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A pathway to peptides in space through the condensation of atomic carbon

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Organic molecules are widely present in the dense interstellar medium, and many have been synthesized in the laboratory on Earth under the conditions typical for an interstellar environment. Until now, however, only relatively small molecules of biological interest have been demonstrated to form experimentally under typical space conditions. Here we prove experimentally that the condensation of carbon atoms on the surface of cold solid particles (cosmic dust) leads to the formation of isomeric polyglycine monomers (aminoketene molecules). Following encounters between aminoketene molecules, they polymerize to produce peptides of different lengths. The chemistry involves three of the most abundant species (CO, C and NH₃) present in star-forming molecular clouds, and proceeds via a novel pathway that skips the stage of amino acid formation in protein synthesis. The process is efficient, even at low temperatures, without irradiation or the presence of water. The delivery of biopolymers formed by this chemistry to rocky planets in the habitable zone might be an important element in the origins of life.

The origin of life has always been one of the most intriguing questions throughout human history. Biomolecules delivered to early Earth by asteroids, meteorites or comets during the period of heavy bombardment about four billion years ago have been proposed to play a role in the origin of life^{1,2}. Similar processes may apply to rocky exoplanets (or their moons). Analysis of meteoritic material led to the identification of amino acids, sugars and nucleobases, among other complex organic molecules of extraterrestrial origin^{3,4}. The simplest amino acid, glycine, has been discovered in comets⁵. The widespread hypothesis of the formation of organic molecules in space suggests that they are synthesized in the icy mantle that covers the refractory particles of cosmic dust^{6,7}. At later stages, these dust grains form asteroids and comets. These bodies may have a liquid phase at low temperatures, where water mixes with gases such as CO₂ and NH₃ (refs. 8–10). The chemistry in the liquid phase may further enhance the molecular complexity. Experimentally, the formation of various amino acids, and even their dimers (as well as other organic molecules) has been detected following energetic processing of different molecular ices^{11,12}. More recently, non-energetic pathways were also found for small organic molecules^{13,14} and glycine^{15,16}. However, the formation of biopolymers under space conditions has not been demonstrated, although several chemical pathways in terrestrial conditions have been widely accepted^{17–19}.

It is generally assumed that the prebiotic synthesis of peptides occurs in two stages. The first stage is the formation of amino acids, and the second is their polymerization²⁰. The polymerization process requires the condensation of amino acids accompanied by the loss of water. This has a high energy barrier and therefore proceeds only at high temperatures or requires energetic processing of the material^{21,22}. Consequently, each of these stages has a relatively low probability. Instead of first synthesizing amino acids to subsequently dehydrate them for the polymerization process, we suggest a much simpler and direct formation of the aminoketene (NH₂CH=C=O) and its polymerization forming peptides under astrophysically relevant conditions. Quantum chemical calculations predict that the

CO + C + NH₃ $\xrightarrow{\text{surface}}$ NH₂CH=C=O reaction is barrierless, and therefore occurs on the cold dust grains without involving external energy²³. Although isolated aminoketene is theoretically predicted to be stable^{24,25}, it has never been observed in experimental studies to the best of our knowledge. This can be related to the extremely high reactivity of the aminoketene, which could also lead to its efficient polymerization. The polymerization of NH₂CH=C=O via the nucleophilic attack of the nitrogen on the carbonyl carbon and an intramolecular proton transfer from NH₂ to CH results in the formation of peptide chains. Therefore, in contrast to the polymerization of amino acids, the formation of polyglycine via polymerization of aminoketene is a much simpler process.

To test this reaction pathway experimentally, we performed the co-deposition of CO, C and NH₃ on the surface of both Si and KBr substrates cooled to 10 K and placed inside an ultrahigh vacuum (UHV) chamber to mimic the conditions in dense molecular clouds.

Chemistry at 10 K

Infrared (IR) absorption spectra of the ice produced after the co-deposition of CO + NH₃ and CO + C + NH₃ at low temperature ($T = 10$ K) are shown in Fig. 1. The additional control experiments, involving only two reactants C + CO and C + NH₃, are shown in Extended Data Figs. 1 and 2.

The IR spectrum representing the deposition of all three reactants (CO, C and NH₃) on the substrate at 10 K reveals several new absorption features, which are absent in reference spectra involving only two reactants (for example, C + CO, C + NH₃ and CO + NH₃). Therefore, these bands have to be assigned to a product formed by reactions of all three reactants at 10 K. In addition, the warming up of the CO/C/NH₃ ice layer to room temperature leaves a residue on the substrate, which is also observed for the reaction of C atoms with only CO or NH₃, but in much smaller amounts. We conclude that the product formed in the reaction of all three reactants at 10 K is required for the formation of the non-volatile products at 300 K. As C atoms are the limiting reactant (that is, the fluence of C atoms was at least ten times less than that of NH₃ or CO), the reactions of

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Fig. 1 | IR absorption spectra of the ice mixtures produced at 10 K. The red line shows the IR spectrum of CO/NH₃ (1:1) ice and the black line the spectrum of CO/NH₃/C (1:1:0.1) ice. Stretching and bending vibrations are denoted ν and Δ , respectively.

C atoms with other C atoms or with its reaction products are negligible. Consequently, the observed product is expected to originate from the reaction of CO + C + NH₃ and contains CH, CN, C=O and NH_n groups, as shown in the IR spectrum (black line in Fig. 1). Given that NH₃ does not react with CO at 10 K, the addition of C atoms must act as a chemical trigger to initiate such solid-state reactions. The formation of the C=C=O functional group is demonstrated by the appearance of the absorption bands in the ~1,965–2,275 cm⁻¹ range. It implies the transformation of the CO triple bond (C≡O) to a CO double bond (C=O) species by attaching a C atom to the C atom of carbon monoxide. Similarly, the detection of CH and NH_n (2,500–3,000 cm⁻¹) and CN (1,320–1,500 cm⁻¹) absorption bands also suggests covalent bond formation between the added C atom and ammonia: C-atom addition to N atoms of NH₃ followed by H-atom rearrangement. On the basis of the above considerations, we can conclude that O=C=CH_nNH_n molecules are formed.

To better understand the chemistry involving all three reactants at 10 K, we performed quantum chemical calculations (results are shown in Fig. 2 and Extended Data Fig. 3). As shown in Extended Data Fig. 3, C atoms initially prefer to react with ammonia molecules, forming the prereactive complex CO + CNH₃, which corresponds to the first energy well in Fig. 2. The C + NH₃ reaction pathway has been studied experimentally¹⁶ using the recently developed calorimetric technique²⁶. The proton transfer from nitrogen to carbon was observed to form NH₂CH before the energy dissipation¹⁶. This mechanism also applies to the reaction investigated here, as shown by the appearance of NH_n and CH absorption bands (~3,048 and ~2,820 cm⁻¹, Fig. 1) after C-atom addition. The formed NH₂CH product subsequently reacts barrierlessly with CO, leading to the formation of NH₂CH=CO. There are thus two possible products of this reaction: NH₂CH=CO and CH₂NH + CO. The latter is the same as the product of the reaction with only two reactants C + NH₃ → CH₂NH. Therefore, the reaction product observed upon the addition of all three reactants at 10 K can only be NH₂CH=CO, the functional groups of which are observed in the IR spectrum. The formation of this molecule was confirmed by temperature-programmed desorption in combination with quadrupole mass spectrometer analyses monitoring the ions with masses of 57 and 56 unified atomic mass units (u) as shown in Extended Data Fig. 4. At the same time, C atoms could react with either CO or NH₃, and a smaller number of CCO or H₂CNH, or even NH₂CH₂NH₂ molecules can also be formed.

Chemical transformation during temperature rise

After deposition, the substrate was heated at a rate of 2 K min⁻¹. The IR spectra measured at selected temperatures are shown in Fig. 3.

Fig. 2 | Energy level diagram for the reaction involving CO, C and NH₃ reactants. The reaction starts with the triplet state. The transition states (TS) and the singlet states (S) are labelled and the dashed lines show the possible reaction pathways, with the most probable pathway based on computational and experimental results in red.

A comparison of the spectra at 10 K and 70 K shows that the sublimation of CO, occurring mainly from 25 to 40 K, does not initiate a new chemistry. However, with the sublimation of ammonia (above 110 K), the products of the C-atom reactions can finally encounter each other and react. We observed the formation and growth of new IR absorption bands, while the intensity of the ν NH_n and ν C=O bands as shown in Fig. 3 decreased significantly. With the mass spectrometer, we did not detect any considerable desorption from the substrate besides CO and NH₃. Therefore, the disappearance of these IR bands was due to chemical transformation rather than sublimation. The decrease of band intensities in the 2,800–3,050 cm⁻¹ range is a common indicator of glycine polymerization²⁷. This occurs due to the transformation of the NH₃⁺ or NH₂ groups of glycine into the NH groups present in peptides. Simultaneously with the transformation of NH_n groups, we observed the formation of the peptide I band at 1,670 cm⁻¹ and peptide II band with a peak maximum ranging from 1,548 to 1,569 cm⁻¹ (Fig. 3 and Extended Data Fig. 5), which are the characteristic features of peptide bonds. The peptide bond is therefore formed at 100–120 K simultaneously with the decrease in the intensity of the IR bands associated with the molecules present in the ice at 10 K (Extended Data Fig. 6). The IR spectrum did not change substantially above 150 K; the residual material present on the substrate after warming up to 300 K (R300K) must therefore have formed at low temperature during ammonia sublimation.

Characterization of the room-temperature residue

The in situ characterization was performed by IR spectroscopy. After removing the substrate from the UHV chamber and exposing it to air, we did not observe any noticeable modification of the IR spectrum of R300K. Therefore, an ex situ analysis could also be performed. The high-resolution ex situ mass spectrum of R300K is shown in Fig. 4. To better characterize the mass spectrum, we selected a threshold shown by the horizontal dashed line in Fig. 4. The peaks with intensities higher than this threshold were analysed to determine the series. We could identify many series of mass peaks, which are separated by the same mass of about 57.0215 u. This value exactly matches the mass of the NH₂CH=CO molecule, which was found to be the main product formed at 10 K. The intensities of the peaks in these series drop exponentially with increasing mass. A series is formed if the same molecule is continuously added to an initial species. Therefore, the presence of these series with

Fig. 3 | IR absorption spectra obtained during annealing of the material produced by co-deposition of CO + C + NH₃. The labels for the vertical dashed lines indicate the positions of the peaks appearing during heating at the temperatures indicated. All CO is sublimated at 70 K and most NH₃ is sublimated at 120 K.

such an intensity distribution unambiguously indicates the polymer formation with the mass of the monomer unit equal to the distance between the peaks. The complete list of all series found is given in Extended Data Fig. 7. As shown in Fig. 4, most of the analysed mass peaks belong to the series.

We can therefore conclude that the polymerization of the NH₂CH=C=O molecule formed at 10 K, is the main chemical pathway leading to the formation of R300K. The formation of a variety of different series (Extended Data Fig. 7) can be understood assuming that other molecules formed at 10 K could be added during the polymer formation.

The IR spectrum of R300K is shown in Fig. 5 (see also details in Extended Data Fig. 5). It highlights the absorption features characteristic for the peptide bonds. As the NH₂CH=C=O molecule does not contain the peptide bond, the appearance of this bond in the formed polymer indicates the type of the bond between the monomeric units. Upon sublimation of volatile species, the monomeric units of NH₂CH=C=O are assembled through peptide bonds, resulting in glycine oligomers, which differ from canonical peptides only by the groups at their C termini. This proceeds by the polymerization of NH₂CH=C=O molecules, involving proton transfer from nitrogen to the carbon atom of the CH group. However, our density functional theory calculations show that the NHCH₂C=O (as well as NH₂CHCO–NHCH₂C=O) molecules are not stable; they are expected to fragment with the loss of the CO molecule. Therefore, peptide chains produced by this mechanism are terminated by NH₂ on one side, which corresponds to the canonical structure of the peptide. The other side of the peptide chain is terminated by a CH₂ group instead of a classical COOH group. However, the observed series is not the most intense one. The most intense peak (189.0983 u) shown in Fig. 4 suggests that the favoured oligopeptides are terminated by amino groups on both sides, which assumes an efficient involvement of ammonia in the polymerization process. In spite of the differences in length and terminal functional groups between the canonical Gl₃ peptide and the peptides identified in our experiment, there is a good match between the two spectra shown in Fig. 5. The main discrepancy is the width of the IR absorption bands; this is expected owing to the formation of oligopeptides with a wide distribution of lengths. The band broadening is caused by the change in the positions of the IR absorption bands with the number of units in the glycine oligomers²⁸.

Fig. 4 | Ex situ mass spectrometric analysis of the R300K residue.

Top: the mass spectra of R300K obtained from ex situ analysis using an ESI-Orbitrap mass spectrometer. The horizontal orange dashed line shows the threshold used to select the analysed peaks. The peaks that belong (or do not belong) to the series are displayed in red (blue). Black peaks lie below threshold and were not analysed for series. The molecular structures display the Gl₄ peptides detected in the mass spectra as protonated species [analyte + H]⁺. Bottom: the intensities of the bands derived from two observed series corresponding to glycine peptides are plotted as a function of the numbers of monomeric units (*n*). Error bars are standard deviations of corresponding peak intensities in the measured mass spectra.

Finally, to confirm the proposed molecular structure of the polymers, we performed the higher-energy C-trap dissociation²⁹ of the most abundant ion from Fig. 4. The result of this measurement is shown in Fig. 6. This analysis explicitly confirms the suggested molecular structure of this ion with mass 189.0984 u. The efficient loss of two ammonia molecules demonstrated by peaks b3 and b3* clearly shows the terminal location of both amino groups. The a3 peak shows that the side amino group is linked with the CO unit. The b2 and y1 peaks demonstrate that the side H₂NCO unit is connected to the CH₂NH unit. The same analysis can also be performed from the other side, where the terminal amino group is joined to the CH₂CO group, as demonstrated by the y2 fragmentation peak. The fragmented ion (189.0983 u) belongs to the series of peaks that considerably exceeds the intensities of other series. Therefore, the mass peaks from this series can be separated by both mass and intensity. This shows that the formation of the ions from this series happens via polymerization of the NH₂CH=C=O molecule. All of this unambiguously confirms the suggested molecular structures of the ions from this series. Moreover, it also strongly suggests that ions from other series shown in Fig. 4 have a similar molecular structure and are modified peptides. We evaluated the entire mass spectrum, comparing the integrated intensities of mass peaks in series and the total integrated intensities of all analysed peaks. This analysis determined a polyglycine concentration in our sample of about 85%, given the same ratio between the polyglycine peaks and other mass peaks among non-analysed peaks below the threshold.

Fig. 5 | Comparison between the IR absorption spectra obtained from R300K and GI₃. The vertical dashed lines are drawn to visualize the coincidences between the peaks in the two spectra. The Gaussian profiles beneath R300K spectrum show the deconvolution of the experimental spectrum (Extended Data Fig. 5). The spectrum of triglycine (GI₃) is adopted from ref. ²⁸.

If all non-analysed (weak) peaks did not belong to polyglycine, the value would be reduced to 19%. Considering that the R300K material is completely solvated in water and that the ratio of the intensities of the peptide bands towards other IR bands in the spectrum of our sample is comparable to that in the spectrum of pure oligoglycine, we concluded that the predominantly formed molecules in our experiments are polyglycine.

Implications

The reactants used in this work are among the most abundant species present in the interstellar medium (ISM). The fractional abundances of NH₃ and CO in the ice mantles covering refractory dust particles are 10% and 40%, respectively³⁰. In the ISM, more than half of all carbon exists in the form of atomic gas³¹. The formation of dense clouds, where new stars and planets are formed, goes through a stage of translucent clouds, where a notable portion of carbonaceous dust is expected to be formed due to the accretion of C atoms. In translucent molecular clouds, the dust temperature is low enough to allow the formation of molecular ices while carbon is dominantly present in the atomic or CO forms³². Therefore, reactions between accreted CO, C and NH₃ could be very common in this region. At the low temperatures of dust in translucent molecular clouds ($T = 10\text{--}20\text{ K}$)³³, this leads to the formation of organics³⁴. As demonstrated here, a portion of these organics could be in the form of peptides. At later stages, such dust becomes the building blocks of comets or meteorites. The formed organics could therefore have been delivered to the early Earth during the period of heavy bombardment. The survival of organics at all delivery stages is thus a relevant question. In this sense, comets have the advantage of being less mechanically stable than asteroids, because their fragmentation after entering the atmosphere may allow strong impacts to be avoided due to the loss of kinetic energy in the atmosphere as the meteor breaks up². Comets might deliver intact organics produced in the ISM at a rate of at least $10^6\text{--}10^7\text{ kg yr}^{-1}$ (ref. ²). However, even the direct impact of larger objects may allow the partial survival of molecules, including amino acids^{35,36}. The biggest issue with this extraterrestrial scenario is the stability of the newly formed organics in the harsh energetic environment of the ISM, filled with X-ray and ultraviolet photons. The non-energetic formation of peptides may be advantageous in this regard: if gas-to-solid phase transitions and reactions occur in regions with a low ultraviolet flux, the formed peptides could survive long enough to be incorporated into bulk solids, which would shield them from further destruction. This is in line with the potential detection of proteins in meteorites^{37,38}.

Fig. 6 | The higher-energy C-trap dissociation of the 189.0983 u ion. The fragmentation peaks are annotated based on the accepted nomenclature for the peptide fragments^{29,44}. The loss of an additional ammonia molecule is indicated with an asterisk. The inset shows the molecular structure of the investigated ion. The coloured marks (a, b, y) determine the fragments that correspond to the mass peaks identified in the spectrum.

Peptides also play a key role in the origin of life³⁰. The delivery of peptides to Earth should therefore expand the opportunities for evolutionary chemistry and biology. Moreover, the peptides with two amino terminals dominantly formed in our experiments are capable of efficient self-assembly³⁹, which offers the prospect of new and interesting opportunities for abiogenesis.

Methods

In situ experiments. The experiments were performed using the UHV INterStellar Ice Dust Experiment (INSIDE) set-up described elsewhere⁴⁰. We performed the co-deposition of CO, C and NH₃ on the surface of KBr substrates cooled down to 10 K and centred in the UHV chamber. The deposition of reactants was performed for about 1 h. Low-energy carbon atoms were generated by an atomic carbon source⁴¹. The source generated a pure flux of low-energy carbon atoms in the triplet ground state $C(^3P_2)$ and about 1% of CO/CO₂ molecules. The background pressure inside the vacuum chamber (1×10^{-10} mbar) and the temperature of the substrate (10 K) allowed us to mimic the chemistry under dense molecular cloud conditions. We used approximately equal amounts of CO and NH₃ molecules, while the number of C atoms was at least ten times smaller. The flux of C atoms from the source was estimated in previous work⁴². This small amount of C atoms allows us to exclude their reactions with each other, as well as with their reaction products. The gases were introduced through two separated leak valves. The ice thicknesses on substrates were monitored by infrared (IR) spectroscopy using a Fourier transform IR spectrometer (Vertex 80 v, Bruker) in the transmission mode. In situ mass spectrometry was performed using a quadrupole mass spectrometer (HXT300M, Hosi-trad) attached to the same UHV chamber. After the deposition, the substrate was heated at a rate of 2 K min^{-1} . As the material was warmed, IR absorption spectra and mass spectra were measured to monitor the residue and sublimated gas-phase species, respectively. The evolution of IR spectra during temperature rise (shown in Extended Data Fig. 6) was obtained by integrating the corresponding bands in the IR spectra.

Ex situ mass spectrometry analysis. For the mass spectrometry analysis, we used silicon substrates and performed a 4 h deposition of the reactants CO, C and NH₃ under the same conditions as the in situ experiments. We obtained very similar IR spectra at all stages of the experiment, using both Si and KBr substrates. After warming up, the substrates were removed and analysed. The residue was extracted with a water–methanol mixture (70:30) and used for the mass spectral analysis. Complete solubility of the residue in water and water–methanol mixtures was observed. There was also no chemical reactivity of the residue with water, as shown by the unchanged IR spectrum of the material after prolonged exposure to wet air. The mass spectrometry was performed using the hybrid linear trap/Orbitrap mass spectrometer (Thermo Fisher QExactive plus mass spectrometer with a heated ESI source). The accuracy of the mass determination was higher than 5 ppm, which resulted in a possible error only in the fourth decimal place for the investigated

mass range. Considering the limited number of elements used in the experiments (C, O, N and H), this high resolution allowed us to unambiguously determine the elemental composition of the detected ions in the mass range shown in Fig. 4. To ensure that the observed mass peaks were generated by the material formed in our experiment, we checked for the possible presence of contaminants (both on the substrate and in the mass spectrometer) using the same procedure with clean silicon substrates.

The same mass spectrometer was used for tandem mass spectrometry measurements. For this purpose, the quadrupole filter was set to preselect ions with the mass range of 188.9–189.3 u (the most abundant Gl_3^+ ion observed in our mass spectra). This cation was then a subject for a higher-energy C-trap dissociation procedure, as described in ref. ²⁹. The collisional energy was set to 30 eV. In test measurements with 10 and 50 eV collisional energies, we did not observe any principal difference in the fragmentation pattern. Two impurity peaks (189.0734 and 97.0373 u) were subtracted from measured spectra. These peaks were assigned to the impurities on the basis of the presence of this ion signal during the tuning of the collisional energies when all other mass peaks were missing, and the fact that ions with these masses cannot simply be fragments of the parent ion.

Quantum chemical computations. Quantum chemical calculations were performed using the GAUSSIAN16 software⁴³. To find out the outcome of the reaction of C atoms landing on CO/NH_3 ice, we performed a two-dimensional potential energy scan of the $\text{CO} + \text{C} + \text{NH}_3$ reaction at the MP2/6-311+G (d, p) level. In this scan, we varied the C–C and C–N distances and allowed complete optimization of molecular geometries without any further restrictions. This computation allowed us to identify the formation of the first prereactive complex used for the calculation of the energy level diagram, which is shown in Fig. 2. The geometries of the molecules in the energy level diagram were determined at the B3LYP/6-311+G (d, p) level. The reaction energies were determined from the difference between the sum of the energies of the reactants and the energy of the product molecules with vibrational zero-point energy corrections.

Data availability

Source data are provided with this paper.

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Author contributions

S.A.K. initiated and led the project. S.A.K. and K.-J.C. recorded and analysed the experimental data. S.A.K. performed the computations and wrote the manuscript. N.U. performed ex situ mass spectrometry. Th.H. and C.J. participated in data interpretation and discussion. All authors contributed to the writing of the manuscript.

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Extended Data Fig. 1 | The IR absorption spectra of 300 K residues. a - C + CO + NH₃, b - C + NH₃, and c - C + CO control experiments.

Extended Data Fig. 2 | The comparison of IR absorption spectra in the range of vibrations of CO and NH_n groups of the materials produced by codeposition of different reactants on the substrate at 10 K. The bands that are observed only when three reactants are deposited are marked by arrows.

Extended Data Fig. 3 | Potential energy surface for the formation of the first energy well structure in Fig. 2. The color of the surface is linked to the energy scale. Big red data points show the barrierless pathway for the formation of the H_3NCCO molecule, while the most probable barrierless path leads to the left minimum and results in the formation of the $\text{H}_3\text{NC} + \text{CO}$ complex, which corresponds to the first energy well structure in Fig. 2. The formation of the H_3NCCO molecule does not notably change the chemical pathway as shown in Fig. 2. In this case, the addition of CO molecule takes place one step earlier, which has very little influence on the energy barrier for the proton transfer.

Extended Data Fig. 4 | The TPD curves measured by quadrupole mass spectrometer monitoring the mass 57 u, corresponding to the $\text{NH}_2\text{CH}=\text{C}=\text{O}$ molecule, and its expected fragment on the mass 56 u. The partial pressure is given around ionizer of QMS considering the same ionization cross section as for nitrogen.

Extended Data Fig. 5 | The deconvolution of IR spectra of R300K in the range of the peptide bond vibrations. The dots show the experimental spectrum, the individual Gaussian peaks used for the fit are displayed with thin lines of individual colors, and the cumulative fits are shown by bolder red lines. To demonstrate that there are several possibilities to fit the IR spectrum of R300K, we are providing several alternative deconvolutions of the experimental spectrum. The fitting was performed using Origin software without fixation of any band parameter. Both band positions and widths were optimized to get the closest match to the experimental spectrum. The peptide II band's position varies depending on whether the absorption peak marked as NH_2 (1592 or 1604 cm^{-1}) is included in the fit. Our calculations predict the presence of a notably intense NH_2 scissor vibration band between peptide I and peptide II bands in the spectra of short peptides terminated by amino groups on both sides. Moreover, the match between the experimental spectrum and the fit is much better when the NH_2 absorption band is included. Therefore, the deconvolution shown in lower frames seems to be more accurate.

Extended Data Fig. 6 | The evolution of the intensities of IR absorption bands during the temperature rise. The curves show the band intensities. The peak positions of the bands and the assignment (if available) are given in the figure. The horizontal dashed line shows the 50% intensity. The temperatures of its crossing with the TPD curves are given. The band at 1081 cm⁻¹ marked as NH₃ is expected to have a contribution from the solid residue at higher temperatures.

MASS	COUNTS	ASSIGNMENT	MASS	COUNTS	ASSIGNMENT
NH ₂ CH ₂ ends			NH ₂ NH ₂ ends + H ₂		
201.0983	219568	Gl ₃ C ₇ H ₁₃ N ₄ O ₃	191.114	13708	Gl ₃ C ₆ H ₁₅ N ₄ O ₃
258.1195	165487	Gl ₄	248.135	82003	Gl ₄
315.1412	105818	Gl ₅	305.1567	339660	Gl ₅
372.1627	56078	Gl ₆	362.178	44675	Gl ₆
429.1842	21739	Gl ₇	419.199	13724	Gl ₇
486.2057	15781	Gl ₈	NH ₂ NH ₂ ends, -O +C		
NH ₂ NH ₂ ends			185.1031	657156	Gl ₃ C ₇ H ₁₃ N ₄ O ₂
189.098	1388446	Gl ₃ C ₆ H ₁₃ N ₄ O ₃	242.1245	337857	Gl ₄
246.1194	509908	Gl ₄	299.146	123962	Gl ₅
303.141	121422	Gl ₅	356.167	60163	Gl ₆
360.1627	50370	Gl ₆	413.188	34643	Gl ₇
417.1842	18616	Gl ₇	470.211	15248	Gl ₈
474.2057	11917	Gl ₈	NH ₂ NH ₂ ends, -O		
Exact polymer			173.1031	378772	Gl ₃ C ₆ H ₁₃ N ₄ O ₂
172.0714	362672	Gl ₃ C ₆ H ₁₀ N ₃ O ₃	230.125	138257	Gl ₄
229.093	175596	Gl ₄	287.146	115564	Gl ₅
286.114	93262	Gl ₅	344.167	33097	Gl ₆
343.136	55089	Gl ₆	401.189	18335	Gl ₇
400.157	35490	Gl ₇	NH ₂ NH ₂ ends, -O +CH ₂		
457.178	16591	Gl ₈	183.0874	379942	Gl ₃ C ₇ H ₁₁ N ₄ O ₂
Exact polymer + H ₂			240.109	148396	Gl ₄
174.0871	367120	Gl ₃ C ₆ H ₁₂ N ₃ O ₃	297.13	83123	Gl ₅
231.108	262322	Gl ₄	354.152	31393	Gl ₆
288.13	119467	Gl ₅	411.173	13063	Gl ₇
345.151	65474	Gl ₆	-		
402.173	28327	Gl ₇	157.0606	339455	C ₆ H ₉ N ₂ O ₃
459.194	14628	Gl ₈	214.082	117609	
Exact polymer + CH			271.151	54266	
184.0715	457345	Gl ₃ C ₇ H ₁₀ N ₃ O ₃	328.125	34221	
241.0929	250906	Gl ₄	385.147	27692	
298.114	87861	Gl ₅	-		
355.136	49995	Gl ₆	253.0904	327530	C ₇ H ₁₅ N ₃ O ₇
412.157	25223	Gl ₇	310.112	45682	
469.179	13877	Gl ₈	367.133	7652	
Exact polymer + C ₂ H ₃			-		
198.0871	238076	Gl ₃ C ₈ H ₁₂ N ₃ O ₃	204.109	20844	C ₆ H ₁₄ N ₅ O ₃
255.108	97056	Gl ₄	261.1305	277321	
312.13	77725	Gl ₅	318.152	18823	
369.151	44642	Gl ₆	375.173	15283	
426.173	17160	Gl ₇	NH ₂ CH ₂ ends + Na		
NH ₂ NH ₂ ends + C ₂			196.069	733000	Gl ₃ C ₆ H ₁₁ N ₃ O ₃ Na
156.0766	383597	Gl ₂ C ₆ H ₁₀ N ₃ O ₂	253.0904	327530	Gl ₄
213.098	295559	Gl ₃	310.112	45681	Gl ₅
270.119	128775	Gl ₄	-		
327.141	80091	Gl ₅	154.0585	426849	C ₃ H ₁₀ N ₂ O ₅
384.162	30402	Gl ₆	211.08	285845	
NH ₂ NH ₂ ends + C ₃ H			268.101	100122	
168.0766	239784	Gl ₂ C ₇ H ₁₀ N ₃ O ₂	325.123	22401	
225.0981	259786	Gl ₃			
282.119	112662	Gl ₄			
339.141	60541	Gl ₅			
396.162	37179	Gl ₆			
453.183	15029	Gl ₇			

Extended Data Fig. 7 | The table of mass peaks found in series in the ex situ analysis of the 300 K residue. Masses correspond to protonated species [analyte+H]⁺ as detected in experiment, while assignment is given for the analyte.