# ARTICLE OPEN (Check for updates) Detection of molecular transitions with nitrogen-vacancy centers and electron-spin labels

C. Munuera-Javaloy (1,2<sup>22</sup>, R. Puebla (3,4, B. D'Anjou (5, M. B. Plenio (5) and J. Casanova (1,2,6)

We present a protocol that detects molecular conformational changes with two nitroxide electron-spin labels and a nitrogenvacancy (NV) center in diamond. More specifically, we demonstrate that the NV can detect energy shifts induced by the coupling between electron-spin labels. The protocol relies on the judicious application of microwave and radiofrequency pulses in a range of parameters that ensures stable nitroxide resonances. Furthermore, we demonstrate that our scheme is optimized by using nitroxides with distinct nitrogen isotopes. We develop a simple theoretical model that we combine with Bayesian inference techniques to demonstrate that our method enables the detection of conformational changes in ambient conditions including strong NV dephasing rates as a consequence of the diamond surface proximity and nitroxide thermalization mechanisms. Finally, we counter-intuitively show that with our method the small residual effect of random molecular tumbling becomes a resource that can be exploited to extract inter-label distances.

npj Quantum Information (2022)8:140; https://doi.org/10.1038/s41534-022-00653-w

# INTRODUCTION

A precise knowledge of molecular structure and dynamics would shed new light on the mechanisms by which, e.g., proteins interact with their environment<sup>1</sup>. This finds important applications in distinct fields such as biochemistry and biology, since macromolecular conformational changes underlie numerous biological processes. In this regard, different techniques have been proposed to study molecular structure and dynamics<sup>2–4</sup>. In particular, methods based on nuclear magnetic resonance (NMR)<sup>5,6</sup> and on electron-spin resonance (ESR)<sup>7,8</sup> stand out as they are central to investigate a wide variety of molecules, with ESR being more sensitive due to the much larger gyromagnetic ratio of electrons<sup>9</sup>. One commonly used ESR method is the Double Electron-Electron Resonance (DEER) in which synchronized and/or delayed pulses are delivered to, e.g., estimate average distances among free radicals in molecular ensembles<sup>10</sup>. However, traditional ESR based on inductive detection with macroscopic coils requires macroscopic samples with a high chemical purity to produce a detectable response. This rules out ESR investigations of static and dynamic properties of individual molecules.

Detectors based on NV centers in diamond<sup>11,12</sup> are promising candidates to overcome the limitations of inductive detection. Indeed, such detectors enable ESR and NMR detection of molecular samples down to the micron- and nanoscale and even down to the level of single molecules. The potential of NV centers in diamond originates from a number of appealing properties and early demonstrations. They can be initialized and read out using laser light in the visible spectrum, thereby removing thermal fluctuations from the measurement process<sup>13–15</sup>. Moreover, NV hyperfine states can be manipulated with microwave radiation in such a way that the NV couples coherently to nearby nuclear and electron spins while at the same time being decoupled from environmental fluctuations<sup>16–20</sup>. These concepts have enabled shallowly implanted NV centers to detect electron spins<sup>21</sup> and even nuclear spins <sup>22,23</sup> above

the diamond surface with single-spin sensitivity. Detection protocols have also been developed allowing for nanoscale NMR<sup>24-27</sup> and ESR detection<sup>28-30</sup> with spectral resolution approaching that of macroscopic ESR and NMR based on inductive detection. Fluctuating electron spins of biological samples<sup>31-34</sup> and small groups of statistically polarized nuclear spins in a target protein<sup>35</sup> have been detected. Finally and particularly relevant to the present work, an NV center has also been used to detect a single nitroxide electron-spin label<sup>36</sup> attached to a protein. The use of nitroxide spin labels is particularly attractive because it enables the study of nonparamagnetic molecules<sup>37</sup>. In particular, ref. <sup>38</sup> reported the detection of an effective inter-label coupling in molecular ensembles using advanced multi-frequency spectroscopy techniques. Attaching several nitroxides to a single target protein could enable the study of its internal dynamics at room temperature with an NV center. However, this approach is challenging because spin labels have resonance frequencies and interaction strengths that strongly depend on their orientation with respect to the external magnetic field<sup>39</sup>. Therefore, the identification of these parameters is complicated by the unavoidable molecular motion and environmental noise affecting the NV and the electron-spin labels.

In this work, we show how even a simple pulse sequence applied to a single pair of electron-spin labels and a shallow NV [see Fig. 1(a)] can detect single-molecule conformational transitions at room temperature. Our carefully engineered DEER sequence asymmetrically delivers microwave (radiofrequency) pulses to electron spins of the NV center (nitroxide spin labels) to resolve small energy shifts in the NV spectrum owing to the coupling between labels. Crucially, we find an energy-transition branch of the labels that presents an excellent robustness against unavoidable tumbling motion in ambient conditions. In addition, we show that this branch is especially robust when each nitroxide label contains a distinct nitrogen isotope. Finally, we develop a simple but accurate model which enables us to estimate not only the coupling but also the

<sup>&</sup>lt;sup>1</sup>Department of Physical Chemistry, University of the Basque Country UPV/EHU, Apartado 644, 48080 Bilbao, Spain. <sup>2</sup>EHU Quantum Center, University of the Basque Country UPV/ EHU, Leioa, Spain. <sup>3</sup>Instituto de Física Fundamental, IFF-CSIC, Calle Serrano 113b, 28006 Madrid, Spain. <sup>4</sup>Centre for Theoretical Atomic, Molecular, and Optical Physics, School of Mathematics and Physics, Queen's University, Belfast BT7 1NN, United Kingdom. <sup>5</sup>Institut für Theoretische Physik and IQST, Universität Ulm, Albert-Einstein-Allee 11, D-89069 Ulm, Germany. <sup>6</sup>IKERBASQUE, Basque Foundation for Science, Plaza Euskadi 5, 48009 Bilbao, Spain. <sup>IM</sup>email: carlosmunueraj@gmail.com

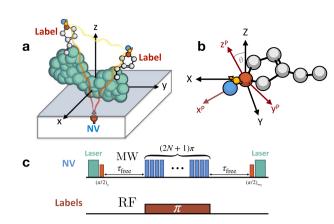


Fig. 1 System schematics and employed sequence. a A protein with two attached labels is near a diamond surface that contains a shallow NV. **b** Nitroxide molecule including the electron spin (yellow). The principal axes are shown in red, while the laboratory frame is shown in black. The azimuth  $\theta$  (the angle between the z principal axis and the z laboratory axis) determines the energy-transition branches of the label. **c** MW and RF driving scheme. This includes initialization and read out with laser and microwave pulses, two free evolution stages of duration  $\tau_{\text{freer}} 2N + 1$  MW  $\pi$ -pulses on the NV, and a simultaneous RF  $\pi$ -pulse on the labels.

distance between labels using Bayesian inference. In particular, we counter-intuitively show that the small residual effect of tumbling, instead of being detrimental, enables the extraction of the distance between labels.

# RESULTS

## The system

The Hamiltonian of the NV, the two nitroxide electron-spin labels, and the microwave (MW) and radiofrequency (RF) driving fields reads

Here,  $\sigma^z$  is the Pauli *z* operator for the NV two-level system and  $\mathbf{J}_i$  are spin-1/2 operators for each label (*i* = 1, 2). The Hamiltonian of the *i*th nitroxide label is  $H_{n_i}$  and the coupling of each label to the NV is mediated by the vectors  $\mathbf{a}_i$ . Moreover,  $J_i^{\pm} = J_i^x \pm i J_i^y$  are electron-spin ladder operators and  $g_{12}$  is the coupling constant between labels. The last line in Eq. (1) describes a MW field resonant with the NV, leading to the term  $\frac{\Omega_{MW}}{2}\sigma^x$  in a suitable interaction picture. In addition, an RF field of frequency  $\omega_{RF}$  and Rabi frequency  $\Omega_{RF}$  excites the electronic resonances of the nitroxide labels. Equation (1) is derived in the Supplementary Note 1 and a sketch of the system is presented in Fig. 1(a).

The term  $H_{n_i}$  models each nitroxide label as<sup>39,40</sup>

$$H_{n_i} = \mu_{\mathsf{B}} B^{\mathsf{z}} \hat{\mathsf{z}} \cdot \mathbb{G}_i \cdot \mathsf{J}_i + \gamma_{\mathsf{N}} B^{\mathsf{z}} I_i^{\mathsf{z}} + \mathsf{I}_i \cdot \mathbb{Q}_i \cdot \mathsf{I}_i + \mathsf{J}_i \cdot \mathbb{A}_i \cdot \mathsf{I}_i,$$
(2)

where  $\mu_{\rm B}$  is the Bohr magneton,  $B^z$  is a magnetic field applied along the *z* axis,  $\gamma_{\rm N}$  is the nuclear gyromagnetic ratio equal to  $2\pi \times 3.077$  kHz mT<sup>-1</sup> for <sup>14</sup>N and to  $2\pi \times -4.316$  kHz mT<sup>-1</sup> for <sup>15</sup>N,  $\mathbf{I}_i$  is the nuclear spin operator for the *i*th nitroxide label, and  $\mathbb{G}_i$ ,  $\mathbb{Q}_i$ , and  $\mathbb{A}_i$  are respectively the Landé, quadrupolar, and electron-nucleus interaction tensors<sup>40</sup>. We note that  $\mathbf{I}_i$  is a spin-1 (spin-1/2) operator when describing a <sup>14</sup>N (<sup>15</sup>N) nuclear spin. The components of the  $\mathbb{G}_i$ ,  $\mathbb{Q}_i$ , and  $\mathbb{A}_i$  tensors in a general reference frame depend on the frame's relative orientation with respect to the principal axes of the nitroxide [see Fig. 1(b)]. In the principal frame,  $\mathbb{G}_i$ ,  $\mathbb{Q}_i$ , and  $\mathbb{A}_i$  are diagonal<sup>40</sup>. In particular, the Landé tensor in the principal frame is  $\mathbb{G}_i^{(P)} = \operatorname{diag}(G^x, G^y, G^z)$ , with  $G^x \approx G^y \approx 2.007 \equiv G^{\perp}$  and  $G^z \approx 2.002 \equiv G^{\parallel}$ . The quadrupolar tensor for <sup>14</sup>N is  $\mathbb{Q}_i^{(P)} = \operatorname{diag}(Q^x, Q^y, Q^z)$ , with  $Q^x \approx 2\pi \times 1.26$  MHz,  $Q^y \approx 2\pi \times 0.53$  MHz, and  $Q^z \approx 2\pi \times -1.79$  MHz. The quadrupolar tensor vanishes for <sup>15</sup>N. Finally, the tensor that mediates electron-nucleus interactions in each nitroxide is  $\mathbb{A}_i^{(P)} = \operatorname{diag}(A^x, A^y, A^z)$ , where  $A^x \approx A^y \approx 2\pi \times 14.7$  MHz  $\equiv A^{\perp}$  and  $A^z \approx 2\pi \times 101.4$  MHz  $\equiv A^{\parallel}$  for <sup>14</sup>N, while  $A^x \approx A^y \approx 2\pi \times 27$  MHz  $\equiv A^{\perp}$  and  $A^z \approx 2\pi \times 141$  MHz  $\equiv A^{\parallel}$  for <sup>15</sup>N<sup>40</sup>.

We obtain approximate nitroxide transition frequencies via a perturbative treatment of the nuclear degrees of freedom in Eq. (2), see Supplementary Note 2. For a nitroxide hosting <sup>14</sup>N, we find

$$H_{n_i} \approx \left[ E_1^i \big| \widetilde{1} \big\rangle \big\langle \widetilde{1} \big|_i + E_0^i \big| \widetilde{0} \big\rangle \big\langle \widetilde{0} \big|_i + E_{-1}^i \big| - \widetilde{1} \big\rangle \big\langle - \widetilde{1} \big|_i \right] J_i^z.$$
(3)

The three *energy-transition branches* exhibit transition energies given by  $E_{1,-1}^{i} = \mu_{B}B^{z}G(\theta_{i}) \pm \frac{1}{\sqrt{2}}\sqrt{\left[\left(A^{\parallel}\right)^{2} - \left(A^{\perp}\right)^{2}\right]\cos(2\theta_{i}) + \left(A^{\perp}\right)^{2} + \left(A^{\parallel}\right)^{2}}}$ and  $E_{0}^{i} = \mu_{B}B^{z}G(\theta_{i}) + \frac{1}{2\mu_{B}B^{z}G(\theta_{i})}\frac{2\left(A^{\perp}A^{\parallel}\right)^{2} + \left[\left(A^{\perp}\right)^{4} - \left(A^{\perp}A^{\parallel}\right)^{2}\right]\sin^{2}(\theta_{i})}{\left(A^{\perp}\right)^{2}\sin^{2}(\theta_{i}) + \left(A^{\parallel}\right)^{2}\cos^{2}(\theta_{i})}}$ . Here,  $\theta_{i}$ is the azimuth angle between the applied magnetic field and the principal *z* axis of the nitroxide [see Fig. 1(b)],  $G(\theta_{i}) = \frac{1}{2}[\left(G^{\parallel} - G^{\perp}\right)\cos(2\theta_{i}) + G^{\perp} + G^{\parallel}]$ , and  $\left|\tilde{1}\rangle_{i}$ ,  $\left|\tilde{0}\rangle_{i}$ , and  $\left|-\tilde{1}\rangle_{i}$  are the states of the *i*th nitroxide nucleus dressed by the hyperfine interaction with the nitroxide electron, see Supplementary Note 2. Similarly, for a nitroxide hosting <sup>15</sup>N, we find

$$H_{n_i} \approx \left[ E_{1/2}^i | \widetilde{1/2} \rangle \langle \widetilde{1/2} |_i + E_{-1/2}^i | - \widetilde{1/2} \rangle \langle -\widetilde{1/2} |_i \right] J_i^z.$$
(4)

There are now two energy-transition branches with transition energies given by  $E_{1/2,-1/2}^{i} = \mu_{\rm B}B^{z}G(\theta_{i}) \pm \frac{1}{2\sqrt{2}}\sqrt{(A^{\perp})^{2} + (A^{\parallel})^{2} + [(A^{\parallel})^{2} - (A^{\perp})^{2}]\cos(2\theta_{i})}$ . Figure 2a compares the energy-transition branches in Eqs. (3) and (4) (dashed lines) with numerical diagonalization of Eq. (2) (solid lines). Note that since we will target the  $E_{0}^{i}$  branch for label detection, we have further developed its functional form to include second-order corrections in the electron-nitrogen coupling, see Supplementary Note 2. Figure 2(a) focuses on a single nitroxide (the index i = 1 is removed for clarity) and shows that our expression for  $E_{0}$  is in excellent agreement with numerical diagonalization.

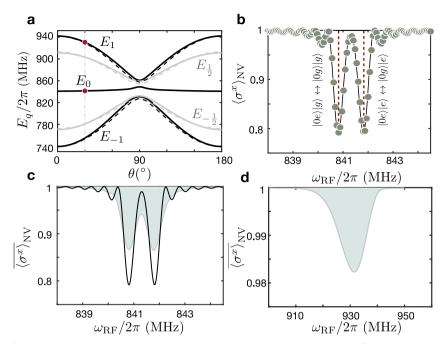
The inter-label coupling leads to an additional splitting  $\propto g_{12}$  of the energy-transition branches in Eqs. (3), (4). This term typically simplifies as  $g_{12}[J_1^2J_2^2 - \frac{1}{4}(J_1^+J_2^- + J_1^-J_2^+)] \longrightarrow g_{12}J_1^2J_2^2$ , since labels with different orientations have different resonance frequencies, thus eliminating the flip-flop contribution. Note that while this simplification is frequently (but not always) valid when using two <sup>14</sup>N, it is guaranteed to be valid when using nitroxides hosting different nitrogen isotopes since  $E_0$  (for <sup>14</sup>N) differs from  $E_{1/2,-1/2}$  (for <sup>15</sup>N) for any value of the azimuth [see Fig. 2(a)]. For <sup>14</sup>N, the available resonances of, e.g., the first label are then determined by the modified nitroxide Hamiltonian  $H'_{n_1} = H_{n_1} + g_{12}J_1^2$ 

$$\begin{aligned} H'_{n_1} &= \left[ E_1^1 |\tilde{1}\rangle \langle \tilde{1}|_1 + E_0^1 |\tilde{0}\rangle \langle \tilde{0}|_1 + E_{-1}^1| - \tilde{1}\rangle \langle -\tilde{1}|_1 \right] J_1^z + g_{12} J_1^z J_2^z \\ &= \sum_{q=1,0,-1} \left[ \left( E_q^1 + \frac{g_{12}}{2} \right) |\tilde{q}e\rangle \langle \tilde{q}e| + \left( E_q^1 - \frac{g_{12}}{2} \right) |\tilde{q}g\rangle \langle \tilde{q}g| \right] J_1^z. \end{aligned}$$

$$(5)$$

That is, any energy-transition branch  $E_q^1$  (corresponding to the  $|\tilde{q}\rangle$  nuclear state of the first nitroxide) splits as

$$E_q^1 \longrightarrow \left( E_q^1 + \frac{g_{12}}{2}, E_q^1 - \frac{g_{12}}{2} \right)$$
(6)



**Fig. 2** Energy branches of the spin labels and associated spectra. a Energy-transition branches of an electron spin label at  $B^z = 30$  mT for <sup>14</sup>N (<sup>15</sup>N). Solid black (gray) lines show the transition energies as a function of the azimuth  $\theta$  obtained via numerical diagonalization of Eq. (2). Dashed black (gray) lines correspond to  $E_{1,0,-1}$  in Eq. (3)  $[E_{1/2,-1/2}$  in Eq. (4)]. The vertical line highlights the case  $\theta_i = 30^\circ$ , with red dots indicating the transition energies relevant in the following plots. **b** NV spectrum  $\langle \sigma^x \rangle_{NV}$  as a function of the RF frequency applied near  $E_0^1$ . The spectrum was obtained by unitary propagation of Eq. (1) with  $H_{n_i}$  given by Eq. (2) (black line) and by  $E_0^i J_i^z$  (gray circles). The first nitroxide has azimuth  $\theta_1 = 30^\circ$  while the second nitroxide has azimuth  $\theta_2 = 91.7^\circ$ . The splitting of the peaks is induced by the inter-label coupling  $g_{12} \approx 2\pi \times 1$  MHz. The positions of the resonances are indicated by the dashed red lines. In (**c**, **d**), shaded areas show the average NV spectra  $\langle \sigma^x \rangle_{NV}$  in the presence of tumbling of the first (near-resonant) nitroxide. Tumbling was mimicked by averaging over a random azimuth following a Gaussian distribution centered at the equilibrium value  $\theta_{1,eq} = 30^\circ$ . The orientation of the other nitroxide was kept fixed for simplicity. The standard deviation was chosen to be  $\sigma_{\theta} = 6.25^\circ$ . In (**c**), we observe that tumbling leads to a loss of contrast and to signal broadening near  $E_0^1$ . Nevertheless, the spectrum  $\langle \sigma^x \rangle_{NV}$  near  $E_1^1$ . Here, the energy splitting cannot be resolved due to nitroxide tumbling in (**b**) is superimposed (black line). **d** Average NV spectrum  $\langle \sigma^x \rangle_{NV}$  near  $E_1^1$ . Here, the energy splitting cannot be resolved due to nitroxide tumbling. Also note that tumbling severely reduces contrast. For that reason, the spectrum in the absence of tumbling has much higher contrast and is not shown.

depending on the electronic state  $|e\rangle$ ,  $|g\rangle$  of the second nitroxide. Supplementary Note 2 includes a complete energy diagram of an electron spin in a nitroxide.

# The protocol

Our DEER protocol simultaneously delivers MW and RF pulses to detect energy shifts in the NV spectrum due to inter-label coupling. The sequence contains two free-evolution stages of duration  $\tau_{\rm free}$ separated by a driving stage [see Fig. 1(c)]. The NV is prepared in the + 1 eigenstate of  $\sigma^x$  and the nitroxides are assumed to be in a fully thermalized state. During free evolution, the NV accumulates a phase due to the term  $\sigma^{z}(\mathbf{a}_{1} \cdot \mathbf{J}_{1} + \mathbf{a}_{2} \cdot \mathbf{J}_{2})$  in Eq. (1). This term can be approximated as  $\sigma^{z}(a_{1}^{z}J_{1}^{z} + a_{2}^{z}J_{2}^{z})$  in the rotating-wave approximation (RWA) since the  $J_{i}^{x,y}$  contributions are suppressed by the large electronic precession frequencies of the nitroxides. Indeed, the energies  $E_{1,0,-1}^{i}$  reach several hundreds of MHz even for moderate values of  $B^{z}$  (we use  $B^{z} = 30$  mT in our simulations). By contrast, the NV-label coupling takes values of hundreds of kHz for typical NVlabel distances of several nanometers. During MW/RF irradiation, the NV undergoes an overall flip  $\sigma^z \rightarrow -\sigma^z$  via 2N+1 contiguous  $\pi$ pulses arising from the same continuous MW drive. At the same time, a single weaker RF  $\pi$ -pulse on the labels induces  $J_i^z \rightarrow -J_i^z$  if it is near-resonant with an electronic transition (i.e., with  $E_{1,0,-1}^i$  for <sup>14</sup>N or with  $E_{1/2,-1/2}^i$  for <sup>15</sup>N). The simultaneous flipping of the NV and spin label ensures a constructive and coherent accumulation of the NV phase during free evolution. Meanwhile, the use of  $2N + 1\pi$ pulses on the NV ensures that the Rabi frequencies of the two drives are well separated, which effectively averages out spurious NV-label interactions during irradiation. Consequently, scanning the RF frequency near nitroxide transitions yields clean resonance peaks in the NV response. As long as  $\Omega_{\rm RF} < g_{12}$ , the resonance peaks are split by  $g_{12}$ . Figure 2(b) shows the NV spectrum (the expectation value of  $\sigma^x$ ) associated with the transitions  $|0g\rangle|g\rangle \leftrightarrow |0e\rangle|g\rangle$  and  $|0q\rangle|e\rangle \leftrightarrow |0e\rangle|e\rangle$ . The resonances are split by ~  $q_{12}$  as expected. The solid black line in Fig. 2(b) was obtained by numerically propagating Eq. (1) with  $H_{n_i}$  given by Eq. (2), while gray circles were obtained assuming the simplified  $H_{n_i}$  in Eq. (3). The two spectra are in excellent agreement. Simulations in Fig. 2(b) assume  $B^{z} = 30 \text{ mT}$ , which is in the range of values that ensure the stability of the  $E_0^i$ energy-transition branch discussed below, see Supplementary Note 2. Moreover, the parameters of the pulse sequence are  $\tau_{\rm free} = 1.3 \ \mu s$ ,  $\Omega_{RF} = 2\pi \times 250$  kHz, and  $\Omega_{MW} = 31 \times \Omega_{RF}$ , for a total sequence time of 4.6 µs. In addition, the labels are separated from the NV by the distances  $d_1 \approx 6.9$  nm and  $d_2 \approx 7.3$  nm, leading to  $\mathbf{a}_1 \approx$  $2\pi \times (128, -132, -223)$  kHz and  $\mathbf{a}_2 \approx 2\pi \times (-22, -16, -264)$ kHz, while the distance between labels is  $d_{12} \approx 3.297$  nm.

The expressions for the energy-transition branches in Eqs. (3) and (4) reveal a dependence on the azimuth angle, i.e.,  $E_q^i \equiv E_q^i(\theta_i)$ . Consequently, unavoidable protein motion during the irradiation stage leads to a distortion in the spectrum and to a difficult interpretation of the signal. However, it is important to note the distinct nature of the  $E_{0}^i$  branch, which shows a much weaker dependence on  $\theta_i$  than  $E_{1,-1}^i$ . This makes the  $E_0^i$  branch particularly well suited for robust detection of the energy splitting [see Fig. 2(c, d)]. To maximize the robustness, we chose the

magnetic field to be large enough to energetically suppress the effect of the anisotropic hyperfine interaction, but small enough that the anisotropy of the Landé tensor does not become significant, see Supplementary Note 2.

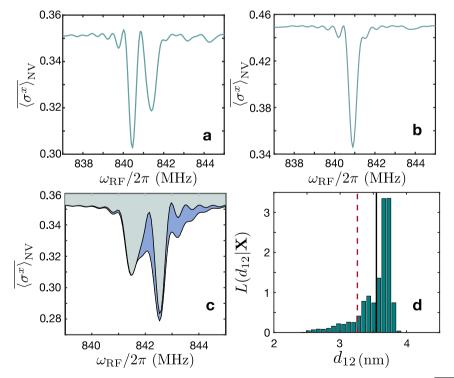
## Numerical simulations

We now illustrate our method in realistic ambient conditions including decoherence leading to NV dephasing with decoherence time  $T_2 = 20 \,\mu s$  (for a 4 nm NV depth<sup>36</sup>), electron-spin label relaxation with  $T_1 = 4 \,\mu s^{36}$ , and molecular tumbling. The dissipative model used in this section is described in the Supplementary Note 3 and captures the main decoherence mechanisms identified in ref. <sup>36</sup>. Note that for the specific protocol considered here, decoherence mainly manifests as a reduced line contrast and not as an increased linewidth. This is because we keep the sequence duration fixed and sweep  $\omega_{\rm RF}$  to find the resonances, as opposed to varying the sequence length at fixed  $\omega_{\rm RF}$  to observe an echo signal. The molecular tumbling is modelled as a random rigid rotation of both nitroxides around an axis parallel to the laboratory x axis, see Supplementary Note 3. Our simulations use a rotation angle  $\delta$  that is Gaussian-distributed with standard deviation  $\sigma_{\delta} = 6.25^{\circ}$ . This is somewhat smaller but comparable to the fluctuations observed in ref. <sup>36</sup>. Note that immobilizing proteins in more rigid matrices<sup>41,42</sup> or attaching them to the diamond surface<sup>35</sup> through multiple rigid covalent bonds could further reduce tumbling. The resulting tumbling-averaged spectra  $\langle \sigma^x \rangle_{NV}$  are shown in Fig. 3. In Fig. 3(a, b, c), the parameters  $\tau_{\text{free}}$  $\Omega_{MW}, \Omega_{RF}$ , and **a**<sub>i</sub> are the same as in Fig. 2. In Fig. 3(a), the labels are separated by  $d_{12} \approx 3.297$  nm and the inter-label coupling is  $q_{12} \approx 2\pi \times 1$  MHz. When the frequency of the RF field is set close to  $E_0^1$ , we clearly identify two resonance peaks in spite of tumbling. We have verified that the two resonances are still visible even if we choose  $\sigma_{\delta}$  to be four times larger. Since  $g_{12} \propto d_{12}^{-3}$ , a change in the relative position of the labels significantly modifies the observed spectrum. This is shown in Fig. 3(b) where the second label was displaced such that  $d_{12} \approx 4.03$  nm, leading to the disappearance of the splitting. This change in the spectrum certifies a change in the relative position between labels and, by extension, a conformational change in the host molecule.

So far, all simulations were performed for nitroxides hosting <sup>14</sup>N and having distinct transition energies. If both nitroxides have similar transition energies (e.g., due to similar azimuths), the spectra of the two nitroxides overlap and the inter-label flip-flop terms cannot be neglected. This complicates the interpretation of the spectrum. As shown in Fig. 3(c), this problem is avoided by using distinct <sup>14</sup>N and <sup>15</sup>N isotopes in each nitroxide. In this case, the  $E_0$  branch of the <sup>14</sup>N nitroxide never overlaps with the branches of the <sup>15</sup>N nitroxide and flip-flop terms can be safely neglected. As a result, the interpretation of the spectrum is much simpler. We emphasize that the purpose of using such orthogonal labels is not merely to resolve their distinct spectral signatures<sup>43</sup>: rather, it is to simplify the form of the inter-label interaction itself for all label orientations.

#### **Bayesian inference**

Finally, we show how to infer the inter-label distance  $d_{12}$  under realistic ambient conditions. We first simulate the experimental acquisition of  $\langle \sigma^x \rangle_{\text{NV}}$ . The simulated dataset has the form  $\mathbf{X} = \{X_1, ..., X_M\}$ , where  $X_j$  is an experimental estimate of the probability of measuring  $\sigma^x = +1$  after  $N_m$  measurements at RF frequency  $\omega_{\text{RF},j}$ . Second, we use a simplified model to efficiently extract information from  $\mathbf{X}$ . The model captures the relevant features of the tumbling-averaged spectrum  $\overline{\mathcal{S}}(\omega_{\text{RF}})$ . More precisely, we derive the approximate expression  $\mathcal{S}(\omega_{\text{RF}}, \theta, \beta) = \mathcal{S}_0 - \sum_{s=+,-} C_s \left[ \frac{\Omega_{\text{RF}}}{\Omega_{\alpha}(\omega_{\text{RF}}, \theta, \beta)} \right]^2 \sin^2 \left[ \frac{\pi}{2} \frac{\Omega_s(\omega_{\text{RF}}, \theta, \beta)}{\Omega_{\text{RF}}} \right]$  for a specific nitroxide



**Fig. 3** Simulated spectra and Bayesian inference of the inter-label distance. a Simulated average NV spectrum  $\overline{\langle \sigma^x \rangle}_{NV}$ . Two resonance peaks due to  $g_{12}$  are observed. **b** Similar to (**a**) but with lower  $g_{12}$ . **c** Simulated average spectrum  $\overline{\langle \sigma^x \rangle}_{NV}$  for overlapping label resonances with two <sup>14</sup>N isotopes (blue shaded area) and with distinct <sup>14</sup>N and <sup>15</sup>N isotopes (gray shaded area). **d** Marginal distribution  $L(d_{12}|\mathbf{X})$  of the inter-label distance for simulated data acquisition using the spectrum shown in (**a**). The true value of the distance and its posterior expectation, 3.297 nm and 3.54(25) nm, are indicated with vertical lines. The error is the standard deviation of the marginal posterior.

azimuth  $\theta$  and a specific angle  $\beta$  between the magnetic field and the line joining the nitroxides. Here,  $\Omega^2_+(\omega_{RF},\theta,\beta) = \Omega^2_{RF} +$  $[\omega_{\text{RF}} - E_0(\theta) \pm g_{12}(\beta)/2]^2$  and  $\mathcal{S}_0$  and  $\mathcal{C}_{\pm}$  are parameters that adjust the baseline and contrast, respectively (see Supplementary Note 3). Moreover,  $E_0(\theta)$  is the <sup>14</sup>N energy-transition branch, while  $g_{12}(\beta) \propto d_{12}^{-3}(1 - 3\cos^2\beta)$  is the inter-label coupling. Both  $\theta$  and  $\beta$ depend on the tumbling angle  $\delta$ . Averaging  $S[\omega_{\text{RF}}, \theta(\delta), \beta(\delta)]$  over a Gaussian distribution for  $\delta$  gives the tumbling-averaged spectrum  $\overline{\mathcal{S}}(\omega_{\text{RF}})$ . Assuming that the baseline  $\mathcal{S}_0$  is known, our model contains eight free parameters denoted by V. These include  $d_{12}$  and the unknown standard deviation  $\sigma_{\delta}$  of the tumbling. It must be emphasized that the light tumbling dependence of the  $E_0$ branch enables the determination of the inter-label distance. Indeed, a simple measurement of the line splitting  $g_{12} \propto d_{12}^{-3}(1 3\cos^2\beta$  cannot give independent access to the distance  $d_{12}$  and the angle  $\beta$ . This is because  $\beta$  is unknown a priori. However, light tumbling fluctuations allow the NV to probe different geometric configurations of the nitroxides. This results in a small distortion of the line shape that yields information beyond that contained in the splitting  $q_{12}$ . This in turn enables the independent extraction of the distance  $d_{12}$  and of the angle  $\beta$ . With the help of our model, we can therefore obtain the posterior probability of the parameters **V** using Markov Chain Monte Carlo sampling<sup>44</sup>. Assuming a uniform the posterior probability for V prior for V,  $\mathcal{L}(\mathbf{V}|\mathbf{X}) = \prod_{j=1,M} e^{-[X_j - (\overline{\mathcal{S}}(\omega_{RF,j}) + 1)/2]^2/2\sigma_m^2} / \sqrt{2\pi\sigma_m^2}.$  Here, the noise variance  $\sigma_m^2 \approx \left(1 - \overline{\langle \sigma^x \rangle_{NV}^2}\right) / 4N_m$  is assumed to be approximately constant and known. In Fig. 3(d) we show the resulting marginal  $L(d_{12}|\mathbf{X})$  of  $d_{12}$  for a dataset **X** simulated from the spectrum shown in Fig. 3(a). Here, **X** was obtained by taking  $N_m = 2 \times 10^4$  ideal measurements for each of M = 25 frequencies ranging from  $\omega_{\rm RF}$ /  $2\pi = 839-843$  MHz (we estimate that the same accuracy is achieved with  $N_m \approx 5 \times 10^5$  for imperfect NV detection efficiency<sup>45</sup>). The expectation of the marginal posterior is  $d_{12} = 3.54(25)$  nm, close to the real value 3.297 nm [see Fig. 3(d)].

## DISCUSSION

Our results open many interesting avenues for future investigation. In particular, it would be of great interest to extend our scheme to molecules with more than two attached spin labels. This would yield a stronger NV response, leading to a more detailed observation of conformational changes as well as to an enhancement in the range of inter-label distances to be estimated. The scheme could also be improved through the use of multiple NVs to better triangulate the label positions. In addition, our analytical understanding of the NV response could enable fast Bayesian inference of molecular dynamical properties. Our results pave the way for the observation of conformational dynamics of individual proteins using tools from magnetic resonance.

#### DATA AVAILABILITY

All the data relevant to this work are available upon request to the authors.

# CODE AVAILABILITY

All the codes employed are available upon request to the authors.

Received: 13 September 2022; Accepted: 15 November 2022; Published online: 30 November 2022

#### REFERENCES

 Stetz, M. A. et al. Characterization of internal protein dynamics and conformational entropy by nmr relaxation. *Meth. Enzymol.* 615, 237–284 (2019).

- 2. Rhodes, G. Crystallography Made Crystal Clear (Elsevier, 2006).
- Duddeck, H., Dietrich, W. & Tóth, G. Structure Elucidation by Modern NMR. (Steinkopff, Heidelberg, 1998).
- Alonso, J. L. & López, J. C. Microwave spectroscopy of biomolecular building blocks. In *Top. Curr. Chem.*, 335–401 (Springer, Cham, 2014).
- 5. Abragam, A. *The Principles of Nuclear Magnetism*. (Oxford University Press, London, 1961).
- Levitt, M. H. Spin Dynamics: Basics of Nuclear Magnetic Resonance (Wiley, West Sussex, 2008), 2 edn.
- Prisner, T., Rohrer, M. & MacMillan, F. Pulsed EPR spectroscopy: biological applications. Annu. Rev. Phys. Chem. 52, 279–313 (2001).
- Schweiger, A. & Jeschke, G. Principles of pulsed electron paramagnetic resonance. (Oxford University Press, New York, 2001).
- Borbat, P. P., Costa-Filho, A. J., Earle, K. A., Moscicki, J. K. & Freed, J. H. Electron spin resonance in studies of membranes and proteins. *Science* 291, 266–269 (2001).
- Jeschke, G. DEER distance measurements on proteins. Annu. Rev. Phys. Chem. 63, 419–446 (2012).
- Doherty, M. W. et al. The nitrogen-vacancy colour centre in diamond. *Phys. Rep.* 528, 1–45 (2013).
- 12. Wu, Y., Jelezko, F., Plenio, M. B. & Weil, T. Diamond quantum devices in biology. Angew. Chem. Int. Ed. 55, 6586–6598 (2016).
- Manson, N. B., Harrison, J. P. & Sellars, M. J. Nitrogen-vacancy center in diamond: Model of the electronic structure and associated dynamics. *Phys. Rev. B* 74, 104303 (2006).
- Steiner, M., Neumann, P., Beck, J., Jelezko, F. & Wrachtrup, J. Universal enhancement of the optical readout fidelity of single electron spins at nitrogenvacancy centers in diamond. *Phys. Rev. B* 81, 035205 (2010).
- Waldherr, G. et al. Dark states of single nitrogen-vacancy centers in diamond unraveled by single shot NMR. *Phys. Rev. Lett.* **106**, 157601 (2011).
- Cai, J., Jelezko, F., Plenio, M. B. & Retzker, A. Diamond-based single-molecule magnetic resonance spectroscopy. *New J. Phys.* 15, 013020 (2013).
- Cai, J. et al. Robust dynamical decoupling with concatenated continuous driving. New J. Phys. 14, 113023 (2012).
- London, P. et al. Detecting and polarizing nuclear spins with double resonance on a single electron spin. *Phys. Rev. Lett.* **111**, 067601 (2013).
- 19. Wang, Z.-Y. et al. Randomization of pulse phases for unambiguous and robust quantum sensing. *Phys. Rev. Lett.* **122**, 200403 (2019).
- Munuera-Javaloy, C., Puebla, R. & Casanova, J. Dynamical decoupling methods in nanoscale NMR. *Europhys. Lett.* **134**, 30001 (2021).
- 21. Grotz, B. et al. Sensing external spins with nitrogen-vacancy diamond. *New J. Phys.* **13**, 055004 (2011).
- Müller, C. et al. Nuclear magnetic resonance spectroscopy with single spin sensitivity. *Nat. Commun.* 5, 4703 (2014).
- 23. Sushkov, A. O. et al. Magnetic resonance detection of individual proton spins using quantum reporters. *Phys. Rev. Lett.* **113**, 197601 (2014).
- Schmitt, S. et al. Submillihertz magnetic spectroscopy performed with a nanoscale quantum sensor. *Science* 356, 832–837 (2017).
- Glenn, D. R. et al. High-resolution magnetic resonance spectroscopy using a solidstate spin sensor. *Nature* 555, 351–354 (2018).
- Bucher, D. B., Glenn, D. R., Park, H., Lukin, M. D. & Walsworth, R. L. Hyperpolarization-enhanced NMR spectroscopy with femtomole sensitivity using quantum defects in diamond. *Phys. Rev. X* 10, 021053 (2020).
- 27. Arunkumar, N. et al. Micron-scale NV-NMR spectroscopy with signal amplification by reversible exchange. *PRX Quant.* **2**, 010305 (2021).
- Chu, Y. et al. Precise spectroscopy of high-frequency oscillating fields with a single-qubit sensor. *Phys. Rev. Appl.* 15, 014031 (2021).
- 29. Meinel, J. et al. Heterodyne sensing of microwaves with a quantum sensor. *Nat. Commun.* **12**, 2737 (2021).
- Staudenmaier, N., Schmitt, S., McGuinness, L. P. & Jelezko, F. Phase sensitive quantum spectroscopy with high frequency resolution. *Phys. Rev. A* **104**, L020602 (2021).
- Ermakova, A. et al. Detection of a few metallo-protein molecules using color centers in nanodiamonds. *Nano Lett.* 13, 3305–3309 (2013).
- Ziem, F. C., Götz, N. S., Zappe, A., Steinert, S. & Wrachtrup, J. Highly sensitive detection of physiological spins in a microfluidic device. *Nano Lett.* 13, 4093–4098 (2013).
- Rendler, T. et al. Optical imaging of localized chemical events using programmable diamond quantum nanosensors. *Nat. Commun.* 8, 14701 (2017).
- Barton, J. et al. Nanoscale dynamic readout of a chemical redox process using radicals coupled with nitrogen-vacancy centers in nanodiamonds. ACS Nano 14, 12938–12950 (2020).
- Lovchinsky, I. et al. Nuclear magnetic resonance detection and spectroscopy of single proteins using quantum logic. *Science* 351, 836–841 (2016).
- Shi, F. et al. Single-protein spin resonance spectroscopy under ambient conditions. *Science* 347, 1135–1138 (2015).

- Haugland, M. M., Anderson, E. A. & Lovett, J. E. Tuning the properties of nitroxide spin labels for use in electron paramagnetic resonance spectroscopy through chemical modification of the nitroxide framework. In *Electron Paramagnetic Resonance*, vol. 25 of SPR - *Electron Paramagnetic Resonance*, 1–34 (Royal Society of Chemistry, 2016).
- 38. Schlipf, L. et al. A molecular quantum spin network controlled by a single qubit. *Sci. Adv.* **3**, 1701116 (2017).
- Marsh, D. Bonding in nitroxide spin labels from <sup>14</sup>n electric-quadrupole interactions. J. Phys. Chem. A **119**, 919–921 (2015).
- Marsh, D. Spin-Label Electron Paramagnetic Resonance Spectroscopy. (CRC Press, Boca Raton FL, 2019).
- Meyer, V. et al. Room-temperature distance measurements of immobilized spinlabeled protein by DEER/PELDOR. *Biophys. J.* 108, 1213–1219 (2015).
- Lira, R. B., Steinkühler, J., Knorr, R. L., Dimova, R. & Riske, K. A. Posing for a picture: vesicle immobilization in agarose gel. *Sci. Rep.* 6, 25254 (2016).
- Galazzo, L., Teucher, M. & Bordignon, E. Orthogonal spin labeling and pulsed dipolar spectroscopy for protein studies. In *Meth. Enzymol.*, 79–119 (Elsevier, 2022).
- 44. Gilks, W. R., Richardson, S. & Spiegelhalter, D. J. Markov Chain Monte Carlo in *Practice*. (Chapman & Hall/CRC, New York, 1996).
- 45. Wan, N. H. et al. Efficient extraction of light from a nitrogen-vacancy center in a diamond parabolic reflector. *Nano Lett.* **18**, 2787–2793 (2018).

## ACKNOWLEDGEMENTS

The authors acknowledge financial support from Spanish Government via PGC2018-095113-B-I00 (MCIU/AEI/FEDER, UE) and, from Basque Government via IT986-16. C.M.-J. acknowledges the predoctoral MICINN grant PRE2019-088519. R.P. acknowledges support from European Union's Horizon 2020 FET-Open project SuperQuLAN (899354). M.B.P. and B.D. acknowledge the ERC Synergy Grants HyperQ (856432), as well as the BMBF project QSens (03ZU1110FF), and Asteriqs (820394). The authors acknowledge support by the state of Baden-Württemberg through bwHPC and the German Research Foundation (DFG) through grant no INST 40/575-1 FUGG (JUSTUS 2 cluster). J.C. acknowledges the Ramón y Cajal (RYC2018-025197-I) research fellowship, the financial support from Spanish Government via EUR2020-112117 and Nanoscale NMR and complex systems (PID2021-126694NB-C21) projects, the EU FET Open Grant Quromorphic (828826), the ELKARTEK project Dispositivos en Tecnologias Cuánticas (KK-2022/00062), and the Basque Government grant IT1470-22.

## AUTHOR CONTRIBUTIONS

All authors contributed to the main ideas and to the paper writing; C.M.-J. developed the analytical study; C.M.-J., R.P., and B.D. produced the numerical simulations; M.B.P. and J.C. led the work jointly.

## **COMPETING INTERESTS**

The authors declare no competing interests.

## ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41534-022-00653-w.

**Correspondence** and requests for materials should be addressed to C. Munuera-Javaloy.

Reprints and permission information is available at http://www.nature.com/ reprints

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http:// creativecommons.org/licenses/by/4.0/.

© The Author(s) 2022