ARTICLE OPEN Check for updates Ethnicity- and sex-specific genome wide association study on Parkinson's disease

Kye Won Park^{1,2}, Ho-Sung Ryu³, Eunsoon Shin⁴, YoonGi Park⁴, Sang Ryong Jeon⁵, Seong Yoon Kim⁶, Jae Seung Kim⁷, Seong-Beom Koh⁸ and Sun Ju Chung ^{1⊠}

Most previous genome-wide association studies (GWASs) on Parkinson's disease (PD) focus on the European population. There are several sex-specific clinical differences in PD, but little is known about its genetic background. We aimed to perform an ethnicity-specific and sex-specific GWAS on PD in the Korean population. A total of 1050 PD patients and 5000 controls were included. For primary analysis, we performed a GWAS using a logistic additive model adjusted for age and sex. The same statistical models were applied to sex-specific analyses. Genotyping was performed using a customized microarray chip optimized for the Korean population. Nine single nucleotide polymorphisms (SNPs) including four in the *SNCA* locus and three from the *PARK16* locus were associated with PD in Koreans. The rs34778348 in the *LRRK2* locus showed a strong association, though failed to pass cluster quality control. There were no notable genome-wide significant markers near the *MAPT* or *GBA1* loci. In the female-only analysis, rs34778348 in *LRRK2* and the four other SNPs in the *SNCA* showed a strong association with PD. In the male-only analysis, no SNP surpassed the genome-wide significance threshold under Bonferroni correction; however, the most significant signal was rs708726 in the *PARK16* locus. This ethnicity- and sex-specific GWAS on PD implicate the pan-ethnic effect of *SNCA*, the universal but East-Asian inclined effect of *PARK16*, the East Asian-specific role of *LRRK2* G2385R variants, and the possible disproportionate effect of *SNCA* and *PARK16* between sexes for PD susceptibility. These findings suggest the different genetic contributions to sporadic PD in terms of ethnicity and sex.

npj Parkinson's Disease (2023)9:141; https://doi.org/10.1038/s41531-023-00580-3

INTRODUCTION

Parkinson's disease (PD) is one of the most common neurodegenerative disorders, clinically characterized by resting tremor, bradykinesia, rigidity, and postural instability¹. Its neuropathologic hallmark is a progressive loss of dopaminergic neurons in the substantia nigra caused by pathologic accumulation of α synuclein, resulting in the formation of Lewy bodies and Lewy neurites².

For the past two decades, genome-wide association studies (GWASs) have shed light on the genetic background of various common diseases including PD³. The largest up-to-date metaanalysis of GWASs on PD identified 90 genome-wide significant risk signals across 78 genomic regions which collectively account for 16–36% of the heritable risk of sporadic PD⁴. These PD-related genes were found to be involved in common biological pathways, where several critical cellular routes including mitochondrial dysfunction and lysosomal membrane trafficking pathways lead to pathologic α -synuclein accumulation².

The inveterate problem in the current field of GWASs is the disproportionate focus on European populations⁵. A recent study found that almost 90% of the participants in the National Human Genome Research Institute GWAS Catalog were of European Ancestry⁶. Genetic variants show a high ethnicity-specific heterogeneity in their distribution and functional activity⁷. Thus, the results of previous GWASs targeting European populations cannot readily be generalized to populations with different ethnic or racial backgrounds. Most precedent GWASs on PD have also

focused on European descents^{4,8}, raising the necessity for the diversity of the target population⁹. Koreans have a distinct genetic makeup in the peninsula owing to their unique geographical and cultural background. Moreover, the prevalence of PD in Korea is expeditiously rising, as it is the world's most rapidly aging society¹⁰. Despite this, there is no GWAS data on the Korean PD population.

There are differences in the clinical characteristics of PD according to sex¹¹. For example, the prevalence of PD, age at onset, and the susceptibility to progression to dementia differ between males and females. However, little attention has been paid to the genetic differences between male and female patients with PD. A recent sex-specific GWAS on PD conducted in a European population showed no sex-specific differences¹².

In this context, we aimed to identify the genetic variants associated with PD focusing on a genetic isolate, Koreans, by applying a microarray chip that is optimized for Koreans (Korean Chip) and determine the genomic risk variants for PD in a sexspecific manner.

RESULTS

Demographics

A total of 1070 patients with PD and 5000 age- and sex-matched healthy controls were initially recruited in the study. Of them, a sample of 20 patients with PD was excluded due to low sample quality. The mean age at sample collection of the patient group





¹Department of Neurology, Asan Medical Center, University of Ulsan College of Medicine, Seoul, Korea. ²Pacific Parkinson's Research Centre, Djavad Mowafaghian Centre for Brain Health, University of British Columbia, Vancouver, Canada. ³Department of Neurology, Kyungpook National University Hospital, Daegu, South Korea. ⁴DNA Link Inc, Seoul, Korea. ⁵Department of Neurosurgery, Asan Medical Center, University of Ulsan College of Medicine, Seoul, Korea. ⁶Department of Psychiatry, Asan Medical Center, University of Ulsan College of Medicine, Seoul, Korea. ⁷Department of Nuclear Medicine, Asan Medical Center, University of Ulsan College of Medicine, Seoul, Korea. ⁸Department of Neurology, Korea University Guro Hospital, Seoul, Korea. ⁸Department of Neurology, Kr

Characteristics	Patients (<i>n</i> = 1050)	Controls (<i>n</i> = 5000)	P-value	
Female sex	554 (53)	2610 (52)	0.740	
Age at sample collection, years	64.0 ± 9.7	64.0 ± 10.0	1.000	
Age at onset of Parkinsonism symptoms, years	58.7 ± 10.2	-		
Disease duration at sample collection, years	5.3 ± 4.4	_		
Education duration, years	8.6 ± 6.0	_		
MMSE	26.0 ± 3.5	_		
Disease duration from PD onset to MMSE, years	5.2 ± 4.1	_		

was 64.0 years, ranging from 31 to 89 years. The mean disease duration at study enrollment was 5.3 years. Among them, 554 were female patients. The baseline demographics of the participants including their mini-mental status examination are depicted in Table 1. Power calculation of the sample showed 80% power to detect variants exerting a risk for PD with odds ratio (OR) as low as 1.25 and minor allele frequency (MAF) of 10% (Supplementary Table 1).

Primary analysis

In the primary analysis between 1050 PD patients and 5000 healthy controls, 492,970 single nucleotide polymorphisms (SNPs) passed the marker quality control (OC). The Quantile-Quantile (O-Q) plot and Manhattan plot of the analysis are shown in Supplementary Fig. 1 and Fig. 1A, respectively. Nine SNPs surpassed the Bonferroni-corrected genome-wide significance, the threshold being 1.01×10^{-7} (Table 2). The most strongly associated SNP was rs3796661 ($P = 3.79 \times 10^{-13}$) in the SNCA. Three additional SNPs (rs356203, rs11931074, and rs12640100) in the SNCA locus showed significant association with PD. Two SNPs in the SLC41A1 (rs708726 and rs947211) and RAB29 (rs708723), which are all located within the PARK16 locus, were also genomewide significant. The regional association plots of the SNCA locus (index SNP rs3796661) and PARK16 locus (index SNP rs708725) showed multiple SNPs within the loci in linkage disequilibrium (LD) with the index SNPs (Fig. 2). There were no further SNPs with statistical significance in LD with rs34779348 and rs2451713. Notably, the rs34778348, an exonal missense variant (G2385R) of the LRRK2 gene, showed a strong association ($P = 4.77 \times 10^{-13}$). However, the SNP failed to pass the cluster QC that was manually performed after the marker QC steps (Fig. 3). Moreover, when we examined all other markers with P < 0.05 in the primary analysis, we could not observe genome-wide significance in the markers near the MAPT or GBA1, except for the only SNP in the MAPT locus (rs374460, $P = 2.32 \times 10^{-3}$).

Sex-specific analysis

In the female-only analysis, 554 female PD patients and 2610 female controls were analyzed. Of the 486,510 SNPs which passed marker QC in the female-only analysis, five SNPs surpassed genome-wide significance threshold under Bonferroni correction ($P < 1.03 \times 10^{-7}$ (0.05/486,510)) (Table 3, Fig. 1B). The most significant SNP was the rs34778348 in *LRRK2* locus ($P = 1.25 \times 10^{-9}$). The other four significant SNPs were in the *SNCA*, the most significant being rs3796661 ($P = 4.89 \times 10^{-9}$). None of the variants in the *PARK16* locus, including those of the *SLC41A1* and *RAB29* genes, had significance under $P < 1.0 \times 10^{-5}$ in the female-only analysis.

In the male-only analysis, 496 male PD patients and 2390 male controls were included. A total of 488,631 SNPs passed the marker

QC. None of the SNPs surpassed the genome-wide significance threshold under Bonferroni correction ($P < 1.02 \times 10^{-7}$ (0.05/488,631); Table 3, Fig. 1C). However, when the top signals' *P*-value under 1.0×10^{-6} were reviewed, the most significant signal was the rs708726 in the *SLC41A1* ($P = 8.23 \times 10^{-6}$), with four others in the *PARK16* locus with $P < 1.0 \times 10^{-6}$. Meanwhile, SNPs within the *SNCA* locus did not show associations with *P*-value < 10^{-4} except for rs3796661 ($P = 5.25 \times 10^{-5}$), indicating its small effect on male patients compared to female patients. The demographics, power calculation, Q-Q plots, and regional association plots of these sex-specific analyses can be found in the supplementary materials (Supplementary Tables 2, 3, Supplementary Figs. 2, 3). The least OR to satisfy the statistical power of 80% at MAF of 10% in the female and male subgroups was 1.34 and 1.36, respectively.

DISCUSSION

In this ethnicity-specific GWAS on PD, variants in the *SNCA* and *PARK16* loci showed the strongest association with PD in the Korean population. We further found that the *LRRK2* G2385R variant was associated with Korean PD, although the variant did not pass cluster QC. Variants in *MAPT* or *GBA1*, the two genes commonly associated with PD in GWASs from Western countries¹³, were not replicated in our GWAS. There were disproportionate effects of *SNCA* and *PARK16* variants on Korean PD according to sex. Although we did not identify any novel loci specific to Korean ethnicity for PD susceptibility, our results suggest that there is a gradient in genetic contribution according to ethnicity and sex to the risk of PD.

Our dominant SNPs being in the SNCA locus demonstrates the universal strong effect of the SNCA variants on the risk of PD across ethnicities. Abnormal accumulation of the α-synuclein protein, which normally regulates synaptic vesicle trafficking and the subsequent neurotransmitter release in neurons, is the pathological hallmark of PD^{14,15}. The SNCA gene, which encodes the α -synuclein protein, was identified as the risk loci from the first large-scale GWAS on PD¹⁶. The following GWASs on PD targeting various populations and subsequent meta-analyses consistently reported strong effects of the loci variants on the risk of PD, regardless of the target population^{4,17,18}. In contrast, the effect of the second leading loci in our study, PARK16, including the SNP rs708726, has been particularly highlighted in GWASs on PD targeting East Asians. The locus spans across five genes, including SLC45A3, NUCKS1, RAB29/RAB7L1, SLC41A1, and PM20D1¹⁹. Among these regions, RAB29 is the master regulator of the LRRK2 protein, controlling its activation, localization, and phosphorylation²⁰. The locus was designated with the name PARK16 after the discovery of its association with PD in a Japanese GWAS²¹. The largest GWAS on PD in the East Asian population thus far by Foo and colleagues, which was performed on more than 30,000 participants across six populations of East Asia, found that PARK16 is a dominant locus in



Fig. 1 Manhattan plots of the study. Primary analysis (a), female-only analysis (b), and male-only analysis (c). The red lines denote the Bonferroni threshold.

Table	Table 2. Genomic variants with genome-wide significance of the primary analysis.						
Chr	SNP	Gene	Region	Allele (minor/major)	OR (95% CI)	Minor allele frequency (patients/controls)	P-value
4	rs3796661	SNCA	Intron	C/T	0.69 (0.62–0.76)	0.37/0.46	3.79 × 10 ⁻¹³
12	rs34778348	LRRK2	Missense, exon	A/G	2.56 (1.99–3.31)	0.05/0.02	4.77×10^{-13}
4	rs356203	SNCA	Intron	T/C	0.71 (0.64–0.79)	0.39/0.46	2.32×10^{-11}
4	rs11931074	SNCA, GPRIN3	Intron, downstream, upstream	G/T	0.72 (0.65–0.79)	0.39/0.47	5.29×10^{-11}
4	rs12640100	SNCA, GPRIN3	Intron, downstream, upstream	G/A	0.72 (0.65–0.79)	0.39/0.47	5.45×10^{-11}
1	rs708726	SLC41A1	Intron	T/G	0.75 (0.68–0.83)	0.43/0.50	1.61×10^{-8}
1	rs947211	SLC41A1, RAB29	Downstream, upstream	A/G	0.75 (0.68–0.83)	0.43/0.50	2.50×10^{-8}
6	rs2451713	ZNF322, LOC101929855, GUSBP2	Upstream, downstream	C/G	1.88 (1.50–2.36)	0.05/0.04	5.39×10^{-8}
1	rs708723	RAB29	UTR-3	C/T	0.76 (0.69–0.84)	0.43/0.49	6.69×10^{-8}

а



Fig. 2 Regional association plots of the primary genome-wide association study. Plots around (a) rs3796661 and (b) rs708726.

East Asian PD, along with the *SNCA* and *LRRK2* loci¹⁸. In line with these previous studies, our study suggests that the effect of PARK16 variants on PD susceptibility is pan-ethnic but particularly stands out in East Asians.

In this study, the strong association of rs34778348 with PD in *LRRK2* was another notable finding. SNP rs34637584, known as the *LRRK2* G2019S variant, is a well-known variant that is strongly associated with PD risk in the Caucasian and Jewish populations²². In contrast to this, rs34778348, which is the *LRRK2* G2385R variant, is mainly found in Asian populations²³. This variant was found to be a genetic risk factor for sporadic PD in Chinese, Japanese, and Korean populations^{23–25}. A previous study on *LRRK2* G2395R in Korean PD included only a small number of participants²⁴, and our study provides a replication of their findings in a larger sample

size. However, a careful interpretation is warranted because this variant failed to pass the cluster QC and no other markers in LD with the variant were shown to be significant in our analysis. The kinase overactivity and downregulation of the *LRRK2* function with kinase inhibitors caused by the *LRRK2* G2019S variant has been suggested as a potential therapeutic target of PD^{26,27}. It has been proposed that *LRRK2* G2385R results in partial loss-of-function of the kinase activity in vitro²⁸, in contrast to its G2019S counterpart. The discrepancy in the *LRRK2* variants and subsequent protein dysfunction between Caucasian and Asian populations is of great importance, warranting different therapeutic approaches according to ethnicity.

We could not observe evidence for associations with the *GBA1* or *MAPT* loci, which are the two important genes highlighted in



Fig. 3 An example of cluster quality control. In general, the three genotypes denoted as blue, purple, and red dots, are clearly clustered (a). The markers were excluded if the three genotypes were not clearly separated as in (b).

Chr	SNP number	Gene	Region	Allele (minor/major)	OR (95% CI)	Minor allele frequency (Patients/Controls)	P-value
Fema	le-only analysis						
12	rs34778348	LRRK2	Missense, exon	A/G	3.53 (2.35–5.29)	0.05/0.02	1.25×10^{-9}
4	rs3796661	SNCA	Intron	C/T	0.63 (0.54–0.74)	0.37/0.46	4.89×10^{-9}
4	rs12640100	SNCA, GPRIN3	Intron, downstream, upstream	G/A	0.64 (0.55–0.75)	0.38/0.47	1.13 × 10 ⁻⁸
4	rs356203	SNCA	Intron	T/C	0.64 (0.55–0.75)	0.38/0.47	1.25×10^{-8}
4	rs11931074	SNCA, GPRIN3	Intron, downstream, upstream	G/T	0.65 (0.56–0.75)	0.38/0.47	2.11 × 10 ⁻⁸
Male	only analysis						
1	rs708726	SLC41A1	Intron	T/G	0.67 (0.57–0.79)	0.40/0.49	8.23 × 10 ⁻⁶
5	rs139422381	ANKRD33B, LOC101929412	Downstream, upstream	A/G	4.02 (2.25–7.16)	0.03/0.01	2.46×10^{-6}
1	rs947211	SLC41A1, RAB29	Downstream, upstream	A/G	0.69 (0.59–0.8)	0.41/0.50	3.24×10^{-6}
12	rs10746109	WSCD2, LOC728739	Upstream	A/G	1.46 (1.24–1.71)	0.51/0.44	3.93 × 10 ^{−6}
1	rs708723	RAB29	UTR-3	C/T	0.69 (0.59–0.81)	0.40/0.49	4.56×10^{-6}
1	rs1775145	SLC41A1, RAB29	Downstream, upstream	C/A	0.69 (0.59–0.81)	0.42/0.50	4.77×10^{-6}
1	rs12748961	NUCKS1, SLC45A3	Downstream, upstream	T/C	0.69 (0.59–0.81)	0.42/0.50	5.12×10^{-6}
7	rs1949132	GNAI1, LOC101927269	Intron, downstream	C/T	1.78 (1.39–2.29)	0.12/0.09	6.90 × 10 ⁻⁶

both sporadic and familial PD in Western countries²⁹. One possible explanation is the high homogeneity of *MAPT* in the East Asian population. In East Asia, *MAPT* is genetically homogenous with only the H1 haplotype in the population, whereas the European population has both H1 and H2 haplotypes³⁰. However, multiple variants exist even in the H1 haplotype, reflecting the greater diversity of *MAPT* than explained by the H1 and H2 clades alone³¹. Thus, the lack of association with *MAPT* or *GBA1* in this Korean-specific analysis may suggest the difference in the susceptibility to PD by the variants within these genes. The associations of the two loci were also not replicated by other Asian GWASs on PD, including those in Japanese²³ Han-Chinese²⁵ and in pan-East Asian GWASs on PD^{18,32}, supporting our findings.

Investigation regarding the genes associated with PD in a sexspecific manner has been limited. In our analysis, rs34778348 of the *LRRK2* locus and four SNPs of the *SNCA* locus showed genomewide significance in females, but the significance was not replicated in males. The most significant SNPs in males were those in the *PARK16* locus whereas they did not surpass the significance threshold under the Bonferroni correction. However, a recent investigation on autosomal genetic and sex-specific differences in PD found no significant genetic differences between male or female PD patients¹². PD is more prevalent in men worldwide but is more prevalent in women in Asian populations, including the Korean population¹¹. The discrepancy between the European and Korean sex-specific GWASs on PD may implicate such ethnicity-specific differences in the sex ratio of PD.

There are some limitations in our study. First, the analysis was conducted without principal component adjustment under the assumption of genetic homogeneity of the Korean population. Ample evidence supports that many of the Far East Asian population groups, especially the Korean, have their own distinct genetic cluster without population admixture^{33,34}. Although we presented the genetic homogeneity of our dataset in Supplementary Fig. 4, without adjusting the principal components in the analyses, potential stratification at the sub-populational level cannot be ruled out. Second, the number of total subjects was relatively small for a GWAS hence underpowering the results, especially when stratified by sex. Nevertheless, it is an inevitable limitation for a genetic study targeting a minor genomic cluster. Third, our study lacks functional validation of the discovered variants and a separate replication analysis. On the other hand,

there have been several well-designed meta-analyses of GWASs on PD with various methods of functional validations³⁵. Our study did not reveal any novel marker specific to Korean PD. Thus, in a way, our work itself could be interpreted as a Korean validation of the worldwide level meta-analyses. To identify the ethnic-sexgenetic interaction suggested in this study, further functional validation specific to ethnicity and sex should be encouraged.

This ethnicity- and sex-specific GWAS on PD in the Korean population suggests the pan-ethnic effect of *SNCA*, the standing out significance of *PARK16* in East Asians, the East Asian-specific role of the *LRRK2* G2385R variants, and the possible disproportionate effect of *SNCA* and *PARK16* between sexes for PD susceptibility. These findings implicate the gradient in genetic contribution to PD susceptibility across ethnicities and sex.

METHODS

Participants

We recruited patients with PD in Asan Medical Center, Seoul, South Korea from January 2011 to April 2016. A total of 1070 ethnically Korean patients who were diagnosed with sporadic PD by movement disorder specialists according to the United Kingdom Parkinson's Disease Brain Bank Criteria were enrolled³⁶. Baseline demographics including age at sample collection, age at the onset of PD, sex, and family history of Parkinsonism were collected. We defined the age at onset as the time when one of the motor cardinal symptoms (resting tremor, rigidity, bradykinesia, stooped posture, or postural instability) was noted by the patient or caregiver. Exclusion criteria were as follows: not being ethnically Korean, genetically confirmed hereditary Parkinsonism, and signs of atypical Parkinsonism (cerebellar signs, Parkinsonism not-responsive to levodopa, supranuclear gaze palsy, early severe autonomic dysfunction, early severe dementia with disturbances of memory, language, and praxis, and otherwise-unexplained pyramidal signs). For controls, we obtained the samples of 5000 age- and sex-matched healthy controls from the Korea Biobank Project. Informed consent was obtained from every participant as per the locally approved protocols. The study was approved by the Institutional Review Board of Asan Medical Center.

Genotyping and quality control

All patients underwent peripheral blood sampling for DNA extraction, and 200 ng of genomic DNA was genotyped for each patient. All samples were genotyped on a Korean Chip (K-CHIP) obtained from the K-CHIP Consortium, Center for Genome Science, Korea National Institute of Health³⁷. The K-CHIP is an SNP microarray chip developed to standardize the genotypic platform optimal for Koreans. All samples were assayed on Affymetrix Axiom[®] 2.0 Reagent Kit (Affymetrix, Santa Clara, CA, USA). Manual target preparation for the assay was processed according to the manufacturer's protocol. Low-quality samples and low-quality SNPs were excluded through the following QC steps. Samples with call rates lower than 97%, sex discrepancy, excessive heterozygosity, or cryptic relatedness were excluded. SNPs with minor allele frequency <1% in patients or controls, markers with a low call rate <95% in patients or controls, and SNPs with significant deviation from the Hardy-Weinberg equilibrium permutation test ($P < 10^{-4}$) were excluded. Lastly, all markers with $P < 10^{-4}$ were visually inspected by one of the investigators for cluster QC (Fig. 3). For each marker, the genotypes (AA, Aa, or aa) were colored red, purple, and blue, respectively. If the genotypes of a marker were not clearly clustered into the three colors, we considered it as genotyping error and the marker was excluded. All QC steps were performed using PLINK software version 1.90 (Free Software Foundation Inc., Boston, MA, USA)^{38,39}.

Statistical analysis

We performed primary analysis between PD patients and healthy controls by multiple logistic additive models with age and sex as the covariates. Age at onset of Parkinsonism for the PD patients and age at sample collection for the healthy controls were adjusted as the age covariate. Secondary analyses, the sex-specific GWASs, were performed in the male and female population separately using the same model and covariates. Principal component analysis (PCA) was performed to verify the populational homogeneity of our dataset (Supplementary Fig. 4). PLINK software version 1.90 (https://www.cog-genomics.org/plink2/) was used for the association analysis^{38,39}. Q–Q plots and Manhattan plots were contrived using the R software (version 3.5.2, R Core Team (2018). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/). Regional association plots were generated using the LocusZoom software (version 0.4.8, http:// locuszoom.sph.umich.edu/)⁴⁰. Power calculations were performed using Quanto software (version 1.2.4.)⁴¹. Regional association plots were generated and visually inspected in all SNPs with $P < 10^{-4}$. Bonferroni corrections were applied to correct multiple tests.

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this article.

DATA AVAILABILITY

The summary statistics of this GWAS are openly available in GWAS Catalog (https:// www.ebi.ac.uk/gwas/downloads/summary-statistics, study accession: GCST90278092).

CODE AVAILABILITY

The related codes and scripts for the study will be made available upon reasonable request to the corresponding author.

Received: 2 March 2023; Accepted: 13 September 2023; Published online: 07 October 2023

REFERENCES

- 1. Poewe, W. et al. Parkinson disease. Nat. Rev. Dis. Primers 3, 17013 (2017).
- Bloem, B. R., Okun, M. S. & Klein, C. Parkinson's disease. The Lancet 397, 2284–2303 (2021).
- 3. Tam, V. et al. Benefits and limitations of genome-wide association studies. *Nat. Rev. Genet.* 20, 467–484 (2019).
- Nalls, M. A. et al. Identification of novel risk loci, causal insights, and heritable risk for Parkinson's disease: a meta-analysis of genome-wide association studies. *Lancet Neurol.* 18, 1091–1102 (2019).
- Sirugo, G., Williams, S. M. & Tishkoff, S. A. The missing diversity in human genetic studies. *Cell* 177, 26–31 (2019).
- Mills, M. C. & Rahal, C. A scientometric review of genome-wide association studies. Commun. Biol. 2, 9 (2019).
- Popejoy, A. B. & Fullerton, S. M. Genomics is failing on diversity. *Nature* 538, 161–164 (2016).
- Day, J. O. & Mullin, S. The genetics of Parkinson's disease and implications for clinical practice. *Genes (Basel)* 12, 1006 (2021).
- Wojcik, G. L. et al. Genetic analyses of diverse populations improves discovery for complex traits. *Nature* 570, 514–518 (2019).
- Park, J.-H. et al. Trends in the incidence and prevalence of Parkinson's disease in Korea: a nationwide, population-based study. BMC Geriatr. 19, 320 (2019).
- Gillies, G. E., Pienaar, I. S., Vohra, S. & Qamhawi, Z. Sex differences in Parkinson's disease. Front. Neuroendocrinol. 35, 370–384 (2014).
- 12. Blauwendraat, C. et al. Investigation of autosomal genetic sex differences in Parkinson's disease. Ann. Neurol. 90, 35–42 (2021).
- Blauwendraat, C., Nalls, M. A. & Singleton, A. B. The genetic architecture of Parkinson's disease. *Lancet Neurol.* 19, 170–178 (2020).
- Wong, Y. C. & Krainc, D. α-synuclein toxicity in neurodegeneration: mechanism and therapeutic strategies. *Nat. Med.* 23, 1–13 (2017).

- Siddiqui, I. J., Pervaiz, N. & Abbasi, A. A. The Parkinson disease gene SNCA: evolutionary and structural insights with pathological implication. *Sci. Rep.* 6, 24475–24475 (2016).
- 16. Simon-Sanchez, J. et al. Genome-wide association study reveals genetic risk underlying Parkinson's disease. *Nat. Genet.* **41**, 1308–1312 (2009).
- Chang, D. et al. A meta-analysis of genome-wide association studies identifies 17 new Parkinson's disease risk loci. *Nat. Genet.* 49, 1511–1516 (2017).
- Foo, J. N. et al. Identification of risk loci for Parkinson disease in Asians and comparison of risk between Asians and Europeans: a Genome-Wide Association Study. JAMA Neurol. 77, 746–754 (2020).
- 19. Tucci, A. et al. Genetic variability at the PARK16 locus. *Eur. J. Hum. Genet.* 18, 1356–1359 (2010).
- Gan-Or, Z., Dion, P. A. & Rouleau, G. A. Genetic perspective on the role of the autophagy-lysosome pathway in Parkinson disease. *Autophagy* **11**, 1443–1457 (2015).
- Satake, W. et al. Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. Nat. Genet. 41, 1303–1307 (2009).
- Liu, X. et al. Genome-Wide association study identifies candidate genes for Parkinson's disease in an Ashkenazi Jewish population. BMC Med. Genet. 12, 104 (2011).
- 23. Zabetian, C. P. et al. LRRK2 mutations and risk variants in Japanese patients with Parkinson's disease. *Mov. Disord.* **24**, 1034–1041 (2009).
- Kim, J. M. et al. The LRRK2 G2385R variant is a risk factor for sporadic Parkinson's disease in the Korean population. *Parkinsonism Relat. Disord.* 16, 85–88 (2010).
- 25. Tan, E. K. et al. Multiple LRRK2 variants modulate risk of Parkinson disease: a Chinese multicenter study. *Hum. Mutat.* **31**, 561–568 (2010).
- West, A. B. et al. Parkinson's disease-associated mutations in LRRK2 link enhanced GTP-binding and kinase activities to neuronal toxicity. *Hum. Mol. Genet.* 16, 223–232 (2007).
- Taymans, J. M. & Greggio, E. LRRK2 kinase inhibition as a therapeutic strategy for Parkinson's disease, where do we stand? *Curr. Neuropharmacol.* 14, 214–225 (2016).
- Carrion, M. D. P. et al. The LRRK2 G2385R variant is a partial loss-of-function mutation that affects synaptic vesicle trafficking through altered protein interactions. *Sci. Rep.* 7, 5377 (2017).
- Davis, A. A. et al. Variants in GBA, SNCA, and MAPT influence Parkinson disease risk, age at onset, and progression. *Neurobiol. Aging* 37, 209.e201–209.e207 (2016).
- Evans, W. et al. The tau H2 haplotype is almost exclusively Caucasian in origin. Neurosci. Lett. 369, 183–185 (2004).
- Pittman, A. M. et al. Linkage disequilibrium fine mapping and haplotype association analysis of the tau gene in progressive supranuclear palsy and corticobasal degeneration. J. Med. Genet. 42, 837–846 (2005).
- Foo, J. N. et al. Genome-wide association study of Parkinson's disease in East Asians. *Hum. Mol. Genet.* 26, 226–232 (2017).
- Gaspar, H. A. & Breen, G. Probabilistic ancestry maps: a method to assess and visualize population substructures in genetics. *BMC Bioinform.* 20, 116 (2019).
- Wang, Y., Lu, D., Chung, Y. J. & Xu, S. Genetic structure, divergence and admixture of Han Chinese, Japanese and Korean populations. *Hereditas* 155, 19 (2018).
- Lichou, F. & Trynka, G. Functional studies of GWAS variants are gaining momentum. *Nat. Commun.* 11, 6283 (2020).
- Gibb, W. & Lees, A. The relevance of the Lewy body to the pathogenesis of idiopathic Parkinson's disease. J. Neurol. Neurosurg. Psychiatry 51, 745–752 (1988).
- Moon, S. et al. The Korea Biobank Array: design and identification of coding variants associated with blood biochemical traits. *Sci. Rep.* 9, 1382 (2019).
- Purcell, S. et al. PLINK: a tool set for whole-genome association and populationbased linkage analyses. Am. J. Hum. Genet. 81, 559–575 (2007).
- 39. Purcell, S. PLINK 1.9. https://www.cog-genomics.org/plink2/ (2014).

- Pruim, R. J. et al. LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics* 26, 2336–2337 (2010).
- Gauderman, W. J. Sample size requirements for matched case-control studies of gene–environment interaction. *Stat. Med.* 21, 35–50 (2002).

ACKNOWLEDGEMENTS

This study was funded by the Korea Healthcare Technology R&D Project, Ministry of Health & Welfare, Republic of Korea (H19C0256) and Asan Institute for Life Sciences, Asan Medical Center, Seoul, Republic of Korea (2022IT0012). The funders played no role in study design, data collection, analysis, and interpretation of data, or the writing of this manuscript.

AUTHOR CONTRIBUTIONS

K.W.P. organized and executed the project, designed, and reviewed the statistical analysis, and wrote the first draft. H.-S.R. reviewed the statistical analysis and the manuscript. E.S. and Y.G.P. designed, executed, and reviewed the statistical analysis. S.R.J., S.Y.K., J.S.K., and S.-B.K. reviewed and critiqued the statistical analysis and the manuscript. S.J.C. conceptualized the research project, designed, and reviewed the statistical analysis, and reviewed and critiqued the manuscript. All authors made final approval of the completed version.

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41531-023-00580-3.

Correspondence and requests for materials should be addressed to Sun Ju Chung.

Reprints and permission information is available at http://www.nature.com/ reprints

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http:// creativecommons.org/licenses/by/4.0/.

© The Author(s) 2023