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OPEN Long-term (2001–2013) observations of water-soluble dicarboxylic acids and related compounds over the western North Pacific: trends, seasonality and source apportionment

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To better understand the impact of East Asian pollutants on the molecular composition of marine organic aerosols, we conducted long-term (2001–2013) observations of water-soluble dicarboxylic acids and related compounds in total suspended particulate samples collected at Chichijima Island in the western North Pacific (WNP). Seasonal variations of all the diacids and related compounds showed maxima in winter and spring and minima in summer, except for azelaic acid (C_o), which maximized in summer to autumn. The overall annual concentrations of the total diacids, ω -oxoacids and α -dicarbonyls showed an increase during 2001–2013. We found a significant (p < 0.05) decadal increase in the inter-annual trends of pyruvic and glyoxylic (p > 0.05) acids, and methylglyoxal (MeGly). In contrast, phthalic acid (p < 0.05) and glyoxal (Gly) showed a decrease in their trends. We also found a significant decrease in the trend of the Gly/MeGly mass ratios. These results demonstrate that the enhanced concentrations of diacids over the WNP are majorly attributed to the aqueousphase photooxidation of biogenic volatile organic compounds from East Asia followed by long-range atmospheric transport. Further, positive matrix factorization analysis showed a biogenic photochemical contribution (42%) was the dominant source of oxalic acid in the WNP.

Low molecular weight dicarboxylic acids (diacids) are ubiquitous and important constituents of atmospheric organic aerosols¹. Due to the low vapor pressures (approximately less than 10^{-7} Pa)^{2,3}, these compounds are almost exclusively partitioned into a particulate phase and contribute significantly to the water-soluble fraction^{4,5}. Thus, particles enriched with diacids can act as cloud condensation nuclei⁶⁻⁸ and have an impact on the earth's radiative forcing⁹. Diacids also play an important role in many biological processes in the ocean¹⁰. Despite the great importance, they continuously represent a challenge to atmospheric scientists due to their complexity, although the first unambiguous observations of diacids in the atmosphere were made three decades ago¹¹.

Diacids are the late photooxidation products of hydrocarbons and other organics¹². Therefore, photochemical chain reactions are the most important sources of diacids^{13,14}. Aqueous-phase oxidation and cloud processing of aerosols are potentially important sources of organic aerosols and could explain the high abundances of oxalic acid in the atmosphere^{15–25}. High levels of diacids were observed in motor exhausts and ambient aerosols influenced by forest fires^{26, 27}, indicating important primary sources of diacids. However, there is still a knowledge gap on the relative contribution of primary and secondary sources of diacids in the atmosphere. On the other hand, iron-carboxylate complexes and their photochemical degradation in the water mediated atmosphere^{28–32} may be an important sink of diacids in addition to wet and dry deposition.

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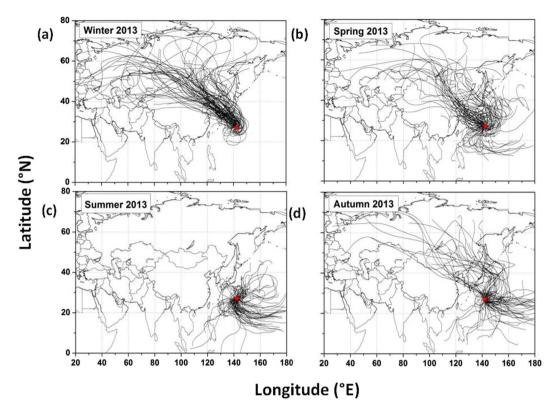


Figure 1. Daily 7-day HYSPLIT backward air mass trajectories for different seasons over the WNP for the year 2013. The star (*) indicates the sampling site, Chichijima Island. Trajectory data downloaded from the NOAA ARL website (http://www.arl.noaa.gov) and plotted using origin lab (v.8) software (http://www.originlab.com/).

A large fraction of secondary organic aerosol (SOA) is derived from the oxidation of isoprene, monoterpenes and sesquiterpenes of biogenic origin. Global aerosol modelling studies suggest an increase in isoprene emissions of 22 to 55% by 2100 in response to the temperature increase following a global warming^{33–35}. Recent modelling studies³⁶ simulate an increased trend of isoprene emissions with 0.16% per year over Asia and this trend is more pronounced over China with 0.52% per year during 1979 to 2012. All these models, however, have significant uncertainties in the projection of SOA formation due to a series of fundamental assumptions and lack of long-term observational studies. The models assume that particles are liquids and are exposed to changes in the surrounding atmosphere by rapid condensation or evaporation and by in-particle mixing during long-range atmospheric transport³⁷.

East Asia is one of the most populated regions on the globe with a population density of 133 inhabitants per square kilometer, which is about three times higher than the world population density (45 per km²). Increasing levels of air pollutants in East Asia and their outflows to the Pacific have received a significant attention due to the potential impacts on regional and global climate and atmospheric circulations^{38–43}. However, the effects of East Asian pollutants and their outflows over the western North Pacific (WNP) are still unclear, especially with regard to the sources and formation of SOA during long-range transport^{44–46}.

In this study, we carried out long-term (2001–2013) measurements of diacids and related compounds in total suspended particulate (TSP) aerosols collected at Chichijima Island in the WNP. Chichijima is a remote island 2000 km away from East Asia and exists in the outflow region of Asian dusts and pollutants from East Asia, especially China. Here, we discuss the inter-annual trends and seasonality of diacids over the WNP. We also discuss the sources and formation pathways of the diacids using a positive matrix factorization (PMF) model.

Results and Discussion

Air mass origin and meteorological parameters. Figure 1(a–d) presents daily 7-day isentropic air mass back trajectories at an altitude of 500 m above the ground level using the HYSPLIT model⁴⁷ for different seasons during the year 2013 over the WNP as an example. Air mass transport from East Asia to the sampling site in the Pacific is stronger during winter (December to February) and spring (March to May) than during summer and autumn to deliver continental air masses via long-range atmospheric transport. The continental air mass transport is almost absent in summer (June to August). Air masses mostly come from the central Pacific carrying pristine air masses to the observation site in summer, whereas in autumn (September to November) the air mass pathway shifts from southeasterly to northwesterly towards winter. More detailed information of air mass transport over the WNP is described elsewhere⁴⁸.

The meteorological parameters such as air temperature (°C), wind speed (ms⁻¹), solar radiation (MJ m⁻²), precipitation (mm) and cloud cover are downloaded from the Japan Meteorological Agency (JMA) for Chichijima

Island during 2001–2013 (Figure S1). We found a clear seasonal variation in air temperature, solar radiation and precipitation with summer maxima and winter minima. Wind speed and cloud cover were higher in spring and lower in summer. However, no significant decadal trends were observed in any meteorological parameters throughout the sampling period, therefore, we used meteorological parameters in this study to better understand the seasonal variations of organic compounds but not to discuss the trends.

Molecular distributions. In order to better understand the sources, formation pathways, and physicochemical properties of organic aerosols, we studied the molecular distributions of the diacids and related compounds at Chichijima Island between 2001 and 2013. Molecular distributions are shown in Figure S2. Throughout the observation period, we found the predominance of oxalic acid (C_2) followed by malonic (C_3) and/or succinic (C_4) acids. This molecular distribution is consistent with our previous study for 1990–1993 at the same observation site⁴⁴ and other East Asian sites such as Okinawa Island^{49, 50}, the Gosan site, Jeju Island in South Korea⁵¹, Mt. Tai in North China⁵², urban sites in China^{53, 54}, as well as different sites in the world including Tanzania, East Africa⁵⁵, remote western European continental sites⁵⁶, west-east transect in the European atmosphere⁵⁷, and Los Angeles²⁶.

Glyoxylic acid (ωC_2) was the dominant species among all ω -oxoacids and the fourth most abundant species detected, whereas methylglyoxal (MeGly) was more abundant than glyoxal (Gly) in the WNP aerosols. Throughout the observation period, we found the molecular distributions of the diacids and related compounds at Chichijima as $C_2 > C_3 > C_4 > \omega C_2 > \text{MeGly} > \text{Ph} > \omega C_7 > C_5 > \omega C_8 > \text{Gly} > C_6 > \omega C_9 > C_9$. However, seasonal molecular distributions provided a different picture depending on the source strengths and formation mechanisms.

Trends. Table 1 summarizes the regression statistics such as range, mean \pm SD, and the trend (slope) for water-soluble diacids, ω -oxocarboxylic acids, pyruvic acid, α -dicarbonyls, and their diagnostic ratios during the period of 2001 to 2013. Figure 2 presents the temporal trends of the major diacids and related compounds and diagnostic mass ratios during the whole study period.

 C_2 , C_3 , and C_4 acids are the end or near-end products of photochemical reaction chains of hydrocarbons and biogenic unsaturated fatty acids, accounting for approximately 74%, 11%, and 6% of the total diacids, respectively, during the 13-year study period at Chichijima. From Table 1, it is obvious that C_4 shows an increasing trend $(+0.003\% \ yr^{-1})$, whereas those of C_5 – C_{12} diacids (particularly, C_5 , C_7 , C_8 , C_9 , and C_{12}) show significant (p < 0.05) decreasing trends during the study period. These results suggest a photochemical conversion of higher to lower molecular weight diacids during the long-range atmospheric transport over the WNP. This point is further supported by significant increasing trends of diagnostic mass ratios of diacids, as discussed below.

It has been suggested that maleic acid (cis configuration) (M), a photo-oxidation product of aromatic hydrocarbons such as benzene and toluene⁵⁸, may be isomerized to fumaric acid (trans configuration) (F) under a high solar radiation in the atmosphere¹. Therefore, the F/M ratio may be a good indicator of photochemical processing. In our study, we found a significant (p < 0.05) increasing trend (+0.016% yr⁻¹) of F/M ratios during 13 years of the study period (Fig. 2g), indicating an intensive photochemical aging or increased oxidant levels over the WNP. Further, it has been suggested that C_2 and C_3 are likely produced in the marine atmosphere by the photooxidation of C_4 through intermediates such as hydroxylsuccinic or malic acid (hc_4) and ketomalonic acid (hc_5)^{59,60}. In this study, we found that hc_3/C_4 ratios showed an increasing trend, although it is not significant (hc_5) solveyer, hc_5/C_4 and hc_5/C_4 ratios show increases (hc_5/C_4) and hc_5/C_4 ratios that hc_5/C_4 ratios show increases (hc_5/C_4) and hc_5/C_4 ratios degradation of hc_5/C_4 ratios of the largely produced by the photochemical degradation of hc_5/C_4 and hc_5/C_4 ratios over the remote marine aerosols (Table 1). All these results suggest that the production of diacids over the WNP may be closely linked with an increased photochemical oxidation of biogenic and anthropogenic precursors that are delivered from the Asian continent by long-range atmospheric transport.

Previous studies suggested that oxidation of aromatic hydrocarbons such as naphthalene and o-xylene, which originate from an incomplete combustion of fossil fuel²⁶, is one of the major sources of phthalic acid (Ph) in the atmosphere^{61, 62}. High abundances of aromatic hydrocarbons are reported over China during winter^{63, 64}. Similarly, adipic acid (C_6) is probably produced through the oxidation of cyclohexene by ozone in the atmosphere and has been proposed as an anthropogenic tracer⁴⁶. It is also well documented that glyoxal (Gly) is largely produced in the atmosphere by the oxidation of many anthropogenic aromatic hydrocarbons^{65–67}, although it has small contribution from biogenic and marine origin^{20,68,69}. In this context, we found a decreasing trend in the concentrations of Ph (p < 0.05; -0.007% yr⁻¹), C_6 (p > 0.05; -0.003% yr⁻¹) and Gly (p > 0.05; -0.006% yr⁻¹) (Fig. 2b,e, and Table 1). These results suggest that combustion (fossil fuel) derived aerosols have declined (or constant) over the WNP during 2001–2013. This point is further supported by the study of Boreddy, *et al.*⁷⁰, who reported that declined concentrations of elemental carbon (EC) over WNP occurred during 2001–2012 over the WNP. However, it should be noted that although Gly concentrations are decreased, its processing to C_2 is not decreased over the WNP, as evidenced by the increasing trend (+0.002% yr⁻¹; p > 0.05) of C_2 /Gly ratios (Table 1).

On the other hand, a modeling study by Stavrakou, *et al.*³⁶ observed a continuous increase of isoprene emissions over Asia during 1979–2012. They found a strong correlation (r > 0.90) between isoprene emissions and above-canopy solar radiation, suggesting that enhanced solar radiation intensifies isoprene emissions from terrestrial higher plants over Asia (particularly in China). Similarly, Zhang, *et al.*⁷² have recently reported an increase of biogenic isoprene emissions in northern China during 1982–2010 using the biogenic emission model. It has also been documented that 79% of MeGly may come from biogenic isoprene emissions globally, as inferred from modeling studies⁷³. Further, pyruvic (Pyr) and ωC_2 have been suggested as in-cloud oxidation products of isoprene, which are subsequently oxidized to C_2^{18} . Therefore, it may be possible that biogenic isoprene derived volatile organic precursors (e.g., MeGly) over East Asia/China are taken up by aqueous-phase aerosol particles in the atmosphere and transported to the WNP.

Dearboy(s) acids (og m²)	Organic compounds	Range (mean ± SD)	Slope (m, diacid year ⁻¹)	Uncertainty (σm)	Trend (% year ⁻¹
Oxalic C2	Dicarboxylic acids (ng m ⁻³)				
Malonic, C ₂	Normal chain saturated diacids				
Saccinic, C ₁ 0.05-52.4 (6.12 ± 6.41) + 0.0002* 0.0002 + 0.003 Glustric, C ₂ 0.748 (1.11 ± 1.19) -0.00003 0.00003 -0.0002 Adipic, C ₂ 0.01-588 (9.95 ± 0.00) -0.00002 0.0000017 -0.003 Pimelic, C ₂ 0.1-124 (0.13 ± 0.14) -0.000014* 0.0000017 -0.009 Suberic, C ₃ 0.1-124 (0.13 ± 0.14) -0.00003* 0.00001 -0.005 Decanedioic, C ₁₁ 0.8-38 (0.05 ± 0.34) -0.00002 0.000009 -0.005 Dedecanedioic, C ₁₂ 0.9-34 (0.01 ± 0.04) -0.00002 0.000001 -0.005 Branched chair sutrated discids 0.000004 0.000007 -0.000 Methylascinic, C ₅ 0.313 (0.52 ± 0.48) -0.00001 0.00001 +0.000 Methylascinic, C ₆ 0.313 (0.52 ± 0.48) +0.00001 0.00001 +0.0009 Methylascinic, C ₆ 0.313 (0.52 ± 0.48) +0.00001 0.00001 +0.0009 Methylascinic, C ₆ 0.313 (0.52 ± 0.48) +0.00001 0.00001 +0.0009 Methylascinic, C ₆	Oxalic, C ₂	2.21-514 (73.9 ± 66.8)	+0.0032	0.0020	+0.004
Glataric, C ₂	Malonic, C ₃	0.28-55.6 (11.5 ± 9.4)	+0.00005	0.0002	+0.0004
Adipic, C _a	Succinic, C ₄	$0.05 – 52.4 (6.12 \pm 6.41)$	+0.0002*	0.0002	+0.003
Pimelic, C _c	Glutaric, C ₅	$0 - 7.48 (1.11 \pm 1.19)$	-0.00003*	0.00003	-0.0027
Suberic, C₀ a 0-1.24 (0.13 ± 0.14) -0.0003* 0.000038 -0.023 Azelaic, C₀ a 0.01-250 (0.53 ± 0.34) -0.00003* 0.00001 -0.005 Undecanedioic, C₀ a 0-0-95 (0.06 ± 0.08) -0.00002 0.0000098 -0.002 Dodecanedioic, C₀ a 0-0-80 (0.08 ± 0.34) -0.000002 0.0000098 -0.002 Branched chian strutted diactis Branched chian strutted diactis -0.00009 0.000074 -0.0029 Methylsaccinic, iC₀ a 0-3.13 (0.52 ± 0.48) +0.000012 0.000014 +0.0019 Methylsylaccinic, iC₀ a 0-3.03 (0.52 ± 0.48) +0.000012 0.000014 +0.0019 Multi functional saturated diactis -0.0028 +0.00002 0.000012 -0.0075 Multi functional saturated diactis -0.12 (0.06±1.12) -0.000111* 0.000022 -0.0005 Hydroxyaccinic, ibC₀ a 0-5.40 (0.38±0.49) +0.00002 0.000012 -0.0014 Maleic, M 0-2 (0.43±0.40) -0.00007* 0.00017 -0.011 Maleic, M 0-2 (0.43±0.40) -0.00002* 0.00001 -0.015<	Adipic, C ₆	$0.01 – 5.08 \ (0.59 \pm 0.60)$	-0.00002	0.0000017	-0.0033
Azelaic, C ₀	Pimelic, C ₇	$0-1.34~(0.15\pm0.16)$	-0.000014*	0.0000047	-0.009
Decanedioic, C ₁₀	Suberic, C ₈	$0-1.24~(0.13\pm0.14)$	-0.00003*	0.0000038	-0.023
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Azelaic, C ₉	0.01 – 2.50 (0.53 ± 0.34)	-0.00003*	0.00001	-0.005
Dodecanedioic, C ₁₂ 0-0.34 (0.01±0.04) -0.000004* 0.000001 -0.04 Branched chain saturated diacids Branched Chain saturated diacids Methylsalonic, iC ₄ 0-1.45 (0.31±0.25) -0.000009* 0.0000074 -0.0019 Methylsalonic, iC ₆ 0-3.13 (0.52±0.48) +0.000012 0.000014 +0.0019 Methylsalutaric, iC ₆ 0-5.13 (0.52±0.48) +0.000012 0.000014 +0.0019 Methylsalutaric, iC ₆ 0-5.12 (0.26±1.12) -0.000111* 0.000032 -0.038 Methylsalutaric, iC ₆ 0-15.2 (0.26±1.12) -0.000111* 0.000032 -0.038 Methylsalutaric, iC ₆ 0-5.40 (0.38±0.54) +0.00002 0.000015 +0.0052 Methylsalutaric, iC ₆ 0-5.40 (0.38±0.54) +0.00002 0.000017 -0.014 Drissalurated aliphatic diacids Wethylsalutaric, iC ₇ 0-2.27 (0.41±0.32) -0.00002* 0.000019 -0.001 Pumaric, F 0-2.27 (0.41±0.32) -0.00002* 0.000012 -0.031 Aromatic diacids Wethylsalic, iRh 0.766 (0.22±0.44) -0.00007* 0.000012 -0.031 Aromatic diacids Philaic, Ph 0.01-12.8 (1.31±1.49) -0.00009* 0.000043 -0.0068 Isophthalic, iPh 0.11.1 (0.14±0.49) +0.00001 0.000014 +0.00071 Drissal diacids 2.93-555 (99.2±86.4) +0.003 0.000015 +0.003 Drissal diacids 2.93-555 (99.2±86.4) +0.003 0.0025 +0.003 Drissal diacids 0.09-29 (4.91±5.13) +0.00007 0.000015 +0.001 Drissal diacids	Decanedioic, C ₁₀	0-0.95 (0.06 ± 0.08)	-0.000003	0.0000023	-0.005
Branched chain saturated diacids Methylmalonic, IC_c	Undecanedioic, C ₁₁	0-8.03 (0.08 ± 0.34)	-0.000002	0.0000098	-0.002
	Dodecanedioic, C ₁₂	0-0.34 (0.01 ± 0.04)	-0.000004*	0.000001	-0.04
Methylsuccinic, iC ₅ 0-3.13 (0.52 ± 0.48) +0.000012 0.000014 +0.0019 Methylglutaric, iC ₆ 0-0.96 (0.08 ± 0.09) -0.00006* 0.00002 -0.0075 Multi functional saturated diacids "Hydroxysuccinic, hC ₆ 0-15.2 (0.26 ± 1.12) -0.000111* 0.000032 -0.038 Ketomalonic, kC ₅ 0-5.40 (0.38 ± 0.54) +0.00002 0.000015 +0.0052 Ketopinelic, kC ₆ 0-3.67 (0.50 ± 0.57) -0.00007 0.000017 -0.0014 Unsaturated aliphatic diacids "William According to the property of the p	Branched chain saturated diacids	I		I	
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Multi functional saturated diacids	· · · · · · · · · · · · · · · · · · ·				
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Hydroxysuccinic, hC ₄ 0-15.2 (0.26 ± 1.12) -0.00011* 0.000032 -0.038 Ketomalonic, kC ₂ 0-540 (0.38 ± 0.54) +0.00002 0.000015 +0.0052 Ketopimelic, kC ₇ 0-367 (0.50 ± 0.57) -0.00007 0.000017 -0.0014 Unsaturated allphatic diacids *** Unsaturated allphatic diacids Maleic, M 0-2.07 (0.41 ± 0.32) -0.00002* 0.000012 -0.011 Fumaric, F 0-2.27 (0.41 ± 0.32) -0.00002* 0.000012 -0.031 Arcmatic diacids **** University of the property of		(1	
Ketomalonic, kC ₃ $0.540 (0.38\pm0.54)$ $+0.00002$ 0.00015 $+0.0052$ Ketopimelic, kC ₇ $0-3.67 (0.50\pm0.57)$ -0.00007 0.00017 -0.0014 Unsaturated aliphatic diacids Maleic, M $0-22 (0.41\pm0.32)$ -0.00002^* 0.00001 -0.015 Methylmaleic, mM $0-7.66 (0.22\pm0.44)$ -0.00007^* 0.000012 -0.031 Aromatic diacids 0.00001 0.000012 -0.001 Phthalic, Ph $0.01-12.8 (1.31\pm1.49)$ -0.00009^* 0.000043 -0.0081 Sophthalic, iPh $0-11.1 (0.14\pm0.49)$ $+0.00001$ 0.000043 $+0.0007$ Terephthalic, iPh $0-7.18 (0.39\pm0.52)$ $+0.00002$ 0.000015 $+0.0001$ Terephthalic, iPh $0-7.18 (0.39\pm0.52)$ $+0.0002$ 0.00015 $+0.0051$ Total diacids $2.93-555 (9.2\pm86.4)$ $+0.0002$ 0.00015 $+0.0031$ Glyox planci, ωC_2 $0.09-29 (4.91\pm5.13)$ $+0.00007$ 0.00015 $+0.003$ Glyox planci, ωC_2 $0.09-22 (4.91\pm5.13)$		0-15.2 (0.26 + 1.12)	-0.000111*	0.000032	-0.038
Retopimelic, RC7	· · · · · · · · · · · · · · · · · · ·				
Unsaturated aliphatic diacids Alaleic, M 0-2 (0.43 ± 0.40) −0.00005* 0.00001 −0.01 Fumaric, F 0-2.27 (0.41 ± 0.32) −0.00002* 0.000092 −0.005 Methylmaleic, mM 0-7.66 (0.22 ± 0.44) −0.00007* 0.000012 −0.03 Aromatic diacids −0.0008 −0.0009* 0.000043 −0.0068 Isophthalic, iPh 0-11.1 (0.14 ± 0.49) +0.00001 0.000014 +0.0071 Terephthalic, tPh 0-7.18 (0.39 ± 0.52) +0.00002 0.000015 +0.001 Total diacids 2.93 − 555 (9.92 ± 86.4) +0.003 0.0025 +0.001 Total diacids (ing m⁻³) −0.008 0.0025 +0.001 Glyoxylic, ωC₂ 0.09 − 29 (4.91 ± 5.13) +0.0007 0.0015 +0.001 3 - Oxopropanoic, ωC₂ 0.09 − 29 (4.91 ± 5.13) +0.00007 0.00015 +0.001 3 - Oxopropanoic, ωC₂ 0.09 − 29 (4.91 ± 5.13) +0.00007 0.00015 +0.001 3 - Oxoportanoic, ωC₂ 0.09 − 22.4 (0.26 ± 0.94) +0.00006* 0.000027 +0.01 5 - Oxopora					
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Aromatic diacids Phthalic, Ph					
Phthalic, Ph $0.01-12.8 (1.31\pm 1.49)$ -0.00009^* 0.000043 -0.0068 Isophthalic, iPh $0-11.1 (0.14\pm 0.49)$ $+0.00001$ 0.000014 $+0.0071$ Terephthalic, tPh $0-7.18 (0.39\pm 0.52)$ $+0.00002$ 0.000015 $+0.003$ Total diacids $2.93-555 (99.2\pm 86.4)$ $+0.003$ 0.0025 $+0.003$ ω-Oxocarboxylic acids (ng m ⁻³) 0.00027 0.00015 $+0.003$ Glyoxylic, ωC ₂ $0.99-29 (4.91\pm 5.13)$ $+0.00007$ 0.00015 $+0.001$ 3 -Oxopropanoic, ωC ₃ $0-22.4 (0.26\pm 0.94)$ $+0.00006*$ 0.000027 $+0.023$ 4 -Oxobutanoic, ωC ₃ $0-22.4 (0.26\pm 0.94)$ $+0.00006*$ 0.000027 $+0.023$ 4 -Oxopatranoic, ωC ₃ $0-6.56 (0.99\pm 0.89)$ $+0.00004*$ 0.000021 $+0.013$ 7 -Oxoheptanoic, ωC ₃ $0-0.65 (0.99\pm 0.89)$ $+0.00001*$ 0.000024 $+0.001$ 7 -Oxobeptanoic, ωC ₇ $0.2-11.6 (1.13\pm 1.27)$ $+0.00002$ 0.000026 $+0.0027$ 8 -Oxocatanoic, ωC ₃ $0-5.72 (0.59\pm 0.68)$ $+0.00002$ <td></td> <td>0-7.66 (0.22 ± 0.44)</td> <td>-0.0000/*</td> <td>0.000012</td> <td>-0.031</td>		0-7.66 (0.22 ± 0.44)	-0.0000/*	0.000012	-0.031
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Terphthalic, tPh 0-7.18 (0.39 ± 0.52) +0.00002 0.000015 +0.0051 Total diacids 2.93-555 (99.2 ± 86.4) +0.003 0.0025 +0.003 ω-Oxocarboxylic acids (ng m $^{-3}$) Glyoxylic, $ωC_2$ 0.09-29 (4.91 ± 5.13) +0.00007 0.00015 +0.001 3-Oxopropanoic, $ωC_3$ 0-22.4 (0.26 ± 0.94) +0.00006* 0.000027 +0.023 4-Oxobutanoic, $ωC_4$ 0-5.56 (0.39 ± 0.47) +0.00004* 0.000013 +0.010 5-Oxopentanoic, $ωC_5$ 0-0.655 (0.09 ± 0.08) +0.00001* 0.0000024 +0.01 5-Oxopentanoic, $ωC_7$ 0.02-11.6 (1.13 ± 1.27) +0.000006 0.000037 +0.00027 9-Oxononanoic, $ωC_6$ 0-6.32 (0.72 ± 0.89) +0.00002 0.000026 +0.00027 9-Oxononanoic, $ωC_6$ 0-5.72 (0.59 ± 0.68) +0.00006* 0.000037 +0.0005 9-Oxononanoic, $ωC_9$ 0-5.72 (0.59 ± 0.68) +0.00006* 0.00002 +0.010 Total $ω$ -oxoacids 0.21-52.7 (8.10 ± 8.20) +0.0003 0.00024 +0.0037 Ketoacid (ng m $^{-3}$) Pyruvic, Pyr 0-6.16 (0.79 ± 0.90) +0.0002* 0.000025 +0.025 α-Dicarbonyls (ng m $^{-3}$) Glyoxal, Gly 0-4.69 (0.80 ± 0.78) -0.00005 0.000036 -0.006 Methylglyoxal, MeGly 0-23.9 (1.73 ± 2.28) +0.00014* 0.000067 +0.008 Total α-dicarbonyls 0.04-26.4 (2.34 ± 2.74) +0.0003* 0.00006 +0.0005 Total α-dicarbonyls 0.00006 0.00003 +0.012 Ratios F/M 0-20.8 (1.83 ± 2.47) +0.0003* 0.00007 +0.006 C ₂ /C ₃ 1.46-33.1 (6.55 ± 2.86) +0.00034* 0.000083 +0.002 C ₂ /C ₃ 0.00006 0.00057 +0.0005 C ₃ /C ₄ 0.50-52.7 (2.76 ± 1.93) +0.00006 0.000057 +0.0005 C ₃ /C ₄ 0.51-52.7 (2.76 ± 1.93) +0.00002 0.000057 +0.0005 C ₃ /C ₄ 0.01-77.0 (3.47 ± 5.10) -0.00006 0.00015 -0.00006 C ₃ /C ₄ 0.01-77.0 (3.47 ± 5.10) -0.000006 0.00017 +0.0010 C ₃ /C ₄ C ₂ 5.48-131 (22.2 ± 14.7) +0.0013* 0.0004 0.0004 +0.0058 C ₃ /Gly 0.004 +0.0058 0.0004 +0.0058 0.0004 +0.0058 0.0004 +0.0058 0.0004 +0.0058 0.00069 0.00017 +0.0010 0.00060 0.00017 +0.0010 0.00060 0.00017 +0.0010 0.00060 0.00017 +0.0010 0.00060 0.00018 +0.0010 0.00060 0.00019 0.00060 0.00019 0.00060 0.00019 0.00060 0.00019 0.00060 0.00019 0.00060 0.00019 0.00060 0.00019 0.00060 0.00060					
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Glyoxylic, $ωC_2$ $0.09-29 (4.91\pm5.13)$ $+0.00007$ 0.00015 $+0.001$ 3 -Oxopropanoic, $ωC_3$ $0-22.4 (0.26\pm0.94)$ $+0.00006*$ 0.000027 $+0.023$ 4 -Oxobutanoic, $ωC_4$ $0-5.56 (0.39\pm0.47)$ $+0.00004*$ 0.000013 $+0.010$ 5 -Oxopentanoic, $ωC_5$ $0-0.65 (0.09\pm0.08)$ $+0.00001*$ 0.000024 $+0.01$ 7 -Oxoheptanoic, $ωC_7$ $0.02-11.6 (1.13\pm1.27)$ $+0.000006$ 0.000024 $+0.0005$ 8 -Oxocotanoic, $ωC_8$ $0-6.32 (0.72\pm0.89)$ $+0.00002$ 0.000026 $+0.0027$ 9 -Oxononanoic, $ωC_9$ $0-5.72 (0.59\pm0.68)$ $+0.00006*$ 0.00002 $+0.010$ $Total ω-oxoacids$ $0.21-52.7 (8.10\pm8.20)$ $+0.0003$ 0.00024 $+0.0037$ N -Catacomore, $ωC_9$ $0-5.72 (0.59\pm0.68)$ $+0.0003*$ 0.00024 $+0.0037$ N -Catacomore, $ωC_9$ $0-5.72 (0.59\pm0.68)$ $+0.0003*$ 0.00024 $+0.0037$ N -Catacomore, $ωC_9$ $0-5.72 (0.59\pm0.68)$ $+0.0002*$ 0.000025 $+0.025$ $ω$ -Catacomore, $ω$ -Catacomore, $ω$ -Cataco		2.93-555 (99.2 ± 86.4)	+0.003	0.0025	+0.003
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5-Oxopentanoic, $ω_5$ $0-0.65 (0.09\pm0.08)$ $+0.00001^*$ 0.0000024 $+0.01$ 7-Oxoheptanoic, $ω_7$ $0.02-11.6 (1.13\pm1.27)$ $+0.000006$ 0.000037 $+0.0005$ 8-Oxooctanoic, $ω_7$ $0-6.32 (0.72\pm0.89)$ $+0.00002$ 0.000026 $+0.0027$ 9-Oxononanoic, $ω_7$ $0-5.72 (0.59\pm0.68)$ $+0.00006^*$ 0.00002 $+0.010$ Total $ω$ -oxoacids $0.21-52.7 (8.10\pm8.20)$ $+0.0003$ 0.00024 $+0.0037$ Ketoacid (ng m ⁻³) -0.0003 -0.00024 $+0.0037$ Pyruvic, Pyr $0-6.16 (0.79\pm0.90)$ $+0.0002^*$ 0.000025 $+0.025$ $α$ -Dicarbonyls (ng m ⁻³) -0.00005 0.000036 -0.006 Methylglyoxal, MeGly $0-23.9 (1.73\pm2.28)$ $+0.00014^*$ 0.000067 $+0.008$ Total $α$ -dicarbonyls $0.04-26.4 (2.34\pm2.74)$ $+0.0003^*$ 0.00006 $+0.008$ Ratios F/M $0-20.8 (1.83\pm2.47)$ $+0.0003^*$ 0.000072 $+0.016$ C_2/C_3 $1.46-33.1 (6.55\pm2.86)$ $+0.0003^*$ 0.000072			+0.00006*	0.000027	+0.023
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	<u> </u>	$0-5.56 \ (0.39 \pm 0.47)$	+0.00004*	0.000013	+0.010
8-Oxooctanoic, $ωC_8$	5-Oxopentanoic, ωC_5	$0-0.65 \ (0.09 \pm 0.08)$	+0.00001*	0.0000024	+0.01
9-Oxononanoic, $ωC_9$	7-Oxoheptanoic, ωC_7	$0.0211.6 \ (1.13 \pm 1.27)$	+0.000006	0.000037	+0.0005
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	8-Oxooctanoic, ωC_8	$0-6.32~(0.72\pm0.89)$	+0.00002	0.000026	+0.0027
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	9-Oxononanoic, ωC_9	$0-5.72~(0.59\pm0.68)$	+0.00006*	0.00002	+0.010
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Total ω-oxoacids	$0.21-52.7 \ (8.10 \pm 8.20)$	+0.0003	0.00024	+0.0037
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Glyoxal, Gly 0-4.69 (0.80 ± 0.78) -0.00005 0.000036 -0.006 Methylglyoxal, MeGly 0-23.9 (1.73 ± 2.28) +0.00014* 0.000067 +0.008 Total α-dicarbonyls 0.04-26.4 (2.34 ± 2.74) +0.0003* 0.00008 +0.012 Ratios F/M 0-20.8 (1.83 ± 2.47) +0.0003* 0.000072 +0.016 C_2/C_3 1.46-33.1 (6.55 ± 2.86) +0.00034* 0.000083 +0.0045 C_2/C_4 0.80-154 (17.0 ± 12.3) +0.001* 0.00036 +0.005 C_3/C_4 0.15-27.7 (2.76 ± 1.93) +0.00002 0.00057 +0.0007 Ph/C_9 0.01-77.0 (3.47 ± 5.10) -0.000006 0.00015 -0.0002 $C_2/\Sigma(C_2$ - C_{12}) 0.38-0.94 (0.77 ± 0.06) +0.00008* 0.0000017 +0.0010 $C_2/\omega C_2$ 5.48-131 (22.2 ± 14.7) +0.0013* 0.0004 +0.0058 C_2/Gly 20.3-4050 (135 ± 332) +0.0022 0.01 +0.002	Pyruvic, Pyr	0-6.16 (0.79 ± 0.90)	+0.0002*	0.000025	+0.025
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	α-Dicarbonyls (ng m ⁻³)				
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Glyoxal, Gly	$0-4.69~(0.80\pm0.78)$	-0.00005	0.000036	-0.006
Ratios $F/M \qquad 0-20.8 \ (1.83\pm 2.47) \qquad +0.0003^* \qquad 0.000072 \qquad +0.016 \\ C_2/C_3 \qquad 1.46-33.1 \ (6.55\pm 2.86) \qquad +0.00034^* \qquad 0.000083 \qquad +0.0045 \\ C_2/C_4 \qquad 0.80-154 \ (17.0\pm 12.3) \qquad +0.001^* \qquad 0.00036 \qquad +0.005 \\ C_3/C_4 \qquad 0.15-27.7 \ (2.76\pm 1.93) \qquad +0.00002 \qquad 0.000057 \qquad +0.0007 \\ Ph/C_9 \qquad 0.01-77.0 \ (3.47\pm 5.10) \qquad -0.000006 \qquad 0.00015 \qquad -0.0002 \\ C_2/\sum (C_2-C_{12}) \qquad 0.38-0.94 \ (0.77\pm 0.06) \qquad +0.000008^* \qquad 0.0000017 \qquad +0.0010 \\ C_2/\omega C_2 \qquad 5.48-131 \ (22.2\pm 14.7) \qquad +0.0013^* \qquad 0.0004 \qquad +0.0058 \\ C_2/Gly \qquad 20.3-4050 \ (135\pm 332) \qquad +0.0022 \qquad 0.01 \qquad +0.002$	Methylglyoxal, MeGly	0-23.9 (1.73 ± 2.28)	+0.00014*	0.000067	+0.008
Ratios $F/M \qquad 0-20.8 \ (1.83\pm 2.47) \qquad +0.0003^* \qquad 0.000072 \qquad +0.016 \\ C_2/C_3 \qquad 1.46-33.1 \ (6.55\pm 2.86) \qquad +0.00034^* \qquad 0.000083 \qquad +0.0045 \\ C_2/C_4 \qquad 0.80-154 \ (17.0\pm 12.3) \qquad +0.001^* \qquad 0.00036 \qquad +0.005 \\ C_3/C_4 \qquad 0.15-27.7 \ (2.76\pm 1.93) \qquad +0.00002 \qquad 0.000057 \qquad +0.0007 \\ Ph/C_9 \qquad 0.01-77.0 \ (3.47\pm 5.10) \qquad -0.000006 \qquad 0.00015 \qquad -0.0002 \\ C_2/\sum (C_2-C_{12}) \qquad 0.38-0.94 \ (0.77\pm 0.06) \qquad +0.000008^* \qquad 0.0000017 \qquad +0.0010 \\ C_2/\omega C_2 \qquad 5.48-131 \ (22.2\pm 14.7) \qquad +0.0013^* \qquad 0.0004 \qquad +0.0058 \\ C_2/Gly \qquad 20.3-4050 \ (135\pm 332) \qquad +0.0022 \qquad 0.01 \qquad +0.002$	Total α -dicarbonyls	0.04-26.4 (2.34 ± 2.74)	+0.0003*	0.00008	+0.012
$\begin{array}{cccccccccccccccccccccccccccccccccccc$				1	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		0-20.8 (1.83 ± 2.47)	+0.0003*	0.000072	+0.016
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		1.46-33.1 (6.55 ± 2.86)			
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$C_2/\omega C_2$ $5.48-131 (22.2\pm14.7)$ $+0.0013^*$ 0.0004 $+0.0058$ C_2/Gly $20.3-4050 (135\pm332)$ $+0.0022$ 0.01 $+0.002$				_	_
C_2/Gly 20.3-4050 (135 ± 332) +0.0022 0.01 +0.002					
$C_2/DREGIY$ $0-758 (90.8 \pm 80.5)$ $+0.006/$ 0.0025 $+0.007$					
Gly/MeGly $0-6.71 (0.98\pm0.81)$ $-0.00008*$ 0.000037 -0.008					

Table 1. Regression statistics (range, mean \pm SD, and slope) of water-soluble dicarboxylic acids and related compounds (n = 607) in remote marine TSP aerosols collected at Chichijima Island during the period of 2001 to 2013. The symbol, *, indicates the trends are significant at a 95% (p < 0.05) confidence level.

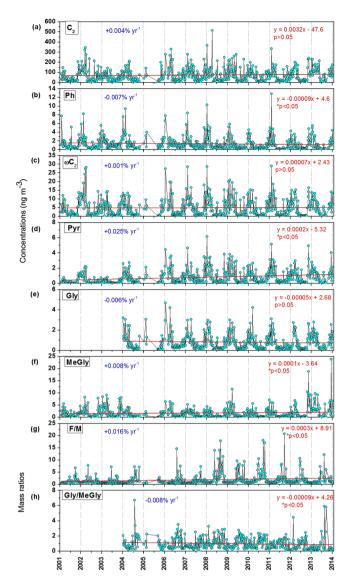


Figure 2. Trends in (a–f) temporal variations of water-soluble dicarboxylic acids (ng m $^{-3}$) and related compounds (ng m $^{-3}$) and (g,h) F/M and Gly/MeGly mass ratios in TSP aerosols collected at Chichijima Island during 2001 to 2013. Linear regression trends are given inset and applied over the whole observation period. The symbol, *, indicates the trends are significant at a 95% (p < 0.05) confidence level. The solid red line represents the trend line.

In this study, we found a significant increasing trend (p < 0.05) in the concentrations of MeGly (+0.008% yr^-l), Pyr (+0.025% yr^-l) (Fig. 2d and f) and C_2 /MeGly ratios (+0.007% yr^-l), suggesting the formation of C_2 from the oxidation of MeGly and Pyr via an intermediate compound, i.e., ωC_2 (MeGly \rightarrow Pyr \rightarrow acetic acid $\rightarrow \omega C_2 \rightarrow C_2$) in the aqueous-phase ¹⁸. These results suggest that enhanced concentrations of the diacids are probably caused by an increase of biogenic isoprene-derived precursors (i.e., MeGly and Pyr)⁷⁴, followed by the subsequent photochemical oxidation during long-range atmospheric transport over the WNP. A concurrent negative trend in Gly/MeGly (-0.008% yr^-l; p < 0.05) (Fig. 2h) also suggests an increase of biogenic precursors over the WNP. This point is further supported by our previous study of Boreddy and Kawamura⁴⁸, which reported that a significant increase (p < 0.05) in the concentrations of methanesulfonic acid (MSA⁻; a tracer for biogenic sources) occurred during 2001–2012 over the WNP.

The observed trends of diacids may be not only reflected by changes in emissions/air masses, but also influenced by changes in oxidant levels. To better understand the variations in the trends of oxidant levels over the WNP, we downloaded the monthly mean levels of total columnar ozone (DU) and daily tropospheric columnar NO $_2$ (cm $^{-2}$) for the periods from 2002 to 2013 and from 2005 to 2013, respectively, from the NASA website (https://giovanni.gsfc.nasa.gov/) (Figure S3). From Figure S3, it is clear that both oxidant levels (O $_3$ and NO $_2$) showed significant (p < 0.05) increase in those trends (+0.0004% yr $^{-1}$ and +0.008% yr $^{-1}$, respectively) during the study period, indicating that increased oxidation processes over the WNP lead to increases in the formation of diacids during long-range atmospheric transport.

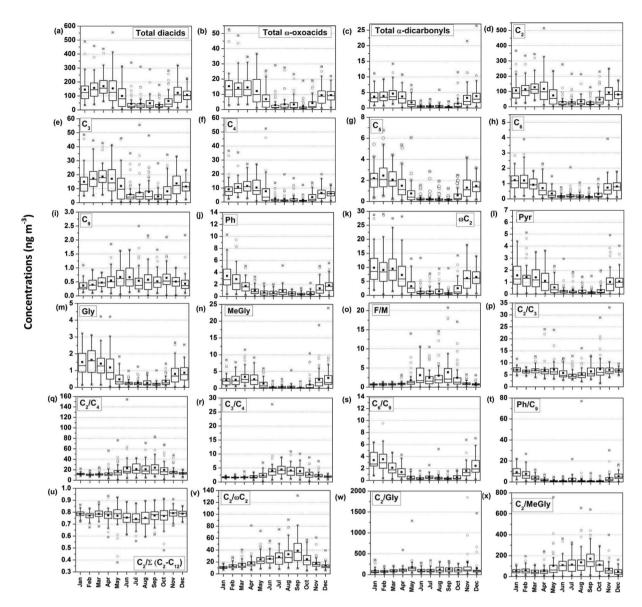


Figure 3. Monthly variations in water-soluble dicarboxylic acids (ng m $^{-3}$) and related compounds (ng m $^{-3}$) and various mass ratios at Chichijima during 2001 to 2013. The horizontal line and dot inside the box indicate maiden and mean, respectively. The vertical hinges represent data points from the lower to the upper quartile (i.e., 25th and 75th percentiles). The whiskers represent data points from the 5th to 95th percentiles. Open circles indicate the outliers.

Based on the above results, we conclude that the enhanced concentrations of diacids over the WNP may be caused due to the increased oxidations of biogenic precursor compounds during long-range atmospheric transport. In contrast, anthropogenic precursors (e.g., Gly) have decreased (or constant) during the study period. It should be noted that all these trends are explained for the whole period (2001–2013); however, seasonal trends may give different results over the WNP because the formations of diacids are very sensitive to the sources of an air mass and the meteorological parameters as reported in Tables S4–S7.

As seen from Tables S4–S7, it is noteworthy that concentrations of total diacids in all seasons showed increases in their trends, except for summer, which showed a decreasing trend (-0.002% yr⁻¹; p > 0.05) (Table S6). The declined concentrations of diacids and related compounds in summer are probably due to the pristine air masses, which suggest the negligible local anthropogenic emissions as well as long-range continental outflow over the WNP. It is also seen from Tables S4–S7 that the trend in concentrations of C_9 showed a decrease in all seasons, except for summer. In summer, the trend of C_9 showed an increase (+0.001% yr⁻¹) during 2001–2013, indicating an importance of oxidation of biogenic unsaturated fatty acids over the WNP, particularly in summer.

Monthly and seasonal variations. Figure 3 presents box and whisker plots of monthly variations of diacids, ω -oxocarboxylic acids, pyruvic acid, α -dicarbonyls, and diagnostic mass ratios at Chichijima Island for the period of 2001 to 2013. Almost all the organic compounds showed clear monthly/seasonal variations with higher

concentrations in winter/spring under the continental outflow from East Asia and lower concentrations in summer/autumn due to the pristine marine air mass, except for C_9 (Fig. 3i). As shown in Fig. 3a, concentrations of total diacids are characterized by a gradual increase from autumn to winter, with a peak in early spring (March), and a decrease with a minimum in summer although a small peak was observed in August. Total ω -oxoacids showed a similar seasonal variation (Fig. 3b). The peak in spring is more significant than in summer. On the other hand, total concentrations of α -dicarbonyls gradually increased from late autumn to early spring and then decreased towards the summer months (Fig. 3c). These seasonal variations can be explained primarily by an enhanced Asian outflow in winter/spring with the heterogeneity of air masses and changes in emission strength and meteorology as discussed below.

Impact of heterogeneity in air masses. As discussed above, there is a clear seasonal difference in the origin of air masses during the study period at Chichijima Island in the WNP. Such changes in air masses may lead to different effects on the formation of diacids and related compounds as well as the characteristics of chemical properties of aerosols. During winter/spring, the air mass transport is stronger; mostly coming from East Asia with westerly winds. The sampling site is influenced by long-range atmospheric transport of continental air masses, whereas the air masses are originating from the central Pacific via easterly winds during summer/autumn. This heterogeneity in air mass origins may clearly reflect the emission strength of organic aerosols and their precursors with higher concentrations during winter/spring and lower concentrations in summer/autumn over the WNP.

Changes in emission strength and formations. Gradual increases of total diacids, ω -oxoacids and α -dicarbonyls during late autumn to early spring are attributable to the combined effect of anthropogenic/biogenic volatile organic compounds (VOCs) emitted from East Asia followed by subsequent oxidation during long-range atmospheric transport. In our previous study, higher concentrations of non-sea salt sulfate (nss-SO₄²⁻), nitrate (NO₃⁻), non-sea salt calcium (nss-Ca²⁺) and MSA⁻ were found in winter/spring over the same site under continental influence during the study period⁴⁸. Being consistent with monthly variations of inorganic ion concentrations, C₂-C₄ diacids show similar trends with a gradual increase from late autumn to early spring and decrease towards summer months. The seasonal variations of C₆, Ph, and Gly are characterized by winter maxima and summer minima (Fig. 3h,j and m). Similarly, concentrations of Pyr and MeGly maximized in winter/spring and minimized in summer (Fig. 3l,n). These results demonstrate that, during late autumn to early spring, East Asian emissions of organic acids and their precursors, followed by long-range atmospheric transport, are important factors in controlling the distributions of diacids and related compounds in the WNP. High speed westerly winds also play an important role in causing the highest concentrations of diacids in spring over the WNP (Figure S1b).

On the other hand, lower concentrations of diacids and related compounds in summer suggest a minor contribution either from local emissions over the sampling site or marine emission of diacids and their precursors in the WNP. Mochida, et al. 4d occumented that local anthropogenic emissions for diacids are insignificant at Chichijima, based on the lower concentration ratios of benzo[a]pyrene (BaP) to (C_2-C_{11}) . Therefore, it is reasonable to believe that the observed concentrations of diacids and their precursors during summer may be associated with marine biological sources and are attributable to the oxidation of unsaturated fatty acids⁷⁵. In this connect, the concentrations of C_9 (aqueous phase photo-oxidation of biogenic unsaturated fatty acids) show a gradual increase from late spring to late autumn with a maximum in June and thereafter show a gradual decrease towards winter and spring months (Fig. 3i). These results suggest that oxidations of biogenic unsaturated fatty acids are important sources of diacids over the WNP in summer/autumn, although their contribution is relatively small compared to those in winter and spring.

Influence of meteorological parameters. Meteorological parameters such as wind speed, solar radiation and cloud cover are crucial for understanding the emission strengths of organic compounds and their oxidation processes in the atmosphere. As inferred from Figure S1, solar radiation was maximized in summer and minimized in winter/spring, indicating a strong photochemical oxidation during summer over the WNP. This result was further discussed in terms of seasonal variations in the mass concentration ratios of diacids. Being consistent with solar radiation, F/M, C₂/C₄, and C₃/C₄ ratios increased gradually from late spring to summer and stayed high until late autumn, and then decreased towards winter (Fig. 30,q,r). These seasonal changes suggest an enhanced photochemical oxidation, superimposed with changing regional biology and meteorology in summer and important sources of the diacids over the WNP. On the other hand, wind speed and cloud cover were higher during spring and/or winter (Figure S1), suggesting the enhanced processing of precursor compounds associated with Asian outflows during atmospheric long-range transport that leads to higher concentrations of diacids during spring and winter over the WNP. Similar seasonal variations have been found in the mass concentration ratios of C₆/ C_9 and Ph/ C_9 with higher values in winter and/or spring (Fig. 3s and t). $C_7/\omega C_7$ and $C_7/MeGly$ ratios showed a gradual increase from early spring to autumn with a peak in early autumn, i.e., September (Fig. 3v and x). These ratios, then decreased towards winter. However, ratios of C2/Gly did not show any clear seasonal trend during the study period, although ratios are slightly higher in summer and autumn (Fig. 3w). Although precipitation occurs throughout the year over the WNP, it was maximized in summer (Figure S1d); therefore, it is also an important sink for the diacids in addition to photochemical decompositions of oxalate-iron complexes, particularly in

Source apportionment. To quantitatively estimate the contribution of different sources to C_2 over the WNP, we performed a positive matrix factorization (PMF) analysis for the different seasons and the whole period (2001–2013) as shown in Fig. 4. PMF (version 5.1) is an effective source apportionment receptor model developed by the United States Environmental Protection Agency (U. S. EPA) and is often used in determining the sources of atmospheric aerosols^{76,77}. A complete description of PMF analysis is discussed elsewhere^{78–80}. The concentrations

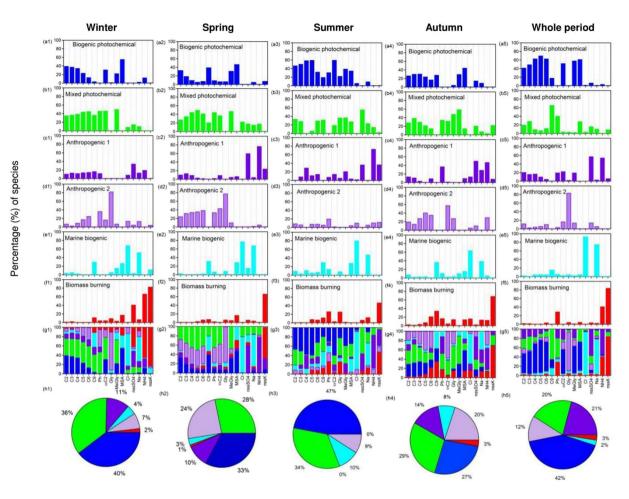


Figure 4. The PMF derived explained variance (%) for the source profiles for the different seasons as well as the whole period (2001-2013) during the study $(\mathbf{a}-\mathbf{f})$. Contributions of each source profile to individual acids (\mathbf{g}) and oxalic acid (\mathbf{h}) for the different seasons over the WNP. Each color indicates the different source as mentioned in plots a-f. Therefore, for interpretation of the colors in plots g and h, the reader is referred to the color version of this article.

of C_2 – C_6 , C_9 , Ph, ωC_2 , Gly, MeGly, and the tracers of water-soluble ions (MSA $^-$, Cl $^-$, nss-SO $_4$ ^{2 $^-$}, Na $^+$, NH $_4$ ⁺, nss-K $^+$) were used as inputs in the PMF analysis. A total of 607 samples were used for this analysis. We identified 6 source profiles such as biogenic photochemical (indicated by blue color), mixed photochemical (green), anthropogenic 1 (purple), anthropogenic 2 (weak purple), marine biogenic (cyan), and biomass burning (red). The derived variations (%) of the species are shown in Fig. 4a–f. The contributions of all sources to the individual diacids for the different seasons as well as for the whole period are shown in Fig. 4g. The detailed descriptions of each PMF resolved-sources are described below for the different seasons over the WNP.

Based on the high variation (%) of loading of MSA $^-$, C_2 , C_3 , ωC_2 , MeGly and weak or no loading of C_6 , Ph, and Gly, we identified source 1 as biogenic photochemical 1. Source 2 was identified as mixed photochemical, which was confirmed by a significant loading of all species, which are majorly associated with photochemical oxidation of longer to shorter chain diacids. Source 3 was identified as anthropogenic 1 as evidenced by high loading of nss-SO₄²⁻ and NH₄ $^+$. Source 4 was attributed to anthropogenic 2, because it may be associated with biomass burning-derived VOCs as evidenced by high loading of Gly. Source 5 was considered as marine biogenic unsaturated fatty acids, as evidenced by high loading of Na $^+$, Cl $^-$, and C₉. Source 6 was attributed to biomass burning, which was majorly associated with primary emissions, as indicated by high loading of nss-K $^+$.

Figure 4h shows the contributions of all sources to C_2 for the different seasons and the whole period. The biogenic photochemical process is a prominent source of C_2 , whose contribution to C_2 is highest in summer (47%), followed by winter (40%) and lowest in autumn (27%). The next prominent source is a mixed photochemical process, whose contribution is more abundant during winter (36%), summer (34%) and autumn (29%) and lowest in spring (28%). The contribution of anthropogenic source 1 is higher in autumn (14%) and winter (11%), whereas those of anthropogenic source 2 are higher in spring (24%), autumn (20%) and lower in summer (less than 9%) and winter (7%). Contributions of biomass burning to C_2 are highest in winter (3%) or autumn (2%) and lowest in summer (<1%), while the contributions of marine biogenic unsaturated fatty acids to C_2 are highest in summer (10%) followed by autumn (8%) and lowest in spring (3%). Overall, for the whole study period (Fig. 4h5), the contribution of biogenic photochemical process (42%) was the dominant source of C_2 followed

by anthropogenic sources (1 plus 2 contribute \sim 32%) and mixed photochemical sources (20%). We found that marine biogenic unsaturated fatty acids are important sources for the formation of diacids over the WNP during winter and summer, respectively.

Conclusions and implications. The 13-year observations of water-soluble diacids and related compounds in marine aerosols in the WNP provided the following findings.

The molecular distributions of diacids showed a predominance of C_2 followed by C_3 and C_4 . Seasonal variations of diacids and their precursor compounds showed maxima in winter to spring and minima in summer, except for C_9 , which was maximized in summer. Annual concentrations of total diacids, ω -oxoacids, pyruvic acid and α -dicarbonyls showed continuous increases toward more recent years.

A decrease in anthropogenic emissions is inferred from a decrease in the trends of anthropogenic tracer compounds such as phthalic acid (Ph), adipic acid (C_6) and glyoxal (Gly), while an increase in biogenic emissions is confirmed from an increase in the concentrations of biogenic tracers including pyruvic acid (Pyr) and methylglyoxal (MeGly) during 2001 to 2013. On the other hand, satellite-derived oxidation levels (total columnar O_3 and tropospheric columnar NO_2) showed significant increases during the study period. These results demonstrate that the increased concentrations of diacids over the WNP are probably due to not only the increased biogenic emissions from East Asia but also increased oxidation processes during atmospheric long-range transport, while anthropogenic precursors of the diacids are decreased or constant during the study period over the WNP. These results further support our previous study, which reported the declined and increased concentrations of nss- SO_4^{2-} and MSA^- , respectively, over the WNP during 2001–2012. We also found increased concentrations of C_9 and ωC_9 in summer, suggesting that marine biogenic unsaturated fatty acids are becoming important sources of diacids over the WNP, particularly, in summer.

These inferences are further supported by PMF analysis, which showed a biogenic photochemical contribution (42%) was a predominant source for C_2 . This is the first study to explain the impact of heterogeneity in air masses on long-term trends of organic acids over the WNP. Therefore, the assessment of future climate effects of East Asian aerosols over the WNP will need continued observations because of the rapid changes in the emission strength of aerosols and their precursors over East Asia. The results of this study should be important for climate modelers, who are interested in radiative forcing calculations over the WNP.

Methodology

Collection of aerosol samples. Aerosol samples were collected on a quartz filter $(20 \times 25 \, \text{cm}, \, \text{Pallflex} \, 2500 \, \text{QAT-UP})$ from 2001 to 2013 using a high volume sampler (HVS) with a flow rate of 1 m³ min⁻¹ at Chichijima Island $(27^{\circ}04' \, \text{N}, \, 142^{\circ}13' \, \text{E})$ in the WNP (Fig. 1). Before sampling, filters were pre-combusted at 450 °C for three hours. The HVS was set up 5 m above ground level at the Satellite Tracking Centre of Japanese Aerospace Exploration Agency (JAXA, elevation 254 m a.s.l.) in Chichijima Island⁴⁵. 4–6 day integrated samples were collected during the study period. After the sampling, filters were put in a pre-baked (450 °C for 6 hrs) glass bottle with a Teflon lined screw cap and stored at $-20 \, ^{\circ} \text{C}$ prior to the analysis of diacids. A total of 607 aerosol and 61 field blank samples were used in this study. A field blank sample was collected every ten aerosol samples by placing a clean filter in the cartridge of the HVS for 10 sec without the pump running.

Determination of diacids and related compounds. The collected filter samples were analyzed for diacids and related compounds using the improved method of Kawamura⁸¹ and Kawamura and Ikushima¹. Briefly, a portion of each filter sample was extracted three times with 5 ml of organic-free ultrapure water (resistivity of >18 $M\Omega$ cm⁻¹, Sartorius arium 611 UV) under ultrasonication. The extracts were filtered through a Pasteur pipette packed with quartz wool to remove the filter debris and insoluble materials and placed in a 50 ml pear-shaped flask. The water extracts were pH-adjusted to 8.5-9.0 using a 0.05 M potassium hydroxide (KOH) solution and then concentrated to almost dryness using a rotary evaporator under vacuum. A 14% borontrifluoride in n-butanol solution was added to the extracts and then heated at 100 °C for 1 hour to derivatize the carboxyl and aldehyde groups. The derived dibutyl esters and dibutoxy acetals were extracted with n-hexane and washed three times with ultrapure water (for the removal of polar compounds, including hydrogen fluoride (HF) and boric acid (H₃BO₄) derived from the borontrifluoride) and concentrated using a rotary evaporator in a vacuum and nitrogen (N2) blow down system. After being dried, a known amount of n-hexane was added to the ester fraction and derivatives were determined using a gas chromatography with a flame ionization detector (GC/FID; Hewlett-Packard, HP6890). Identification of the GC peaks was confirmed by comparing the GC retention times with those of authentic standards and confirmed by mass spectral examination using a GC/mass spectrometry (GC/MS; Thermoquest, Trace MS).

In order to check the recovery, $5\,\mu\text{L}$ of authentic diacids were spiked on the pre-combusted ($450\,^{\circ}\text{C}$ for 6 hrs) quartz filters and analyzed like a real sample using the above-mentioned procedure. The recoveries were 85% for oxalic acid, 90% for malonic acid and more than 90% for succinic, glutaric, and adipic acids. The analytical errors in the duplicate analyses are less than 10%. The concentrations of all diacids and related compounds reported in this study have been corrected using the field blanks. The blank levels were less than 5% for the major species measured in the real samples.

Several studies show that positive and negative artifacts can be significant during filter sampling and organic analysis of aerosols^{82, 83}. They generally arise from adsorption of gaseous species to the substrate surface and from particle losses on the walls and volatilization of semi-volatile species, respectively. In order to evaluate the potential artifacts, including adsorption or evaporation of collected particles to the gas phase, we performed simultaneous measurements of the diacids in marine aerosols using a HVS and a denuder/filter/denuder system⁴³. It is well established that the artifacts due to adsorption or evaporation are to be minor for the denuder system. We found good agreement between the two different techniques, suggesting that the HVS technique is valid for

collection of diacids in marine aerosols⁴³. This is true because, in general, evaporation loss of particulate organic species is minor compared to gases adsorbed on the filter surfaces⁸⁴. On the other hand, adsorption of gases is limited because diacids are predominantly observed in particle phase⁸⁵. However, it is noteworthy that organic compounds originating from continental regions are much more aged during atmospheric transport toward Chichijima Island; therefore the artifacts may be minor for the diacids. More details about the potential sampling artifacts are described in elsewhere⁴⁴.

Statistical analysis. To explore the inter-annual difference in the concentrations of the diacids and related compounds, the statistical regression analysis of variations (ANOVA)⁸⁶ was performed by comparing all the data points during the study period. Differences with p < 0.05 were considered to be statistically significant and indicated by a star (*) in the trend analyses as shown in Table 1. This statistical approach is simple, robust and easy to interpret⁸⁷. For example, the sign of the diacid trend depends on the value of the slope of the regression analysis. In this kind of interpretation, when the slope is greater than zero, the trend is positive (increases) whereas the trend is negative (decreases) when the slope is less than zero. When the slope is equal to zero, there is no trend in the diacid concentrations. Uncertainties in the trends are reported as standard errors in the slope of regression lines and variations in the trends (% yr⁻¹) are also reported for each organic compound in Table 1.

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Author Contributions

All authors contributed to the data interpretation and reviewed the manuscript. K.K. designed and led the research work. E.T. performed the data analysis. S.K.R.B. wrote the manuscript with significant contributions from K.K.

Additional Information

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