## **CORRECTION**

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## Author Correction: A CRISPR/Cas system mediates bacterial innate immune evasion and virulence

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We have been unable to reproduce the RNA stability experiment in Fig. 2a of this Letter, and no longer observe a Cas9-dependent decrease in the relative levels of the FTN\_1103 transcript. The new data reveal that the wild-type and *cas9* deletion mutant ( $\Delta cas9$ ) strains have equivalent levels of FTN\_1103 mRNA at each time point (see Fig. 1 of this Amendment). We do not have an explanation for the discrepancy with the original data, but it may in part be due to the complications derived from measuring the stability of the FTN\_1103 transcript in strains with very different baseline levels of this mRNA. We have also been unable to replicate the results of the immunoprecipitation experiments in Fig. 2g of the original Letter, and no longer observe lower levels of the FTN\_1103 transcript after immunoprecipitation with Cas9(R59A)-Flag as compared with wild-type Cas9-Flag (see Fig. 2 of this Amendment). We do still observe the original trend of higher levels of small, CRISPR-Cas-associated RNA (scaRNA) and transactivating crRNA (tracrRNA) in the wild-type Cas9-Flag immunoprecipitation as compared with the Cas9(R59A)-Flag mutant (Fig. 2e, f of the original Letter). However, we cannot rule out that these new results are simply a reflection of the overall levels of these transcripts in the different strains, owing to the biological lack of a Cas9-regulated control outside the  $FTN_{1103}$  locus. Independently of the immunoprecipitation experiment, there remain robust genetic data indicating that scaRNA and tracrRNA are required for Cas9-dependent repression of  $FTN_{1103}$  mRNA levels.

In addition, in Fig. 2h of this Letter, a tracrRNA mutant (tracrR-NA(rc13–17)) containing substitutions in the putative interaction site with *FTN\_1103* was used to suggest that the expression of *FTN\_1103* was repressed via interaction with the tracrRNA. Although not known at the time, subsequent crystallographic data<sup>1</sup> indicate that the mutation to tracrRNA was in an important stem–loop region that interacts with Cas9, and Cas9 has been shown to be required for stable tracrRNA levels<sup>2</sup>. For these reasons, we can no longer definitively deduce the role of the mutated tracrRNA bases in the repression of *FTN\_1103* and have no evidence to support a role of tracrRNA in direct interaction with *FTN\_1103* RNA.

Together, these results suggest that we do not have evidence to support RNA degradation as the mechanism that underlies Cas9mediated regulation of *FTN\_1103* mRNA expression. The other experiments and conclusions presented in the Letter are unaffected by this Amendment, and we apologize for the previous misleading data and the effect that they may have had on others in the field. We are grateful to Hannah K. Ratner for identifying these errors, conducting the experiments described here along with Siddharth Jaggavarapu, and for writing the majority of this Amendment. The original Letter has not been corrected.

- Deltcheva, E. et al. CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III. Nature 471, 602–607 (2011).
- Hirano, H. et al. Structure and engineering of *Francisella novicida* Cas9. *Cell* 164, 950–961 (2016).



Fig. 1 | The incorrect published Fig. 2a and the corrected Fig. 2a of the original Letter. Stability of  $FTN_{-1103}$  RNA in wild-type (WT) and  $\Delta cas9$  Francisella novicida. Levels of  $FTN_{-1103}$  RNA (relative to uvrD) were measured by quantitative PCR (qPCR) at 0, 15 and



30 min after treatment with rifampicin. Results are indicated as the percentage of  $FTN_{1103}$  mRNA remaining from the levels at t = 0 (n = 8).

## **CORRECTIONS & AMENDMENTS**

## Original 2e-g



Fig. 2 | The incorrect published Fig. 2e–g and the corrected Fig. 2e–g of the original Letter. e–g, qPCR after immunoprecipitation of wild-type Cas9–Flag or mutant Cas9(R59A)–Flag from cell lysates.

Levels of scaRNA (e), tracrRNA (f) and *FTN\_1103* (g) were quantified by qPCR relative to *uvrD* (n = 3). Data are mean and s.e.m. \*\* $P \le 0.005$ ; \*\*\* $P \le 0.001$ , two-tailed Student's *t*-test.