

DATA REPORT

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A novel *PTCH1* mutation in basal cell nevus syndrome with rare craniofacial features

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Abstract

Basal cell nevus syndrome (BCNS) is a rare, multisystem, autosomal dominant disorder that is characterized by various phenotypes, including multiple basal cell carcinomas of the skin, odontogenic keratocysts of the jaws, and occasionally cleft lip and/or palate. In this report, we describe a 6-year-old Japanese girl with a novel heterozygous nonsense mutation in *PTCH1* who exhibited rare craniofacial phenotypes, such as oligodontia and a short-tooth root.

Basal cell nevus syndrome (BCNS) (OMIM #109400), also known as Gorlin syndrome, is a rare, multisystem, autosomal dominant disorder with various phenotypes, including multiple basal cell carcinomas of the skin, palmar, and/or plantar pits and bifid ribs¹. Typical craniofacial phenotypes are also associated with BCNS, such as keratocystic odontogenic tumor (KCOT), Falx calcification, macrocephaly, hypertelorism, hydrocephalus, and cleft lip/palate, which occur at different frequencies¹. Based on these characteristic symptoms, BCNS can be discovered during orofacial examination. The prevalence of BCNS has been reported to be approximately 1 per 235,800 in the Japanese population, with an equal frequency in males and females¹, though the reported frequency varies among studies². Although most cases of BCNS are identified in patients aged between 17 and 35 years, it is sometimes diagnosed in very young children³. Genetic testing can be applied to distinguish BCNS from other syndromes and to obtain a clear prognosis.

Three different genes, *PTCH1* (OMIM *601309)⁴, *PTCH2* (OMIM *603673)⁵, and *SUFU* (*607035)⁶, have

been identified as being responsible for BCNS, with most mutations occurring in the *PTCH1* gene. Despite no genotype–phenotype correlation among the various mutations in *PTCH1*⁷, patients with *SUFU* mutations exhibit a high frequency of medulloblastoma but do not develop KCOT⁸. Treatment of BCNS includes an individualized multidisciplinary therapeutic approach. Keratocysts usually require surgical excision, and aggressive basal cell carcinomas require complete eradication. Surgical excision is supplemented by a number of other possible treatments, including cryotherapy, laser treatment, and photodynamic therapy⁹. With regard to craniofacial defects such as cleft lip/palate, which occur in 2–9.1% of cases, chelioplasty, palatoplasty, and subsequent orthodontic treatment are generally required. Approximately, 90% of patients develop KCOT at various ages, typically requiring enucleation to prevent severe tooth disruption and jaw fracture^{1,10}. Because of the widely divergent expressivity of the BCNS phenotype, continuous investigations and reports of individual cases with physical and genetic information are crucial for expanding our knowledge.

The present patient was a 6-year-old Japanese girl referred to the Osaka University Dental Hospital Department of Orthodontics for the management of malocclusion. She exhibited several symptoms, including ocular hypertelorism (Fig. 1a), bilateral cleft lip and palate

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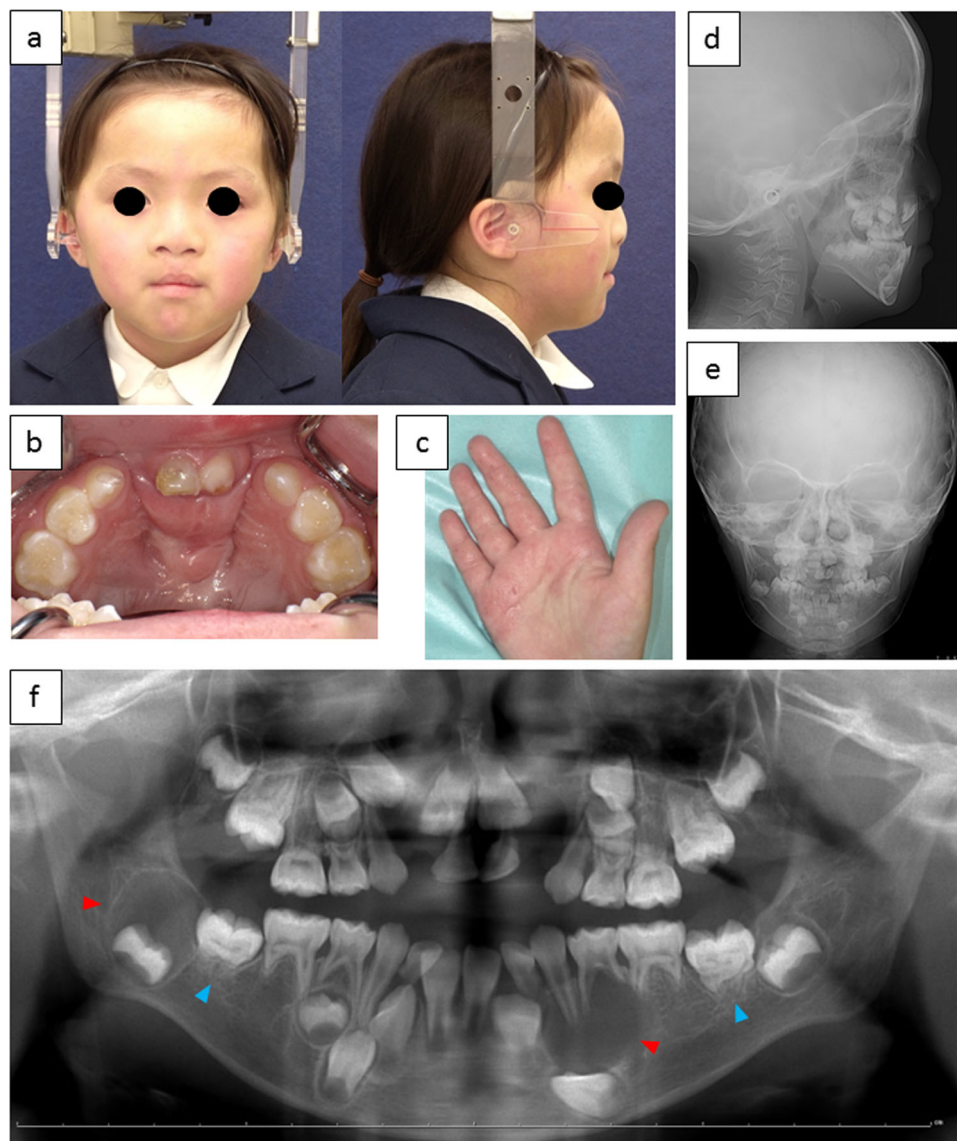
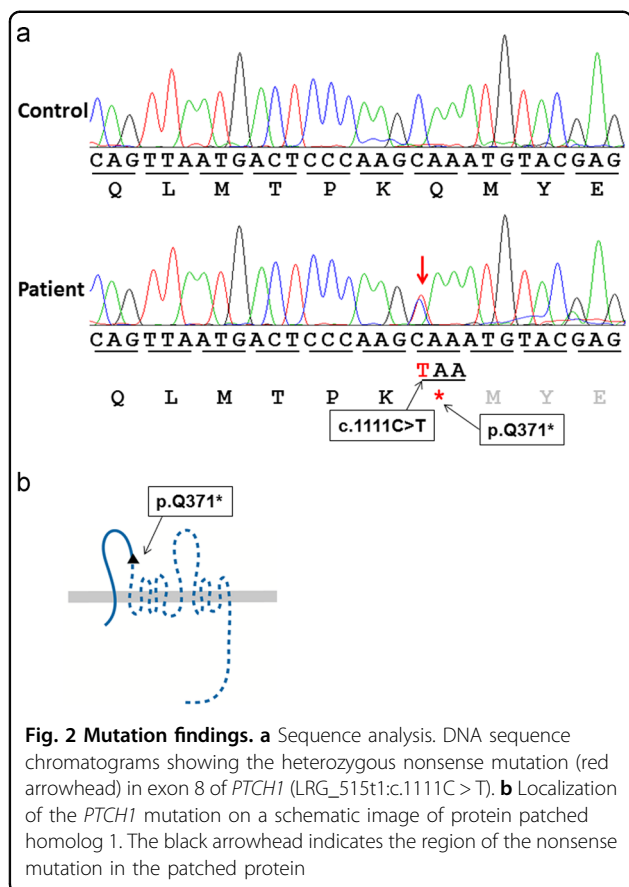


Fig. 1 Clinical features of the present BCNS patient. **a** Facial photographs. **b** Cleft lip and palate. **c** Palmar pits. **d, e** Cephalometric radiograph. **f** Panoramic radiograph. Red arrows show multiple cystic radiolucent areas, and blue arrows show root hypoplasia

(Fig. 1b), and palmar pits (Fig. 1c), with a history of cheiloplasty, palatoplasty, and surgical excision of cardiac fibroma. Craniofacial examination revealed a typical facial profile for cleft lip and palate, with midfacial deficiency and upper lip retrusion. Ensuing cephalometric analysis showed a long-cranial base and a normal jaw-base relationship (Fig. 1d, e). Intraoral examination showed an anterior open bite with bilateral cleft lip and palate, a narrow upper intercanine width and palatally inclined upper incisors. Multiple KCOTs on both sides of the mandible, one around the lower left canine, and another around the lower right second molar were observed by panoramic radiography (Fig. 1f). Interestingly, additional dental anomalies were found, such as

oligodontia (the upper lateral incisors, lower left first premolar, and upper and lower second premolars) and a truncated root of the lower first molars. None of her relatives exhibited any of these symptoms. Based on these clinical findings, we suspected that this patient had BCNS, and we suggested that the patient's family undergo genetic counseling. After explaining all the risks and benefits of genetic testing during counseling, the parents of the patient decided to receive genetic testing for BCNS. Genetic analysis approved by the Institutional Review Board of Chiba University and Kitasato University was planned to determine the gene mutation in this patient. After written informed consent had been obtained, genomic DNA was extracted from peripheral



blood, and the genomic DNA in the samples was amplified with primers targeting all exons of the *PTCH1* gene, as described previously¹¹. The amplified products were gel-purified using a QIAex II Gel Extraction Kit (Qiagen, Hilden, Germany) and cycle sequenced in both directions using a BigDye Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, CA, USA), and the sequences were analyzed using a 3130 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA). This analysis revealed a heterozygous nonsense mutation in exon 8 of *PTCH1* (NM_000264.4:c.1111C > T, p. Q371*) (Fig. 2a, b). Considering the almost complete penetrance of this disorder and the fact that neither parent presented the BCNS phenotype, the patient's mutation is suggested to be de novo. The mutation detected in this patient was not found in the 1000 Genomes or Exome Aggregation Consortium database.

The patient underwent enucleation of KCOTs at the age of 6, and histopathological examination also confirmed the diagnosis. She is now awaiting tooth eruption and subsequent correction of the malocclusion by orthodontic treatment. The present case is one example of BCNS diagnosed at a relatively young age by orofacial examination and demonstration of *PTCH1* mutation.

PTCH1 is one of the best studied receptors of the Sonic Hedgehog (SHH) ligand. In the absence of ligand, *PTCH1* functions as a repressor of SHH signaling⁴. During embryogenesis, SHH signaling plays critical roles in various tissues and organs, and thus, disturbance of this signaling pathway results in a variety of defects¹². Because *PTCH1* plays a repressive role in SHH signaling, loss of *PTCH1* function usually results in elevated activity of the pathway. In some animal experiments, craniofacial defects, including congenital cranial dysinnervation disorders, hypertelorism, cleft lip, and palate due to *PTCH1* loss of function mutations have been reported, in accord with the phenotype of the present patient^{13–15}. Cleft lip and palate is known to be a multifactorial condition, and from the high incidence in BCNS, *PTCH1* mutation is suggested to be one of the contributing genetic causes in these patients. Indeed, some GWAS studies report that the incidence of *PTCH1* mutation is high in the facial cleft population¹⁶. Interestingly, some tooth phenotypes that have not been previously reported in BCNS, such as oligodontia and short root, were observed in the present case. Shh signaling play critical roles during tooth development¹⁷, and previous studies have shown that disturbing Shh signaling results in tooth development arrest and mispatterned dentition¹⁸. Additionally, by affecting cell proliferation in the tissue called the Hertwig epithelial root sheath, which is critical for root elongation, time- and tissue-specific disruption of Shh signaling results in short-tooth roots^{19,20}. However, the mechanism of tooth root development has not been fully elucidated, and further research will be required to explain the possible mechanism of the truncated root in BCNS. Nevertheless, considering the requirement for orthodontic treatment, tooth anomalies should be taken into account if they exist in BCNS patients. The literature suggests variations in the phenotype of BCNS patients and that *SUFU* mutation does not result in KCOT⁸. This might indicate a tissue-specific requirement for different genes, even though both *PTCH1* and *SUFU* regulate the same signaling pathway. In this study, we report one case of BCNS with a novel mutation in *PTCH1* that resulted in a rare craniofacial phenotype combination. Additional reports of cases such as this will be necessary to increase our knowledge and investigate possible genotype–phenotype correlations in BCNS.

HGV database

The relevant data from this Data Report are hosted at the Human Genome Variation Database at <https://doi.org/10.6084/m9.figshare.hgv.2546>.

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Conflict of interest

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References

- Endo, M. et al. Nationwide survey of nevoid basal cell carcinoma syndrome in Japan revealing the low frequency of basal cell carcinoma. *Am. J. Med. Genet. A* **158A**, 351–357 (2012).
- Evans, D. G. et al. Birth incidence and prevalence of tumor-prone syndromes: estimates from a UK family genetic register service. *Am. J. Med. Genet. A* **152A**, 327–332 (2010).
- Kiwilsza, M. & Sporniak-Tutak, K. Gorlin-Goltz syndrome—a medical condition requiring a multidisciplinary approach. *Med. Sci. Monit.* **18**, RA145–RA153 (2012).
- Johnson, R. L. et al. Human homolog of patched, a candidate gene for the basal cell nevus syndrome. *Science* **272**, 1668–1671 (1996).
- Smyth, I. et al. Isolation and characterization of human patched 2 (PTCH2), a putative tumour suppressor gene in basal cell carcinoma and medulloblastoma on chromosome 1p32. *Hum. Mol. Genet.* **8**, 291–297 (1999).
- Pastorino, L. et al. Identification of a SUFU germline mutation in a family with Gorlin syndrome. *Am. J. Med. Genet. A* **149A**, 1539–1543 (2009).
- Wicking, C. et al. Most germ-line mutations in the nevoid basal cell carcinoma syndrome lead to a premature termination of the PATCHED protein, and no genotype-phenotype correlations are evident. *Am. J. Hum. Genet.* **60**, 21–26 (1997).
- Smith, M. J. et al. Germline mutations in SUFU cause Gorlin syndrome-associated childhood medulloblastoma and redefine the risk associated with PTCH1 mutations. *J. Clin. Oncol.* **32**, 4155–4161 (2014).
- Oseroff, A. R. et al. Treatment of diffuse basal cell carcinomas and basaloid follicular hamartomas in nevoid basal cell carcinoma syndrome by wide-area 5-aminolevulinic acid photodynamic therapy. *Arch. Dermatol.* **141**, 60–67 (2005).
- Mui Lee, L., Ma, L. & Jun, Li, T. Clinical finding and management of 12 orofacial clefts cases with nevoid basal cell carcinoma syndrome. *Cleft. Palate Craniofac. J.* **54**, 338–342 (2017).
- Fujii, K. et al. Mutations in the human homologue of Drosophila patched in Japanese nevoid basal cell carcinoma syndrome patients. *Hum. Mutat.* **21**, 451–452 (2003).
- Varjosalo, M. & Taipale, J. Hedgehog: functions and mechanisms. *Genes Dev.* **22**, 2454–2472 (2008).
- Kurosaka, H., Iulianella, A., Williams, T. & Trainor, P. A. Disrupting hedgehog and WNT signaling interactions promotes cleft lip pathogenesis. *J. Clin. Invest.* **124**, 1660–1671 (2014).
- Kurosaka, H. The roles of Hedgehog signaling in upper lip formation. *Biomed. Res. Int.* **2015**, 901041 (2015).
- Kurosaka, H., Trainor, P. A., Leroux-Berger, M. & Iulianella, A. Cranial nerve development requires co-ordinated Shh and canonical Wnt signaling. *PLoS ONE* **10**, e0120821 (2015).
- Mansilla, M. A. et al. Contributions of PTCH gene variants to isolated cleft lip and palate. *Cleft Palate Craniofac. J.* **43**, 21–29 (2006).
- Dassule, H. R., Lewis, P., Bei, M., Maas, R. & McMahon, A. P. Sonic hedgehog regulates growth and morphogenesis of the tooth. *Development* **127**, 4775–4785 (2000).
- Cho, S. W. et al. Interactions between Shh, Sostdc1 and Wnt signaling and a new feedback loop for spatial patterning of the teeth. *Development* **138**, 1807–1816 (2011).
- Nakatomi, M., Morita, I., Eto, K. & Ota, M. S. Sonic hedgehog signaling is important in tooth root development. *J. Dent. Res.* **85**, 427–431 (2006).
- Liu, Y. et al. An Nfic-hedgehog signaling cascade regulates tooth root development. *Development* **142**, 3374–3382 (2015).