

# Characterization of glycosyl dioxolenium ions and their role in glycosylation reactions

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Controlling the chemical glycosylation reaction remains the major challenge in the synthesis of oligosaccharides. Though 1,2-*trans* glycosidic linkages can be installed using neighboring group participation, the construction of 1,2-*cis* linkages is difficult and has no general solution. Long-range participation (LRP) by distal acyl groups may steer the stereoselectivity, but contradictory results have been reported on the role and strength of this stereoelectronic effect. It has been exceedingly difficult to study the bridging dioxolenium ion intermediates because of their high reactivity and fleeting nature. Here we report an integrated approach, using infrared ion spectroscopy, DFT computations, and a systematic series of glycosylation reactions to probe these ions in detail. Our study reveals how distal acyl groups can play a decisive role in shaping the stereochemical outcome of a glycosylation reaction, and opens new avenues to exploit these species in the assembly of oligosaccharides and glycoconjugates to fuel biological research.

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The principle challenge in chemical oligosaccharide synthesis is the stereoselective installation of glycosidic bonds<sup>1–4</sup>. Glycosidic bonds connecting monosaccharides can either exist as 1,2-*trans* or 1,2-*cis* diastereomers and the nature of the linkage has a profound influence on the structure and function of the glycans. The most common approach to chemically create glycosidic bonds is a nucleophilic substitution reaction between a glycosyl donor carrying an anomeric leaving group, and a glycosyl acceptor containing a nucleophilic alcohol. The stereochemical outcome of glycosylation reactions can be controlled using neighboring group participation (NGP)<sup>5,6</sup>. Acyl groups at the C-2 position of glycosyl donors can engage in NGP affording bicyclic C-1,C-2 dioxolenium ion intermediates that react in a stereospecific manner with glycosyl acceptors to afford 1,2-*trans* products (Fig. 1a)<sup>7</sup>. NGP of an *O*- or *N*-acyl functionality at C-2 applies to a wide variety of monosaccharides, has enabled the stereoselective synthesis of oligosaccharides both in solution and on solid support and is one of the pillars upon which chemical oligosaccharide synthesis stands<sup>8,9</sup>.

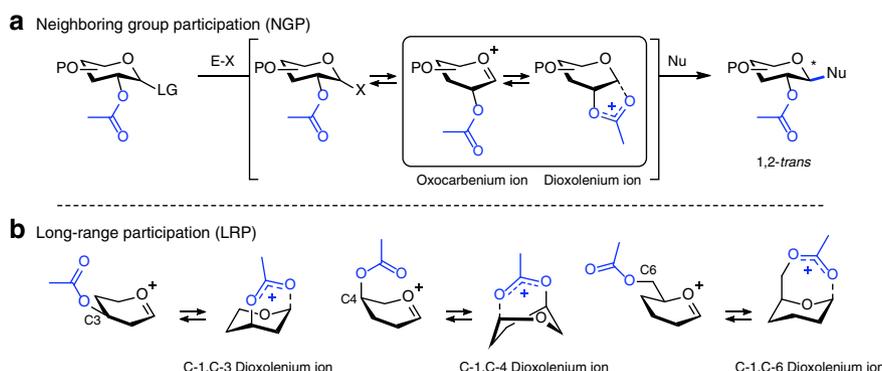
By definition, NGP by a C-2 acyl group only allows access to 1,2-*trans* glycosides and is not applicable to the synthesis of C-2-deoxy or 1,2-*cis* glycosides. Long-range participation (LRP) of acyl functionalities farther away from the anomeric center, i.e., placed on the C-3, C-4, or C-6 hydroxyl groups, has also been suggested to direct the stereoselectivity of glycosylation reactions (Fig. 1b). Importantly, LRP potentially allows for the utilization of the relative stereochemistry of C-3, C-4, or C-6 groups to control the facial selectivity in glycosylation reactions thereby enabling the stereoselective synthesis of C-2-deoxy and 1,2-*cis* glycosides. However, contradictory results have been reported and there is an ongoing debate as to the role and strength of this stereoelectronic effect<sup>10–24</sup>. Indirect proof for LRP has been derived from the stereochemical outcome of glycosylation reactions and studies using model systems<sup>24</sup>. The ability of acyl substituents positioned at C-3, C-4, or C-6 to engage in LRP likely depends on their distance and stereochemical orientation with respect to the cationic center. In addition, the relative configuration of the neighboring substituents may influence LRP by steric and electronic effects. Hence, differences in LRP of acyl groups in glycosyl donors are to be expected. Due to the instability of the intermediate dioxolenium ions and their short lifetime in solution, it is exceedingly difficult to detect and characterize them and has only been reported with respect to NGP<sup>25,26</sup>. Although glycosyl cations have been characterized in superacid solution by nuclear magnetic resonance (NMR), protonation of the acetyl groups under these conditions prevents the assessment of their ability to engage in NGP and LRP<sup>27,28</sup>. This hampers our fundamental understanding of LRP and prevents its systematic development to advance stereoselective oligosaccharide synthesis<sup>27–30</sup>.

Herein, we report the use of infrared ion spectroscopy (IRIS) to characterize glucosyl, mannosyl, and galactosyl dioxolenium ions formed via LRP and show how the stability and reactivity of these species depend on the position and configuration of the acyl group. Using density-functional theory (DFT) calculations, the conformational energy landscape (CEL) of these glycosyl cations was systematically mapped in gas and solution-phase. Finally, through a series of glycosylation reactions, employing a set of model nucleophiles of gradually decreasing nucleophilicity, we mapped the importance of the remote dioxolenium ions in glycosylation reactions. The combination of these techniques established the strength of LRP as 3-Ac-Man » 4-Ac-Gal > 3-Ac-Glu ~ 3-Ac-Gal > 4-Ac-Glu > 4-Ac-Man ~ 6-Ac-Glc/Gal/Man. The establishment of dioxolenium ion intermediates as possible reactive intermediates in glycosylation reactions opens up avenues to exploit these species for more stereoselective glycosylations.

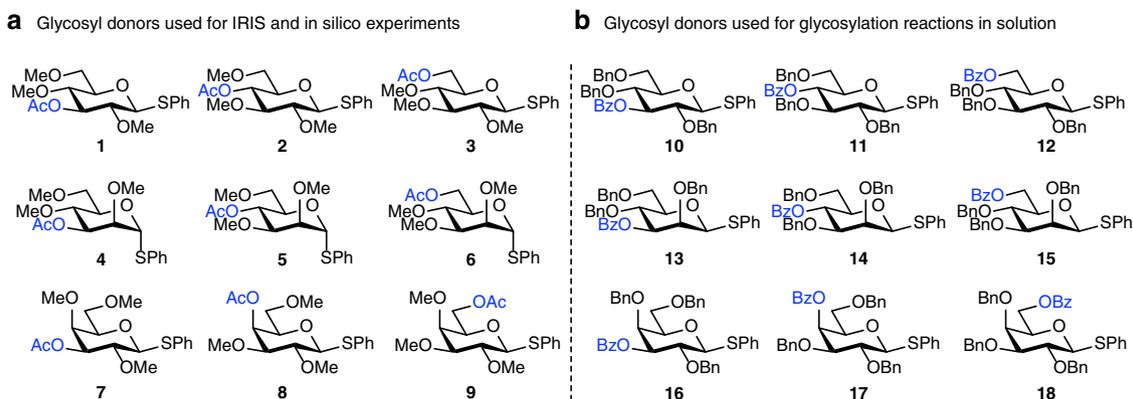
## Results

**IRIS of glycosyl cations.** Recently, we and others have employed IRIS to characterize both glycosyl oxocarbenium and dioxolenium ions in the gas-phase<sup>31–34</sup>. In this method, glycosyl donors are introduced into the mass spectrometer via electrospray ionization (ESI) and, in a tandem-mass spectrometric (MS<sup>2</sup>) scheme, glycosyl cations are formed from the isolated donors by collision-induced dissociation. This allowed us to generate the “naked” glycosyl cations in the absence of a counter ion and solvent molecules and characterized them using multi photon IRIS<sup>32,33</sup>. The IR spectra showed diagnostic vibrational bands and were used to characterize both C-1,C-2 dioxolenium ions and oxocarbenium ions<sup>32</sup>. In addition, we provided the first example of dioxolenium ions formed by LRP in uronic acid derivatives<sup>33</sup>. To systematically investigate whether LRP plays a role in glycosylation reactions we assembled two sets of glycosyl donors, derived from the most commonly used pyranosides, D-glucose, D-mannose, and D-galactose donors, and equipped these with an acyl group at either the C-3, C-4, or C-6 hydroxyl group. The first set (Fig. 2a) comprises *S*-phenyl donors equipped with methyl ethers and acetyl esters (**1–9**) used for the IRIS studies and computational studies to minimize computing costs. The second set (Fig. 2b) features benzyl ethers and benzoate esters (**10–18**) used in a matrix of model glycosylation reactions, as these represent the most commonly used protecting groups in synthetic carbohydrate chemistry. The benzyl ether and benzoate esters are structurally very similar, while differing significantly in electronic properties and their ability to stabilize an oxocarbenium ion<sup>35,36</sup>.

Glycosyl cations derived from **1** to **9** were formed from precursor ions using tandem-MS (see Supplementary Figs. 1–9).



**Fig. 1** NGP and LRP in glycosylation reactions offers an opportunity to control the stereoselectivity of glycosylations. Schematic representation of possible reactive intermediates in NGP (**a**) and LRP (**b**). P protection group, E-X promoter system, Nu nucleophile.



**Fig. 2** 2-phenyl donors equipped with ester protection groups on systematically varied positions on the ring. **a** Glycosyl donors used for IRIS experiments and DFT computations. Donors **3–9** and **13–15** were converted into their corresponding sulfoxides prior for the IRIS experiments to improve the yield of the glycosyl cation generation<sup>33</sup>. **b** Glycosyl donors used for chemical glycosylation reaction in solution.

IRIS of the glycosyl cations was carried out using the FELIX infrared free electron laser (IR-FEL) operating in the 700–1850  $\text{cm}^{-1}$  frequency range which is well suited to assign oxocarbenium and dioxolenium ion structures (Fig. 3a)<sup>37</sup>. For example, oxocarbenium ions derived from **1** to **9** can be assigned on the basis of their characteristic  $\text{C}_1=\text{O}_5^+$  stretch ( $\sim 1600 \text{ cm}^{-1}$ ) and preservation of the acetyl  $\text{C}=\text{O}$  stretch near  $1800 \text{ cm}^{-1}$ . Conversely, the formation of a dioxolenium ion leads to the absence of the acetyl  $\text{C}=\text{O}$  stretch and the  $\text{C}_1=\text{O}_5^+$  stretch and appearance of a dioxolenium ion  $\text{O}-\text{C}=\text{O}^+$  stretch ( $\sim 1550 \text{ cm}^{-1}$ ) and bending mode ( $\sim 1500 \text{ cm}^{-1}$ ). Accurate spectral assignments were made by comparing the experimental IR spectra with computed IR spectra obtained by high-level DFT calculations (B3LYP/6–31+ +G(d,p))<sup>38</sup>. The experimental IR spectra of **1–9** are presented in Fig. 3b–j (black line) together with the best matching calculated spectra (blue filled).

The IR spectra of the gluco-, manno-, and galacto-C-3 acetyl derivatives **1**, **4**, and **7** all confirm LRP of the C-3 acetyl group (Fig. 3b, e, h), as indicated by a characteristic dioxolenium  $\text{O}-\text{C}=\text{O}^+$  stretch ( $\sim 1550 \text{ cm}^{-1}$ ) and the absence of an oxocarbenium  $\text{C}_1=\text{O}_5^+$  or acetyl  $\text{C}=\text{O}$  stretch. The DFT calculated IR spectra of the formed dioxolenium ions matched well with the experimentally obtained spectra, while the computational spectra of the possible oxocarbenium ions derived from **1**, **4**, and **7** did not (Supplementary Figs. 1, 4, and 7). The calculated energy difference between the dioxolenium ions and the corresponding oxocarbenium ions also indicate LRP to be favorable for the C-3 acyl glycosides as all C-1,C-3 dioxolenium ions are lower in energy than their oxocarbenium ion counterparts (vide infra). From these experiments, it is clear that LRP of the equatorial C3-acetyl group of gluco-, manno-, and galactopyranosides is favorable.

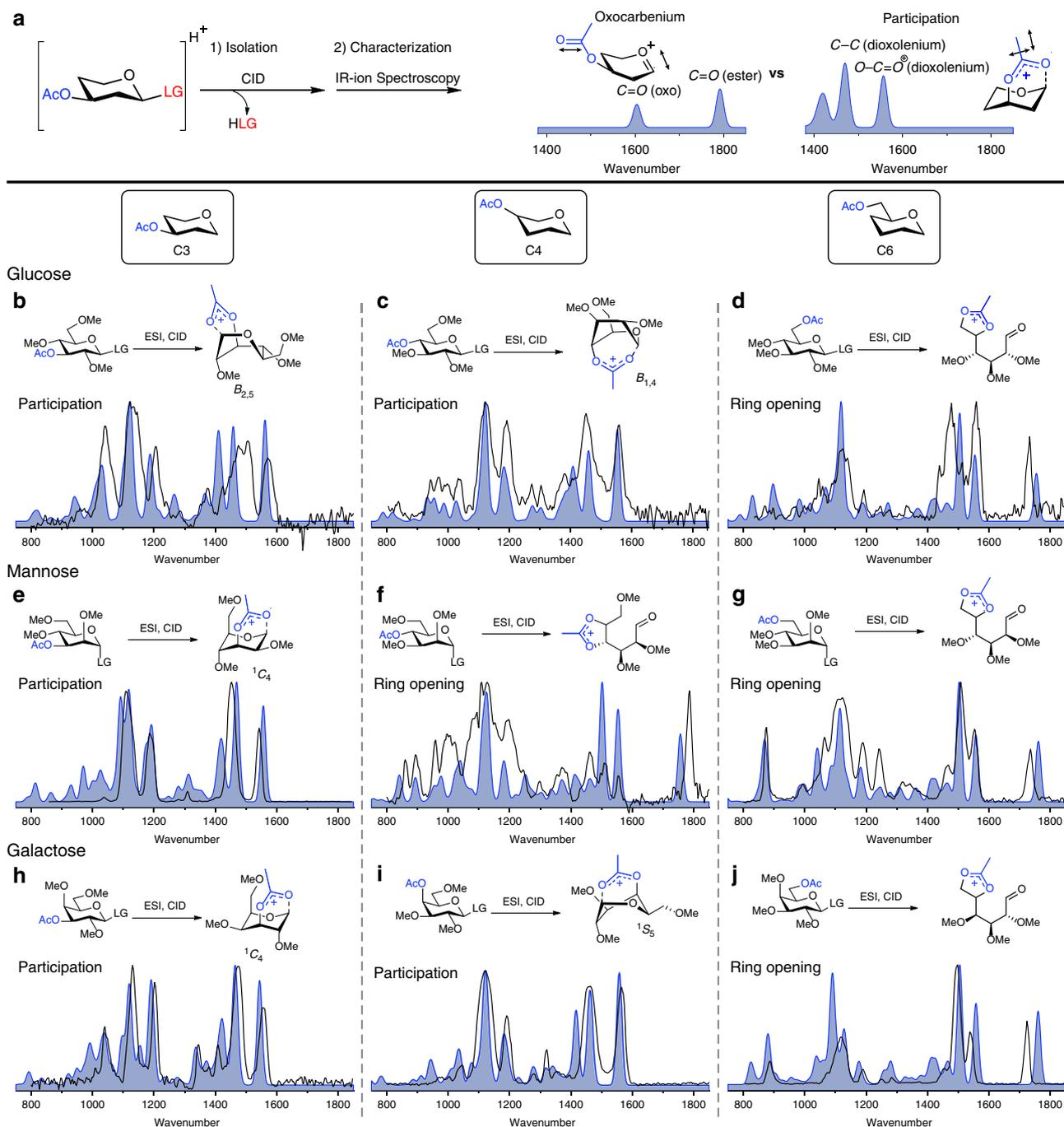
IRIS of C-4 acetyl derivatives **2**, **5**, and **8** also produced characteristic IR spectra (Fig. 3c, f, i, black line). The IRIS spectra of glucoside **2** and galactoside **8** showed an absence of a  $\text{C}=\text{O}$  ester stretch and instead showed diagnostic dioxolenium ion signals. In these cases, the agreement between the experimental and calculated IR spectra clearly indicate LRP. Alternative structures lacking LRP were calculated but did not match the experimental spectrum (Supplementary Figs. 2 and 8). The IRIS spectrum of the ion resulting from C-4 acetyl mannoside donor **5** showed distinctive dioxolenium absorptions at  $1550$  and  $1500 \text{ cm}^{-1}$  and a significant band at  $1790 \text{ cm}^{-1}$  suggesting the presence of a carbonyl functionality. A mixture of dioxolenium ions formed by LRP and oxocarbenium ions could explain this observation. However, comparing and mixing the DFT calculated spectra of these ions did not lead to a good match with the experimental spectrum

(Supplementary Fig. 5). The only structure that was in good agreement with the experimental spectrum is the dioxolenium ion, formed by attack of the C-4 acetyl on the C-5 of the initially formed oxocarbenium ion (Fig. 3f). This leads to ring opening and the formation of the C-4,C-5 dioxolenium ion with an aldehyde functionality at C-1. To the best of our knowledge, this type of rearrangement has not been reported before and the experimental conditions likely promote the formation of this species. Hence, we conclude that the C-4 acetyl in mannoside does not directly engage in LRP at the anomeric center. Taken together, these results show that the axial and equatorial C-4 esters in the glucose and galactose ions can engage in LRP, while the mannoside ion provides a C-4,C-5 dioxolenium ion.

Finally, the cations of C-6 acetyl glycosides **3**, **6**, and **9** were investigated (Fig. 3d, g, j, respectively). In all cases, a strong absorption near  $1730 \text{ cm}^{-1}$  was observed indicating the presence of a carbonyl functionality. However, a clear dioxolenium ion signature ( $\sim 1550 \text{ cm}^{-1}$ ) was also observed. Again, neither the DFT calculated IR spectra of the oxocarbenium ion or LRP dioxolenium ion nor a mixture of the two matched with the experimental spectrum (Supplementary Figs. 3, 6, and 9, respectively). Similar to the C-4 acetyl mannoside donor **5**, the experimental spectrum was matched best with calculated spectra corresponding to the ring-opened structures featuring a C-5,C-6 dioxolenium ion, producing both the aldehyde  $\text{C}=\text{O}$  and the dioxolenium  $\text{O}-\text{C}=\text{O}^+$  stretching bands. This suggests that C-6 acetyls are unlikely to provide LRP.

To verify that LRP in the methyl ether/acetyl ester protected series is similar to their benzyl ether/benzoyl ester counterparts we performed IRIS experiments with the mannosyl set **13–15** (Supplementary Figs. 10–12) and compared their spectra to those of **4–6** (Fig. 3e–g). These experiments confirmed that the LRP behavior of these two sets of glycosyl donors is the same in the IRIS experiments.

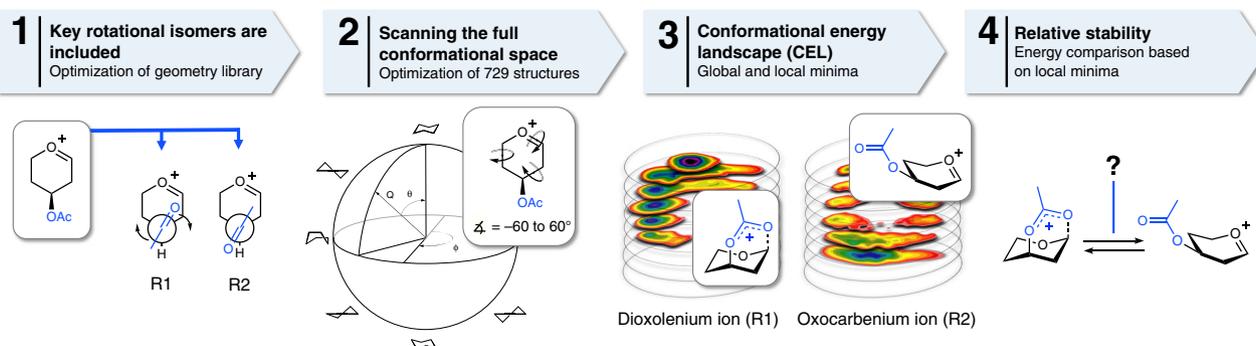
**Computing CEL of glycosyl cations.** To understand why some acetyl esters engage in LRP and lead to the formation of bicyclic dioxolenium ions from the parent oxocarbenium ions, while others do not, we computationally investigated their relative stability. We recently developed a DFT protocol to compute the relative energy of a large ensemble of oxocarbenium ion conformers, filling the complete conformational space these cations can occupy and plotted their relative energy to afford CEL maps<sup>29,39–41</sup>. Employing this method, we are able to find low energy conformers and relevant (conformational) pathways



**Fig. 3 Infrared ion spectroscopy of glycosyl cations.** **a** Oxocarbenium ions and dioxolenium ions give different diagnostic peaks (in blue). Overview of a comparison of the computed IR-ion spectra (filled blue) and the measured IR-ion spectra (black line) of the glycosyl cations derived from glucosyl (**b–d**), mannosyl (**e–g**) and galactosyl (**h–j**) donors **1–9**. Ring-opening of donors **3**, **5**, **6**, and **9** have been presented as accessible structures, their exact conformation is presented in Supplementary Figs. 3, 5, 6, and 9 and coordinates presented in the Supplementary Data file.

which connect these on the CEL. We adopted this method here (Fig. 4) to evaluate the relative stability of the oxocarbenium and dioxolenium ions derived from **1** to **9** (Fig. 5). Two rotamers of the acetyl ester were taken into account and separately visualized as a CEL map: rotamer 1 (R1) in which the acetyl is pointing toward C-1 making LRP geometrically feasible, and rotamer 2 (R2) in which the acetyl points away from C-1 making the inspection of the oxocarbenium ion possible. The geometry of all the conformers was optimized by DFT using the hybrid functional B3LYP and the basis set 6–311 G(d,p), which presents a practical trade-off between computing time and accuracy (for

more information see Supplementary Methods). To probe the difference in glycosyl cation structure between the IRIS gas-phase experiments devoid of solvent and glycosylation experiments performed in solution (vide infra), CEL maps were generated for ions formed in the gas-phase at room temperature and in solvent (computationally evaluated using a polarizable continuum model, see SI) at  $-60\text{ }^{\circ}\text{C}$  at which the experimental glycosylations take place, respectively. Figure 5a–i depicts the maps for the solution-phase ions, while Fig. 5j summarizes the relative energy of the structures found in the solution-phase and the gas-phase (all gas-phase CEL maps can be found in Supplementary Fig. 25).



**Fig. 4 Overview of the workflow to map the relative stability of glycosyl dioxolenium- and oxocarbenium ions.** (1) Two rotamers are used to probe long-range participation: R1 makes it geometrically feasible to form dioxolenium ions, where R2 generates the free oxocarbenium ion. (2) The complete conformational space of the 6-membered rings was scanned by computing 729 prefixed structures per rotamer; A few canonical conformations (Chair, half-chair, envelope, and boat) are depicted. (3) The associated energies were graphed on slices dividing the Cremer–Pople sphere; the CEL map of the R1 rotamer and CEL map of the R2 rotamer. (4) Based on the CEL maps of R1 and R2 the relative stability of both intermediates can be evaluated.

Figure 5a–c shows the solution-phase CEL maps of the C-3 acetyl protected glucosyl, mannosyl, and galactosyl dioxolenium (R1) and oxocarbenium ions (R2). The computed local minima (in dark and purple), show that formation of the dioxolenium ions is energetically favorable. For the gas-phase, the maps are similar but with larger energy differences between the lowest energy dioxolenium and oxocarbenium ions (Fig. 5j). The comparison shows that the energy difference is largest for the C-3 acetyl mannose system, with the most favorable mannose C-1,C-3 dioxolenium ion adopting a  ${}^1C_4$ -conformation. The glucose and galactose ions benefit from LRP with the  $E_2$ -glucose and  ${}^1C_4$ -galactose C-1,C-3 dioxolenium ions being 2–3 kcal/mol more stable than the corresponding lowest energy  ${}^4H_3$ -oxocarbenium ions. In the mannose system the lowest energy oxocarbenium and dioxolenium ions are close in conformational space, indicating that only a small conformational change is required for the formation of the dioxolenium ion from the oxocarbenium ion. In the glucose and galactose systems, the lowest energy oxocarbenium and dioxolenium ions are found in different regions of the conformational space. Hence, transition from the initially formed oxocarbenium ions into the more stable dioxolenium ions requires a conformational change accompanied by the crossing of a significant energy barrier. Overall the CEL maps suggest that participation by a C-3 acyl group can be favorable for all three diastereoisomeric ions studied, which is corroborated by the IRIS experiments and is most beneficial in the mannose configured C-3 acetyl system.

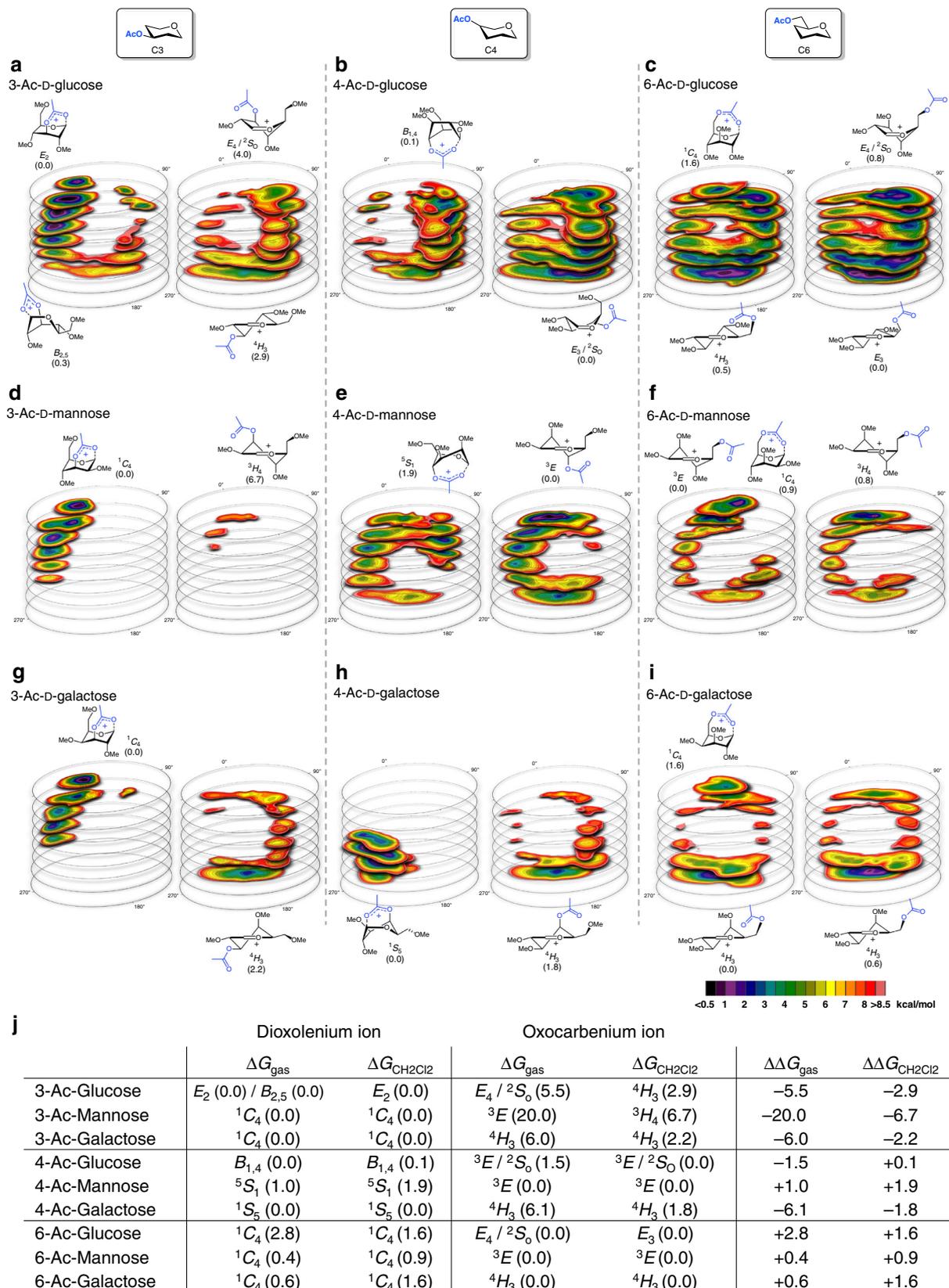
A similar analysis of the C-4 acetyl systems (Fig. 5d–f) reveals important differences between the glucose, mannose and galactose systems, again consistent with the IRIS experiments. The participation of the C-4 acetyl in the mannose ion is unfavorable, while the C-4 acetyl glucose and galactose systems benefit from LRP. The formation of the galactosyl C-1,C-4 dioxolenium ion, adopting a  ${}^1S_5$ -like structure, from the  ${}^4H_3$ -oxocarbenium ion requires only a minimal adjustment of the sugar ring conformation and is therefore facile. For the glucosyl C-4 acetyl case this requires significantly more structural rearrangement from the  $E_3$ -oxocarbenium ion to the lowest energy  $B_{1,4}$ -structure. Furthermore, the computations indicate that the glucose C-1,C-4 dioxolenium ion is more stable than the oxocarbenium ion in the gas-phase, while the relative energy of both ions is similar in solution. The relative instability of the mannosyl C-1,C-4 dioxolenium ion may be due to the *pseudo*-axial orientation of all substituents in the  ${}^5S_1$ -like structure, with the C-2 and C-3 substituents experiencing unfavorable eclipsing interactions that are not present in the glucose and galactose dioxolenium ions.

Finally, the C6-acetyl systems were probed. As can be seen from Fig. 5g–i, the energy difference between the two acyl rotamers (R1 and R2) is small, indicating that LRP does not lead to significant stabilization of the ions.

**Model glycosylation reactions.** Finally, to correlate the IRIS and CEL map findings to solution-phase experiments, we probed the influence of LRP in glycosylation reactions using 10–18 (Table 1b, entry 2–4, 6–8, and 10–12, respectively). To this end, we performed a matrix of glycosylation reactions with a set of model alcohol nucleophiles of gradually decreasing nucleophilicity<sup>43,44</sup>. The trends observed relate to changes from an  $S_N2$ -type substitution reaction of the covalent intermediate (e.g., a glycosyl triflate) for the most nucleophilic alcohols, to reactions involving more oxocarbenium character for the poorest nucleophiles (Table 1a). The glycosylation reactions were performed under pre-activation conditions using diphenyl sulfoxide ( $Ph_2SO$ )/triflic anhydride ( $Tf_2O$ ) as an activator and the results compared to donors bearing solely benzyl ether protecting groups (Table 1b, entry 1, 5, and 9)<sup>45</sup>.

The glycosylations of the C-3 benzoyl donors reveal a shift in stereoselectivity with respect to their C-3 benzyl counterparts towards the side of the  $\alpha$ -products, formed on the  $S_N1$ -side of the reaction mechanism spectrum. The change in stereoselectivity between the benzoyl/benzyl donors can be explained as arising from the LRP of the C-3 acyl groups. This shift is most pronounced in the mannose series, where all glycosylations proceed to give solely the  $\alpha$ -product (this is also observed in the glycosylation with the methyl/acetyl protected donor analog, Supplementary Table 1). These observations are in excellent agreement with the IRIS and the CEL maps results which indicate that C-1,C-3 dioxolenium ion formation is possible and most pronounced in the mannose system. For the glucose and galactose donors (Table 1b, entry 2 and 6), C-3 LRP provides less stabilization, which is reflected by the smaller impact on the  $\alpha$ -selectivity in these cases.

In contrast, the stereoselectivity of the C-4 benzoate glucose and mannose donors is virtually identical to the selectivity of the C-4 benzyl glucose and mannose donors, revealing little influence of the group present at C-4 position. The IRIS experiments and CEL maps revealed that the formation of the mannosyl C-1,C-4 dioxolenium ion is not favorable. The C-4 acyl group in mannose thus has relatively little effect on the position of the mechanistic continuum at which the substitution reactions take place and LRP can be excluded. While IRIS and the CEL maps have shown that the formation of the glucosyl C-1,C-4 dioxolenium ion is favorable in the gas-phase, the glycosylation reactions of the



**Fig. 5** CEL maps of selected glycosyl cations in which the local minima identified are shown with their respective energy<sup>42</sup>. Two acetyl ester rotamers (R1 and R2) were considered for all computed glycosyl cations generating two separate CEL maps. All energies are as computed at PCM(CH<sub>2</sub>Cl<sub>2</sub>)-B3LYP/6-311G(d,p) at 213.15 K and expressed as solution-phase Gibbs free energy. CEL maps for C3-acetyl pyranosyl ions (**a-c**), C4-acetyl pyranosyl ions (**d-f**), C6-acetyl pyranosyl ions (**g-i**). **j** Table summarizing the relative energy of the dioxolenium and oxocarbenium ion conformers in the gas- and solution-phase.

**Table 1 Experimentally found stereoselectivities for model glycosylation reactions.**

**a**

**b**

	>98:2 (41%)	75:25 (80%)	48:52 (58%)	36:64 (75%)	15:85 (70%)
	>98:2 (28%)	76:24 (91%)	58:42 (97%)	44:56 (95%)	40:60 (88%)
	>98:2 (19%)	79:21 (76%)	48:52 (70%)	34:66 (82%)	15:85 (76%)
	>98:2 (30%)	95:5 (82%)	77:23 (82%)	35:65 (83%)	17:83 (94%)
	>98:2 (39%)	>98:2 (84%)	80:20 (65%)	60:40 (75%)	33:67 (70%)
	>98:2 (61%)	>98:2 (79%)	>98:2 (87%)	>98:2 (87%)	>98:2 (94%)
	>98:2 (50%)	88:12 (77%)	71:29 (81%)	60:40 (83%)	31:69 (88%)
	>98:2 (64%)	>98:2 (56%)	78:22 (89%)	51:49 (99%)	35:65 (69%)
	>98:2 (33%)	87:13 (79%)	66:34 (69%)	31:69 (84%)	17:83 (73%)
	>98:2 (54%)	86:14 (77%)	66:34 (81%)	40:60 (82%)	41:59 (79%)
	>98:2 (55%)	97:3 (79%)	87:13 (65%)	63:37 (84%)	45:55 (86%)
	>98:2 (57%)	89:11 (74%)	61:39 (81%)	33:67 (83%)	15:85 (88%)

>90:10   >80:20   >60:40   >50:50   <50:50   <40:60   <20:80   <10:90   (α:β)

Experimental conditions: pre-activation based glycosylation conditions; nucleophile (2 eq.), Tf<sub>2</sub>O (1.3 eq.), Ph<sub>2</sub>SO (1.3 eq.), TTBP (2.5 eq.), DCM (0.05 M), -80 to -60 °C. The stereoselectivity of the reaction is expressed as alpha:beta and based on <sup>1</sup>H-NMR of the purified compounds. In all cases, the NMR spectra for both the crude and purified compounds are compared to analyze whether the measured stereoselectivity did not alter upon purification.

glucosyl donor appear to be unaffected by the nature of the C-4 substituent. The solution-phase CEL maps have revealed the C-1, C-4 dioxolenium ion, and oxocarbenium ions to be of similar energy. This may account for the moderate effect of the C-4 acetyl group on the stereochemical outcome of the glycosylation reactions. In contrast to the C-4 acyl glucose and mannose series, the C-4 acyl group in the galactose donor is capable of LRP. Both

IRIS and the CEL maps provide support for the formation of the bridged C-1,C-4 dioxolenium ion. The stability of this ion translates into the formation of more α-product in the glycosylations of the C-4 acyl galactosides.

Finally, the IRIS spectra of the C-6 acyl gluco-, manno-, and galactosyl donors provide no evidence for LRP of this functional group and also the CEL maps indicate that the formation of the

dioxolenium ion bridging the C-1 and C-6 positions does not lead to significant stabilization of the ions. The matrix of glycosylation reactions indeed shows little influence of the C-6 acyl groups. Based on this combined dataset, LRP by C-6 acyl appears to have little influence on glycosylation reactions.

In conclusion, we report a systematic evaluation of LRP in glucosyl, mannosyl and galactosyl donors bearing an acyl protecting group at their C-3, C-4, or C-6 hydroxyl group functionality. A three-pronged approach consisting of IRIS, CEL computations, and glycosylation reactions was used to assess the effect of LRP in these glycosyl donors. These studies confirm that LRP can play a decisive role in shaping the stereochemical outcome of a glycosylation reaction. LRP plays a major role in glycosylations of C-3 acyl mannosides and to a somewhat lesser extent C-4 acyl galactosides. C-3 acyl groups in glucose and galactosyl donors can engage in LRP but this anchimeric assistance has relatively little influence on the stereochemical course of glycosylations of these donors. No important role for C-6 acyl LRP has been found. The strength of LRP thus follows the order: 3-Ac-Man » 4-Ac-Gal > 3-Ac-Glu ~ 3-Ac-Gal > 4-Ac-Glu > 4-Ac-Man ~ 6-Ac-Glc/Gal/Man. The establishment of dioxolenium ion intermediates as possible reactive intermediates in glycosylation reactions opens up avenues to enhance and exploit this effect to gain stereocontrol<sup>14,46,47</sup>. These are expected to accelerate the assembly of glycoconjugates to fuel biological research.

## Methods

**Ion spectroscopy in a modified ion trap mass spectrometer.** The experimental apparatus is based on a modified 3D quadrupole ion trap mass spectrometer (Bruker, AmaZon Speed ETD) coupled to the beamline of the FELIX infrared free electron laser (IR-FEL)<sup>37</sup>. Glycosyl precursor cations ( $[M + NH_4]^+$  or  $[M + H]^+$ ) were generated by ESI from  $10^{-6}$  M solutions (in 50:50 acetonitrile:water) containing 2% ammonium acetate. An  $m/z$  of interest was isolated and collisionally activated for 40 ms in order to generate the relevant oxonium ions. Following the fragmentation reaction, the product ion  $m/z$  of interest was isolated and subsequently irradiated by the tunable mid-infrared beam. Following the absorption of a sufficient number of photons, typically occurring during a single macropulse, unimolecular dissociation results and is monitored as a frequency-dependent fragmentation (in this case, MS<sup>3</sup>) signal in the mass spectrum. Relating the precursor  $m/z$  intensity to the sum of all fragment  $m/z$  intensities in the mass spectrum recorded at each frequency position generates an infrared vibrational spectrum. IR spectra were linearly corrected for frequency-dependent variation of the laser pulse energy and the frequency was calibrated using a grating spectrometer.

**Simulation of IR spectra.** Chemical structures of the oxocarbenium, dioxolenium and rearranged ions were defined using SMILES notation for input to our workflow using the cheminformatics toolbox RDKit<sup>48</sup>. A conformational search was performed using a distance geometry algorithm, yielding 500 random 3D-conformations, which were minimized using a classical forcefield<sup>49</sup>. A maximum of 40 conformations was selected based on root-mean-squared deviation of atomic positions between geometries. Selected conformations were then submitted to Gaussian 16<sup>50</sup> for geometry optimization and vibrational analysis at a semi-empirical PM6 level. Based on the comparison of relative energies, structures were filtered using an energy cut-off of 40 kJ mol<sup>-1</sup> (for pyranosyl cations, the cut-off was increased to 80 kJ mol<sup>-1</sup>). Remaining structures were reoptimized at the B3LYP/6-31 + +G(d,p) level of theory and thereafter a harmonic frequency calculation was performed (frequencies were scaled by 0.975). In order to facilitate comparison to the experimental spectra, a 20 cm<sup>-1</sup> Gaussian broadening function was applied to the calculated vibrational lines. Finally, for each structure, electronic energies (based on the B3LYP optimized geometries) were calculated using second-order Møller–Plesset perturbation theory (MP2) and combined with enthalpy and entropy contributions resulting from the B3LYP vibrational analysis to give Gibbs free energies ( $T = 298$  K).

**CEL calculations.** A complete survey of the possible conformational space of the glycosyl cation was done by scanning three dihedral angles ranging from  $-60^\circ$  to  $60^\circ$ , including the C1–C2–C3–C4, C3–C4–C5–O, and C5–O–C1–C2<sup>29</sup>. The resolution of this survey is determined by the step size which was set to  $15^\circ$  per puckering parameter, giving a total of 729 prefixed conformations per glycosyl cation spanning the entire conformational landscape. All other internal coordinates were unconstrained. The 729 structures were computed with Gaussian 09 Rev. D.01<sup>51</sup> with DFT/B3LYP/6-311G(d,p). Furthermore, where applicable solvation effects of CH<sub>2</sub>Cl<sub>2</sub> were taken into account with a polarizable continuum model. For this specific study, two rotamers were taken into account of the C–O bond rotamer of the ring carbon and the oxygen of one of the substituents which is protected by an acyl protecting group. The CEL

maps were separately computed and visualized. The final denoted energy is expressed as free Gibbs energy. The thermodynamic corrections were computed using the quasi-harmonic approximation in the gas-phase<sup>52,53</sup>. All optimized structures were checked for the absence of imaginary frequencies. To visualize the energy levels of the conformers on the Cremer–Pople sphere, we have generated slices dissecting the sphere that combine closely associated conformers. The OriginPro software was employed to produce the energy heat maps, contoured at 0.5 kcal/mol<sup>42</sup>.

**Pre-activation Tf<sub>2</sub>O/Ph<sub>2</sub>SO-based O-glycosylation experiments.** A solution of the donor (100 μmol), Ph<sub>2</sub>SO (26 mg, 130 μmol, 1.3 eq.) and TTBP (62 mg, 250 μmol, 2.5 eq.) in DCM (2 mL, 0.05 M) was stirred over activated 3 Å molecular sieves (rods, size 1/16 in., Sigma-Aldrich) for 30 min under an atmosphere of N<sub>2</sub>. The solution was cooled to  $-80^\circ\text{C}$  and Tf<sub>2</sub>O (22 μl, 130 μmol, 1.3 eq.) was slowly added to the reaction mixture. The reaction mixture was allowed to warm to  $-60^\circ\text{C}$  in approximately 45 min, followed by cooling to  $-80^\circ\text{C}$  and the addition of the acceptor (200 μmol, 2 eq.) in DCM (0.4 mL, 0.5 M). The reaction was allowed to warm up to  $-60^\circ\text{C}$  and stirred for an additional 4–18 h at this temperature until full reaction completion was observed. The reaction was quenched with sat. aq. NaHCO<sub>3</sub> at  $-60^\circ\text{C}$  and diluted with DCM (5 mL). The resulting mixture was washed with H<sub>2</sub>O and brine, dried over MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. Purification by column chromatography yielded the corresponding O-coupled glycoside.

## Data availability

The authors declare that the data supporting the findings of this study are available within the paper and its supplementary method and data files.

Received: 31 December 2019; Accepted: 22 April 2020;

Published online: 29 May 2020

## References

- Leng, W.-L., Yao, H., He, J.-X. & Liu, X.-W. Venturing beyond donor-controlled glycosylation: new perspectives toward anomeric selectivity. *Acc. Chem. Res.* **51**, 628–639 (2018).
- Zhu, X. & Schmidt, R. R. New principles for glycoside-bond formation. *Angew. Chem. Int. Ed.* **48**, 1900–1934 (2009).
- Nigudkar, S. S. & Demchenko, A. V. Stereocontrolled 1,2-cis glycosylation as the driving force of progress in synthetic carbohydrate chemistry. *Chem. Sci.* **6**, 2687–2704 (2015).
- Bohé, L. & Crich, D. A propos of glycosyl cations and the mechanism of chemical glycosylation; the current state of the art. *Carbohydr. Res.* **403**, 48–59 (2015).
- Crich, D. & Li, M. Revisiting the armed–disarmed concept: the importance of anomeric configuration in the activation of s-benzoxazolyl glycosides. *Org. Lett.* **9**, 4115–4118 (2007).
- Poulsen, L. T., Heuckendorff, M. & Jensen, H. H. Effect of 2-O-benzoyl para-substituents on glycosylation rates. *ACS Omega* **3**, 7117–7123 (2018).
- Pittman, C. U., McManus, S. P. & Larsen, J. W. 1,3-Dioxolan-2-ylum and related heterocyclic cations. *Chem. Rev.* **72**, 357–438 (1972).
- Plante, O. J., Palmacci, E. R. & Seeberger, P. H. Automated solid-phase synthesis of oligosaccharides. *Science* **291**, 1523–1527 (2001).
- Wu, Y., Xiong, D.-C., Chen, S.-C., Wang, Y.-S. & Ye, X.-S. Total synthesis of mycobacterial arabinogalactan containing 92 monosaccharide units. *Nat. Commun.* **8**, 1–7 (2017).
- Cheng, Y.-P., Chen, H.-T. & Lin, C.-C. A convenient and highly stereoselective approach for α-galactosylation performed by galactopyranosyl dibenzyl phosphite with remote participating groups. *Tetrahedron Lett.* **43**, 7721–7723 (2002).
- Baek, J. Y., Lee, B.-Y., Jo, M. G. & Kim, K. S. β-Directing effect of electron-withdrawing groups at O-3, O-4, and O-6 positions and α-directing effect by remote participation of 3-O-acyl and 6-O-acetyl groups of donors in mannopyranosylations. *J. Am. Chem. Soc.* **131**, 17705–17713 (2009).
- Crich, D., Vinod, A. U. & Picione, J. The 3,4-O-carbonate protecting group as a β-directing group in rhamnopyranosylation in both homogeneous and heterogeneous glycosylations as compared to the chameleon-like 2,3-O-carbonates. *J. Org. Chem.* **68**, 8453–8458 (2003).
- Lei, J.-C., Ruan, Y.-X., Luo, S. & Yang, J.-S. Stereodirecting effect of C3-ester groups on the glycosylation stereochemistry of l-rhamnopyranose thioglycoside donors: stereoselective synthesis of α- and β-l-rhamnopyranosides. *Eur. J. Org. Chem.* **2019**, 6377–6382 (2019).
- Demchenko, A. V., Rousson, E. & Boons, G.-J. Stereoselective 1,2-cis-galactosylation assisted by remote neighboring group participation and solvent effects. *Tetrahedron Lett.* **40**, 6523–6526 (1999).
- Baek, J. Y. et al. Directing effect by remote electron-withdrawing protecting groups at O-3 or O-4 position of donors in glycosylations and galactosylations. *Tetrahedron* **71**, 5315–5320 (2015).

16. Ayala, L., Lucero, C. G., Romero, J. A. C., Tabacco, S. A. & Woerpel, K. A. Stereochemistry of nucleophilic substitution reactions depending upon substituent: evidence for electrostatic stabilization of pseudoaxial conformers of oxocarbenium ions by heteroatom substituents. *J. Am. Chem. Soc.* **125**, 15521–15528 (2003).
17. Ma, Y., Lian, G., Li, Y. & Yu, B. Identification of 3,6-di-O-acetyl-1,2,4-O-orthoacetyl- $\alpha$ -D-glucopyranose as a direct evidence for the 4-O-acyl group participation in glycosylation. *Chem. Commun.* **47**, 7515–7517 (2011).
18. Komarova, B. S., Orekhova, M. V., Tsvetkov, Y. E. & Nifantiev, N. E. Is an acyl group at O-3 in glucosyl donors able to control  $\alpha$ -stereoselectivity of glycosylation? The role of conformational mobility and the protecting group at O-6. *Carbohydr. Res.* **384**, 70–86 (2014).
19. Frechet, J. M. & Schuerch, C. Solid-phase synthesis of oligosaccharides. II. Steric control by C-6 substituents in glucoside syntheses. *J. Am. Chem. Soc.* **94**, 604–609 (1972).
20. Dejter-Juszynski, M. & Flowers, H. M. Studies on the koenigs-knorr reaction: Part IV: the effect of participating groups on the stereochemistry of disaccharide formation. *Carbohydr. Res.* **28**, 61–74 (1973).
21. Tokimoto, H., Fujimoto, Y., Fukase, K. & Kusumoto, S. Stereoselective glycosylation using the long-range effect of a [2-(4-phenylbenzyl)oxycarbonyl] benzoyl group. *Tetrahedron Asymmetry* **16**, 441–447 (2005).
22. Meo, C. D., Kamat, M. N. & Demchenko, A. V. Remote participation-assisted synthesis of  $\beta$ -mannosides. *Eur. J. Org. Chem.* **2005**, 706–711 (2005).
23. Hagen, B., Ali, S., Overkleef, H. S., van der Marel, G. A. & Codée, J. D. C. Mapping the reactivity and selectivity of 2-azidofucosyl donors for the assembly of n-acetylglucosamine-containing bacterial oligosaccharides. *J. Org. Chem.* **82**, 848–868 (2017).
24. Crich, D., Hu, T. & Cai, F. Does neighboring group participation by non-vicinal esters play a role in glycosylation reactions? Effective probes for the detection of bridging intermediates. *J. Org. Chem.* **73**, 8942–8953 (2008).
25. Paulsen, H., Trautwein, W.-P., Espinosa, F. G. & Heyns, K. Darstellung stabiler 1,2-acetoxonium-salze acetylierter hexosen und pentosen. *Tetrahedron Lett.* **7**, 4131–4135 (1966).
26. Crich, D., Dai, Z. & Gastaldi, S. On the role of neighboring group participation and ortho esters in  $\beta$ -xylosylation: 13C NMR observation of a bridging 2-phenyl-1,3-dioxalenium Ion. *J. Org. Chem.* **64**, 5224–5229 (1999).
27. Martin, A. et al. Catching elusive glycosyl cations in a condensed phase with HF/SbF<sub>5</sub> superacid. *Nat. Chem.* **8**, 186–191 (2016).
28. Lebedel, L. et al. Structural and computational analysis of 2-halogeno-glycosyl cations in the presence of a superacid: an expansive platform. *Angew. Chem. Int. Ed.* **58**, 13758–13762 (2019).
29. Hansen, T. et al. Defining the SN1 side of glycosylation reactions: stereoselectivity of glycopyranosyl cations. *ACS Cent. Sci.* **5**, 781–788 (2019).
30. Frihed, T. G., Bols, M. & Pedersen, C. M. Mechanisms of glycosylation reactions studied by low-temperature nuclear magnetic resonance. *Chem. Rev.* **115**, 4963–5013 (2015).
31. Mucha, E. et al. Unravelling the structure of glycosyl cations via cold-ion infrared spectroscopy. *Nat. Commun.* **9**, 1–5 (2018).
32. Elferink, H. et al. Direct experimental characterization of glycosyl cations by infrared ion spectroscopy. *J. Am. Chem. Soc.* **140**, 6034–6038 (2018).
33. Elferink, H. et al. The glycosylation mechanisms of 6,3-uronic acid lactones. *Angew. Chem. Int. Ed.* **58**, 8746–8751 (2019).
34. Marianski, M. et al. Remote Participation during Glycosylation Reactions of Galactose Building Blocks: Direct Evidence from Cryogenic Vibrational Spectroscopy. *Angew. Chem. Int. Ed.* **59**, 6166–6171 (2020).
35. Zhang, Z. et al. Programmable one-pot oligosaccharide synthesis. *J. Am. Chem. Soc.* **121**, 734–753 (1999).
36. Chang, C.-W. et al. Establishment of guidelines for the control of glycosylation reactions and intermediates by quantitative assessment of reactivity. *Angew. Chem. Int. Ed.* **58**, 16775–16779 (2019).
37. Martens, J., Berden, G., Gebhardt, C. R. & Oomens, J. Infrared ion spectroscopy in a modified quadrupole ion trap mass spectrometer at the FELIX free electron laser laboratory. *Rev. Sci. Instrum.* **87**, 103108 (2016).
38. Rijs, A. M. & Oomens, J. Gas-Phase IR Spectroscopy and Structure of Biological Molecules. (Springer International Publishing, 2015).
39. Madern, J. M. et al. Synthesis, reactivity, and stereoselectivity of 4-thiofuranosides. *J. Org. Chem.* **84**, 1218–1227 (2019).
40. van der Vorm, S. et al. Furanosyl oxocarbenium ion conformational energy landscape maps as a tool to study the glycosylation stereoselectivity of 2-azidofuranoses, 2-fluorofuranoses and methyl furanosyl uronates. *Chem. Eur. J.* **25**, 7149–7157 (2019).
41. van Rijssel, E. R. et al. Furanosyl oxocarbenium ion stability and stereoselectivity. *Angew. Chem. Int. Ed.* **53**, 10381–10385 (2014).
42. OriginPro, 9.0.0. (OriginLab Corporation, Northampton, MA, USA). <https://www.originlab.com/index.aspx?go=Company&pid=1130>.
43. Vorm, S., van der, Hansen, T., Overkleef, H. S., Marel, G. Avander & Codée, J. D. C. The influence of acceptor nucleophilicity on the glycosylation reaction mechanism. *Chem. Sci.* **8**, 1867–1875 (2017).
44. Vorm, Svander et al. Acceptor reactivity in glycosylation reactions. *Chem. Soc. Rev.* **48**, 4688–4706 (2019).
45. Codée, J. D. C. et al. Ph<sub>2</sub>SO/Tf<sub>2</sub>O: a powerful promotor system in chemoselective glycosylations using thioglycosides. *Org. Lett.* **5**, 1519–1522 (2003).
46. Liu, H. et al. Dual-participation protecting group solves the anomeric stereocontrol problems in glycosylation reactions. *Org. Lett.* **21**, 8713–8717 (2019).
47. Hahm, H. S., Hurevich, M. & Seeberger, P. H. Automated assembly of oligosaccharides containing multiple cis- glycosidic linkages. *Nat. Commun.* **7**, 1–8 (2016).
48. Landrum, G. *RDKit: Open-Source Cheminformatics* (2006). <https://www.bibsonomy.org/bibtex/28d01fcccdd6bf2486e47d7c4207b108/salotz>.
49. Tosco, P., Stiefl, N. & Landrum, G. Bringing the MMFF force field to the RDKit: implementation and validation. *J. Cheminformatics* **6**, 37 (2014).
50. Gaussian 16, Revision A.03, Frisch, M. J. et al. (Gaussian, Inc., Wallingford, CT, 2016). <https://gaussian.com/g09citation/>.
51. Gaussian 09, Revision D.01, Frisch, M. J. et al. (Gaussian, Inc., Wallingford, CT, 2009). <https://gaussian.com/g09citation/>.
52. Ribeiro, R. F., Marenich, A. V., Cramer, C. J. & Truhlar, D. G. Use of solution-phase vibrational frequencies in continuum models for the free energy of solvation. *J. Phys. Chem. B* **115**, 14556–14562 (2011).
53. Funes-Ardoiz, I. & Paton, R. S. *GoodVibes: GoodVibes v1.0.2*. (2016). <https://github.com/bobbypaton/goodvibes>

## Acknowledgements

We gratefully acknowledge the expert technical support by the FELIX staff. Quantum-chemical calculations were performed at the SurfSARA HPC center in Amsterdam; access was provided via NWO-Rekentijd grant 17603 (to J.O.) and 17569 (to T.H. and J.D.C.C.). This work was supported by an ERC-STG (758913) and NWO VIDI grant awarded to T.J.B. and an ERC-CoG (726072) and NWO VICI (VICI.182.020) grant awarded to J.D.C.C. We kindly acknowledge Mark Somers for technical support.

## Author contributions

T.H., H.E., J.O., J.D.C.C., and T.J.B. designed the experiments. T.H., H.E., J.M.A.H., K.J.H., W.A.R., A.K., G.B., S.V., and J.M., carried out the experimental work. T.H., H.E., J.M.A.H., K.J.H., W.A.R., A.K., G.B., S.V., A.M.R., H.S.O., D.V.F., F.P.J.T.R., G.A.M., J.M., J.O., J.D.C.C., and T.J.B. were involved in scientific discussions and critically reviewed the paper.

## Competing interests

The authors declare no competing interests.

## Additional information

**Supplementary information** is available for this paper at <https://doi.org/10.1038/s41467-020-16362-x>.

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**Peer review information** *Nature Communications* thanks Christian Pedersen and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

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