported that the French-Canadian (FC) population has higher IA/SAH incidence and that patients usually aggregate in large pedigrees¹⁰, with 30% of IA patients having a family history (fIA)¹¹. Similarly, to the Finnish people, French-Canadians are also descended from a relatively small founder population and have population specific variants due to the population bottleneck and genetic drift. Therefore, we hypothesized that population specific and median/low frequency variants may play an important part in the disease risk in FC fIA.

Results

GWAS discovery phase. After data QC and sample pruning, 621,983 SNPs with 173 FC IA cases and 1,772 FC controls remained in the analysis. The genome-wide threshold for significance after Bonferroni correction was set to 5×10^{-8} . Marker-wise P values of Cochran–Armitage trend test were performed using PLINK 1.9¹² for genotyped variants. Genomic inflation factor $\lambda = 1.02$ indicated that there was little inflation of excessive significant markers, as shown in quantile-quantile (QQ) plot (Supplementary Figure S1).

The result of the initial trend test showed 3q13.2 as the most significant locus: rs2705520 ($p=6.93 \times 10^{-8}$), an intronic SNP in *ATG3 (autophagy 3*), followed by rs1877362 ($p=1.16 \times 10^{-7}$) in *CCDC80 (coiled-coil domain containing 80*) and rs1472107 ($p=1.18 \times 10^{-7}$) in *SLC35A5 (Solute Carrier Family 35 member A5)* (Supplementary Figure S2).

After imputation using the Haplotype Reference Consortium (HRC) and the exclusion of low quality and low MAF variants, 7,614,484 remaining variants were included in the test of associations using SNPtest¹³. The results were shown in the Manhattan plot (Figure 1). One locus reached genome-wide significant level after imputation: 3p14.2 (rs1554600, OR 0.26, $p = 4.66 \times 10^{-9}$) located in gene *FHIT*. TaqMan validation of rs1554600 on IA cases and controls suggested the imputation was accurate (maf.case = 0.0838 and maf.control = 0.026). The 28 most significant SNPs, representing 26 distinct loci that each reached suggestive level ($p < 5 \times 10^{-6}$), were prioritized for further validation (Table 1). A meta-analysis of 138 SNPs (Supplementary Table S1) located in 23 out of 26 loci comprising the current study and FIA cohort⁸ showed that three SNPs: rs76308736 (8p23.1), rs7084131 (10p14) and rs4867356 (5p13.3) are validated with decreased p-value (Table 2).

Using imputed SNPs, the two loci 3p14.2 and 3q13.2 were estimated by GCTA-GREML¹⁴⁻¹⁶ (Genome-wide Complex Trait Analysis) to account for approximately 3% of the heritability in the FC cohort (standard error (SE) = 0.026).

Replication in exome data and Inuit cohort. We looked into the exome sequencing data of the aforementioned 28 genes containing the top GWAS SNPs in our FC WES cohort; of the 23 genes that had exonic variants, a total of 177 exonic and splicing variants were found in 138 FC cases and controls. Sequence Kernel Association Test (SKAT)¹⁷ results showed excessive exonic variation burden in IA cases in four genes *SLC35F3* (p = 0.002), *DTNB* (p = 0.003), *CCDC80* (p = 0.0005) and *PABPC3* (p = 0.0001) (Table 3). However, the first two genes were less convincing with the limited number of variants in the testing (two variants for each). *PABPC3* was unlikely to be a risk gene for IA due to its human testis-specific expression. Thresholds Test (VT)¹⁸ focused on the selected genes revealed that only *CCDC80* (p = 0.01) in 3q13.2 reached the statistical significance after accounting for multiple testing.

In the Nunavik Inuit IA cohort, after performing Family Based Association Test (FBAT)¹⁹ of the 2,429 SNPs within the 28 aforementioned genes and neighboring regions, 50 SNPs located in the *FHIT* gene region were with p < 0.05 (Supplementary Table S2), the most significant one being rs780365 (p = 0.002839). Although the associations were no longer significant after corrections of multiple testing (p < 0.00014, 2,429 variants in 353 independent tests), it could be due to the limited sample size, which still suggested that *FHIT* variants may likely still be associated with IA.

Replication of previous GWAS risk loci. We also attempted to replicate the 12 IA risk loci identified in previous GWAS in our FC IA cohorts. 825 distinct LD blocks were established in these 12 loci in the FC

loci	top SNP	Position (hg19)	Gene (or nearby)	Frequency in cases	Frequency in controls	OR	P-value	SE	Beta	INFO
3p14.2	rs1554600	61157774	FHIT	0.080925	0.022291	0.258936	4.66E-09	0.338309	-1.98215	0.94
7p22.2	rs12535623	2951412	CARD11	0.260116	0.154345	0.519156	1.15E-07	0.157048	-0.83267	0.95
4p15.32	rs116130729	16012786	PROM1	0.072254	0.023138	0.304125	1.80E-07	0.343542	-1.79302	0.86
3q13.2	rs78125721	112325677	CCDC80	0.043353	0.009029	0.201063	4.77E-07	0.492736	-2.4811	0.97
21q22.13	rs111610752	38151254	HLCS	0.052023	0.016366	0.303181	5.39E-07	0.430726	-2.15882	0.68
8p23.1	rs117537300	10743939	XKR6	0.118497	0.052765	0.414387	6.64E-07	0.244413	-1.21512	0.88
13q12.13	rs7989887	25681112	PABPC3	0.16185	0.087472	0.4964	6.93E-07	0.203382	-1.00943	0.99
13q14.3	rs114375292	54413853	LINC00558	0.080925	0.030756	0.360387	7.25E-07	0.313554	-1.55348	0.89
1q42.2	rs150148362	234191488	SLC35F3	0.040462	0.008183	0.195651	8.16E-07	0.517978	-2.55442	0.74
16p13.2	rs138031402	8960325	CARHSP1	0.043353	0.01044	0.232811	8.87E-07	0.489044	-2.40373	0.77
7p13	rs146929064	44346336	CAMK2B	0.060694	0.020598	0.325486	1.32E-06	0.38035	-1.83941	0.85
1q41	rs12058987	217421015	ESRRG	0.083815	0.035553	0.402958	1.33E-06	0.305576	-1.47758	0.98
8q24.21	rs7011138	127922200	PCAT1	0.092486	0.19526	2.38087	1.35E-06	0.149667	0.72319	0.99
9p23	rs2039332	9138642	PTPRD	0.049133	0.011005	0.215339	1.48E-06	0.42766	-2.05874	0.79
16p13.2	rs982855	8297685	RBFOX1/TMEM114	0.572254	0.437641	0.581703	1.78E-06	0.116681	-0.55742	0.99
6p21.1	rs6911069	43617687	RSPH9	0.069364	0.025113	0.34561	1.92E-06	0.346004	-1.6476	0.99
20p13	rs56040592	1611918	SIRPG	0.104046	0.051072	0.463459	1.98E-06	0.257248	-1.22327	0.98
3p24.1	rs142836448	29398714	RBMS3	0.063584	0.153217	2.66475	2.26E-06	0.166286	0.786315	0.95
2p23.3	rs77639126	25598423	DTNB	0.072254	0.023984	0.315525	2.35E-06	0.335271	-1.58283	0.91
2p23.2	rs13028204	28853137	PLB1	0.086705	0.041479	0.455814	2.39E-06	0.29275	-1.38092	0.98
3q22.1	rs114999403	131692910	CPNE4	0.049133	0.014955	0.293814	2.67E-06	0.428374	-2.01095	0.97
20p13	rs77402555	876841	ANGPT4	0.046243	0.012415	0.259286	2.69E-06	0.456509	-2.14253	0.89
5p13.3	rs72753560	31626036	PDZD2	0.086705	0.178612	2.29048	2.75E-06	0.14898	0.698537	0.97
22q12.3	rs150551568	33136068	SYN3	0.046243	0.011851	0.247359	3.12E-06	0.444976	-2.07483	0.95
14q24.2	rs117272176	72902205	RGS6	0.046243	0.014108	0.295149	3.23E-06	0.449474	-2.09269	0.77
2p33.3	rs1207426	206154365	PARD3B	0.16763	0.096501	0.530358	4.10E-06	0.189958	-0.87503	0.99
15q25.1	rs8032417	78569930	DNAJA4	0.393064	0.275959	0.588522	4.82E-06	0.129826	-0.59364	1
10p14	rs7084131	8399121	GATA3	0.011561	0.068002	6.23842	4.83E-06	0.226803	1.03693	0.99

Table 1. 26 loci (28 genes) with top SNPs reached promising level of association.

.....

LOCI	Marker	CHR	POS	Allele1	Allele2	Zscore	P-value	Direction	P.FC	SampleSize.FC	P.FIA	SampleSize.FIA
8p23.1	rs76308736	8	10572792	a	g	-4.753	2.01E-06	_	4.5E-06	1945	0.0093	4033
10p14	rs7084131	10	8399121	a	g	4.686	2.78E-06	++	4.83E-06	1945	0.0114	4033
5p13.3	rs4867356	5	31623146	t	с	-4.608	4.07E-06	_	4.91E-06	1945	0.0148	4033

Table 2. Three loci with SNPs validated in the FIA cohort.

.....

population, the level of significance was therefore $p < 6.06 \times 10^{-5}$ after multiple correction. As a result, only one SNP rs35127791 located in 18q11.2 was replicated (maf = 0.157, p = 5.05×10^{-5} , beta = -0.66, SE = 0.16) (Figure 2). LocusZoom plots covering the rest of the 23 genome-wide significant SNPs from previous GWAS in these 12 loci were shown in Supplementary Figure S3.

We further looked into the FC WES data in locus 18q11.2, which comprised 4 protein coding genes: *GATA6*, *CTAGE1*, *RBBP8* and *CABLES1*, a variant burden test revealed exonic variants of *CABLES1* seemed to be associated with IA in FC (p = 0.022, corrected) (Supplementary Table S3).

Discussion

In this study, we discovered a new IA associated region on 3p14.2 which encompasses the *FHIT* gene in the French-Canadians (Figure 3), intronic variants in *FHIT* also suggested an association with IA in the Nunavik Inuit population. Additionally, we found evidence suggesting exonic variants in *CCDC80* within the 3q13.2 locus to be associated with the French-Canadian IA. Collectively, SNPs in *FHIT* and *CCDC80* could explain approximately 3% of the heritability of IA in French-Canadians, higher than that was reported in the Finnish study (2.1%)⁹ and in the GWAS replication (2.5%)⁶. The underlying reason for this might be that French-Canadians are a more homogenous population and this study has mainly included familial cases. On the other hand, we also replicated a previous GWAS risk locus 18q11.2²⁰, the top SNP rs35127791 located approximately 200 kb upstream of *GATA6*, close to a H3K24Ac region in HUVEC cells. *GATA6* regulates the differentiative state of vascular smooth muscle cells, and an important candidate for cardiovascular development. *CABLES1* proves to have an

Gene	Loci	Number of tested variants	P-value (SKAT)
ESRRG	1q41	2	0.7073712
SLC35F3	1q42.2	1	0.002189217
DTNB	2p23.3	2	0.002747022
PLB1	2p23.2	20	0.1122009
PARD3B	2p33.3	13	0.9205563
RBMS3	3p24.1	1	0.354907
CCDC80	3q13.2	8	0.0005070405
CPNE4	3q22.1	1	0.7073712
PROM1	4p15.32	12	0.05065522
PDZD2	5p13.3	19	0.7212919
RSPH9	6p21.1	2	0.8510591
CARD11	7p22.2	4	0.8172528
CAMK2B	7p13	3	0.09754801
PTPRD	9p23	7	0.2814774
PABPC3	13q12.13	54	0.0001097577
RGS6	14q24.2	1	0.7073712
DNAJA4	15q25.1	2	0.5637608
RBFOX1	16p13.2	5	1
CARHSP1	16p13.2	2	0.3250554
ANGPT4	20p13	4	0.09886071
SIRPG	20p13	5	0.7073707
HLCS	21q22.13	6	0.6115834
SYN3	22q12.3	3	0.5083075

 Table 3. Exonic and splicing variants of 23 GWAS suggestive genes in 138 FC cases and controls from WES.



Figure 2. Regional association signals of the previous GWAS risk locus 18q11.2. LocusZoom plot showing the regional association of chr18.19.7-20.75 mb, including the most significant SNP rs35127791 and previous GWAS SNP rs11661542. Purple line indicates the genetic recombination rate (cM/Mb). SNPs in linkage disequilibrium with rs35127791 are shown in color gradient indicating r2 levels (hg19, 1KGP, Nov 2014, EUR).

important role in cancer and development of neurons²¹, a recent study also highlighted its function vascular cell senescence and inflammation through p21 regulation²².

Unfortunately, we could not replicate the most significant SNP in *FHIT* in another IA cohort. However, this was expected, as *FHIT* variants have higher MAFs in French-Canadians compared to other European populations. The signal may only exist in French-Canadian populations and may correlate with IA cases with





hypertension. Among the three SNPs which were validated in the FIA cohort, rs76308736 is located in the promoter region of *SOX7*, with its crucial function in angiogenesis and establishment of arteriovenous identity. rs76308736 only showed suggestive significance in our discovery cohort and did not pass multiple correction in the FIA data, the relationship of this SNP and risk for IA remains to be explored.

Although the number of cases in this study was limited, we tried improving the power by doing the following: 1) targeting IA patients with family history, 2) focusing on individuals only with the French-Canadian ethnicity, and 3) validating the findings in exome sequencing results and in another founder population. Because French-Canadians originated from a small founder population, we also included intermediate variants after the imputation. The top SNPs that we discovered in *FHIT* and *CCDC80* had rare or intermediate frequency (2-4%). The MAF of rs1554600 was higher in French-Canadians (3.3%) compared to Europeans (1.9%) and was significantly higher when compared with East Asians (0.1%), which suggest a bottleneck and/or drift may be the reason for the accumulation of low-frequency variants that potentially associated with the risk of IA in the French-Canadians.

FHIT (fragile histidine triad) is a tumor suppressor gene that regulates DNA replication and signals stress responses²³, and encompasses the most active of the common human chromosomal fragile regions (FRA3B). Its expression has an important role in response to oxidative damage²⁴. On the other hand, oxidative stress is known to be a key contributor to IA formation and rupture²⁵. Interestingly, a study highlighted that the SNPs in *FHIT* have been associated with hypertensive traits in populations from Saguenay-Lac-St-Jean region, mainly French-Canadians²⁶. As over 40% of our French-Canadian IA cases were also affected with hypertension (Table 4), we considered that rs1554600 in *FHIT* is possibly more likely to be a risk for hypertensive IA in French-Canadians. Further test of *FHIT* SNPs between IA patients with and without hypertension revealed several SNPs in LD with rs1554600 to be significantly associated with this trait (ie. rs73098963, p = 0.002611, GWAS p value = 2.42×10^{-8}). *FHIT* has been reported in other GWAS studies to associate with hypertension: rs6782531, which located at approximately 160 kb upstream and in high LD of rs1554600 have also been reported to be significantly associated with blood pressure²⁷.

The 3q13.2 locus is a gene-rich region with many functions in inflammatory responses. *BTLA* (B- and T-lymphocyte attenuator) is involved in inflammatory responses²⁸ and homeostasis of the immune system²⁹. A previous study showed *BTLA* expression to be up-regulated in organs after a hemorrhagic shock³⁰. The 3q13.2 locus also comprises *ATG3*, which encodes a protein known to induce apoptosis³¹ and also to act as a regulator of oxidant and inflammatory balance that regulates endothelial cell stress response³². The top SNP rs2705520 in *ATG3* was also reported in a GWAS to be associated with asthma³³, suggesting its role in inflammatory diseases.

The most interesting gene in the 3q13.2 locus is *CCDC80*, also known as *SSG1* (steroid-sensitive gene 1), which is a cGMP signalling effector and was reported to be widely express in vascular smooth muscle cells³⁴. A previous study also showed that it has a role as a modulator of glucose and energy homeostasis³⁵. Another study highlighted that the product of fibroblast growth factor (*FGF*) regulates the expression of *CCDC80*³⁶, which is in turn also involved in cell adhesion during differentiation of fibroblast³⁷. *CCDC80* was reported as a tumor suppressor as well³⁸. These evidences suggest the critical function of *CCDC80* in vascular formation. *CCDC80* harbor a large number of rare variants in the French-Canadians (Supplementary Table S4); which also showed a

Sample demographic of cases and controls						
	Cases	Controls				
Number	257	1192				
Mean age of recruit (SD)	53.9 (11.2)	54.4 (16.8)				
% Male	31.5%	60.6%				
Clinical information of cases (212)						
	With family history	Sporadic				
With clinical information	147	65				
Multiple lesions (%)	55 (37.4)	19 (29.2)				
SAH (%)	51 (34.7)	25 (38.5)				
Hypertension (%)	60 (40.8)	31 (47.7)				
Drinking (%)	33 (22.4)	26 (40.0)				
Smoking (%)	89 (60.5)	50 (76.9)				
Hypercholesterolemia (%)	29 (19.7)	24 (36.9)				

Table 4. FC sample demographics and the clinical information of IA cases.

.....

significant difference in the variation burden in cases and controls. Both ATG3 and CCDC80 are dosage sensitive genes^{39,40}, therefore the potential different expression levels of those genes may affect the risk of IA.

In conclusion, we have provided evidence for four new loci associated with IA in French-Canadian IA cohort recruited from Montréal and Québec city, which could explain 3% of the disease heritability. Based on the findings of this study and the functions of their encoded products, two genes (*FHIT* and *CCDC80*) are potentially relevant to IA with strong aggregation of familial IA cases with high blood pressure in the French-Canadian population. *FHIT* is more particularly associated with hypertensive IA cases and this may be the consequence of a bottleneck and/or drift that affected the French-Canadian founder population. *CCDC80* was shown to have a large number of rare variations in the French-Canadian cohort and with a significantly different variation burdens between IA cases and controls. Both the lack of association of SNPs in *FHIT* and *CCDC80* and the replication of the only 18q11.2 locus of the previous GWAS hits suggests a genetic heterogeneity in IA, and thus additional studies targeting other high-risk populations are needed. However, the limited number of cases available in our study calls for a validation study that will have access to a larger cohort from the same founder population, thus to increase the power of detection.

Methods

Discovery cohort. The discovery cohort included 257 French-Canadian IA patients, a majority of them were with family history, which were recruited in Montréal and Québec City, Canada. The diagnoses were confirmed either by magnetic resonance angiography (MRA), or by surgical confirmation (clipped or coiled). An additional 1,992 controls, mainly comprised of unrelated FC individuals without cerebrovascular diseases were also included in the analysis. Their demographic information is listed in Table 4. Written informed consent were obtained from all participants, and this manuscript contain no identifying information for any participant. This study was approved by Comité d'éthique de la recherche du Centre hospitalier de l'Université de Montréal and McGill University ethics, all methods were performed in accordance with the relevant guidelines and regulations of McGill University (REB NEU-14-051).

Genotyping and quality control. All patients and controls were genotyped using the Illumina NeuroX SNP-chip, which contains 719,885 markers and is comprised of the backbone of Illumina HumanOmniExpress-v24 BeadChip. Raw data was processed by Illumina GenomeStudio software before the genotypes were generated. Both markers and samples were passed through a series of quality control (QC) steps. Samples were first removed if duplicated or if they had one of the following issues: 1) sex discrepancies; 2) exceeded a missing rate of 0.02; 3) ethnical admixture determined by PCA; or 4) with cryptic relateness determined by PLINK. Markers were removed if they meet one of the following criteria: 1) exceeding a missing rate of 0.02; 2) having a minor allele frequency (MAF) lower than 0.01; 3) deviating from Hardy Weinberg Equilibrium (p < 0.0001).

Principal Component Analysis (PCA) impelemented in the package EIGENSOFT 6.0⁴¹ was performed to assess the ethnicity of the samples. Three distinct populations CEU, CHB and YRI from 1000 Genome (1KGP) Phase III were used for clustering and CEU outliners were removed from further analysis. The remaining homogeneous population was also adjusted for the principal components in the subsequent tests for associations.

Imputation. Imputation was done by the Sanger Imputation Server (https://imputation.sanger.ac.uk/) using Haplotype Reference Consortium $r1.1^{41}$, and were pre-phased using SHAPEIT2⁴³. Imputed variants were included in the further analyses only with MAF >0.01 and with imputation quality score >0.3.

Association analysis. Frequentist additive association implemented in SNPtest¹³ was used to test for association of the imputed dataset, between FC IA cases and controls. Five major principal components were used as covariates for ancestry adjustment along with the sex of samples. Only autosomal SNPs were analyzed.

Regional association of suggestive loci were plotted using LocusZoom⁴⁴ with LD data from 1KGP CEU population.

Heritability estimation. We estimated the heritability from the original and imputed variants within the most promising loci, using methods of Estimation of Variance explained by SNPs (GREML)¹⁴ and GREML-LDMS¹⁶ programs implemented in the package of the Genome-wide Complex Trait Analysis (GCTA)¹⁵.

Meta-analysis. SNP identified in the current study that reached suggestive level of association ($p < 5 \times 10^{-6}$) were compared with the previous FIA GWAS summary statistics from 2,617 IA cases and 1,416 controls⁸. METAL was used to conduct meta-analysis of the two GWAS results for these selected SNPs.

FC and Inuit IA WES data. We examined the loci from the GWAS signals that have reached suggestive level of association in the exome sequencing results of 32 selected FC IA cases and 106 FC controls. Variable Thresholds Test (VT)¹⁸ implemented in Variant Tools (Vtools)⁴⁵ and Sequence Kernel Association Test (SKAT)¹⁷ were performed to test the exonic variation burden in the genes located in the GWAS significant regions.

Thirty-four Nunavik Inuit (Québec, Canada) families comprised of 49 IA patients and 124 family controls were also used to follow up the regions of significance. The samples were genotyped on Illumina HumanOmniExpress-v24 Beadchip which contains 730,525 SNPs. We looked into all the suggestive loci of the FC IA discovery cohort, and performed family-based association analysis (fbat) implemented in FBAT package¹⁹ in the Inuit SNP-chip data to test the case-control association in related individuals.

Replication of previous IA GWAS loci. 12 loci from previous GWA studies (Supplementary Figure S3) with 23 SNPs that reached genome-wide significance were selected to examine if they could be replicated in our study. 500 kb upstream/downstream of the first and last genome-wide significant SNP located in each of the 12 loci were investigated in our GWAS data. Multiple correction was performed on the number of independent tests, which was defined by the number of LD blocks within these 12 regions. LocusZoom was used for data plotting. The validated loci will further be examined in FC WES data using Genome Analysis Toolkit (GATK)⁴⁶ and VTools.

References

- Rinkel, G. J. Natural history, epidemiology and screening of unruptured intracranial aneurysms. J Neuroradiol 35, 99–103, https:// doi.org/10.1016/j.neurad.2007.11.004 (2008).
- Vernooij, M. W. et al. Incidental findings on brain MRI in the general population. N Engl J Med 357, 1821–1828, https://doi. org/10.1056/NEJMoa070972 (2007).
- 3. Wiebers, D. O. *et al.* Unruptured intracranial aneurysms: natural history, clinical outcome, and risks of surgical and endovascular treatment. *Lancet* **362**, 103–110 (2003).
- 4. Bilguvar, K. *et al.* Susceptibility loci for intracranial aneurysm in European and Japanese populations. *Nat Genet* **40**, 1472–1477, https://doi.org/10.1038/ng.240 (2008).
- Foroud, T. et al. Genome-wide association study of intracranial aneurysms confirms role of Anril and SOX17 in disease risk. Stroke 43, 2846–2852, https://doi.org/10.1161/STROKEAHA.112.656397 (2012).
- Akiyama, K. et al. Genome-wide association study to identify genetic variants present in Japanese patients harboring intracranial aneurysms. J Hum Genet 55, 656–661, https://doi.org/10.1038/jhg.2010.82 (2010).
- Low, S. K. et al. Genome-wide association study for intracranial aneurysm in the Japanese population identifies three candidate susceptible loci and a functional genetic variant at EDNRA. Hum Mol Genet 21, 2102–2110, https://doi.org/10.1093/hmg/dds020 (2012).
- Foroud, T. et al. Genome-wide association study of intracranial aneurysm identifies a new association on chromosome 7. Stroke 45, 3194–3199, https://doi.org/10.1161/STROKEAHA.114.006096 (2014).
- Kurki, M. I. *et al.* High risk population isolate reveals low frequency variants predisposing to intracranial aneurysms. *PLoS Genet* 10, e1004134, https://doi.org/10.1371/journal.pgen.1004134 (2014).
- Halal, F., Mohr, G. & Toussi, T. & Napoleon Martinez, S. Intracranial aneurysms: a report of a large pedigree. *Am J Med Genet* 15, 89–95, https://doi.org/10.1002/ajmg.1320150112 (1983).
- 11. Mathieu, J. *et al.* Epidemiological study of reptured intracranial aneurysms in the Saguenay-Lac-Saint-Jean region (Quebec, Canada). *Can J Neurol Sci* 23, 184–188 (1996).
- Chang, C. C. et al. Second-generation PLINK: rising to the challenge of larger and richer datasets. Gigascience 4, 7, https://doi. org/10.1186/s13742-015-0047-8 (2015).
- Marchini, J., Howie, B., Myers, S., McVean, G. & Donnelly, P. A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat Genet* 39, 906–913, https://doi.org/10.1038/ng2088 (2007).
- 14. Yang, J. *et al.* Common SNPs explain a large proportion of the heritability for human height. *Nat Genet* **42**, 565–569, https://doi.org/10.1038/ng.608 (2010).
- Yang, J., Lee, S. H., Goddard, M. E. & Visscher, P. M. GCTA: a tool for genome-wide complex trait analysis. Am J Hum Genet 88, 76–82, https://doi.org/10.1016/j.ajhg.2010.11.011 (2011).
- Yang, J. et al. Genetic variance estimation with imputed variants finds negligible missing heritability for human height and body mass index. Nat Genet 47, 1114–1120, https://doi.org/10.1038/ng.3390 (2015).
- 17. Wu, M. C. *et al.* Rare-variant association testing for sequencing data with the sequence kernel association test. *Am J Hum Genet* **89**, 82–93, https://doi.org/10.1016/j.ajhg.2011.05.029 (2011).
- Price, A. L. et al. Pooled association tests for rare variants in exon-resequencing studies. Am J Hum Genet 86, 832–838, https://doi.org/10.1016/j.ajhg.2010.04.005 (2010).
- 19. Laird, N. M., Horvath, S. & Xu, X. Implementing a unified approach to family-based tests of association. *Genet Epidemiol* **19**(Suppl 1), S36–42, https://doi.org/10.1002/1098-2272(2000)19:1+<::AID-GEPI6>3.0.CO;2-M (2000).
- 20. Yasuno, K. *et al.* Genome-wide association study of intracranial aneurysm identifies three new risk loci. *Nat Genet* **42**, 420–425, https://doi.org/10.1038/ng.563 (2010).
- Huang, J. R., Tan, G. M., Li, Y. & Shi, Z. The Emerging Role of Cables1 in Cancer and Other Diseases. *Mol Pharmacol* 92, 240–245, https://doi.org/10.1124/mol.116.107730 (2017).
- 22. Pu, Z. *et al.* Cables1 Inhibits Proliferation and Induces Senescence by Angiotensin II via a p21-Dependent Pathway in Human Umbilical Vein Endothelial Cells. *J Vasc Res* 54, 13–21, https://doi.org/10.1159/000452409 (2017).
- Barnes, L. D. et al. Fhit, a putative tumor suppressor in humans, is a dinucleoside 5',5''' -P1,P3-triphosphate hydrolase. Biochemistry 35, 11529–11535, https://doi.org/10.1021/bi961415t (1996).
- Karras, J. R., Paisie, C. A. & Huebner, K. Replicative Stress and the FHIT Gene: Roles in Tumor Suppression, Genome Stability and Prevention of Carcinogenesis. *Cancers (Basel)* 6, 1208–1219, https://doi.org/10.3390/cancers6021208 (2014).
- 25. Starke, R. M. et al. The role of oxidative stress in cerebral aneurysm formation and rupture. Curr Neurovasc Res 10, 247–255 (2013).

- Nikpay, M. *et al.* Genetic mapping of habitual substance use, obesity-related traits, responses to mental and physical stress, and heart rate and blood pressure measurements reveals shared genes that are overrepresented in the neural synapse. *Hypertens Res* 35, 585–591, https://doi.org/10.1038/hr.2011.233 (2012).
- Simino, J. et al. Gene-age interactions in blood pressure regulation: a large-scale investigation with the CHARGE, Global BPgen, and ICBP Consortia. Am J Hum Genet 95, 24–38, https://doi.org/10.1016/j.ajhg.2014.05.010 (2014).
- 28. Chemnitz, J. M., Lanfranco, A. R., Braunstein, I. & Riley, J. L. B and T lymphocyte attenuator-mediated signal transduction provides a potent inhibitory signal to primary human CD4 T cells that can be initiated by multiple phosphotyrosine motifs. *J Immunol* **176**, 6603–6614 (2006).
- Steinberg, M. W. et al. BTLA interaction with HVEM expressed on CD8(+) T cells promotes survival and memory generation in response to a bacterial infection. PLoS One 8, e77992, https://doi.org/10.1371/journal.pone.0077992 (2013).
- Cheng, T. et al. Enhanced Innate Inflammation Induced by Anti-BTLA Antibody in Dual Insult Model of Hemorrhagic Shock/ Sepsis. Shock 45, 40–49, https://doi.org/10.1097/SHK.00000000000479 (2016).
- Yoo, B. H. *et al.* Upregulation of ATG3 contributes to autophagy induced by the detachment of intestinal epithelial cells from the extracellular matrix, but promotes autophagy-independent apoptosis of the attached cells. *Autophagy* 11, 1230–1246, https://doi.or g/10.1080/15548627.2015.1056968 (2015).
- Bharath, L. P. et al. Impairment of autophagy in endothelial cells prevents shear-stress-induced increases in nitric oxide bioavailability. Can J Physiol Pharmacol 92, 605–612, https://doi.org/10.1139/cjpp-2014-0017 (2014).
- Ding, L. et al. Rank-based genome-wide analysis reveals the association of ryanodine receptor-2 gene variants with childhood asthma among human populations. Hum Genomics 7, 16, https://doi.org/10.1186/1479-7364-7-16 (2013).
- Wang, G. R. et al. Steroid-sensitive gene 1 is a novel cyclic GMP-dependent protein kinase I substrate in vascular smooth muscle cells. J Biol Chem 288, 24972–24983, https://doi.org/10.1074/jbc.M113.456244 (2013).
- Tremblay, F. et al. Loss of coiled-coil domain containing 80 negatively modulates glucose homeostasis in diet-induced obese mice. Endocrinology 153, 4290–4303, https://doi.org/10.1210/en.2012-1242 (2012).
- Jarrin, M., Pandit, T. & Gunhaga, L. A balance of FGF and BMP signals regulates cell cycle exit and Equarin expression in lens cells. Mol Biol Cell 23, 3266–3274, https://doi.org/10.1091/mbc.E12-01-0075 (2012).
- Song, X., Sato, Y., Sekiguchi, K., Tanaka, H. & Ohta, K. Equarin is involved in cell adhesion by means of heparan sulfate proteoglycan during lens development. Dev Dyn 242, 23–29, https://doi.org/10.1002/dvdy.23902 (2013).
- Ferraro, A. et al. Tumor suppressor role of the CL2/DRO1/CCDC80 gene in thyroid carcinogenesis. J Clin Endocrinol Metab 98, 2834–2843, https://doi.org/10.1210/jc.2012-2926 (2013).
- Huang, N., Lee, I., Marcotte, E. M. & Hurles, M. E. Characterising and predicting haploinsufficiency in the human genome. PLoS Genet 6, e1001154, https://doi.org/10.1371/journal.pgen.1001154 (2010).
- Karavitakis, E. et al. Microduplication 3q13.2q13.31 identified in a male with dysmorphic features and multiple congenital anomalies. Am J Med Genet A 164A, 666–670, https://doi.org/10.1002/ajmg.a.36346 (2014).
- Price, A. L. et al. Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet 38, 904–909, https://doi.org/10.1038/ng1847 (2006).
- McCarthy, S. et al. A reference panel of 64,976 haplotypes for genotype imputation. Nat Genet 48, 1279–1283, https://doi. org/10.1038/ng.3643 (2016).
- Delaneau, O., Marchini, J. & Zagury, J. F. A linear complexity phasing method for thousands of genomes. Nat Methods 9, 179–181, https://doi.org/10.1038/nmeth.1785 (2012).
- Pruim, R. J. et al. LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics 26, 2336–2337, https:// doi.org/10.1093/bioinformatics/btq419 (2010).
- San Lucas, F. A., Wang, G., Scheet, P. & Peng, B. Integrated annotation and analysis of genetic variants from next-generation sequencing studies with variant tools. *Bioinformatics* 28, 421–422, https://doi.org/10.1093/bioinformatics/btr667 (2012).
- McKenna, A. et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. Genome Res 20, 1297–1303, https://doi.org/10.1101/gr.107524.110 (2010).

Acknowledgements

P.X. and Z.G.O. are the recipient of fellowship from the Canadian Institutes of Health Research. G.A.R. funded this study from the Canadian Heart and Stroke Foundation. G.A.R. holds the Canada's Research Chair in Neurogenetics and the Wilder Penfield Chair in Neuroscience.

Author Contributions

S.Z. designed the study, performed the analyses and wrote the manuscript; Z.G.O., C.V.B. and N.D. provided samples and data; D.L. and T.M.F. provided validation data for the meta-analysis; A.A., P.X., A.D.L. and D.S. maintained the bioinformatics pipeline and helped the analyses; S.S. and J.P.R. co-performed the experiments; L.X. and P.A.D. provided suggestions to the study design and co-wrote the manuscript; G.A.R. led and oversaw the study, collected the samples and revised the manuscript.

Additional Information

Supplementary information accompanies this paper at https://doi.org/10.1038/s41598-018-21603-7.

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2018