

## ARTICLE OPEN



# Lifespan and healthspan benefits of exogenous H<sub>2</sub>S in *C. elegans* are independent from effects downstream of *eat-2* mutation

Li Theng Ng<sup>1,2,4</sup>, Li Fang Ng<sup>1</sup>, Richard Ming Yi Tang<sup>3,4,5</sup>, Diogo Barardo<sup>1,3</sup>, Barry Halliwell<sup>3,4</sup>, Philip Keith Moore<sup>2,4</sup> and Jan Gruber<sup>1,3</sup>✉

Caloric restriction (CR) is one of the most effective interventions to prolong lifespan and promote health. Recently, it has been suggested that hydrogen sulfide (H<sub>2</sub>S) may play a pivotal role in mediating some of these CR-associated benefits. While toxic at high concentrations, H<sub>2</sub>S at lower concentrations can be biologically advantageous. H<sub>2</sub>S levels can be artificially elevated *via* H<sub>2</sub>S-releasing donor drugs. In this study, we explored the function of a novel, slow-releasing H<sub>2</sub>S donor drug (FW1256) and used it as a tool to investigate H<sub>2</sub>S in the context of CR and as a potential CR mimetic. We show that exposure to FW1256 extends lifespan and promotes health in *Caenorhabditis elegans* (*C. elegans*) more robustly than some previous H<sub>2</sub>S-releasing compounds, including GYY4137. We looked at the extent to which FW1256 reproduces CR-associated physiological effects in normal-feeding *C. elegans*. We found that FW1256 promoted healthy longevity to a similar degree as CR but with fewer fitness costs. In contrast to CR, FW1256 actually enhanced overall reproductive capacity and did not reduce adult body length. FW1256 further extended the lifespan of already long-lived *eat-2* mutants without further detriments in developmental timing or fertility, but these lifespan and healthspan benefits required H<sub>2</sub>S exposure to begin early in development. Taken together, these observations suggest that FW1256 delivers exogenous H<sub>2</sub>S efficiently and supports a role for H<sub>2</sub>S in mediating longevity benefits of CR. Delivery of H<sub>2</sub>S *via* FW1256, however, does not mimic CR perfectly, suggesting that the role of H<sub>2</sub>S in CR-associated longevity is likely more complex than previously described.

npj Aging and Mechanisms of Disease (2020)6:6; <https://doi.org/10.1038/s41514-020-0044-8>

## INTRODUCTION

The fraction of aged individuals in many populations around the world is increasing more rapidly than at any time in human history<sup>1</sup>. As a result of this unprecedented demographic change, there is a growing need to identify efficacious, low-risk interventions that promote healthy longevity. A major challenge in biomedical research today is to develop safe, efficacious therapies that can extend human healthspan by delaying or preventing ageing-associated diseases.

Caloric restriction (CR), restricting the amount of calories consumed without causing malnutrition, is one of only a few interventions that have been shown to extend lifespan and healthspan in a wide range of model organisms, ranging from unicellular yeast<sup>2</sup> to multicellular organisms including flies<sup>3</sup>, nematodes<sup>4</sup> and rodents<sup>5</sup>. While there are no direct data on longevity effects of CR in humans, findings from rhesus monkeys imply that CR likely also benefits lifespan and certainly healthspan in primates<sup>6–8</sup>. Evidence in humans also supports such health benefits, for example, CALERIE (Comprehensive Assessment of the Long-term Effects of Reducing Intake of Energy), a two year study of caloric restriction in humans conducted by the National Institute of Aging (NIA), has confirmed that CR is both feasible in humans and benefits a number of health indicators<sup>9</sup>. However, restricting food intake in humans is notoriously difficult and fraught with physiological and psychological challenges<sup>10</sup>. CR diets are extremely difficult to maintain over long periods of time and outside of carefully controlled experiments, there is a

significant risk of inadvertently causing malnutrition<sup>11</sup>. This is of particular concern during human ageing, where malnutrition is already known to be a common problem<sup>12</sup>. Encouraging CR as a means of reducing morbidity in ageing populations is therefore probably not a viable strategy. However, a recent study has shown that extended daily fasting periods can benefit lifespan, regardless of dietary composition or total calories consumed, opening up potential alternative approaches to improve healthspan<sup>13</sup>. Insights into the mechanisms by which CR delays or prevents ageing-associated diseases and extends lifespan may provide another alternative strategy. One promising approach to extending healthspan would be to identify compounds that reproduce CR-associated benefits without the need of adhering to an actual CR regime<sup>14–16</sup>. In this respect, hydrogen sulfide (H<sub>2</sub>S) has recently been reported to act as a potential mediator of CR-associated benefits<sup>17</sup>.

H<sub>2</sub>S is a poisonous, water-soluble gas with a pungent odor of rotten eggs. At levels above 50ppm in air, H<sub>2</sub>S is acutely toxic to humans, whilst levels in excess of 300ppm are potentially fatal<sup>18</sup>. However, it is now clear that H<sub>2</sub>S is produced endogenously and that low endogenous levels may play a role in many biological processes<sup>19,20</sup>. H<sub>2</sub>S is synthesized endogenously in mammalian tissues by three types of enzymes: cystathionine-γ-lyase (CSE), cystathionine β-synthase (CBS) and 3-mercaptopyruvate sulfurtransferase (3-MST). Importantly, H<sub>2</sub>S has been shown to be involved in several physiological and pathophysiological

<sup>1</sup>Ageing Research Laboratory, Science Division, Yale-NUS College, Singapore 138527, Singapore. <sup>2</sup>Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore 117600, Singapore. <sup>3</sup>Department of Biochemistry, Yong Loo Lin School of Medicine, National University of Singapore, Singapore 117596, Singapore. <sup>4</sup>Neurobiology Programme, Life Sciences Institute, National University of Singapore, Singapore 117456, Singapore. <sup>5</sup>NUS Graduate School for Integrative Sciences & Engineering, National University of Singapore, Singapore 117456, Singapore. ✉email: yncjg@nus.edu.sg

processes closely associated with ageing, including inflammation<sup>21,22</sup>, cancer<sup>23</sup> and atherosclerosis<sup>24</sup>.

Interestingly, recent evidence in *Caenorhabditis elegans* (*C. elegans*), yeast, flies and mice suggest that H<sub>2</sub>S may also play an evolutionarily conserved role in lifespan determination and in mediating CR benefits. Miller and Roth first showed that exposure to H<sub>2</sub>S gas significantly extends lifespan and improves thermo-tolerance in *C. elegans*<sup>25</sup>. Later, Hine et al. reported that one of the *C. elegans* orthologues of the mammalian CBS enzyme, *cbs-1*, is required for the extended lifespan of *C. elegans eat-2* mutants<sup>17</sup>. CR also significantly induces endogenous H<sub>2</sub>S production in fruit flies, nematodes, yeast and mice<sup>17</sup>. In addition, *cbs-1* has been reported to be required for the long-lifespan of germline-deficient *glp-1* nematodes<sup>26</sup>. Reducing *cbs-1* expression in *glp-1* mutants was found to decrease H<sub>2</sub>S levels and shorten their lifespan<sup>26</sup>. H<sub>2</sub>S has therefore been implicated in lifespan modulation not only in CR, but also in response to germline signaling. However, whether either mechanism is related to the lifespan effects following lifelong exposure to exogenous H<sub>2</sub>S remains to be shown. In mice, CR has been shown to transcriptionally induce the CSE enzyme and this has been suggested to be a mechanism underlying protection against ischemia-reperfusion injury, as evidenced by a failure in protection against ischemia-reperfusion injury following CR in CSE knockout mice<sup>17</sup>. It therefore seems that increased H<sub>2</sub>S synthesis may be an evolutionarily conserved mechanism, involved in mediating physiological benefits under CR conditions<sup>17</sup>. However, whether H<sub>2</sub>S acts as a signaling molecule or plays a more direct role in modulating CR-associated benefits remains to be determined. Nevertheless, there is increasing evidence that enhancing endogenous H<sub>2</sub>S levels can be physiologically beneficial. A particularly intriguing idea is that H<sub>2</sub>S donor drugs, or modulators of pathways involved in H<sub>2</sub>S metabolism, might represent avenues for the development of CR mimetics<sup>27</sup>.

Several inorganic compounds can be utilized to generate H<sub>2</sub>S exogenously. For example sodium hydrosulfide (NaHS) releases H<sub>2</sub>S rapidly upon hydrolysis and can be used to deliver H<sub>2</sub>S in vivo<sup>28</sup>. However, during CR, physiological exposure to H<sub>2</sub>S is likely at low-levels and chronic, unlike exposure resulting from the rapid release of H<sub>2</sub>S from NaHS. Several novel synthetic organic H<sub>2</sub>S donors have therefore been developed. Such compounds are designed to release H<sub>2</sub>S slowly over an extended period of time; an approach which likely better approximates endogenous H<sub>2</sub>S release in vivo. One of the first was GYY4137 (morpholin-4-ium-4-methoxyphenyl [morpholino] phosphinodithioate), a compound that releases H<sub>2</sub>S slowly both in vitro and in vivo<sup>29</sup>. Since it was first described, GYY4137 has been studied extensively and shown to exert biological effects in mice, rats and nematodes<sup>29–33</sup>. In recent years, AP39, a H<sub>2</sub>S donor designed to release H<sub>2</sub>S specifically within mitochondria has been developed<sup>34</sup> and shown to confer cytoprotective activity and to attenuate the loss of cellular bioenergetics in cells subjected to oxidative stress<sup>35</sup>. AP39 has been reported to elicit protection effects against renal ischemia-reperfusion injury in rats<sup>36</sup> and preserve mitochondrial function in APP/PS1 mice and neurons, indicating a potential role in protecting against Alzheimer's disease<sup>37</sup>.

We have developed a series of novel H<sub>2</sub>S donors. Among these H<sub>2</sub>S donors, 3-dihydro-2-phenyl-sulfanylbenzo[d][1,3,2]-oxazaphosphole (FW1256) exerts superior anti-proliferative effects compared to GYY4137, as evaluated in MCF7 breast cancer cells<sup>38</sup>. FW1256 has also been shown to exert significant anti-inflammatory effects in RAW264.7 macrophages<sup>39</sup>. However, the efficacy of these novel H<sub>2</sub>S donors in the context of ageing remains to be explored.

Here we report the effects of two novel H<sub>2</sub>S donor compounds (FW1251 and FW1256) on lifespan, healthspan and other CR-associated phenotypes in nematodes. We observed that FW1256 extended lifespan and promoted healthy ageing in wild-type (WT) *C. elegans* but did not fully replicate CR-associated effects. We

found that, in *C. elegans*, exogenous H<sub>2</sub>S promoted healthy longevity with less severe detrimental effects on some parameters of fitness than CR as modelled by *eat-2* mutation. FW1256 was also able to further extend the already long lifespan of *eat-2* mutants. Finally, we provide evidence that H<sub>2</sub>S affects mitochondrial function and that exposure to H<sub>2</sub>S during larval development was required for these benefits, suggesting involvement of a developmental adaptation. Our data imply that the function of H<sub>2</sub>S in lifespan regulation and CR is more complex than previously appreciated.

## RESULTS

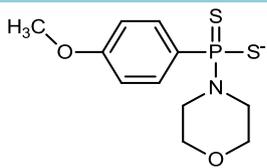
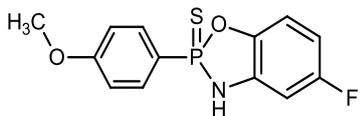
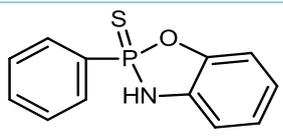
Screen for novel H<sub>2</sub>S donor compounds with lifespan extension effect

It was previously reported that GYY4137 (morpholin-4-ium-4-methoxyphenyl [morpholino] phosphinodithioate), a slow-releasing H<sub>2</sub>S donor, extends lifespan in *C. elegans*<sup>33</sup> but the effect size, even after careful dose optimization, was small compared to that previously reported by Miller and Roth for direct exposure to H<sub>2</sub>S gas<sup>25</sup>. We wondered if a H<sub>2</sub>S donor compound with a more rapid H<sub>2</sub>S release rate might elicit effects more comparable to those achieved by Miller and Roth. For the current study, we therefore selected two compounds from a series of our novel H<sub>2</sub>S donor compounds, FW1251 and FW1256 (Table 1). Both FW1251 and FW1256 release H<sub>2</sub>S more rapidly than GYY4137<sup>38</sup>. Both compounds were designed primarily to deliver exogenous H<sub>2</sub>S to maximize anti-proliferative activity against cancer cells<sup>38</sup>. While GYY4137 releases H<sub>2</sub>S at a rate of about 1% per day under standard assay conditions (acetonitrile [MeCN]:phosphate buffered saline [PBS] at room temperature), FW1251 releases H<sub>2</sub>S twice as fast under the same condition and FW1256 releases H<sub>2</sub>S even more rapidly, releasing about 5% per day<sup>38</sup>. The mechanism for H<sub>2</sub>S release by FW1256 in aqueous environment is shown in schematic form in Supplementary Fig. 1. Release of H<sub>2</sub>S from another H<sub>2</sub>S donor, diallyl trisulfide (DATS), has been shown to be highly dependent on the presence of biological thiols<sup>40</sup>. It is therefore, worth noting that biological thiols might affect the efficacy and actual release rate of H<sub>2</sub>S from donor compounds in vivo. However, previous data suggest that FW1256 is more potent as an anticancer agent than GYY4137 yet is nontoxic to human lung fibroblast cells (WI38) even at 500 μM<sup>38</sup>.

To explore effects of these compounds in *C. elegans*, we first optimized dose-response in terms of lifespan in *C. elegans*. We carried out range finding for optimal dose for both compounds. WT *C. elegans* were exposed to four different concentrations (10 μM, 50 μM, 250 μM and 750 μM) of FW1251 in a series of operator-blinded lifespan studies. We observed no obvious toxicity below 750 μM and found that 50 μM resulted in the largest beneficial effect within this range (log-rank test,  $p < 0.05$ ) (Fig. 1a). However, mean lifespan was increased by only 11% even at the ideal dose (mean lifespan:  $19.9 \pm 0.5$  days, compared to control:  $17.9 \pm 0.5$  days,  $p < 0.05$  (Fig. 1b, Supplementary Table 2). At 750 μM, FW1251 showed some toxic effect on WT *C. elegans* (Supplementary Fig. 2, Supplementary Table 2). FW1251 therefore provided only limited benefits at low concentrations and was ineffective or toxic at high doses (Supplementary Fig. 3).

For our second (and faster-releasing) compound, FW1256, *C. elegans* were exposed to 10 μM, 50 μM and 150 μM during the initial screening. The highest concentration, 150 μM, significantly extended lifespan while the lower concentrations were ineffective (Supplementary Fig. 4, Supplementary Table 2). Thereafter, *C. elegans* were exposed to even higher concentrations of FW1256 (250 μM and 500 μM) to establish an optimal dose and test for possible toxicity at high levels of FW1256. Surprisingly, FW1256 was beneficial at both of these concentrations (Fig. 1c). FW1256 (250 μM) resulted in 42% mean lifespan extension (mean lifespan:

**Table 1.** Chemical structures and H<sub>2</sub>S releasing speeds of H<sub>2</sub>S donor compounds used in this study compared to GYY4137.

Name	Structures	Molecular weight	% of H <sub>2</sub> S released per day (room temperature, pH7.4)
<b>GYY4137</b>		376.47	0.7
<b>FW1251</b>		295.02	1.8
<b>FW1256</b>		247.02	5.0

24.0 ± 0.3 days, compared to control: 16.9 ± 0.4 days,  $p < 0.001$ ) while FW1256 (500 μM) extended lifespan by 51% (mean lifespan: 25.6 ± 0.3 days, compared to control: 16.9 ± 0.4 days,  $p < 0.001$ ) (Fig. 1d, Supplementary Table 2). While 500 μM of FW1256 resulted in the largest overall benefit, doubling concentration from 250 μM to 500 μM of FW1256 resulted in an additional increase of only 9% of mean lifespan. For the remainder of this study, we therefore focused on FW1256 at a dose of 500 μM (Supplementary Fig. 3).

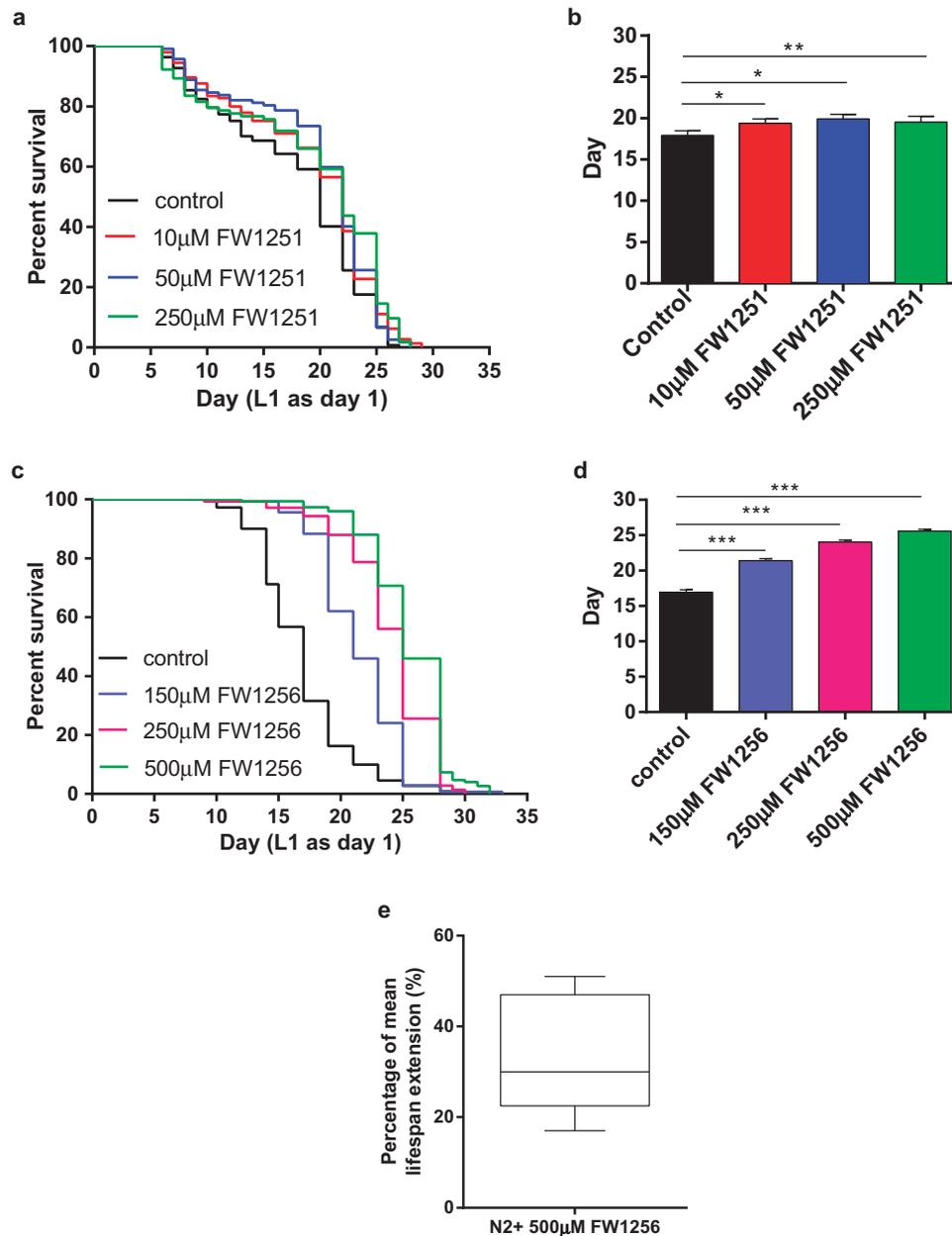
Comparison with over 400 compounds from an exhaustive database of drugs with published evidence for effects on lifespan (DrugAge Build3) puts FW1256 into the top 15 of all 400 drugs (top 4%) in terms of efficacy in *C. elegans*<sup>41</sup>. However, lifespan effects in *C. elegans* can be highly variable<sup>41</sup>. In order to accurately quantify the lifespan extension effect of FW1256, a total of five independent (biologically independent cohorts of WT *C. elegans*), operator-blinded repeats of the lifespan assay at 500 μM were carried out. While lifespan effect size showed variability between repeats, FW1256 consistently extended mean lifespan in all 5 of these independent experiments (mean effect size: 33.8%; 95% confidence interval: 17.3%–50.4%, Supplementary Table 1, Fig. 1e).

#### FW1256 releases H<sub>2</sub>S in vivo

Having established that FW1256 extends lifespan of *C. elegans*, we wondered if this effect was indeed due to the release of H<sub>2</sub>S in *C. elegans*. To test whether FW1256 was able to deliver H<sub>2</sub>S in vivo in *C. elegans*, we first used a fluorescence H<sub>2</sub>S sensor probe, 7-azido-4-methylcoumarin (AzMC)<sup>42</sup>. Fluorescence intensity of AzMC was

significantly increased in WT *C. elegans* exposed to 500 μM FW1256 (Fig. 2a, b), confirming successful delivery of detectable quantities of exogenous H<sub>2</sub>S by FW1256. The amount of increase in H<sub>2</sub>S-related fluorescence seen in *eat-2* mutants is comparable to the FW1256-induced increase as judged by AzMC in WT *C. elegans* exposed to 500 μM of FW1256 (Supplementary Fig. 5). This comparison suggested that the amount of endogenous H<sub>2</sub>S resulting in nematodes exposed to 500 μM of FW1256 was approximately half of the increase seen in *eat-2* mutants (Supplementary Fig. 5).

We next monitored gene expression changes of two H<sub>2</sub>S oxidation enzymes, SQRD-1 and ETHE-1. SQRD-1 is an oxidoreductase that oxidizes H<sub>2</sub>S to polysulfide, which is then further oxidized to sulfate and thiosulfate *via* catalysis by ETHE-1<sup>43</sup>. Both of these enzymes are therefore directly involved in detoxifying/degrading H<sub>2</sub>S, contributing to maintaining intrinsic H<sub>2</sub>S levels<sup>43</sup>. While *sqrd-1* has previously been shown to be transcriptionally upregulated in response to the presence of H<sub>2</sub>S gas, *ethe-1* was not induced significantly under these conditions<sup>43</sup>. We nevertheless tested for potential upregulation of both *sqrd-1* and *ethe-1* mRNA upon exposure to 500 μM of FW1256. Consistent with delivery of physiologically relevant levels of H<sub>2</sub>S gas, mRNA levels of *sqrd-1* were increased by about 3 fold ( $p < 0.05$ ) while mRNA levels of *ethe-1* were not upregulated significantly (about 2 fold change,  $p = 0.1952$ ) in WT *C. elegans* exposed to 500 μM of FW1256 (Fig. 2c). Together, these data supported the notion that FW1256 delivers physiologically significant levels of H<sub>2</sub>S to *C. elegans*.



**Fig. 1** Effects of FW1251 and FW1256 on lifespan of WT *C. elegans*. **a** Survival curves and **b** mean lifespan of WT *C. elegans* exposed to different doses of FW1251,  $n = 103$ –145 animals per condition. **c**, **d** Dose–response experiment of FW1256 on lifespan of WT *C. elegans*, **c** survival curves, **d** mean lifespan,  $n = 111$ –150 animals per condition. **e** Boxplot showing distribution of mean lifespan extension effects on WT *C. elegans* upon exposure to 500  $\mu$ M of FW1256 from the five independent experiments (Supplementary Table 1). In this box plot, the bottom and top of the box represent the 25% percentile and 75% percentile, the center line represents the median and the whiskers encompass the entire range of values for all five independent experiments. (Survival curves were analyzed using log-rank tests while mean lifespans were analyzed using OASIS2, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ ).

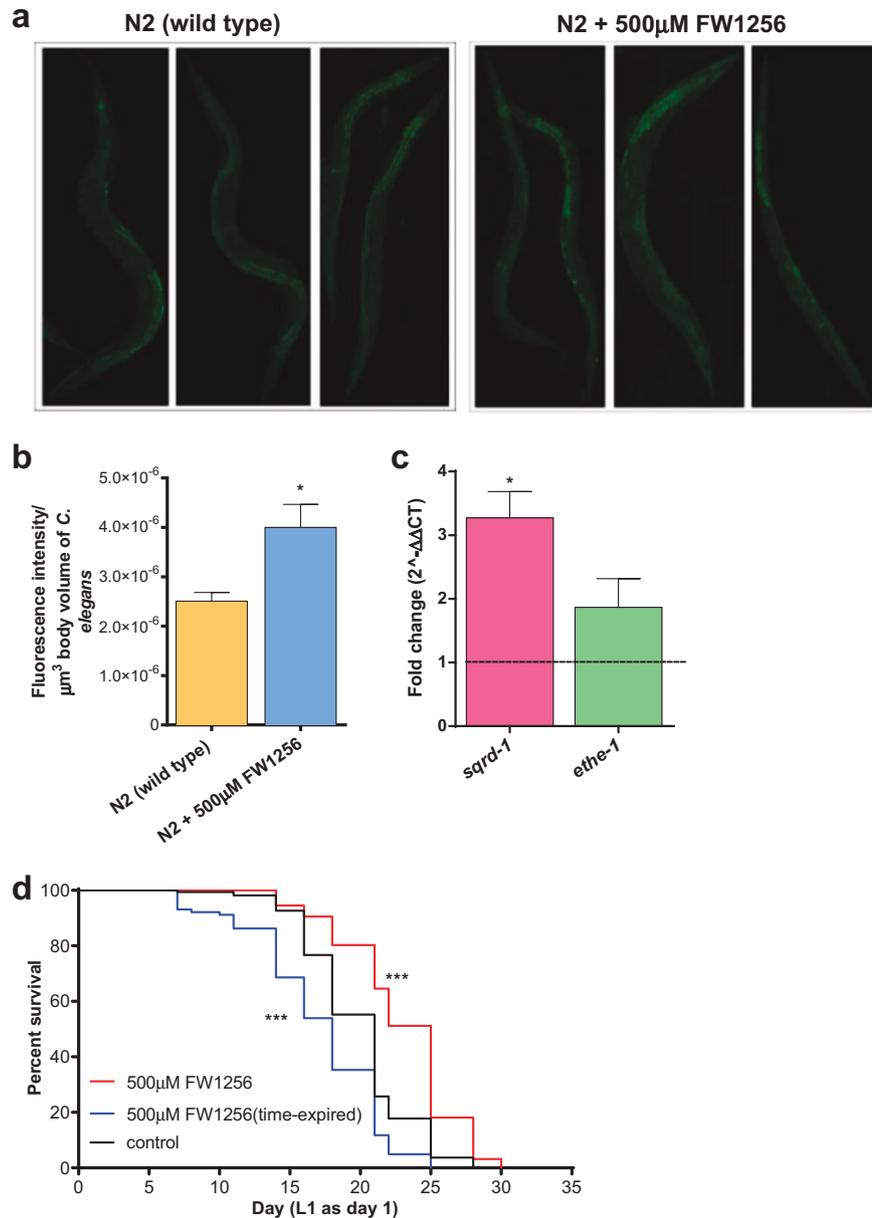
#### The H<sub>2</sub>S moiety is required for FW1256 lifespan benefits

Next, we tested whether the lifespan enhancing effect of FW1256 was dependent on the presence of the H<sub>2</sub>S moiety in the molecule. To rule out possible lifespan benefits related to pharmacological effects of FW1256 independent of H<sub>2</sub>S, we carried out a control lifespan study with time-expired FW1256. Prior to exposing WT *C. elegans* to FW1256, FW1256 was allowed to expire for 4 weeks at room temperature in dimethyl sulfoxide (DMSO) to ensure that H<sub>2</sub>S was quantitatively released<sup>44</sup> (For more information on this approach, see Supplementary Fig. 6). We found that time-expired FW1256 did not extend the lifespan of *C. elegans*. Time-expired FW1256, in fact,

shortened lifespan (log-rank test,  $p < 0.001$ , mean  $\pm$  SEM: 17.1  $\pm$  0.4 days, compared to control: 19.9  $\pm$  0.3 days,  $p < 0.001$ ) (Fig. 2d, Supplementary Table 2), suggesting some toxicity of the expired compound in *C. elegans*. Together these data imply that fresh FW1256 delivers significant amounts of H<sub>2</sub>S exogenously to *C. elegans* and that the ability to release H<sub>2</sub>S is required for its lifespan benefits.

#### FW1256 rescues detrimental effects and extends lifespans of H<sub>2</sub>S mutants

*C. elegans* possesses three families of endogenous H<sub>2</sub>S synthesizing enzymes, comprising two orthologues, each, of the mammalian

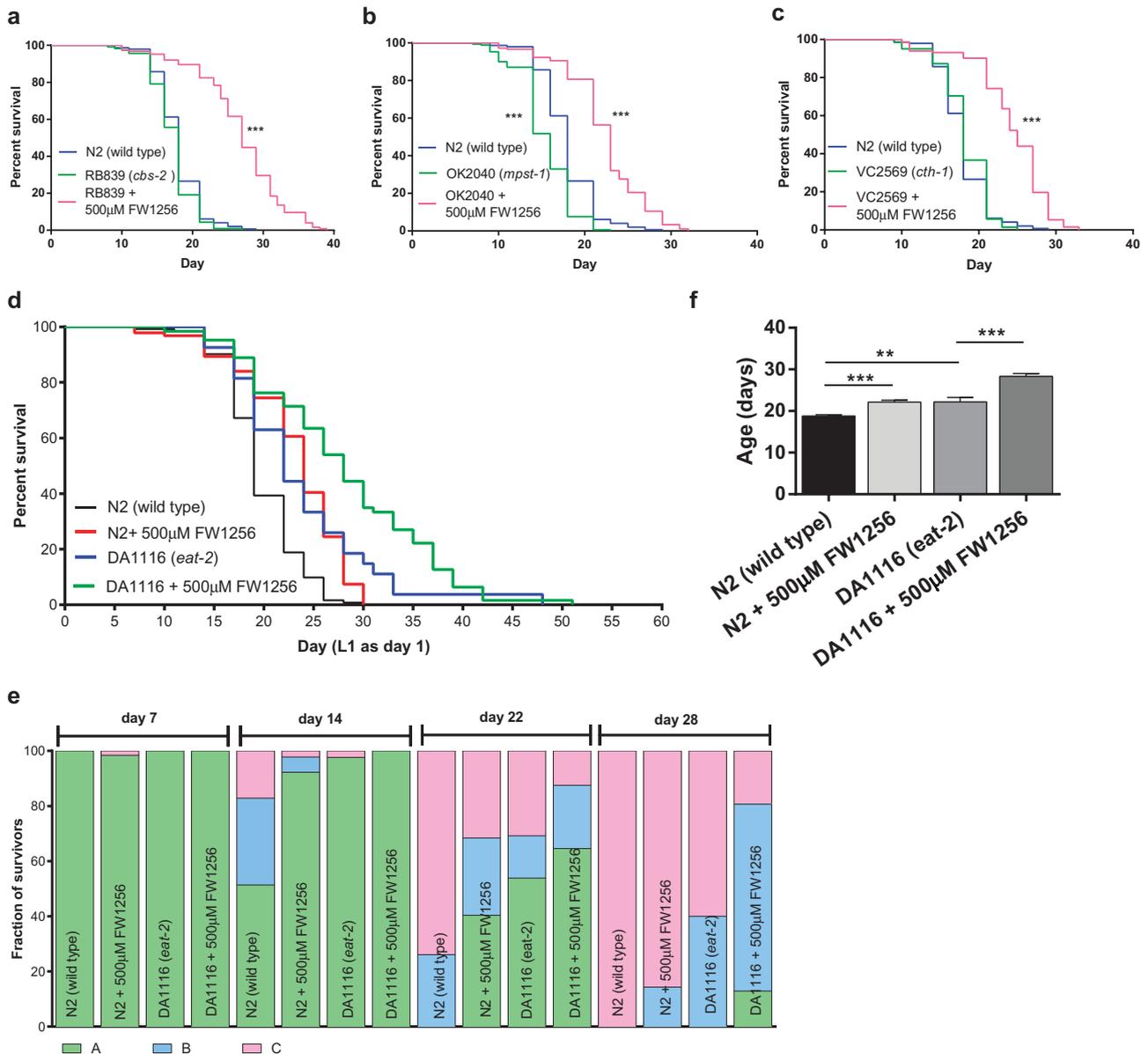


**Fig. 2** H<sub>2</sub>S moiety from FW1256 is the key of longevity increase. **a** Confocal microscopy fluorescence images of H<sub>2</sub>S detection using AzMC (in green) in *C. elegans*. **b** Fluorescence intensity of the AzMC signal, normalized to body volume of *C. elegans* ( $n = 5$  animals per condition,  $t$ -test,  $*p < 0.05$ ). **c** Relative quantification mRNA levels of *sqrd-1* and *ethe-1* encoding for endogenous H<sub>2</sub>S detoxification system in WT *C. elegans* exposed to 500  $\mu$ M FW1256. Data show expression of *sqrd-1* and *ethe-1* relative to WT control ( $n = 3$  repeated experiments, one sample  $t$ -test,  $*p < 0.05$ ). **d** Effect of time-expired FW1256 (500  $\mu$ M, 4 weeks at room temperature) on lifespan of WT *C. elegans* ( $n = 102$ –163 animals per condition, log-rank tests,  $***p < 0.001$ ).

CBS enzyme (*cbs-1*, *cbs-2*) and CSE enzyme (*cth-1*, *cth-2*) as well as seven orthologues of 3-MST (*mpst-1*–*7*)<sup>33</sup>. Both CBS and CSE require L-cysteine as substrate for H<sub>2</sub>S production and act mainly in cytoplasm while 3-MST converts 3-mercaptopyruvate to H<sub>2</sub>S, predominantly in the mitochondria (Supplementary Fig. 7)<sup>45,46</sup>.

We have previously shown that deletion of *mpst-1* shortens the lifespan of *C. elegans*<sup>33</sup>, suggesting a role of H<sub>2</sub>S production in the mitochondria in the context of normal lifespan determination. Whilst there is no prior report regarding the effect of *cbs-2* deletion on lifespan, *C. elegans* with *cbs-2* deletion has been reported to develop normally<sup>47</sup>. By contrast, RNA interference (RNAi) knockdown of *cth-1* has been reported to be detrimental to the lifespan of WT *C. elegans*<sup>17</sup>. These data suggest that H<sub>2</sub>S

production in cytoplasm may also be required for normal lifespan. To quantitatively compare the importance of H<sub>2</sub>S production in different cellular compartments for lifespan regulation, we systematically compared the lifespan of three different H<sub>2</sub>S mutants: RB839 (*cbs-2*), OK2040 (*mpst-1*) and VC2569 (*cth-1*) under the standard conditions used in our laboratory. In our hands, the lifespans of *cbs-2* and *cth-1* mutants were unchanged, while a statistically significant shortened lifespan was observed consistently only in *mpst-1* mutants (log-rank test,  $p < 0.001$ , mean  $\pm$  SEM:  $15.4 \pm 0.2$  days, compared to WT:  $17.9 \pm 0.3$  days,  $p < 0.001$ ) (Fig. 3a–c, Supplementary Table 2), confirming that endogenous H<sub>2</sub>S production, at least in mitochondria, is required for normal lifespan in *C. elegans*.



**Fig. 3** FW1256 not only benefits lifespans of H<sub>2</sub>S mutants and *eat-2* mutants, but also promotes health. **a–c** Comparison of lifespans in WT *C. elegans*, H<sub>2</sub>S mutants and H<sub>2</sub>S mutants exposed to FW1256 (500  $\mu$ M) (data show log-rank tests, \*\*\* $p$  < 0.001). **a** RB839, *cbs-2* mutants,  $n$  = 115–147 animals, **b** OK2040, *mpst-1* mutants,  $n$  = 147–181 animals, **c** VC2569, *cth-1* mutants,  $n$  = 132–147 animals. **d** Effect of FW1256 (500  $\mu$ M) on lifespans of WT and *eat-2* mutants,  $n$  = 27–122 animals per condition. **e** Mobility status of surviving nematodes at different ages,  $n$  = 27–122 animals per condition. Generally, class A nematodes moved constantly, class B nematodes moved only when prodded while class C nematodes showed movement of their head and tails only. **f** Mean healthspan of unexposed and FW1256-exposed WT and *eat-2* mutants ( $n$  = 27–122 animals per condition, one-way ANOVA with Bonferroni's post-test, \*\* $p$  < 0.01, \*\*\* $p$  < 0.001).

To investigate if FW1256 rescues or extends the lifespan of these H<sub>2</sub>S mutants, three H<sub>2</sub>S mutant strains were exposed to FW1256 (500  $\mu$ M). Interestingly, exposure to FW1256 extended lifespan in these mutants beyond that even of untreated WT animals (Fig. 3a–c). This further supports the notion that FW1256 effectively delivers H<sub>2</sub>S in vivo. Interestingly, comparison of FW1256 benefits between the three H<sub>2</sub>S mutant strains showed that lifespan benefits were most pronounced in the *cbs-2* mutant strain (log rank test,  $p$  < 0.001 with mean lifespan extension of 56%, mean  $\pm$  SEM: 26.9  $\pm$  0.5 days, compared to RB839 control: 17.2  $\pm$  0.3 days,  $p$  < 0.001) (Fig. 3a, Supplementary Table 2), perhaps suggesting that loss of cytosolic *cbs* leads to compensatory effects that prime animals for lifespan extension by exogenously delivered H<sub>2</sub>S.

Similar to CR, FW1256 promotes healthy longevity in *C. elegans*. In *C. elegans*, CR can be modelled genetically by mutations that reduce pharyngeal pumping rate, thereby decreasing the amount of food consumed<sup>4</sup>. It has previously been reported that one *C. elegans* endogenous H<sub>2</sub>S synthesizing enzyme, *cbs-1*, is required for the extended lifespan of a commonly used CR mutant (*eat-2*)<sup>17</sup>. To further test this link, we sought to investigate whether delivery of exogenous H<sub>2</sub>S by FW1256 resulted in CR-like benefits and/or tradeoffs and phenotypes in WT and *eat-2* mutants. Comparing the lifespan of unexposed and FW1256-exposed (500  $\mu$ M) WT and *eat-2* mutants, we found that the lifespan of FW1256-exposed WT (mean lifespan: 23  $\pm$  0.5 days) was nearly identical to that of unexposed *eat-2* mutants (mean lifespan: 23.7  $\pm$  1.7 days) (Fig. 3d, Supplementary Table 2). Exogenous H<sub>2</sub>S delivered by donor drugs

was thus capable of producing lifespan benefits in WT animals that were of the same magnitude as CR. However, strikingly, FW1256 also further extended the lifespan of already long-lived *eat-2* mutants (log rank test,  $p < 0.01$ ; mean lifespan:  $28.1 \pm 1.1$  days, compared to unexposed *eat-2* mutants:  $23.7 \pm 1.4$  days,  $p < 0.001$ ) (Fig. 3d, Supplementary Table 2). Lifespan of *eat-2* mutants was extended by 18.6%, statistically within the range of lifespan extension effects observed in WT nematodes (Fig. 1e), suggesting that lifespan benefits of exogenous  $H_2S$  were not significantly blunted in *eat-2* mutants.

CR not only extends lifespan in many organisms but has been shown to delay and prevent ageing-associated diseases in many animals, including non-human primates<sup>6,48</sup>. We therefore compared effects of exposure to FW1256 in WT and *eat-2* mutants with respect to healthspan. Health status of *C. elegans* was assessed using the mobility score of Herndon et al.<sup>49</sup>. Observation of mobility status demonstrated that both exogenous  $H_2S$  and CR retarded the age-dependent deterioration of mobility in *C. elegans*. Both aged WT and *eat-2* mutants exposed to 500  $\mu M$  of FW1256 were healthier, on average, than aged WT control ( $p < 0.001$ ). The differences were more pronounced as nematodes aged and were most notable between day 14 and day 28. For example, on day 22, only 26% of unexposed WT remained healthy, while 68% of FW1256-exposed WT, 69% of unexposed *eat-2* mutants and 91% of FW1256-exposed *eat-2* mutants remained healthy (Fig. 3e). While unexposed WT had an average healthspan of 18.8 days, a statistically significant longer average healthspan was observed in unexposed *eat-2* mutants (22.2 days,  $p < 0.01$ ), FW1256-exposed WT (22.1 Days,  $p < 0.001$ ) and FW1256-exposed *eat-2* mutants (28.3 days,  $p < 0.001$ ) (Fig. 3f). These data suggest that the beneficial effects of FW1256 on both lifespan and healthspan are additive with the *eat-2* mutation.

FW1256 elicits healthy longevity with fewer fitness costs than CR. In addition to beneficial and protective effects, CR is typically associated with developmental tradeoffs, in particular with reduced fecundity<sup>50</sup>, slower growth and smaller adult body size<sup>51</sup> as well as delayed development<sup>52</sup>. To assess if exposure to  $H_2S$  replicates these longevity-associated tradeoffs in *C. elegans*, we determined egg laying profiles, growth and developmental timing for WT and *eat-2* mutants with and without exposure to FW1256. Comparing the egg laying distributions (Fig. 4a), we found that both FW1256 and CR delayed reproduction by 1.7 days (Peak of progeny production:  $6.9 \pm 0.1$  days, compared to unexposed WT:  $5.2 \pm 0.1$  days,  $p < 0.001$ ) and 2.1 days (Peak of progeny production:  $7.3 \pm 0.3$  days, compared to unexposed WT:  $5.2 \pm 0.1$  days,  $p < 0.001$ ) respectively. However, exposure to 500  $\mu M$  of FW1256 did not further delay the peak of progeny reproduction in *eat-2* mutants, compared to unexposed *eat-2* mutants (Fig. 4c). As expected, unexposed *eat-2* mutants laid significantly fewer eggs (Total number of eggs laid:  $65 \pm 15.7$ ) than unexposed WT (Total number of eggs laid:  $207 \pm 11.6$ ,  $p < 0.001$ ). However, to our surprise, we found that FW1256 did not suppress total reproduction capability in either WT (Total number of eggs laid by FW1256-exposed WT:  $245 \pm 15.6$ , compared to unexposed WT:  $207 \pm 11.6$ ,  $p > 0.05$ ) or *eat-2* mutants (Total number of eggs laid by FW1256-exposed *eat-2* mutants:  $93 \pm 23.7$ , compared to unexposed *eat-2* mutants:  $69 \pm 15.7$ ,  $p > 0.05$ ) (Fig. 4d).

Exposure to FW1256 and *eat-2* mutation both resulted in developmental delay, with *eat-2* mutants being somewhat slower to develop than FW1256-exposed WT *C. elegans*, yet the exposure to FW1256 did not further delay developmental timing of *eat-2* mutants (Fig. 4e).

WT *C. elegans* grew rapidly in size between day 1 and day 6 of life, growing at a rate of  $125 \pm 6 \mu m/day$ , on average (Fig. 4f, Table 2). Unexposed *eat-2* mutants initially grew much more slowly and only started growing between day 3 and day 6 at a rate

of  $119 \pm 5 \mu m/day$ . Thereafter, they stopped growing in size and remained small relative to WT for their entire life ( $p < 0.001$ ) (Fig. 4b and f, Table 2). Exposure to FW1256 similarly initially delayed growth of WT, but between day 3 and day 6, FW1256-exposed WT animals actually grew at a faster rate compared to unexposed WT (at a rate of  $142 \pm 6 \mu m/day$ ,  $p < 0.05$ ). FW1256-exposed WT therefore were able to catch up in size and reached the same adult body size as unexposed WT before both stopped growing from day 9 onwards (Fig. 4b and f, Table 2).

This means that, FW1256 only reduced body length of WT significantly during L2 to L4 larval stages ( $p < 0.001$ ) whereas *eat-2* mutants had significant shorter body length than unexposed WT from L2 larval stages onwards and throughout the entire lifespan ( $p < 0.001$ ). Finally, exposure to FW1256 did not further impact growth of *eat-2* mutants (Fig. 4b).

#### FW1256 causes alterations in mitochondrial metabolism

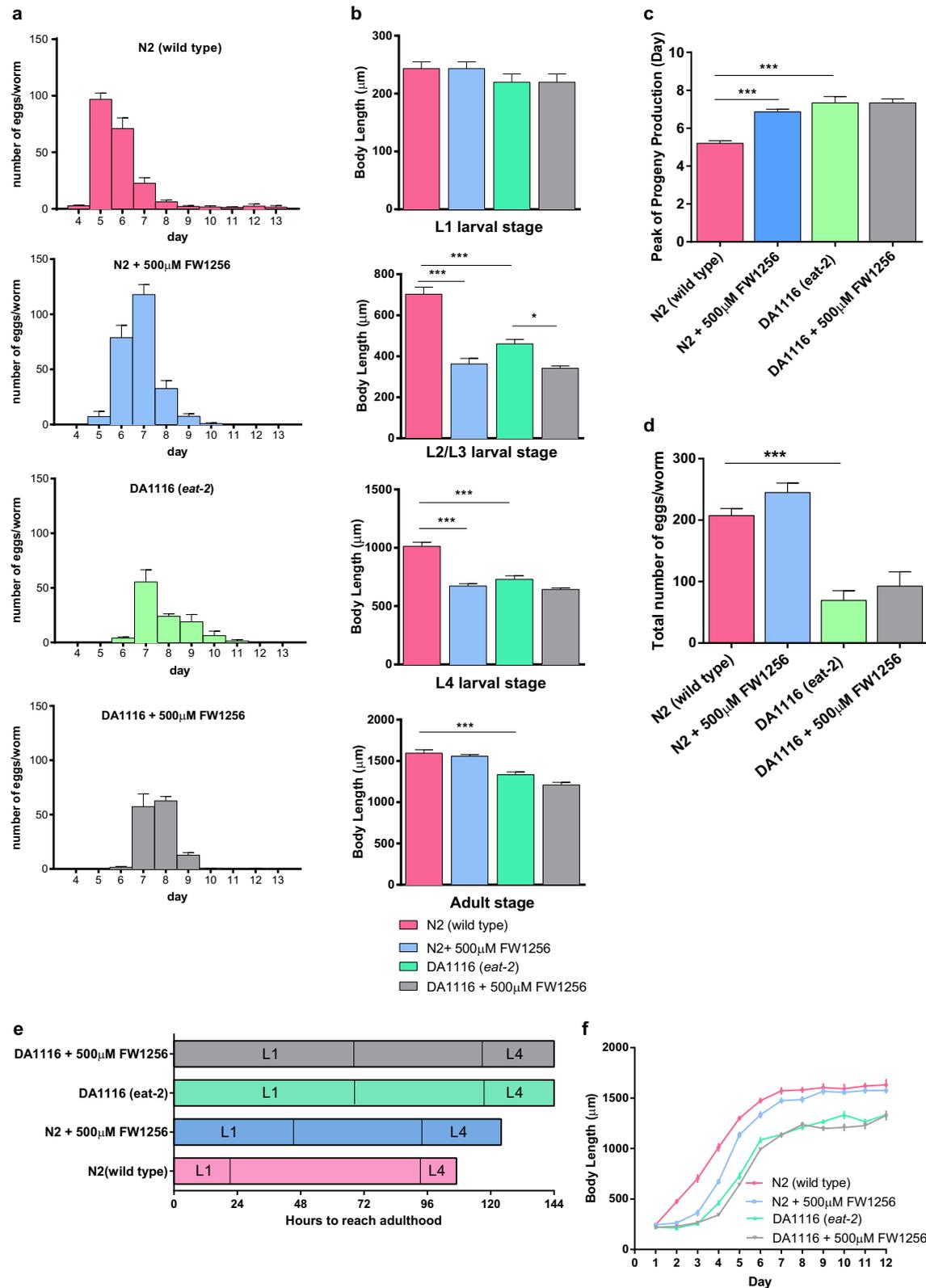
CR has previously been reported to increase mitochondrial respiration in *C. elegans*<sup>53</sup>, *Saccharomyces cerevisiae*<sup>54</sup> and mice<sup>55</sup>. On the other hand,  $H_2S$  has been reported to bind to mitochondrial Cytochrome C Oxidase, thereby inhibiting mitochondrial respiration<sup>56</sup>. We therefore wondered in what way exogenous  $H_2S$  generated by FW1256 would impact respiratory capacity and ATP levels of *C. elegans*. To compare the metabolic effects of CR and FW1256 exposure, we determined both basal respiration and maximal respiration with and without exposure to FW1256 (Supplementary Fig. 8)<sup>57</sup>. WT animals exposed to FW1256 and *eat-2* mutants both showed significantly higher basal respiration (about 40%,  $p < 0.001$ ; about 30%,  $p < 0.01$ , respectively) (Fig. 5a) compared to unexposed WT but we detected no significant change in maximal respiration rates. However, exposure to FW1256 decreased maximal respiration rate of *eat-2* mutants significantly (20%,  $p < 0.05$ ) (Fig. 5b). To determine if significant changes in energy availability were associated with this increase in basal respiration rate, we next quantified ATP levels using the firefly luciferase assay<sup>58</sup>. Interestingly, we found that both WT exposed to FW1256 and *eat-2* mutants had significantly lower ATP levels compared to unexposed WT, with 16% and 24% relative reduction, respectively ( $p < 0.01$ ) (Fig. 5c).

#### Effects of FW1256 on markers of ROS and oxidative damage

The free radical theory of ageing (FRTA) proposes accumulation of free radical-mediated damage as a key driving force of the ageing process<sup>59</sup>. However, this view has been challenged, especially in *C. elegans*<sup>53,60–63</sup>. We therefore attempted to examine to what extent (if any) oxidative stress was correlated with lifespan extension by FW1256 in *C. elegans*. Four different parameters were measured to determine changes in oxidative damage and reactive oxygen species (ROS). We measured oxidative damage to mitochondrial DNA (mtDNA), using quantitative PCR (qPCR)<sup>64</sup>, mitochondrial superoxide via mitoSOX red fluorescence<sup>33</sup> as well as 8-Hydroxydeoxyguanosine (8-OHdG) and 8-hydroxyguanosine (8-OHG) using liquid chromatography–mass spectrometry (LC-MS) as markers of oxidative DNA (8-OHdG) and RNA (8-OHG) damage. While we observed no significant changes in oxidative damage to mtDNA, mitoSOX fluorescence or 8-OHG (Fig. 5d, e, g), we found a significant elevation in 8-OHdG upon exposure to FW1256 (fold change 1.5,  $p < 0.01$ ), suggesting increased DNA damage in animals exposed to FW1256 (Fig. 5f). Exposure to FW1256 therefore does not inhibit mitochondrial respiration and does not elicit an antioxidant effect but instead causes increased respiration and elevates oxidative DNA damage.

#### FW1256 acts differently from CR

Most of the evidence above implies that the effect of exogenous  $H_2S$  as delivered by FW1256, is not directly mimicking CR. Our data

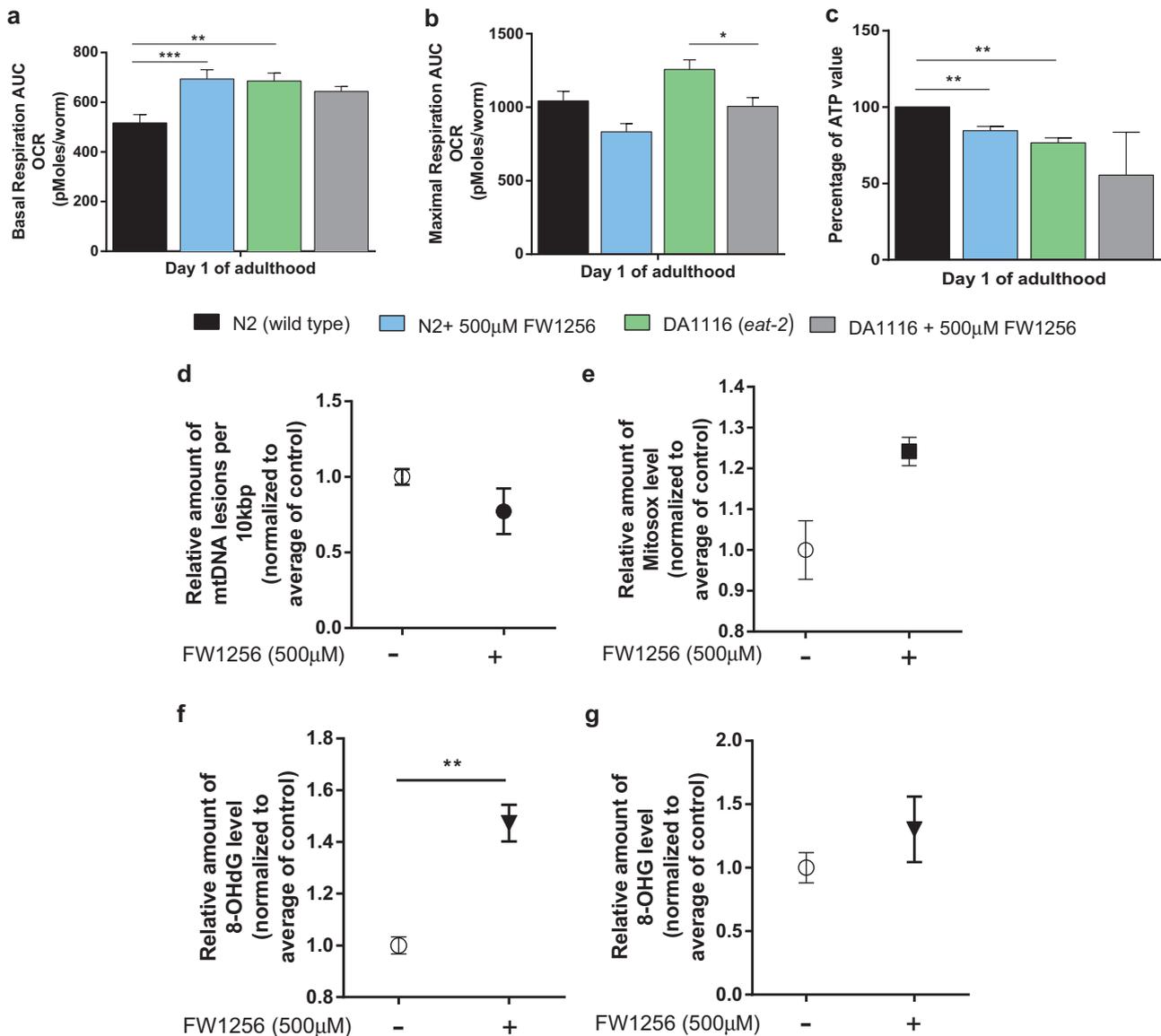


**Fig. 4** Comparing to CR, FW1256 elicits fewer fitness costs in *C. elegans*. **a** Egg laying study reveals reproduction profiles of each group of *C. elegans*. **b** Comparison of body length at L1 larval stage, L2/L3 larval stage, L4 larval stage and adult stage. **c** Day of peak progeny production. **d** Total number of eggs laid by a nematode. **e** Developmental timing and schedules from eggs into adults of WT and *eat-2* mutants (with or without exposure to 500  $\mu$ M of FW1256). **f** Measurement of body length of nematodes from age day 1 (L1 larval stage) to age day 12 (adult stage). ( $n = 10$  animals each, one-way ANOVA with Bonferroni's post-test,  $*p < 0.05$ ,  $***p < 0.001$ ).

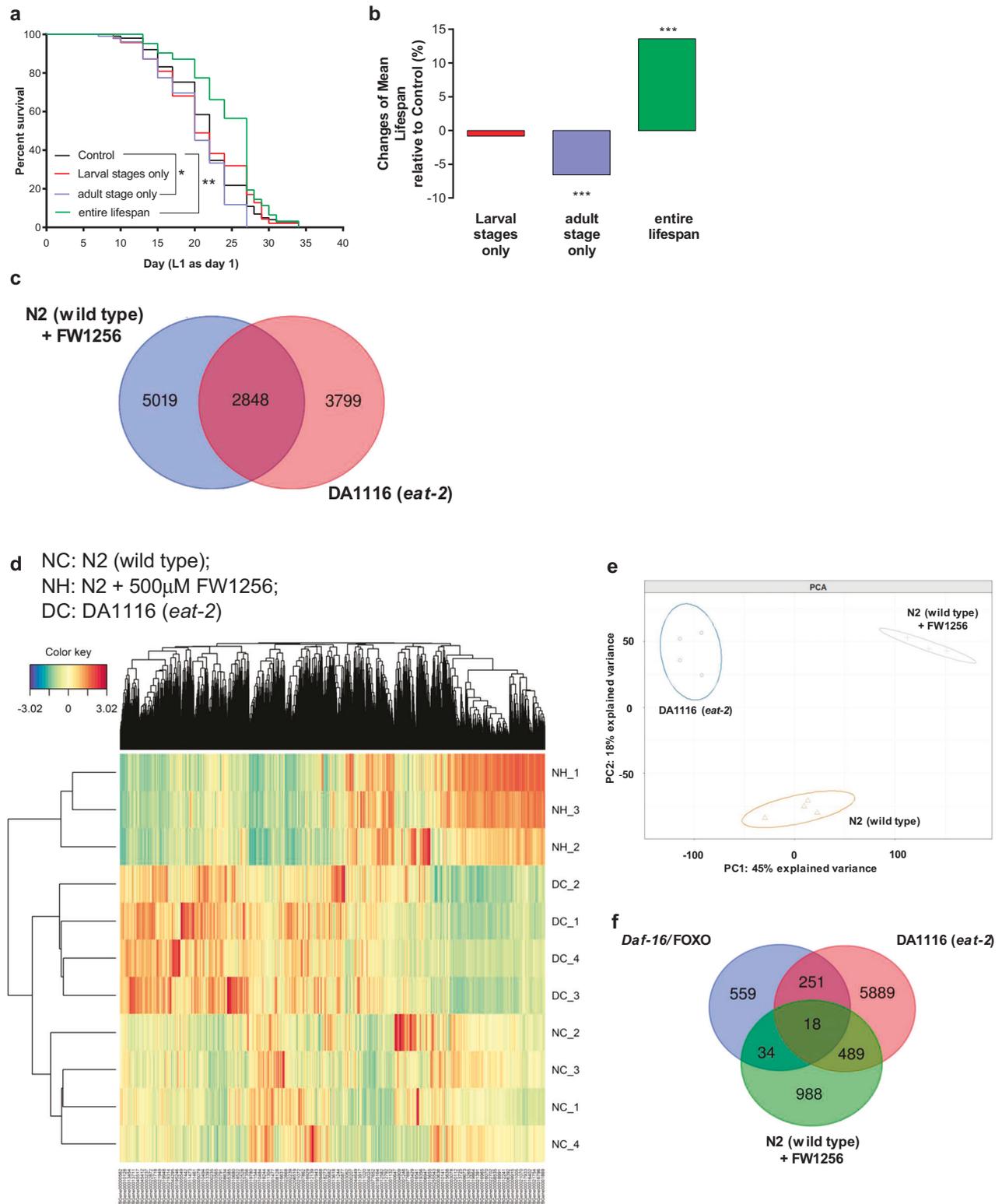
suggest a more complex interplay between endogenous H<sub>2</sub>S production, energy metabolism, oxidative damage, lifespan, healthspan, and developmental- and fitness- tradeoffs. Moreover,

Samples	Rate of development ( $\mu\text{m}/\text{day}$ )
N2 (wild type)	125 $\pm$ 6
N2+ 500 $\mu\text{M}$ FW1256	142 $\pm$ 6
DA1116 ( <i>eat-2</i> )	119 $\pm$ 5
DA1116+ 500 $\mu\text{M}$ FW1256	119 $\pm$ 4

CR is typically initiated in adult animals and CR certainly has been shown to robustly benefit longevity when initiated in adults<sup>65,66</sup>. On the other hand, studies on H<sub>2</sub>S exposure in *C. elegans*, to date, have typically included exposure throughout life, starting from eggs or L1 larvae<sup>25,33,67</sup>. Miller and Roth have reported too that exposure to H<sub>2</sub>S gas, when restricted to adults alone, failed to extend lifespan of *C. elegans*<sup>25</sup>. We therefore wondered if, as would be expected for a true CR mimetic, exposure to FW1256 during adulthood only would still result in lifespan benefits. To systematically explore requirements for H<sub>2</sub>S exposure during different life stages, we compared lifespan effects following exposure to 500  $\mu\text{M}$  of FW1256 during either larval or adult stages only, to benefits seen with lifelong exposure. We found that FW1256 exposure beginning at larval stage (L1) was necessary but



**Fig. 5** Effects of FW1256 on metabolic profiles and oxidative stress in *C. elegans*. **a–c** Comparison of metabolic parameters in WT (unexposed), *eat-2* mutants (unexposed), WT and *eat-2* exposed to FW1256 (500  $\mu\text{M}$ ). **a** Basal respiratory capacity ( $n = 3$  biological repeats, one-way ANOVA with Bonferroni's post-test, \*\* $p < 0.01$ , \*\*\* $p < 0.001$ ), **b** maximal respiratory capacity ( $n = 3$  biological repeats, one-way ANOVA with Bonferroni's post-test, \* $p < 0.05$ ), **c** ATP levels ( $n = 3$  biological repeats,  $t$ -test, \*\* $p < 0.01$ ). **d–g** Measurement of oxidative stress in WT. **d** Mitochondrial DNA oxidative damage marker was measured by quantitative PCR ( $n = 3$  biological repeats, mean of three separate experiments normalized to average of control,  $t$ -test). **e** Reactive oxygen species (ROS) production determined by mitoSOX red fluorescence ( $n = 2$  biological repeats, mean of two separate experiments normalized to average of control,  $t$ -test). **f**, **g** general oxidative DNA damage marker, 8-OHdG (**f**) and oxidative RNA damage marker, 8-OHG (**g**) normalized to average of control ( $n = 3$  biological repeats,  $t$ -test, \*\* $p < 0.01$ ).



**Fig. 6** FW1256 acts differently from CR. **a** Survival curves of WT *C. elegans* upon exposure to 500 μM of FW1256 at different developmental stages: Larval only, adult stage only and entire lifespan ( $n = 47\text{--}102$  animals per condition, log-rank tests,  $*p < 0.05$ ,  $***p < 0.01$ ). **b** Percentage change in mean lifespan relative to control ( $n = 47\text{--}102$  animals per condition, mean lifespans were analyzed using OASIS2,  $***p < 0.001$ ). **c** Venn diagram of genes differentially expressed following exposure to FW1256 or in *eat-2* mutants, relative to WT controls (Hypergeometric test,  $***p < 0.001$ ). **d** Heatmap and cluster analysis of all genes with expression differences in unexposed WT, WT exposed to FW1256 and unexposed *eat-2* mutants. **e** PCA of unexposed WT, WT exposed to FW1256 and unexposed *eat-2* mutants. **f** Venn diagram of genes downstream of *Daf-16/FOXO* that are affected following exposure to FW1256 or in *eat-2* mutants (Hypergeometric test, non-significant,  $p > 0.05$ ).

not sufficient for longevity increase in *C. elegans* (Fig. 6a). In fact, exposure during the adult stage alone (starting after L4) was detrimental to lifespan (Fig. 6a), resulting in a significant reduction of mean lifespan (mean lifespan:  $20.2 \pm 0.5$  days, compared to control:  $21.6 \pm 0.5$  days,  $p < 0.001$ ) (Fig. 6b, Supplementary Table 2) while lifelong exposure to FW1256 showed significant lifespan extension effect (mean lifespan:  $24.5 \pm 0.6$  days, compared to control:  $21.6 \pm 0.5$  days,  $p < 0.001$ ) (Fig. 6b, Supplementary Table 2) and exposure during the larval stages only was neither detrimental nor beneficial ( $p > 0.05$ , Fig. 6a, b, Supplementary Table 2).

FW1256 is different from CR at the transcriptional level

To further compare effects of FW1256 with those of *eat-2* mutation, we used transcriptomic analysis (RNAseq) to determine sets of differentially expressed genes (DEG) for FW1256-exposed WT and *eat-2* mutants, relative to unexposed WT animals. Comparing these DEGs showed that 2848 of 7867 genes (36%) significantly affected by FW1256 were also regulated in *eat-2* mutants, while 43% of genes in the DEG of *eat-2* mutants were also affected by exposure to FW1256 (Fig. 6c). This level of overlap is statistically significantly higher than expected by chance ( $p < 0.001$ ) (Fig. 6c). However, since both interventions extend lifespan, it may not be surprising that they share some of the same genetic targets. A heatmap analysis further revealed that many of the gene expression changes induced by FW1256-exposure were different in direction and magnitudes from the gene expression changes induced by *eat-2* mutation (Fig. 6d). Principal component analysis (PCA) further revealed that gene expression changes following FW1256-exposure and *eat-2* mutation were well-separated, being far from each other in transcriptional space. Interestingly, in the PCA, WT exposed to FW1256 and *eat-2* mutants lie in opposite directions relative to unexposed WT, suggesting that FW1256 does not mimic global transcriptional changes of *eat-2* mutants but affects major parts of the transcriptome in a dissimilar or opposite way as CR (Fig. 6e). One of the key transcription factors thought to mediate CR benefits is *Daf-16/FOXO*<sup>68</sup>. Among 862 genes that were downstream of *Daf-16/FOXO*, 52 genes were regulated by FW1256 while 269 genes were regulated in *eat-2* mutants, with only 18 genes being affected by both FW1256 and *eat-2* mutation (Fig. 6f). This level of overlap is non-statistically significantly higher than expected by chance ( $p > 0.05$ ) (Fig. 6f). Taken together, these data show that, while FW1256 affects some of the same downstream genes as *eat-2* mutation, global gene expression changes are dissimilar between FW1256-exposed animals and *eat-2* mutants, suggesting that FW1256 does not mimic CR at the transcriptional level.

## DISCUSSION

Exposure of *C. elegans* to H<sub>2</sub>S gas at 50ppm extends mean lifespan by up to 70%<sup>25</sup>. Here show that a novel slow-releasing H<sub>2</sub>S donor, FW1256, which releases H<sub>2</sub>S more rapidly than previous compounds, extended mean lifespan by up to 51% (with an average of 33.8% across 5 experimental repeats). The observation that FW1256 is non-toxic at 500  $\mu$ M is consistent with a previous report that high concentrations (up to 500  $\mu$ M) of FW1256 are non-toxic in human lung fibroblasts<sup>38</sup>. Exposure to FW1256 also promoted health and reproduction in *C. elegans*. Although evolutionary theories imply that links between longevity benefit and fitness tradeoffs are obligatory<sup>69,70</sup>, our data show that FW1256 caused fewer fitness costs, at least on those aspects of fitness that we tested.

Given the evidence that H<sub>2</sub>S plays a role in mediating CR-associated benefits<sup>17</sup>, we used FW1256 to elucidate the function of H<sub>2</sub>S in the context of CR and to evaluate the capability of H<sub>2</sub>S donor compounds to replicate CR-associated effects in *C. elegans*.

While FW1256 replicated some of the phenotypes of *eat-2* mutants, our data suggest that exogenous H<sub>2</sub>S does not completely mimic CR. We found that (i) FW1256 did not delay developmental rate substantially, (ii) FW1256 only slightly delayed reproduction but, in contrast to *eat-2* mutation, (iii) FW1256 did not reduce reproductive capacity and (iv) FW1256 did not affect the final body length of nematodes. FW1256 promoted further lifespan and healthspan extension in *eat-2* mutants and FW1256 also promoted healthy longevity with less severe fitness costs than seen in *eat-2* mutants. Furthermore, in contrast to CR, H<sub>2</sub>S exposure throughout the entire lifespan was required for lifespan benefits. These data imply that H<sub>2</sub>S is involved in regulating CR-associated effects, but that its regulatory function is more complex than appreciated previously. Finally, transcriptomics analysis showed that global transcriptional changes between WT exposed to FW1256 and *eat-2* mutants were different. This further confirmed that FW1256 and CR evoke mostly independent or only partially overlapping pathways to elicit their lifespan extension effect.

Our findings suggest that FW1256 elevated rather than reduced oxidative stress in some compartments, although only 8-OHdG was elevated significantly. The effect of FW1256 on mitochondrial ROS as evaluated by mitoSOX contrasts with previous results showing that another H<sub>2</sub>S releasing drug, GYY4137, reduces mitoSOX levels substantially<sup>33</sup>, suggesting that H<sub>2</sub>S does not always act as an antioxidant.

In conclusion, we have validated FW1256, a novel slow-releasing H<sub>2</sub>S donor, as a tool for the delivery of exogenous H<sub>2</sub>S. We found that delivery of exogenous H<sub>2</sub>S by FW1256 extended lifespan and promoted healthy ageing in *C. elegans*. These lifespan and healthspan benefits were of a similar magnitude as those typically resulting from CR. This is consistent with previous reports that H<sub>2</sub>S may be a key mediator of lifespan benefits associated with CR. However, the present work suggests that H<sub>2</sub>S releasing donors do not mimic CR completely. Since the benefits of H<sub>2</sub>S exposure on lifespan and healthspan are associated with fewer fitness tradeoffs, research into the role of H<sub>2</sub>S in longevity may reveal novel ways to modulate ageing.

## METHODS

### Maintenance of *C. elegans*

The following *C. elegans* strains were used in this study: wild type (N2), RB839 (*cbs-2*), VC2569 (*ctt-1*), OK2040 (*mpst-1*), DA1116 (*eat-2*). The nematodes were maintained and exposed to H<sub>2</sub>S donor compounds (FW1251 and FW1256) according to the protocol as described previously<sup>33</sup>. FW1251 and FW1256 were synthesized as described elsewhere<sup>38</sup> and drugs were supplied by Professor Brian William Dymock and Dr. Feng Wei from the Department of Pharmacy, Faculty of Science, National University of Singapore. Briefly, synchronous cultures of nematodes were obtained by hypochlorite treatment of gravid adult nematodes as described previously<sup>71</sup>. Eggs obtained from hypochlorite treatment were allowed to hatch in M9 buffer with rotation at 20 rpm overnight in order to obtain synchronized larval stage 1 (L1) nematodes. Synchronized L1 stage larvae were then maintained in the liquid medium containing M9 buffer, *Escherichia coli* OP50-1, and streptomycin (200  $\mu$ g/ml) in the presence or absence of H<sub>2</sub>S donor compounds for 48 h at 20 °C. Thereafter, nematodes were transferred and cultivated on freshly prepared standard petri-dishes containing nematode growth medium (NGM) agar seeded with *E. coli* OP50-1 (see <sup>58</sup>) (For more information regarding freshly prepared NGM agar and *E. coli* OP50-1, see Supplementary Fig. 9 and Supplementary Fig. 10) with or without addition of H<sub>2</sub>S donor drugs.

### Lifespan determination assay

Lifespan assays were carried out as described elsewhere using randomization and operator blinding<sup>72</sup>. Nematodes were scored as live or dead and surviving nematodes were transferred to fresh NGM agar plates every 1 to 2 days. Death was scored based on failure to respond to gentle prodding with inoculation loop. Nematodes that died due to internal hatching, crawled off plates or lost were censored.

### Mobility status

Locomotion of 200 nematodes in each condition was assessed simultaneously as previously described<sup>49,73</sup>. The locomotion patterns of nematodes were classified into 3 classes. Class A animals moved constantly, class B animals only moved when prodded while class C animals showed movement of their head and tail only. Both Class A and B nematodes were scored as “healthy” whereas class C and dead nematodes were scored as unhealthy<sup>73</sup>. The progression from healthy to non-healthy classes was scored and the rate was utilized as the index of mobility status in each population.

### Egg laying studies

10 larvae from each condition were transferred onto individual NGM agar plates with or without FW1256 (500  $\mu$ M). Thereafter, each nematode was transferred to a new NGM agar plate daily until egg laying ceased. Offspring were allowed to develop at room temperature and the number of progeny was quantified 2 days later.

### Nematode length analysis

For each condition, photographs of 10 nematodes were taken at each timepoint using a calibrated Leica MZ10F microscope. Body length of nematodes was quantified using the free curve tool, provided by the Leica Application Suite software (v2.6.0 R1).

### Developmental rates studies

The times required by about 30 to 50 eggs in each condition to become egg-laying adults (first day of adulthood) were observed.

### H<sub>2</sub>S detection in *C. elegans*

**Body volume:** Images of adult nematodes were taken beforehand, using a calibrated Leica MZ10F microscope. Body length and body width of nematodes were quantified using the free curve tool provided by the Leica Application Suite software (v2.6.0 R1). The body volume of nematodes was determined using the formula for the volume of a cylinder,  $\pi r^2 h$ .

**H<sub>2</sub>S detection:** Adult nematodes were incubated in M9 buffer containing 50  $\mu$ M of H<sub>2</sub>S sensor, 7-azido-4-methylcoumarin (AzMC) (Sigma-Aldrich) at 20 °C for 2 h. AzMC fluorescence signal were visualized using confocal laser scanning microscope (Zeiss LSM800). The fluorescence intensity was then quantified using ImageJ software and normalized to body volume of nematodes.

### Quantification of ATP levels

ATP levels in 300 nematodes were measured as described previously<sup>58</sup>. Briefly, nematodes were collected and flash frozen. Frozen nematodes were lysed in trichloroacetic acid, followed by centrifugation at 15,000 *g* for 5 min at 4 °C. 5  $\mu$ l of ATP standards or supernatants of samples were added into a white 96-well microtiter plate containing arsenite ATP buffer. Thereafter, ATP levels were quantified using a luminometer (Synergy H1, Biotek) preprogrammed to inject firefly lantern extract (2 mg/ml).

### mRNA quantification *via* real time polymerase chain reaction

Total RNA was extracted from day 1 adult nematodes using RNeasy Micro kits from Qiagen and was reverse transcribed into cDNA using oligo(dT) priming according to manufacturer's protocol (GoScript Reverse Transcription System, Promega). Real time PCR was performed using PowerUp SYBR Green Master Mix (Life Technologies) on ViiA7 real time PCR system (Applied Biosystems). Relative fold change was determined by 2<sup>- $\Delta\Delta$ CT</sup> method and normalized to housekeeping gene, *pmp-3*. Primer sequences as listed below were taken from<sup>43,74</sup>.

*sqr-1* Forward primer: GTGATCCTCGCAGAATTTGG  
*sqr-1* Reverse primer: GCTGGTCCATCCAGTATCC

*eth-1* Forward primer: TCAGTGCTCAGTTCAAATCG  
*eth-1* Reverse primer: TGCAGATCTCAATGAATGTTC

*pmp-3* Forward primer: TGGCCGGATGATGGTGTCCG  
*pmp-3* Reverse primer: ACGAACATGCCAAAGGCCAGC

### Mitochondrial DNA oxidative damage determination

Oxidative damage to mitochondrial DNA was determined using XL-PCR as described elsewhere<sup>64</sup>. Mitochondria were extracted and purified using Prepman Ultra Sample Preparation Reagent (Applied Biosystems). Thereafter, real time PCR was performed using GeneAMP XL PCR kit (Applied Biosystems) to assess sequence-specific mitochondrial DNA damage, in which 6.3 kb region of the mitochondrial genome was assessed using SYBR green dye (primer sequences taken from ref.<sup>75</sup>) and 71 bp region was assessed using Taqman probe

(Forward primer: GAGCGTCATTATTGGGAAGAAGA  
 Reverse primer: TGTGCTAATCCCATAAATGTAACCTT).

### ROS quantification

ROS production in *C. elegans* was measured as described<sup>33</sup> by using MitoSOX Red mitochondrial superoxide indicator (Life Technologies). 100 nematodes were transferred manually into each well of a black 96-well microtiter plate containing 100  $\mu$ l of M9 buffer and 100  $\mu$ l of 20  $\mu$ M MitoSOX red reagent. Thereafter, ROS-associated fluorescence levels were measured every 2 min for 5 h using a fluorescence plate reader (Synergy H1 multimode microplate reader, Biotek) at excitation 396 nm and emission 579 nm at room temperature.

### Respiratory capacity determination

Respiratory capacity in live nematodes was determined as described elsewhere<sup>57</sup>. Briefly, 10 nematodes were transferred into each well of a XF96 microplate containing 200  $\mu$ l of M9 buffer. Thereafter, oxygen consumption rates (OCR) were measured using XF96 extracellular flux analyzer (Seahorse Bioscience) according to manufacturer's instruction with the injection of 25  $\mu$ l of 90  $\mu$ M carbonyl cyanide 4-(trifluoromethoxy) phenylhydrazone (FCCP) (Sigma-Aldrich) and 25  $\mu$ l of 500 mM sodium azide (Sigma-Aldrich) sequentially. Basal respiration and maximal respiration were determined *via* quantifying area under curve of OCR with and without addition of FCCP.

### 8-OHdG and 8-OHG measurement

Frozen nematodes were sonicated in lysis buffer and DNA was extracted with phenol-chloroform. 100  $\mu$ g of DNA (Tris pH8) and heavy labelled internal standards were mixed and hydrolyzed according to<sup>76</sup>. Samples were deproteinized with methanol and evaporated under nitrogen. Hydrolyzed nucleosides were dissolved in water for analysis using an Agilent 1200 System coupled to 6460 ESI tandem MS. 10  $\mu$ l of the samples were injected into Accucore C18 (150  $\times$  3.0 mm, Thermo Scientific) at 30 °C. Solvent A was Acetonitrile and Solvent B was 0.1% Formic Acid. Chromatographic separation was carried out at 0.5 ml/min using the following gradient elution: 1.5 minutes of 98% B followed by 4 min gradient decrease to 95% B, wash with 5% A for 2.5 min before 98% B for equilibration. Total run time was 12 minutes, with 8-OHG, dG, and 8-OHdG eluted at 3.6, 3.3 and 4.7 min, respectively. Mass spectrometry was carried out under positive ion ESI and multiple reaction monitoring (MRM) mode, at 3000 V and 50 psi, 350 °C, 12 L/min nitrogen nebulizer. Ultra-high purity nitrogen was used as collision gas. The compound product ion transitions are listed in Supplementary Table 3.

### RNA sequencing

Approximately 1000 of adult nematodes were harvested and total RNA was extracted using RNeasy Micro kits from Qiagen. Extracted RNA was thereafter sent to NovogeneAIT Genomics Singapore for library prep and sequencing using Illumina HiSeq4000 sequencing platform (Illumina) in a paired end read approach at a read length of 150 nucleotides. The RNAseq reads from each sample were mapped to the reference *C. elegans* transcriptome (WBcel235) with *kallisto* (v0.46.0)<sup>77</sup>. The estimated counts were imported from *kallisto* to the R environment (v3.6) and summarized to gene-level in length scaled TPM units using the *tximport* package (v1.12.3)<sup>78</sup>. The *DESeq2* package (v1.24.0)<sup>79</sup> was used to identify differentially expressed genes (DEGs) while the *mixOmics* package (6.8.0)<sup>80</sup> was used to obtain PCA plot and heatmap.

### Statistical analysis

All data were analyzed using GraphPad Prism version 5.02 software except for mean lifespan data. Lifespan curves were plotted using Kaplan–Meier

survival curves and analyzed using log-rank tests while mean lifespans were plotted using GraphPad Prism version 5.02 software and analyzed using OASIS 2 (Online Application for Survival Analysis 2; <https://sbi.postech.ac.kr/oasis2>)<sup>81</sup>. All other data are presented and plotted as mean  $\pm$  SEM of at least three separate experiments, analyzed using one-way ANOVA and Bonferroni's multiple comparisons post-test, unless otherwise noted. Statistical differences with  $p < 0.05$  were considered significant.

## Reporting Summary

Further information on research design is available in the Nature Research Reporting Summary linked to this article.

## DATA AVAILABILITY

The data that support the findings of this study are available from the corresponding author upon request. All RNA-Seq data were deposited in the National Center for Biotechnology Information Gene Expression Omnibus (GEO) (accession number GSE146412).

Received: 18 March 2019; Accepted: 12 March 2020;

Published online: 10 June 2020

## REFERENCES

- United Nations. *World Population Ageing 2017. World Population Ageing 2017* (2017).
- Jiang, J., Jagura, E., Repnevskaya, M. & Jazwinski, S. An intervention resembling caloric restriction prolongs life span and retards aging in yeast. *FASEB J.* **14**, 2135–2137 (2000).
- Chippindale, A. K., Leroi, A. M., Kim, S. B. & Rose, M. R. Phenotypic plasticity and selection in *Drosophila* life-history evolution. I. Nutrition and the cost of reproduction. *J. Evol. Biol.* **6**, 171–193 (1993).
- Lakowski, B. & Hekimi, S. The genetics of caloric restriction in *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. USA* **95**, 13091–13096 (1998).
- Weindruch, R., Walford, R. L., Fligiel, S. & Guthrie, D. The retardation of aging in mice by dietary restriction: longevity, cancer, immunity and lifetime energy intake. *J. Nutr.* **116**, 641–654 (1986).
- Colman, R. J. et al. Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science* **325**, 201–204 (2009).
- Colman, R. J. et al. Caloric restriction reduces age-related and all-cause mortality in rhesus monkeys. *Nat. Commun.* **5**, 1–5 (2014).
- Mattison, J. A. et al. Caloric restriction improves health and survival of rhesus monkeys. *Nat. Commun.* **8**, 14063–14075 (2017).
- Ravussin, E. et al. A 2-year randomized controlled trial of human caloric restriction: feasibility and effects on predictors of health span and longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **70**, 1097–1104 (2015).
- Dirks, A. J. & Leeuwenburgh, C. Caloric restriction in humans: potential pitfalls and health concerns. *Mech. Ageing Dev.* **127**, 1–7 (2006).
- Fontana, L. Aging, adiposity, and calorie restriction. *JAMA* **297**, 986 (2007).
- Hickson, M. Malnutrition and ageing. *Postgrad. Med. J.* **82**, 2–8 (2006).
- Mitchell, S. J. et al. Daily fasting improves health and survival in male mice independent of diet composition and calories. *Cell Metab.* **29**, 221–228.e3 (2019).
- Ingram, D. K. et al. Calorie restriction mimetics: an emerging research field. *Aging Cell* **5**, 97–108 (2006).
- Ingram, D. K. & Roth, G. S. Calorie restriction mimetics: can you have your cake and eat it, too? *Ageing Res. Rev.* **20**, 46–62 (2015).
- Lane, M. A., Ingram, D. K. & Roth, G. S. 2-Deoxy-D-glucose feeding in rats mimics physiologic effects of calorie restriction. *J. Anti. Aging Med.* **1**, 327–337 (1998).
- Hine, C. et al. Endogenous hydrogen sulfide production is essential for dietary restriction benefits. *Cell* **160**, 132–144 (2015).
- Hughes, M. N., Centelles, M. N. & Moore, P. K. Making and working with hydrogen sulfide. The chemistry and generation of hydrogen sulfide in vitro and its measurement in vivo: a review. *Free Radic. Biol. Med.* **47**, 1346–1353 (2009).
- Goodwin, L. R. et al. Determination of sulfide in brain tissue by gas dialysis/ion chromatography: Postmortem studies and two case reports. *J. Anal. Toxicol.* **13**, 105–109 (1989).
- Wang, R. Physiological implications of hydrogen sulfide: a whiff exploration that blossomed. *Physiol. Rev.* **92**, 791–896 (2012).
- Li, L., Hsu, A. & Moore, P. K. Actions and interactions of nitric oxide, carbon monoxide and hydrogen sulphide in the cardiovascular system and in inflammation - a tale of three gases! *Pharmacol. Ther.* **123**, 386–400 (2009).
- Wallace, J. L., Ferraz, J. G. P. & Muscara, M. N. Hydrogen sulfide: an endogenous mediator of resolution of inflammation and injury. *Antioxid. Redox Signal.* **17**, 58–67 (2012).
- Lee, Z. W. et al. Utilizing hydrogen sulfide as a novel anti-cancer agent by targeting cancer glycolysis and pH imbalance. *Br. J. Pharmacol.* **171**, 4322–4336 (2014).
- Mani, S. et al. Decreased endogenous production of hydrogen sulfide accelerates atherosclerosis. *Circulation* **127**, 2523–2534 (2013).
- Miller, D. L. & Roth, M. B. Hydrogen sulfide increases thermotolerance and lifespan in *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. USA* **104**, 20618–20622 (2007).
- Wei, Y. & Kenyon, C. Roles for ROS and hydrogen sulfide in the longevity response to germline loss in *Caenorhabditis elegans*. <https://doi.org/10.1073/pnas.1524727113> (2016).
- Ng, L. T., Gruber, J. & Moore, P. K. Is there a role of H<sub>2</sub>S in mediating health span benefits of caloric restriction? *Biochem. Pharmacol.* **149**, 91–100 (2018).
- Zheng, Y., Ji, X., Ji, K. & Wang, B. Hydrogen sulfide prodrugs—a review. *Acta Pharm. Sin. B* **5**, 367–377 (2015).
- Li, L. et al. Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): new insights into the biology of hydrogen sulfide. *Circulation* **117**, 2351–2360 (2008).
- Lee, Z. W. et al. The slow-releasing hydrogen sulfide donor, GYY4137, exhibits novel anti-cancer effects in vitro and in vivo. *PLoS ONE* **6**, 5–11 (2011).
- Karwi, Q. G., Whiteman, M., Wood, M. E., Torregrossa, R. & Baxter, G. F. Pharmacological postconditioning against myocardial infarction with a slow-releasing hydrogen sulfide donor, GYY4137. *Pharmacol. Res.* **111**, 442–451 (2016).
- Liu, Z. et al. The hydrogen sulfide donor, GYY4137, exhibits anti-atherosclerotic activity in high fat fed apolipoprotein E<sup>−/−</sup> mice. *Br. J. Pharmacol.* **169**, 1795–1809 (2013).
- Qabazard, B. et al. Hydrogen sulfide is an endogenous regulator of aging in *Caenorhabditis elegans*. *Antioxid. Redox Signal.* **20**, 2621–2630 (2014).
- Le Trionnaire, S. et al. The synthesis and functional evaluation of a mitochondria-targeted hydrogen sulfide donor, (10-oxo-10-(4-(3-thioxo-3H-1,2-dithiol-5-yl)phenoxy)decyl)triphenylphosphonium bromide (AP39). *Medchemcomm* **5**, 728 (2014).
- Szczesny, B. et al. AP39, a novel mitochondria-targeted hydrogen sulfide donor, stimulates cellular bioenergetics, exerts cytoprotective effects and protects against the loss of mitochondrial DNA integrity in oxidatively stressed endothelial cells in vitro. *Nitric Oxide* **41**, 120–130 (2014).
- Ahmad, A. et al. AP39, a mitochondrially targeted hydrogen sulfide donor, exerts protective effects in renal epithelial cells subjected to oxidative stress in vitro and in acute renal injury in vivo. *Shock* **45**, 88–97 (2016).
- Zhao, F. L. et al. AP39, a mitochondria-targeted hydrogen sulfide donor, supports cellular bioenergetics and protects against Alzheimer's disease by preserving mitochondrial function in APP/PS1 mice and neurons. *Oxid. Med. Cell. Longev.* **2016**, 8360738 (2016).
- Feng, W. et al. Discovery of new H<sub>2</sub>S releasing phosphordithioates and 2,3-dihydro-2-phenyl-2-sulfanylenebenzo[d][1,3,2]oxazaphospholes with improved antiproliferative activity. *J. Med. Chem.* **58**, 6456–6480 (2015).
- Huang, W. C. et al. A novel slow-releasing hydrogen sulfide donor, FW1256, exerts anti-inflammatory effects in mouse macrophages and in vivo. *Pharmacol. Res.* **113**, 533–546 (2016).
- Benavides, G. A. et al. Hydrogen sulfide mediates the vasoactivity of garlic. *Proc. Natl. Acad. Sci. USA* **104**, 17977–17982 (2007).
- Barardo, D. et al. The DrugAge database of aging-related drugs. *Aging Cell* **16**, 594–597 (2017).
- Chen, B. et al. Fluorescent probe for highly selective and sensitive detection of hydrogen sulfide in living cells and cardiac tissues. *Analyst* **138**, 946–951 (2013).
- Budde, M. W. & Roth, M. B. The response of *Caenorhabditis elegans* to hydrogen sulfide and hydrogen cyanide. *Genetics* **189**, 521–532 (2011).
- Feng, W. Personal Communication. (2017).
- Li, L., Rose, P. & Moore, P. K. Hydrogen sulfide and cell signaling. *Annu. Rev. Pharmacol. Toxicol.* **51**, 169–187 (2011).
- Nagahara, N., Ito, T., Kitamura, H. & Nishino, T. Tissue and subcellular distribution of mercaptopyruvate sulfurtransferase in the rat: Confocal laser fluorescence and immunoelectron microscopic studies combined with biochemical analysis. *Histochem. Cell Biol.* **110**, 243–250 (1998).
- Vozdek, R., Hnízda, A., Krijt, J., Kostrouchová, M. & Kožich, V. Novel structural arrangement of nematode cystathionine  $\beta$ -synthases: characterization of *Caenorhabditis elegans* CBS-1. *Biochem. J.* **443**, 535–547 (2012).
- Mattison, J. A. et al. Impact of caloric restriction on health and survival in rhesus monkeys from the NIA study. *Nature* **489**, 318–321 (2012).
- Herndon, L. A. et al. Stochastic and genetic factors influence tissue-specific decline in ageing *C. elegans*. *Nature* **419**, 808–814 (2002).

50. Moatt, J. P., Nakagawa, S., Lagisz, M. & Walling, C. A. The effect of dietary restriction on reproduction: a meta-analytic perspective. *BMC Evol. Biol.* <https://doi.org/10.1186/s12862-016-0768-z> (2016).
51. Lenaerts, I., Walker, G. A., Hoorebeke, L., Van, Gems, D. & Vanfleteren, J. R. Dietary restriction of *Caenorhabditis elegans* by axenic culture reflects nutritional requirement for constituents provided by metabolically active microbes. *J. Gerontol. A Biol. Sci. Med. Sci.* **63**, 242–252 (2008).
52. Szewczyk, N. J. et al. Delayed development and lifespan extension as features of metabolic lifestyle alteration in *C. elegans* under dietary restriction. *J. Exp. Biol.* **209**, 4129–4139 (2006).
53. Schulz, T. J. et al. Glucose restriction extends *Caenorhabditis elegans* life span by inducing mitochondrial respiration and increasing oxidative stress. *Cell Metab.* **6**, 280–293 (2007).
54. Lin, S. et al. Calorie restriction extends *Saccharomyces cerevisiae* lifespan by increasing respiration. *Nature* **418**, 344–348 (2002).
55. Hempenstall, S., Page, M. M., Wallen, K. R. & Selman, C. Dietary restriction increases skeletal muscle mitochondrial respiration but not mitochondrial content in C57BL/6 mice. *Mech. Ageing Dev.* **133**, 37–45 (2012).
56. Szabo, C. et al. Regulation of mitochondrial bioenergetic function by hydrogen sulfide. Part I. Biochemical and physiological mechanisms. *Br. J. Pharmacol.* **171**, 2099–2122 (2014).
57. Fong, S. et al. Identification of a previously undetected metabolic defect in the Complex II *Caenorhabditis elegans* mev-1 mutant strain using respiratory control analysis. *Biogerontology* **18**, 189–200 (2017).
58. Gruber, J. et al. Mitochondrial changes in ageing *caenorhabditis elegans* - what do we learn from superoxide dismutase knockouts? *PLoS ONE* **6**, e19444 (2011).
59. Harman, D. Free radical theory of aging. *Mutat. Res.* **275**, 257–266 (1992).
60. Ristow, M. & Zarse, K. How increased oxidative stress promotes longevity and metabolic health: The concept of mitochondrial hormesis (mitohormesis). *Exp. Gerontol.* **45**, 410–418 (2010).
61. Cypser, J. R., Tedesco, P. & Johnson, T. E. Hormesis and aging in *Caenorhabditis elegans*. *Exp. Gerontol.* **41**, 935–939 (2006).
62. Ristow, M. & Schmeisser, S. Extending life span by increasing oxidative stress. *Free Radic. Biol. Med.* **51**, 327–336 (2011).
63. Van-Raamsdonk, J., Hekimi, S., Van Raamsdonk, J. M. & Hekimi, S. Reactive oxygen species and aging in *Caenorhabditis elegans*: causal or casual relationship? *Antioxid. Redox Signal* **13**, 1911–1953 (2010).
64. Ng, L. F. et al. The mitochondria-targeted antioxidant MitoQ extends lifespan and improves healthspan of a transgenic *Caenorhabditis elegans* model of Alzheimer disease. *Free Radic. Biol. Med.* **71**, 390–401 (2014).
65. Weinruch, R. & Walford, R. L. Dietary restriction in mice beginning at 1 year of age: effect on life-span and spontaneous cancer incidence. *Science* **215**, 1415–1418 (1982).
66. Lee, G. D. et al. Dietary deprivation extends lifespan in *Caenorhabditis elegans*. *Aging Cell* **5**, 515–524 (2006).
67. Qabazard, B. et al. *C. elegans* aging is modulated by hydrogen sulfide and the sulfhydrylase/cysteine synthase *cysl-2*. *PLoS ONE* **8**, 1–12 (2013).
68. Martins, R., Lithgow, G. J. & Link, W. Long live FOXO: unraveling the role of FOXO proteins in aging and longevity. *Aging Cell* **15**, 196–207 (2016).
69. Williams, G. C. Pleiotropy, natural selection, and the evolution of senescence. *Evolution* **11**, 398–411 (1957).
70. Kirkwood, T. B. L. & Holliday, R. The evolution of ageing and longevity. *Proc. R. Soc. Lond.* **205**, 531–546 (1979).
71. Stiernagle, T. *Maintenance of C. elegans*. (2006).
72. Gruber, J., Ng, L. F., Poovathingal, S. K. & Halliwell, B. Deceptively simple but simply deceptive - *Caenorhabditis elegans* lifespan studies: considerations for aging and antioxidant effects. *FEBS Lett.* **583**, 3377–3387 (2009).
73. Schaffer, S. et al. The effect of dichloroacetate on health- and lifespan in *C. elegans*. *Biogerontology* **12**, 195–209 (2011).
74. Zhang, Y., Chen, D., Smith, M. A., Zhang, B. & Pan, X. Selection of reliable reference genes in *Caenorhabditis elegans* for analysis of nanotoxicity. *PLoS ONE* **7**, (2012).
75. Melov, S., Lithgow, G. J., Fischer, D. R., Tedesco, P. M. & Johnson, T. E. Increased frequency of deletions in the mitochondrial genome with age of *Caenorhabditis elegans*. *Nucleic Acids Res.* **23**, 1419–1425 (1995).
76. Quinlivan, E. P. & Gregory, J. F. III DNA digestion to deoxyribonucleoside: a simplified one-step procedure. *Anal. Biochem.* **15**, 383–385 (2008).
77. Bray, N. L., Pimentel, H., Melsted, P. & Pachter, L. Near-optimal probabilistic RNA-seq quantification. *Nat. Biotechnol.* **34**, 525–528 (2016).
78. Sonesson, C., Love, M. I. & Robinson, M. D. Differential analyses for RNA-seq: transcript-level estimates improve gene-level inferences. *F1000Res.* **4**, 1521 (2015).
79. Love, M. I., Huber, W. & Anders, S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.* **15**, 550 (2014).
80. Rohart, F., Gautier, B., Singh, A. & Cao, K. L. mixOmics: An R package for 'omics feature selection and multiple data integration. *PLoS Comput. Biol.* **13**, e1005752 (2017).
81. Han, S. K. et al. OASIS 2: online application for survival analysis 2 with features for the analysis of maximal lifespan and healthspan in aging research. *Oncotarget* **7**, 56147–56152 (2016).

## ACKNOWLEDGEMENTS

We are grateful to Professor Brian William Dymock and Dr. Feng Wei for providing the H<sub>2</sub>S donor compounds used in this study. We thank the *Caenorhabditis* Genetics Centre, which is funded by NIH Office of Research Infrastructure Programs (P40 OD010440) for the provision of worm strains. This work was funded by the Ministry of Education Singapore (Grant MOE2012-T2-2-003 and MOE2014-T2-2-120).

## AUTHOR CONTRIBUTIONS

L.T.N. and J.G. designed experiments. L.T.N., L.F.N., R.M.Y.T. and D.B. performed experiments and analyzed results. L.T.N., L.F.N., R.M.Y.T., D.B. and J.G. wrote the manuscript. L.T.N., L.F.N., R.M.Y.T., D.B., B.H., P.K.M. and J.G. contributed critical comments and corrections and have approved the manuscript. Work was funded by grants held by P.K.M. and J.G.

## COMPETING INTERESTS

The authors declare no competing interests.

## ADDITIONAL INFORMATION

**Supplementary information** is available for this paper at <https://doi.org/10.1038/s41514-020-0044-8>.

**Correspondence** and requests for materials should be addressed to J.G.

**Reprints and permission information** is available at <http://www.nature.com/reprints>

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020