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OPEN Enantiomeric excess by magnetic circular dichroism in Archaean atmosphere

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Evolution of homochirality requires an initial enantiomeric excess (EE) between right and left-handed biomolecules. We show that magnetic circular dichroism (MCD) of sun's ultraviolet C light by oxygen in Archaean earth's anoxic atmosphere followed by chirally selective damage of biomolecules due to circular dichroism (CD) can generate EE of correct handedness. Our calculation of EE uses published data for CD of biomolecules and accepted magnitude for Archaean earth's magnetic field. Independent of atmospheric oxygen concentration calculated EE has the same sign for all pyrimidine nucleosides which is opposite to that for amino-acids. Purine nucleosides have smaller EE values with opposite sign to pyrimidines but are less susceptible to UV damage. Homochirality is explained by origin of prebiotic life in one hemisphere of earth and its evolution to EE ~ \pm 1 before reversal of terrestrial magnetic field. Chirality of biomolecules is decided by the direction of magnetic field where prebiotic life originated on Archaean earth.

While homochirality is pervasive in molecules which form the building blocks of life¹, results of Urey-Miller experiment² form the basis of hypothesis that life originated on earth with a racemic mixture of prebiotic molecules. It is believed that evolution of homochirality rests on: (i) a mechanism to create initial enantiomeric excess (EE) between right (R) and left (L)-handed biomolecules and (ii) amplification of EE over time to generate 100% homochirality^{1,3}. Two possible mechanisms have been proposed to explain the initial EE which evolved to 100% homochirality over a period of time. One of these has an extra-terrestrial source and rests on the observed small EE of L-amino acids found on chondritic meteorites⁴. However, it is possible that some of the evidence on these meteorites may be due to contamination⁵ by the L-amino acids of our biosphere. Second mechanism is stochastic and the observed homochirality is explained as a "by-chance" phenomenon^{6,7}. Neither of these two mechanisms can explain why all five nucleosides are R-handed and all twenty standard amino acids (except achiral glycine) have the opposite L-handedness. Presence of terrestrial magnetic field can in principle break the chiral symmetry of biomolecules. In an effect labeled "magnetochiral dichroism", unpolarized light propagating in the direction of a magnetic field can produce an EE in some chemical reactions involving metal and organic compounds⁸. However, its role in creating an EE for biomolecules like nucleic and amino acids remains to be demonstrated.

We show that initial EE of correct handedness can be generated by MCD of ultraviolet C (UVC) light by paramagnetic oxygen molecules in the atmosphere of Archaean earth followed by chirally selective photo damage due to CD in biomolecules. Dextro (right-handed) ribose and deoxyribose sugar is primarily responsible for the chirality of nucleic acids. These sugars have strong absorption/CD bands only for vacuum UV wavelengths of less than 190 nm^{9,10}. As shown in this report, UV flux reaching Archean earth was limited to wavelengths above 200 nm making isolated sugar molecules immune to UV photolysis. Nucleosides are the smallest chiral building blocks of nucleic acids with strong circular dichroism for UV wavelengths in the 200-300 nm range and are analyzed together with amino-acids for EE. The model implies that enantiomeric enrichment by asymmetric photolysis with UV light happened after bigger molecules like nucleosides had formed.

Our analysis utilizes published data for CD of L-amino-acids and R-nucleosides and is based on the accepted understanding of paleomagnetism¹¹ and Archaean atmosphere ~4,000 Mya. Evidence from Archaean rocks related to mass independent fractionation of sulphur (MIF-S) isotopes suggests an anoxic atmosphere¹² with partial pressure of oxygen, $p(O_2) < 10^{-2}$ PAL (present atmospheric level) although there are reports of loss of MIF-S and possible oxic atmosphere even 3.8 billion years ago¹³. While MIF-S effect still needs to be understood¹⁴ to make quantitative conclusions, based on photochemical modeling, maximum pressure of atmospheric

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Figure 1. Logarithmic plots of spectral flux of UVC light reaching earth's surface for atmospheric CO2 pressure of 0.25, 1 and 2 bar and of absorption cross section of molecular oxygen by the forbidden transitions of the Herzberg bands. The figure shows that the spectral region outside of 200–300 nm is unimportant for magnetic circular dichroism by Archean atmospheric oxygen.

oxygen¹² that would support MIF-S is $p(O_2) \sim 10^{-2}$ PAL. As shown in this report the sign of EE of nucleosides and amino-acids is independent of oxygen concentration but depends significantly on the pressure of carbon-dioxide in Archaean atmosphere.

Results

Transmission of UVC in Archean atmosphere. For analysis in this report, carbon dioxide and oxygen are the two most important components of Archaean atmosphere when prebiotic life arose around 4 billion years ago. While CO₂ is primarily responsible for attenuating far UV sunlight in wavelength range of 200–300 nm¹⁵, O₂ is the only atmospheric component which is paramagnetic and capable of producing MCD of this UV light. Concentration of O_2 in atmosphere will determine the extent of MCD of UV light and the differential circular polarization (CP) intensity reaching earth's surface. Accepted models¹⁶ for earth's atmosphere around 4,000 Mya require high CO_2 pressure (0.1–10 bar) to compensate for lower solar luminosity of young Sun. Figure 1 is a plot of calculated UVC flux reaching early Archaean earth's surface $(p(CO_2) = 0.25, 1, 2 \text{ bar})$ and shows that wavelengths below 200 nm were strongly attenuated. In plotting these curves, accepted values are used for CO₂ absorption cross-sections, density-altitude profile and the spectral-intensity of sunlight reaching earth's upper atmosphere 4,000 Mya¹⁵. For wavelengths in 200-300 nm, molecular oxygen absorbs weakly by three forbidden transitions^{17,18}, giving rise to the Herzberg continuum which originates in paramagnetic $X^{3}\Sigma^{-}_{\sigma}$ ground state and is important for atmospheric physics. As seen in Fig. 1, total absorption cross section for these transitions in oxygen $(X^3\Sigma^-_g \rightarrow A^3\Sigma^+_u, X^3\Sigma^-_g \rightarrow c^1\Sigma^-_u \text{ and } X^3\Sigma^-_g \rightarrow A^3\Delta_u)$ rapidly decreases from 200 nm to 300 nm¹⁷⁻¹⁹. In laboratory, MCD of O₂ has been observed²⁰ within the Herzberg continuum by a matrix-isolation technique at low temperatures and high magnetic field. Attenuation by CO₂ and absorption cross-section for oxygen in Fig. 1 serves to explain why the effect of MCD by Archaean atmospheric oxygen is unimportant outside the spectral region of 200-300 nm.

Magnetic circular dichroism of UVC in Archean atmosphere. The dipolar geomagnetic field is as old¹¹ as the Earth itself with the dipole axis nearly but not exactly aligned with axis of earth's rotation. Paleomagnetism^{11,21} evidence suggests that the magnitude of magnetic field has not changed significantly since 4,000 Mya. The magnetic poles have wandered by as much as 15–20° but for the purpose of this analysis we will assume that the axis of dipole coincides with the axis of earth's rotation (Fig. 2). We also assume the magnetic field intensity at the equator of 0.3 Gauss which is the present average value²² for this latitude; and the tilt of earth's axis towards the ecliptic plane as $\theta_T = 23.4^\circ$ (current value). For MCD, the component (B_S) of geomagnetic field parallel to the direction of sunlight reaching earth at zenith (midday) varies with the latitude as well as the time of the year and is derived with reference to Fig. 2a. Close to the surface of Earth (radius R), the radial (B_r) component of the dipolar magnetic field and the component parallel (B_H) to earth's surface is given by²²,

$$B_{\rm r} = -2B_0 \operatorname{Sin}(\theta_{\rm L}) \quad \text{and} \quad B_{||} = B_0 \operatorname{Cos}(\theta_{\rm L}) \tag{1}$$

where B_0 is the magnetic field at the equator and θ_L is the latitude of the position on earth. Average value of B_0 on the equator is 0.3 Gauss.

For the geometry shown in Fig. 2a (northern solstice) when the North Pole is most tilted towards the sun, the component of earth's magnetic field in the direction of sunlight at zenith (midday) is:

$$B_{\rm S} = -B_{\rm r} \cos(\theta_{\rm L} - \theta_{\rm T}) + B_{\parallel} \sin(\theta_{\rm L} - \theta_{\rm T})$$
⁽²⁾



Figure 2. (a) Model of terrestrial magnetic dipole for calculating B_s , the component of field in the direction of sunlight, (b). Variation of B_s (component of geomagnetic field intensity in the direction of sunlight at zenith) with latitude for the two extremes of solstices. Dipolar magnetic field intensity of 0.3 Gauss at equator is assumed. Dark gray regions of latitude are relatively important for MCD where B_s does not change sign throughout the year and $|B_s| > 0.25$ Gauss. For light gray regions B_s changes sign in the course of a year and so are not important for MCD.

where $\theta_T = 23.4^\circ$ is the angle of tilt of the earth's axis towards the ecliptic plane. Similarly, for southern solstice when the North Pole is most tilted away from the sun, the component of earth's magnetic field in the direction of sunlight at zenith (midday) is:

$$B_{\rm S} = -B_{\rm r} \cos(\theta_{\rm L} + \theta_{\rm T}) + B_{||} \sin(\theta_{\rm L} + \theta_{\rm T})$$
(3)

Both B_r and $B_{||}$ vary²² with radial distance (r) as $(R/r)^3$ which shows that for distances of up to 100km above earth's surface B_S varies by less than 3%. Atmosphere of this thickness accounts for 99% of all gaseous oxygen around earth²².

Magnetic field B_s is shown in Fig. 2b for two different times of the year, i.e. for northern and southern solstices. For given latitude, B_s varies over period of a year between the two extremes of solstices. As shown by the shaded (dark gray) region in Fig. 2b, only for a small range of latitudes (24°N to 58°N and 24°S to 58°S), B_s remains significant ($|B_s| > 0.25$ Gauss) throughout the year without changing sign. In the light-gray regions (close to equator and the poles) of Fig. 2b, B_s changes sign in the course of a year. With these assumptions, 24°N to 58°N and 24°S to 58°S are the most important geographic regions where the effects of MCD are observed.

The significance of MCD-related transitions²⁰ in molecular oxygen which results in net circular polarization of UVC light is explained with Fig. 3. Of the three forbidden transitions, $X^3\Sigma^-_g \rightarrow A^3\Sigma^+_u$ is the strongest, accounting for most (86%) of the absorption¹⁸ in Herzberg continuum. Zeeman splitting of states by the small (~0.25 Gauss) geomagnetic field and possible transitions with left circularly polarized (LCP, σ^+) and right circularly polarized (RCP, σ^-) light (propagating in the direction of magnetic field) are shown schematically. In the ground $X^3\Sigma^-_g$ state, Boltzmann distribution will result in a small excess of population in $M_S = -1$ over $M_S = +1$. As seen in this figure, for $X^3\Sigma^-_g \rightarrow A^3\Sigma^+_u$ and $X^3\Sigma^-_g \rightarrow c^1\Sigma^-_u$ transitions, $M_S = -1$ in the ground state can only absorb σ^+ light while $M_S = +1$ can only absorb σ^- light. Population difference between $M_S = -1$ and $M_S = +1$ results in slightly greater absorption of σ^+ light over σ^- . This is not possible with $X^3\Sigma^-_g \rightarrow A'^3\Delta_u$ transition (Fig. 3). Net



Figure 3. Energy levels for molecular absorption from the $X^{3}\Sigma^{-}_{g}$ ground state of paramagnetic molecular oxygen. X \rightarrow A accounts for bulk of the absorption in the forbidden Herzberg band and is most important for MCD. Due to Boltzmann distribution in the ground state magnetic sublevels, σ^{+} (LCP) light will be absorbed more than σ^{-} (RCP) light and there will be a small relative excess of σ^{-} light reaching earth's surface for magnetic field in the direction of light propagation.

(albeit very small) circular polarization of light reaching earth's surface is a result of ${}^{3}\Sigma \rightarrow \Sigma$ transitions in molecular oxygen, i.e. UV light reaching earth will be net σ^{-} when propagating in the direction of earth's magnetic field and net σ^{+} when propagating against the direction of magnetic field.

Differential (net) circular polarization (CP) intensity due to MCD by atmospheric O₂ is derived as follows: If $I_+(\lambda)$ and $I_-(\lambda)$ is the spectral intensity (W.m⁻². nm⁻¹) of LCP (σ^+) and RCP (σ^-) circularly polarized solar UV radiation respectively, reaching the earth's surface after passing through the atmosphere,

$$I_{+}(\lambda) = \frac{I_{0}(\lambda)}{2}e^{-\alpha_{+}} \quad \text{and} \quad I_{-}(\lambda) = \frac{I_{0}(\lambda)}{2}e^{-\alpha_{-}}$$
(4)

 $I_0(\lambda)$ is the initial (above atmosphere) unpolarized intensity (equal components of RCP and LCP). Coefficients α_+ and α_- describe bulk of the atmospheric absorption²³ by CO₂ and relatively very weak differential absorption of circularly polarized light (MCD) by atmospheric O₂.

$$\alpha_{+}(\lambda) = \sigma(CO_{2}, \lambda). \int N(CO_{2}, L). \, dL + \sigma_{+}(O_{2}, \lambda). \int N_{-}(O_{2}, L). \, dL$$
(5a)

Likewise,

$$\alpha_{-}(\lambda) = \sigma(CO_{2}, \lambda). \int N(CO_{2}, L). \ dL + \sigma_{-}(O_{2}, \lambda). \int N_{+}(O_{2}, L). \ dL$$
(5b)

The integration is over altitude (L) between 0 and 100 km of atmosphere which adequately²⁴ includes absorption by more than 99% of CO_2 and O_2 .

 $\sigma(CO_2, \lambda)$: Absorption cross-section for CO₂ at wavelength λ

 $N(CO_2, L)$: Number density of CO_2 at altitude L

 $\sigma_+(O_2, \lambda)$: absorption cross-section for O_2 at wavelength λ (LCP light)

 $\sigma_{-}(O_2, \lambda)$: absorption cross-section for O_2 at wavelength λ (RCP light)

 $N_{-}(O_2, L)$: Number density of O_2 in $M_s = -1$ at altitude L

 $N_{+}(O_2, L)$: Number density of O_2 in $M_s = +1$ at altitude L

For small magnetic field, H \approx 0.25 Gauss, the magnetic sublevels of O₂ are degenerate and we assume, $\sigma_+(O_2, \lambda) = \sigma_-(O_2, \lambda) \equiv \sigma(O_2, \lambda)$

Boltzmann distribution gives,

$$\left(\frac{N_{-}(O_{2}, L)}{N_{+}(O_{2}, L)}\right) = e^{\Delta E(H)/kT}$$
(6)

Here, $\Delta E(H) = \mu_B .g. \Delta M_S.H$ is the energy difference between $M_S = -1$ and $M_S = 1$ Zeeman levels for magnetic field H; μ_B is the Bohr magneton, g is the Lande factor, $\Delta M_S = 2$ for the two relevant sublevels of the ground state $X^3 \Sigma_{g}^-$ and k is the Bohrzmann constant. We assume, $H \equiv B_S = 0.25$ Gauss, Lande g factor²⁵ ≈ 2 and T = 300 K giving, $\Delta E/kT = 2.24 \times 10^{-7}$.

Due to three nearly degenerate magnetic sublevels of the ground state of O_2 ,

$$N_{-}(O_2, L) = \frac{N(O_2, L)}{3}$$
 and $N_{+}(O_2, L) = \frac{N(O_2, L)}{3}$. $e^{-\Delta E(H)/kT}$ (7)

 $N(O_2, L)$ is the total number density of O_2 at altitude L. Spectral intensities $I_+(\lambda)$ and $I_-(\lambda)$ are calculated from above equations, published data for absorption cross-sections $\sigma(CO_2, \lambda)^{15}$ and $\sigma(O_2, \lambda)^{17-19}$, and the density-altitude profile¹⁵ for given surface pressure of CO_2 . We assume complete mixing of CO_2 and O_2 , i.e., $N(O_2, L)/N(CO_2, L)$ is independent of altitude L. Differential circular polarization (CP) intensity $\Delta I(\lambda) \equiv (I_-(\lambda) - I_+(\lambda))$



Figure 4. Differential circular polarization intensity for $p(O_2) = 0.002$ bar and $p(CO_2) = 0.5$ bar. (a) $\Delta E/kT = 2.24 \times 10^{-7}$, (b) $\Delta E/kT = 4.48 \times 10^{-7}$.

is calculated from Eqns 4–7 and shown in Fig. 4 for a surface O₂ pressure $p(O_2) = 10^{-2} \text{ PAL} = 0.002 \text{ bar}$ and CO₂ pressure of 0.5 bar. Similar plots can be generated for any other surface O₂ and CO₂ pressure. As seen in this figure, $\Delta I(\lambda)$ is proportional to B_s/T .

Enantiomeric excess for biomolecules. Calculation of EE is made for biomolecules in aqueous medium as is the case with most theories for origin of prebiotic life in marine/freshwater environment^{26,27}. The products of Miller Urey type reactions with atmospheric gases collected over the large surface area of Archean oceans. Concentration of amino acids in Archean oceans is estimated to $vary^{27,28}$ from 4×10^{-3} M to 10^{-7} M. Concentration of five nucleobases in one Miller-Urey experiment²⁹ was measured to be between 1–100 ppm. While these numbers are small, there are several proposed mechanisms for amplifying the concentration of prebiotic molecules on ocean shorelines and drying freshwater ponds³⁰. Additionally, enantiomeric excess depends on the ratio and not absolute values of enantiomeric concentrations. As shown in a recent report³¹, around 4000 Mya the pH of seawater was in the 6.5–7.0 range and not too different from freshwater. Data used in this report is for solutions of pH around 7. While asymmetric photolysis and enantiomeric enrichment of amino acids by circularly polarized light in liquid varies with the acidic pH in the range of 2–6, there is no effect of pH in the 6.5–7.0 range or higher^{32,33}.

Chiral molecules like nucleic acid monomers and amino acids show circular dichroism^{34,35} in the 200–300 nm region of UV light. Starting with a racemic population of these prebiotic building blocks of life, exposure to net circularly polarized UVC light will result in an enantiomeric excess that will be a seed for evolution to homochirality. Such an explanation has been proposed⁴ for the observed enantiomeric excess of L-amino acids in chondritic meteorites and assumes that the UV damage/inactivation rates D_R , D_L of right-/left-handed biomolecules are proportional to light absorption rates. This is also seen from the observed similarity of absorption spectrum and action spectrum for direct photo-damage of nucleic acids by UVC radiation^{36,37}. Using published CD parameter $\Delta \epsilon(\lambda)^{34,35}$, extinction coefficient $\epsilon(\lambda)^{35,38-40}$ and calculated differential CP intensity $\Delta I(\lambda) \equiv (I_-(\lambda) - I_+(\lambda))$, the differential UV damage rate $\Delta D/D \equiv (D_R - D_L)/(D_R + D_L)$ for nucleosides and amino acids is derived below. As shown here, enantiomeric excess is related to $\Delta D/D$.

Cross sections for absorption of UV by biomolecules (amino-acids/nucleosides) are listed below.

 $\sigma_R^+(\lambda)$: Cross-section for absorption of σ^+ light by right-handed (RH) molecule

 $\sigma_R^{-}(\lambda)$: Cross-section for absorption of σ^{-} light by right-handed (RH) molecule

 $\sigma_L^+(\lambda)$: Cross-section for absorption of σ^+ light by left-handed (LH) molecule

 $\sigma_L^{-}(\lambda)$: Cross-section for absorption of σ^- light by left-handed (LH) molecule

For the two kinds of chiral molecules (RH/LH) and two circular light polarizations (σ^+ and σ^-), following rate equations are valid. LCP (σ_+) energy absorbed per sec by one RH molecule is⁴¹

$$\Gamma_{R}^{+} = \int I_{+}(\lambda). \ \sigma_{R}^{+}(\lambda). \ d\lambda$$
(8a)

Similarly, absorption rates for other three combinations,

$$\Gamma_{R}^{-} = \int I_{-}(\lambda). \ \sigma_{R}^{-}(\lambda). \ d\lambda$$
(8b)

$$\Gamma_L^+ = \int I_+(\lambda). \ \sigma_L^+(\lambda). \ d\lambda \tag{8c}$$

$$\Gamma_L^- = \int I_-(\lambda). \ \sigma_L^-(\lambda). \ d\lambda \tag{8d}$$

Since chiral molecules can absorb both σ^+ and σ^- components of UV light, total absorption rates for RH and LH enantiomers are:

$$\Gamma_{\!R} = \Gamma_{\!R}^+ + \Gamma_{\!R}^- \tag{9a}$$

$$\Gamma_L = \Gamma_L^+ + \Gamma_L^- \tag{9b}$$

From chiral symmetry, $\sigma_R^+(\lambda) = \sigma_L^-(\lambda)$ and $\sigma_R^-(\lambda) = \sigma_L^+(\lambda)$. As before, differential circular polarization (CP) intensity due to MCD by atmospheric O₂ is $\Delta I(\lambda) \equiv I_-(\lambda) - I_+(\lambda)$. Define, $\Delta \sigma(\lambda) \equiv \sigma_L^+(\lambda) - \sigma_L^-(\lambda) = \sigma_R^-(\lambda) - \sigma_R^+(\lambda)$. It follows from above rate Eqs 8 and 9,

$$\Gamma_{R} - \Gamma_{L} = \int \Delta I(\lambda). \ \Delta \sigma(\lambda). \ d\lambda$$
(10a)

Similarly,

$$\Gamma_{R} + \Gamma_{L} = \int I_{0}(\lambda). \ \sigma(\lambda). \ d\lambda$$
(10b)

Here, $I_{-}(\lambda) \approx I_{+}(\lambda) = I_{0}(\lambda)/2$ and $\sigma(\lambda) = \sigma_{L}^{+}(\lambda) + \sigma_{L}^{-}(\lambda) = \sigma_{R}^{+}(\lambda) + \sigma_{R}^{-}(\lambda)$.

Molar circular dichroism is defined as $\Delta \varepsilon = \varepsilon^+ - \varepsilon^-$, where ε^+ and ε^- are the molar extinction coefficients $(mol^{-1}. cm^{-1})$ for σ^+ and σ^- light respectively. Absorption cross-section $\sigma(\lambda)$ is proportional⁴² to $\varepsilon(\lambda)$. Thus,

$$\frac{\Gamma_{R} - \Gamma_{L}}{\Gamma_{R} + \Gamma_{L}} = \frac{\int \Delta I(\lambda). \ \Delta \sigma(\lambda). \ d\lambda}{\int I_{0}(\lambda). \ \sigma(\lambda). \ d\lambda} = \frac{\int \Delta I(\lambda). \ \Delta \varepsilon_{L}(\lambda). \ d\lambda}{\int I_{0}(\lambda). \ \varepsilon(\lambda). \ d\lambda} = -\frac{\int \Delta I(\lambda). \ \Delta \varepsilon_{R}(\lambda). \ d\lambda}{\int I_{0}(\lambda). \ \varepsilon(\lambda). \ d\lambda}$$
(11a)

Here, $\Delta \varepsilon_L = \varepsilon_L^+ - \varepsilon_L^-$ for L-handed molecule and $\Delta \varepsilon_R = \varepsilon_R^+ - \varepsilon_R^-$ for R-handed molecule. Since $\Delta \varepsilon(\lambda)$ is typically less than 1% of $\varepsilon(\lambda)$, in the denominator $\varepsilon(\lambda) \approx \varepsilon_L(\lambda) \approx \varepsilon_R(\lambda)$. It is commonly assumed that the UV destruction/damage rate (D) for nucleic acids^{36,37} is proportional to the light energy absorption rate (Γ). Thus,

$$\frac{\Delta D}{D} \equiv \frac{D_R - D_L}{D_R + D_L} = \frac{\int \Delta I(\lambda). \ \Delta \varepsilon_L(\lambda). \ d\lambda}{\int I_0(\lambda). \ \varepsilon(\lambda). \ d\lambda} = -\frac{\int \Delta I(\lambda). \ \Delta \varepsilon_R(\lambda). \ d\lambda}{\int I_0(\lambda). \ \varepsilon(\lambda). \ d\lambda}$$
(11b)

Relative efficiency for photolysis has been studied only for a few amino acids and for some isolated wavelengths. As an example, photolysis of phenylalanine at 206 nm and 254 nm does not show any significant effect of wavelength⁴³. Further, CD of amino acids is significant only over a band-width of 20–30 nm³⁴. Above equation to calculate $\Delta D/D$ is used for nucleosides and amino acids.

In eqn. 11, $\Delta \epsilon_L(\lambda)$, $\Delta \epsilon_R(\lambda)$ is the molar CD parameter for L/R-handed enantiomer. Enantiomeric excess $(N_R - N_L)/(N_R + N_L)$ is the relative excess of right-handed molecular concentration (N_R) over left-handed (N_L) molecules. It is related to $\Delta D/D$ and the sign of EE is opposite to that of $\Delta D/D$ (if $D_R > D_L$, $N_R < N_L$). In a simple model described below, for racemic production and UV damage of biomolecules enantiomeric excess, $EE = -(\Delta D/D)$.

Model for racemic production and UV damage of biomolecules. Correlation between EE and the parameter ($\Delta D/D$) is explained with the help of a simple phenomenological model that describes the racemic production of prebiotic life-molecules and their destruction by Archaean UVC radiation. For number density N_R and N_L of right and left handed molecules,

$$\frac{dN_R}{dt} = -D_R N_R + P \quad \text{and} \quad \frac{dN_L}{dt} = -D_L N_L + P \tag{12}$$

The terms $-D_R N_{R_c} - D_L N_L$ describe destruction by UV light with rate constants D_R and D_L which could be different due to differential circular polarization intensity, $\Delta I(\lambda) \equiv I_-(\lambda) - I_+(\lambda)$. The racemic production rate P is same for both enantiomers. For time, $t \gg D_R^{-1}$, D_L^{-1} the steady-state distribution ($dN_{R,L}/dt \approx 0$) results in an enantiomeric excess,

$$EE \equiv \frac{N_R - N_L}{N_R + N_L} = -\left(\frac{D_R - D_L}{D_R + D_L}\right) \equiv -\frac{\Delta D}{D}$$
(13a)

For time, $t \ll D_R^{-1}$, D_L^{-1}

$$EE \equiv \frac{N_R - N_L}{N_R + N_L} = -(D_R - D_L). \ \frac{t}{4} \equiv -\frac{\Delta D}{D} \left(\frac{Dt}{4}\right)$$
(13b)

L-amino-acids ³⁴ : All CD bands are positive for wavelength in 200–300 nm range								
Alanine	Peak wavelength: 203 nm; Bandwidth (FWHM): 22 nm							
Serine	Peak wavelength: 205 nm; Bandwidth (FWHM): 25 nm							
Lysine	Peak wavelength: 212 nm; Bandwidth (FWHM): 23 nm							
Proline	Peak wavelength: 210 nm; Bandwidth (FWHM): 19 nm							
Phenylalanine	Peak wavelength: 219 nm; Bandwidth (FWHM): 9 nm							
R-nucleosides ³⁵ (pyrimidines)								
Cytidine	Negative band between 200–240 nm; positive bands for 240–300 nm							
Thymidine	Negative band between 208–254 nm; positive bands for 254–300 nm and 180–208 nm							
Uridine	Negative band between 208–250 nm; positive bands for 250–290 nm and 182–208 nm							
R-nucleosides ³⁵ (purines)								
Adenosine	Positive band for 210–240 nm; negative bands for 240–280 nm and 170–210 nm							
Guanosine	Positive band for 206–232 nm; negative bands for 232–290 nm and 170–206 nm							

Table 1. Summary of Circular Dichroism (CD) bands for L-amino-acids and R-nucleosides. Calculated values of EE for different pressures of atmospheric CO₂ and O₂ is given in Table 2 for nucleosides and five randomly selected amino-acids for a scenario where the intensity of σ^- light reaching earth's surface is more than σ^+ intensity ($\Delta I(\lambda) \equiv I_-(\lambda) - I_+(\lambda) > 0$).

		Enantiomeric Excess $(-\Delta D/D)$											
							Nucleosides (EE in unit of 10 ⁻¹⁵)						
$P(CO_2)$	P(O ₂) Bar	Amino Acids (EE in unit of 10 ⁻¹⁰)				Pyrimidines			Purines				
bar		Ala	Ser	Lys	Pro	Phe	Cytidine	Thymidine	Uridine	Adenosine	Guanosine		
0.25	0.002	-0.81	-1.2	-1.3	-0.69	-0.0088	-29.1	-33.7	-35.6	2.2	10.7		
0.50	0.002	-0.77	-1.2	-1.5	-0.78	-0.0041	-0.11	-8.0	-5.5	-1.2	1.2		
0.75	0.002	-0.77	-1.2	-1.7	-0.87	-0.0013	4.5	-1.0	2.4	-1.7	-0.23		
1.00	0.002	-0.81	-1.3	-2.0	-0.97	-0.00039	4.5	0.93	4.4	-1.5	-0.29		
1.25	0.002	-0.91	-1.5	-2.4	-1.1	-0.00012	3.8	1.5	4.6	-1.2	-0.21		
1.25	0.0002	-0.085	-0.014	-0.22	-0.010	-0.000012	0.38	0.14	0.46	-0.12	-0.021		
1.50	0.002	-1.1	-1.7	-2.9	-1.3	-0.000039	3.2	1.5	4.3	-0.94	-0.13		
1.75	0.002	-1.4	-2.0	-3.7	-1.5	-0.000013	2.6	1.4	3.9	-0.72	-0.088		
2.00	0.002	-1.8	-2.5	-4.8	-1.8	-0.0000046	2.2	1.3	3.4	-0.55	-0.060		

Table 2. Calculated Enantiomeric Excess (EE) for atmospheric CO₂ pressure between 0.25–2 bar and an O₂ pressure of 0.002 bar. EE values are for $I_{-}(\lambda) > I_{+}(\lambda)$, i.e. ${}^{\Delta}\Delta I(\lambda) > 0$ and when UVC light reaching earth's surface is preferentially σ^{-} (RCP).

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Equation 11b is used to calculate EE ($-\Delta D/D$) using published values of CD parameter $\Delta \varepsilon(\lambda)^{34,35}$ and extinction coefficient $\varepsilon(\lambda)^{35,38-40}$ for R-nucleosides and L-amino-acids. A summary of reported ^{34,35} CD bands for these biomolecules is given in Table 1.

Discussion

UV damage rates $D_{R,L}$ depend on the action spectrum weighted, integrated UVC flux. For 200–300 nm and CO_2 pressure of 1 bar, total integrated UVC flux is 0.63 W/m² (Fig. 1) and is equivalent to 0.21 W/m² when weighted with the DNA action spectrum³⁷. Several studies have measured wavelength specific damage rate constants for nucleic acids which is largely due to photo-damage of pyrimidine bases. From the measured nucleic acid damage rate constant of 0.061 cm²/mJ for wavelength of 260 nm⁴⁴, $D_{R,L}$ has a value of $1.2 \times 10^{-3} s^{-1}$. When subjected to Archean UVC flux, pyrimidine nucleosides will be damaged in a time of few minutes. The UV damage rate for purine nucleosides is much smaller than for pyrimidine nucleosides ($D_{purine} \approx 0.01 D_{pyrimidine}$)⁴⁵ and the corresponding damage time for purine nucleosides in the Archaean UV flux will be several hours. As seen from Equation 13a,b, for the estimated Archaean UVC flux, EE for pyrimidines will saturate to $-\Delta D/D$ in a time of few minutes but will take several hours for purines to do so. Compared to pyrimidines, the Archaean purine population density is racemic (EE \approx 0).

Relatively fewer studies have measured wavelength specific UV photolysis rate constants for amino acids. Photolysis of amino acids for wavelengths shorter than 200 nm was investigated for space environment^{46,47} outside of terrestrial atmosphere where UV light at these wavelengths is relatively abundant. Photolysis has also been investigated for wavelengths longer than 200 nm⁴⁸ and these are the only ones relevant to our investigation due to absence of vacuum UV light on Archean earth. In one investigation⁴³, a comparison was made of the relative effectiveness for amino-acid photolysis at wavelengths of 147 nm, 206 nm and 254 nm. The results show that photolysis effectiveness at these three wavelengths is comparable. For UV flux of 2.66 W/m² at a wavelength of 254 nm, the measured half-life of phenylalanine was 130 min and for UV flux of 8.34 W/m² at a wavelength of 206 nm, it was 38 min. Without any wavelength specific damage action spectra for amino acids for 200–300 nm, it

is not possible to correlate these numbers with the Archaen UVC flux shown in Fig. 1. However, in the absence of any dramatic variation of photolysis rates with UVC wavelength, amino-acid half-life of a few hours is expected⁴³.

The UV absorption coefficient of water varies from 7 m⁻¹ for a wavelength of 200 nm to 0.7 m⁻¹ for 300 nm and has a value of 1.7 m^{-1} for the central wavelength of 250 nm⁴⁹. Thus UV light in this wavelength range easily penetrates around 0.5 meters of water. The products of Urey Miller type reactions will continue interacting with UV light till the molecules have sunk to depths greater than ~0.5 meters by diffusion. Diffusion coefficient (D) for molecules of the size of nucleosides and amino acids in water has a value⁵⁰ of ~ $1.5 \times 10^{-5} \text{ cm}^2/\text{s}$. Time to diffuse⁵¹ down by a distance (L) of 0.5 m is (T $\approx L^2/4D$) several months. This can be compared to a time of several minutes to a few hours required for photodamage of biomolecules in the Archean UV light flux, as explained above. Thus biomolecules in water get adequately illuminated for the photodamage processes described in this report with near-full UV flux from sun.

As seen from Table 1, $\Delta \epsilon_L$ is positive³⁴ for CD of L-amino-acids in 200–300 nm wavelength region. From equation 11b, $\Delta D/D$ is positive for $\Delta I(\lambda) > 0$. Thus EE is negative $(N_L > N_R)$ for amino-acids and the sign of EE is independent of CO₂ pressure (Table 2). Unlike amino-acids, the CD spectra of nucleosides show both positive and negative bands³⁵ for $\Delta \epsilon_R$ (Table 1). Increasing atmospheric CO₂ shifts the UV pass-band (Fig. 1) and the differential circular polarization (Fig. 4) to longer wavelengths, resulting in a change of sign of $\Delta D/D$ (and EE) for pyrimidine nucleosides. As seen in Table 2, for pyrimidine nucleosides EE is positive ($N_R > N_L$) above a threshold CO₂ pressure of around 1 bar. In these calculations, the pressure of atmospheric O₂ is kept at 0.002 bar (0.01 PAL). At low pressures of O₂, calculated values of EE is approximately proportional to O₂ pressure. This is shown (Table 2) for CO₂ pressure of 1.25 bar and lower O₂ pressure of 0.0002 bar. While magnitude of EE values reduce by a factor of 10, the sign of EE remains unchanged.

Interestingly, the magnitude of EE for both purine nucleosides is smaller and the sign is opposite to that for pyrimidines. However, as explained above (Eqn. 13), compared to pyrimidines EE will take much longer time to reach the saturation value of $-\Delta D/D$ and the population of purines is relatively racemic. Further, heterochiral nucleic acids involving pyrimidine and purine building blocks are significantly more unstable thermally^{52,53} as compared to homochiral counterparts. It is conceivable that smaller EE values for purines and significant thermal stability of homochiral nucleic acids together with $D_{purine} \approx 0.01 (D_{pyrimidine})^{45}$ was a determining factor (not the sign of their enantiomeric excess) in the evolution of chirality of purines. Values of $\Delta D/D$ for aromatic amino-acids are lower by a factor of ~100 due to larger extinction coefficient $\varepsilon(\lambda)^{38}$ and relatively small circular dichroism $\Delta \varepsilon(\lambda)^{34}$. In all of these calculations we assume an ambient temperature (T) of 300 K and a magnetic field (B_S) of 0.25 Gauss. Varying conditions of atmospheric temperature and magnetic field can be accounted by the observation that differential circular polarization intensity $\Delta I(\lambda)$ and EE are proportional to B_S/T (Fig. 4).

Conclusions

We have shown that magnetic circular dichroism of UVC by sparse oxygen in the anoxic Archaean atmosphere results in net circularly polarized light reaching earth's surface. This results in chirally selective damage of prebiotic molecules by circular dichroism and creates an EE which evolved to homochirality of R-nucleosides and L-amino-acids. Irrespective of the partial pressure of oxygen, the sign of enantiomeric excess of pyrimidine nucleosides and the amino-acids is predicted correctly when UVC light reaching earth has net σ^- circular polarization $(\Delta I(\lambda) \equiv I_-(\lambda) - I_+(\lambda) > 0)$ and P(CO₂) > 1 bar. This is consistent with the requirement of large CO₂ pressure for greenhouse effect¹⁶ to balance lower solar luminosity 4,000 Mya. It is also an indication that initial enantiomeric excess was generated by circularly polarized UV light and not by a stochastic event.

Since MCD depends on the direction of magnetic field with respect to the propagation of light, net σ^- light in one hemisphere of earth would also result in net σ^+ light in the other hemisphere. Homochirality in life-molecules can be explained by assuming (i) prebiotic life originated either in the northern or the southern hemisphere of earth which had higher UVC flux of σ^- over σ^+ (south magnetic hemisphere), (ii) evolution of homochirality was complete (EE ~ ±1) before the earth's magnetic field reversed. Recorded length of polarity intervals between reversals varies between 0.1 and several Million years⁵⁴ and (iii) no significant dispersal of prebiotic life molecules happened between hemispheres before the transition to homochirality was complete. For the terrestrial magnetic field in Fig. 2, the most important regions for effective MCD in atmospheric oxygen lie between latitudes of 24° and 58° in both hemispheres and any EE will be nullified by large-scale dispersal of enantiomers between these regions from south to north magnetic hemisphere.

Methods

Calculation of spectral flux I(λ) (W. m⁻². nm⁻¹) of UVC light reaching earth's surface Calculation of differential circular polarization intensity Δ I(λ) (W.m⁻².nm⁻¹) of UVC light reaching earth's surface. Equations 4 and 5 together with published data¹⁵ for the following is used:

1a. Spectral flux $I_0(\lambda)$ (200–300 nm) from model young sun (~4,000 Mya) reaching earth (before entering the atmosphere)

1b. Absorption cross-sections (200–300 nm) for CO₂

1c. Variation of density with altitude (up to 120 km) for CO₂ and O₂. Altitude up 100 km accounts for 99% of these gases.

1d. Absorption cross-sections (200–300 nm) for O_2 is from refs^{17–19}.

Calculation of enantiomeric excess ($-\Delta D/D$ **) from equation 11b.** Published data (200–300 nm) for the following is used:

2a. Extinction coefficient ε (mol-1. cm-1) for the four amino acids (Ala, Ser, Lys, Pro) is significant³⁸ only for 200–230 nm. For aromatic amino acid (Phe), ε is significant between 200–270 nm.

2b. Circular dichroism (milli.deg) for a mino acids³⁴ is converted to $\Delta\epsilon$ (mol -1.cm-1) and is significant only for 200–240 nm

Due to a small range of wavelengths for a mino-acids, published data for $\Delta\epsilon$ and ϵ values is taken in steps of 1 nm.

Following equation (11b) is used to evaluate EE ($-\Delta D/D$) for amino-acids

$$EE = -\frac{\Delta D}{D} = -\frac{\int \Delta I(\lambda). \ \Delta \varepsilon_L(\lambda). \ d\lambda}{\int I_0(\lambda). \ \varepsilon(\lambda). \ d\lambda} = -\frac{\sum_{\lambda=200nm}^{240} \Delta I(\lambda). \ \Delta \varepsilon_L(\lambda)}{\sum_{\lambda=200nm}^{230} I(\lambda). \ \varepsilon_L(\lambda)}$$

Summation is done in steps of 1 nm.

2c. Extinction coefficient 39,40 ϵ (mol⁻¹. cm⁻¹) and circular dichroism $\Delta\epsilon$ (mol⁻¹. cm⁻¹) for the five nucleo-sides 35 in 200–300 nm

Due to a larger range of wavelengths for nucleosides, published data for $\Delta \varepsilon$ and ε values is taken in steps of 2 nm.

Following equation (11b) is used to evaluate EE ($-\Delta D/D$) for five nucleosides

$$EE = -\frac{D}{D} = \frac{\int \Delta I(\lambda). \ \Delta \varepsilon_R(\lambda). \ d\lambda}{\int I_0(\lambda). \ \varepsilon(\lambda). \ d\lambda} = \frac{\sum_{\lambda=200nm}^{300} \Delta I(\lambda). \ \Delta \varepsilon_R(\lambda)}{\sum_{\lambda=200nm}^{300} I(\lambda). \ \varepsilon_R(\lambda)}$$

Summation is done in steps of 2 nm. As seen from Figs 1 and 4, there is very little UV light intensity for wavelengths shorter than 220 nm and there is insignificant contribution to EE from this wavelength region. Likewise, differential circular polarization intensity (Fig. 4) is minimal for wavelength longer than 290 nm and again there is insignificant contribution from this long-wavelength region.

Data availability. The author declares that the main data supporting the findings of this study are available within this article. Additional data used in this study is available publicly in published journal articles. Appropriate references are provided (Methods section) for the sources of this data.

References

- 1. Blackmond, D. G. The origin of biological homochirality. Philos. Trans. R. Soc. Lond. B Biol. 366, 2878-2884 (2011).
- 2. Miller, S. L. & Urey, H. C. Organic compound synthesis on the primitive earth. Science 130, 245-251 (1959).
- Breslow, R. & Cheng, Z.-L. On the origin of terrestrial homochirality for nucleosides and amino acids. PNAS 106, 9144–9146 (2009).
 Cronin, J. R. & Pizzarello., S. Enantiomeric excesses in meteoric amino acids. Science 275, 951–955 (1997).
- Cronin, J. A. & Pizzareno, S. Enantioneric excesses in infectoric animo acids. Science 273, 951–955 (1997).
 Pizzarello, S., Huang, Y. & Alexandre, M. R. Molecular asymmetry in extraterrestrial chemistry: Insights from a pristine meteorite.
- PNAS 105, 3700–3704 (2008).
- Jafarpour, F., Biancalni, T. & Goldenfeld, N. Noise induced symmetry breaking far from equilibrium and the emergence of biological homochirality. *Phys. Rev. E* 95, 032407–18 (2017).
- 7. Mislow, K. Absolute asymmetric synthesis: A commentary. Collect. Czech. Chem. Commun. 68, 849-864 (2003).
- 8. Hattori, S., Yamamoto, Y., Miyatake, T. & Ishii, K. Magneto chiral dichroism measurements using a pulsed electromagnetic. *Chem. Phys. Lett.* **674**, 38–41 (2017).
- 9. Ito, A. & Ito, T. Absorption spectra of deoxyribose, ribosephosphate, ATP and DNA by direct transmission measurements in the vacuum-UV (150–190 nm) regions using synchrotron radiation as a light source. *Photochem. Photobiol.* 44, 355–358 (1986).
- Nielson, S. B., Chakraborty, T. & Hoffman, S. V. Synchrotron radiation circular dichroism spectroscopy of ribose and deoxyribose sugars, AMP and dAMP nucleotides. *ChemPhysChem* 6, 2619–2624 (2005).
- 11. Tarduno, J. A., Cottrell, R. D., Davis, W. J., Nimmo, F. & Bono, R. K. A Hadean to Paleoarchean geodynamo recorded by single zircon crystals. *Science* 349, 521–524 (2015).
- 12. Farquhar, J. & Wing, B. A. Multiple sulphur isotopes and the evolution of the atmosphere. Earth Planet, Sci. Lett. 213, 1–13 (2003).
- Ohmoto, H., Watanabe, Y., Ikemi, H., Poulson, S. R. & Taylor, B. E. Sulphur isotope evidence for an oxic Archean atmosphere. *Nature* 442, 908–911 (2006).
- 14. Babikov., D., Semenov, A. & Teplukhin, A. One possible source of mass-independent fractionation of sulphur isotopes in the Archean atmosphere of Earth. *Geochimica et Cosmochimica Acta* 204, 388–406 (2017).
- 15. Cnossen, I. et al. The habitat of early life: Solar X-ray and UV radiation at Earth's surface 4-3.5 billion years ago. J. Geophys. Res. Atmosph. 112, E02008–10 (2007).
- 16. Kasting, J. F. Earth's early atmosphere. Science 259, 920-926 (1993).
- Amoruso, A., Crescentini, L., Cola, M. S. & Fiocco, G. Oxygen absorption cross-section in the Herzberg continuum. J. Quant. Spectrosc. Radiat. Transfer 56, 145–152 (1996).
- Yoshino, K., Esmond, J. R. & Parinson, W. H. Fourier transform spectroscopy and cross section measurements of the Herzberg III bands of O₂ at 295 K. J. Chem. Phys. 112, 9791–9801 (2000).
- Merienne, M.-F. et al. Fourier transform spectroscopy of O₂ Herzberg bands II. Band oscillator strengths and transition moments. J. Mol. Spectrosc. 202, 171–193 (2000).
- Douglas, I. N., Grinter, R. & Thomson, A. J. The magnetic circular dichroism spectrum of matrix-isolated oxygen. *Chem. Phys. Lett.* 28, 192–196 (1974).
- 21. Tarduno, J. A. et al. Geodynamo, solar wind, and magnetopause 3.4 to 3.45 billion years ago. Science 327, 1238-1240 (2010).
- Merrill, R. T., McElhinny, M. W. & McFadden, P. L. In *The magnetic field of the Earth*, pp. 19–68 (Academic Press, New York, 1996).
 Andrews, D. G. In *Introduction to atmospheric physics*, pg. 9 (Cambridge, 2000).
- 24. Coe, H. Atmospheric energy and the structure of the atmosphere. Atmospheric science for environmental scientists (Wiley Blackwell, Editors: Hewitt, C. N. and Jackson, A. V.), pg. 61 (2009).
- 25. Christensen, H. & Veseth, L. On the high precision Zeeman effect in O₂ and SO. J. Mol. Spectrosc. 72, 438-444 (1978).
- Pollack, G. H., Figueroa, X. & Zhao, Q. Molecules, water and radiant energy: New clues for the origins of life. Int. J. Mol. Sci. 10, 1419–1429 (2009).
- Stribling, R. & Miller, S. L. Energy yields for hydrogen cyanide and formaldehyde synthesis: the HCN and amino acid concentrations in the primitive ocean. Orig. Life Evol. Biosph. 17, 261–273 (1987).
- Lahav, N. & Chang, S. The possible role of solid surface area in condensation reactions during chemical evolution: reevaluation. J. Mol. Evol. 8, 357–380 (1976).
- 29. Ferus, M. et al. Formation of nucleobases in a Miller-Urey reducing atmosphere. PNAS 114, 4306-4311 (2017).

- Breslow, R. & Levine, M. S. Amplification of enantiomeric concentrations under credible prebiotic conditions. PNAS 103, 12979–12980 (2006).
- 31. Halevy, I. & Bachan, A. The geologic history of seawater pH. Science 355, 1069-1071 (2017).
- Nishino, H. et al. Mechanism of pH-dependent photolysis of aliphatic amino acids and enantiomeric enrichment of racemic leucine by circularly polarized light. Org. Lett. 3, 921–924 (2001).
- Nishino, H. et al. Absolute asymmetric photoreactions of aliphatic amino acids by circularly polarized synchrotron radiation: critically pH-dependent photobehavior. J. Am. Chem. Soc. 124, 11618–11627 (2002).
- Amdursky, M. & Stevens, M. M. Circular dichroism of amino acids: following the structural formation of phenylalanine. Chem Phys Chem 16, 2768–2774 (2015).
- 35. Sprecher, C. A. & Johnson, W. C. Jr. Circular dichroism of nucleic acid monomers. Biopolymers 16, 2243-2264 (1977).
- 36. Jagger, J. Introduction to research in ultraviolet photobiology, pg. 87 (Prentice Hall, N. J., 1967).
- Matsunaga, T., Hieda, K. & Nikaido, O. Wavelength dependent formation of thymine dimers and (6-4) photoproducts in DNA by monochromatic ultraviolet light ranging from 150 to 365 nm. *Photochem Photobiol* 54, 403–410 (1991).
- Saidel, L. J., Goldfarb, A. R. & Waldman, S. The absorption spectra of amino acids in the region of two hundred to two hundred and thirty millimicrons. J. Biol. Chem. 197, 285–291 (1952).
- Talrose, V. et al. UV/Visible Spectra in NIST Chemistry WebBook, NIST Standard Reference Database Number 69, Eds Linstrom, P. J. and Mallard, W. G. National Institute of Standards and Technology, Gaithersburg MD, 20899, https://doi.org/10.18434/T4D303.
- Ploeser, J. M. T. & Loring, H. S. The ultraviolet absorption spectra of the pyrimidine Ribonucleotides and ribonucleosides. J. Biol. Chem. 178, 431–437 (1949).
- 41. Demtroeder, W. In Laser Spectroscopy (3rd Edition, Springer, Berlin), pg. 572 (2003).
- 42. Lakowicz, J. R. In Principles of fluorescence spectroscopy (Plenum Press, N. Y.), pg. 48 (1986).
- Johnson, P. V., Hodyss, R., Chernow, V. F., Lipscomb, D. M. & Goguen, J. D. Ultraviolet photolysis of amino acids on the surface of icy solar system bodies. *Icarus* 221, 800–805 (2002).
- Beck, S. E. et al. Comparison of UV-induced inactivation and RNA damage in MS2 phage across the germicidal Uvspectrum. Appl. Environmen. Microbiol. 82, 1468–1474 (2016).
- 45. Davies, R. J. H. Ultraviolet radiation damage inDNA. Biochemical Soc. Transact. 23, 407-418 (1995).
- Meierhenrich, U. J. et al. Circular dichroism of amino acids in the vacuum-ultraviolet region. Angew. Chem. Int. Ed. 49, 7799–7802 (2010).
- 47. Ehrenfreund, P., Bernstein, M. P., Dworkin, J. P., Sandford, S. A. & Allamandola, L. J. The photostability of amino acids in space. Astrophys. Jour. 550, L95–L99 (2001).
- 48. Ten Kate, I. L. et al. Amino acid photostability on the Martian surface. Meteoritics & Planetary Sc. 40, 1185-1193 (2005).
- 49. Hale, G. M. & Querry, M. R. Optical constants of water in the 200-nm to 200-µm wavelength region. Appl. Opt. 12, 555-563 (1973).
- 50. Bonoli, L. & Witherspoon, P. A. Diffusion of aromatic and cycloparaffin hydrocarbons. J. Phys. Chem. 72, 2532–2534 (1968).
- 51. Jost. W. Diffusion in solids, liquids, gases (Academic Press, NY), pg. 16 (1960).
- 52. Ogawa, S., Wada, S. & Urata, H. Base recognition by L-nucleotides in heterochiralDNA. RSC Advances 2, 2274-2275 (2012).
- Blommers, M. J. J., Tondelli, L. & Garbesi, A. Effects of the introduction of L-nucleotides into DNA. Solution structure of the heterochiral duplex d(G-C-G-(L)T-G-C-G).d(C-G-C-A-C-G-C) studied by NMR spectroscopy. *Biochemistry* 33, 7886–7896 (1994).
- 54. Merrill, R. T., McElhinny, M. W. & McFadden, P. L. In The magnetic field of the Earth, pp. 163–215 (Academic Press, New York, 1996).

Author Contributions

A.S. conceived this study and wrote the paper.

Additional Information

Competing Interests: The authors declare that they have no competing interests.

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