



Curvulaide A, a bicyclic polyketide with anti-anaerobic bacteria activity from marine-derived *Curvularia* sp.

Wei-He Liu¹ · Yi Ding¹ · Xiang Ji¹ · Fa-Liang An¹ · Yan-Hua Lu¹

Received: 9 April 2018 / Revised: 16 September 2018 / Accepted: 21 September 2018 / Published online: 26 October 2018
© The Author(s) under exclusive licence to the Japan Antibiotics Research Association 2018

Abstract

Curvulaide A (**1**), a new bicyclic polyketide, together with its known congener preussilide D, were produced by solid-state fermentation with *Curvularia* sp. IFB-Z10. The structure of **1** was elucidated using a combination of spectral methods, including extensive one-dimensional (1D) and two-dimensional (2D) nuclear magnetic resonance (NMR) data, high resolution electrospray ionization mass spectrometry (HRESIMS), experimental and calculated electronic circular dichroism (ECD) spectra. Curvulaide A (**1**) exhibited moderate anti-anaerobic bacteria activity against periodontal pathogen *Porphyromonas gingivalis* with the MIC value of 62.5 μM , and showed moderate cytotoxicity against human hepatoma cell lines BEL7402/5-Fu with IC₅₀ values of 12.46 μM .

Introduction

The rapid development of drugs resistance in human diseases such as cancers and invasive fungal infections calls for a class of structurally unique lead drug compounds and beneficial therapeutic agents with new pharmacological mechanisms¹. Marine endophytic fungi may play an important role in the discovery of unique secondary metabolites with chemical and biological properties unlike those found in terrestrial ones in recent years². Bicyclic polyketides, a class of compounds with a 4,6-dimethylhepta-2,4,6-trienoic acid substructure and decalin derivative motif from acetyl Co-A and malonyl Co-A interdispersed with units of methylmalonyl Co-A, are moderate inhibitors of angiogenesis, and inhibit proliferation of filamentous fungi and plant pathogens³. Bicyclic polyketides had been discovered from terrestrial fungal metabolites, such as the antarones

from *Penicillium antarcticum*⁴, embellistarin from *Embellisia chlamydospora*⁵, and hamigerone from *Hamigera avellanea*⁶. To discover bioactive compounds from marine fungi, a new bicyclic polyketide curvulaide A (**1**) (Fig. 1) was isolated from the culture of the marine fungus *Curvularia* sp. IFB-Z10.

Curvularia sp. IFB-Z10 was isolated from the gut flora of fish (white croaker *Argyrosomus argentatus*); it produces novel alkaloids named curvulammine⁷ and curindolizine⁸, when cultured in shake flask in the Czapek's medium. These compounds exhibited significant antibacterial and anti-inflammatory activities. In this study, we aimed to explore the "new fate" of polyketide analogs from the metabolites of *Curvularia* sp. IFB-Z10 through solid-state fermentation⁹. A new bicyclic polyketide, curvulaide A (**1**), together with its known congener preussilide D³ were produced by solid-state cultivated *Curvularia* sp. IFB-Z10. Curvulaide A (**1**) was evaluated for antitumor and anti-anaerobic bacteria activities, and showed moderate cytotoxicity against human hepatoma cell lines BEL7402/5-Fu with IC₅₀ value of 12.46 μM , and exhibited moderate anti-anaerobic bacteria activity against periodontal pathogen *Porphyromonas gingivalis* with an MIC value of 62.5 μM . Hence, we reported the isolation, structural elucidation, and biological activity evaluation of **1**.

Curvulaide A (**1**) was isolated as a pale-yellow gum, and its molecular formula was determined to be C₂₅H₃₀O₄ (11 degrees of unsaturation) based on the HRESIMS ion at *m/z* 417.2035 [M+Na]⁺ (calcd. 417.2036). In the ¹H NMR

Electronic supplementary material The online version of this article (<https://doi.org/10.1038/s41429-018-0110-7>) contains supplementary material, which is available to authorized users.

- ✉ Fa-Liang An
flan2016@ecust.edu.cn
- ✉ Yan-Hua Lu
luyanhua@ecust.edu.cn

¹ State Key Laboratory of Bioreactor Engineering, East China University of Science and Technology, Shanghai, PR China

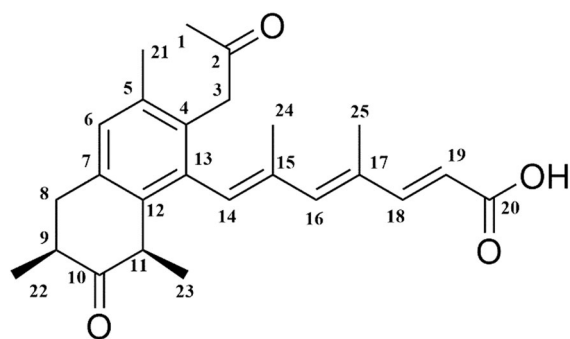


Fig. 1 The chemical structure of **1**

Table 1 The ^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) of **1** in CD_3OD

No.	δ_{H} mult. (J in Hz)	δ_{C}	No.	δ_{H} (J in Hz)	δ_{C}
1	2.08 s	29.9	14	6.47 s	132.0
2		208.8	15		138.6
3	3.88 d (18.0) 3.82 d (18.0)	46.4	16	6.48 d (4.0)	142.5
4		137.7	17		134.6
5		132.5	18	7.41 dd (15.5, 4.0)	151.6
6	7.07 s	130.0	19	5.94 d (15.5)	118.8
7		136.3	20		170.9
8	2.95 m	36.3	21	2.21 s	20.2
9	2.43 m	44.1	22	1.23 d (6.5)	15.7
10		217.9	23	1.23 d (6.5)	19.3
11	3.48 m	47.5	24	1.55 s	18.1
12		136.9	25	2.09 s	14.1
13		136.3			

spectra (Table 1), six methyl groups including four singlet methyl groups (δ_{H} 2.08, 2.21, 2.09, 1.55, each 3H), two doublet methyl groups (δ_{H} 1.23, d, $J = 6.5$ Hz, 6H) were observed. It also displayed a pair of characteristic *trans*-coupled olefinic protons (δ_{H} 7.41, d, $J = 15.5$ Hz; 5.94, d, $J = 15.5$ Hz), a doublet olefinic proton (δ_{H} 3.82, d, $J = 4.0$ Hz), and two specific singlet olefinic protons (δ_{H} 7.07, 6.47). The ^{13}C NMR spectrum (Table 1) showed two carbonyl carbons (δ_{C} 217.9, 208.8), a conjugated carbonyl carbon (δ_{C} 170.9), and twelve alkenic carbons. Thus, the aforementioned data suggested that compound **1** was a bicyclic polyketide like preussilide **C**³.

The ^1H NMR data analyses of **1** showed much similarity with those of preussilide **C**, except for the loss of two specific methine protons (δ_{H} 1.76, d, $J = 12.5$ Hz; 3.21, d, $J = 10.5$ Hz) in preussilide **C**, and the presence of two additional alkenic carbons (δ_{C} 136.3, 136.9) in **1**. The HMBC correlations from H-18 (δ_{H} 7.41, d, $J = 15.5$ Hz) to C-20 (δ_{C} 170.9)

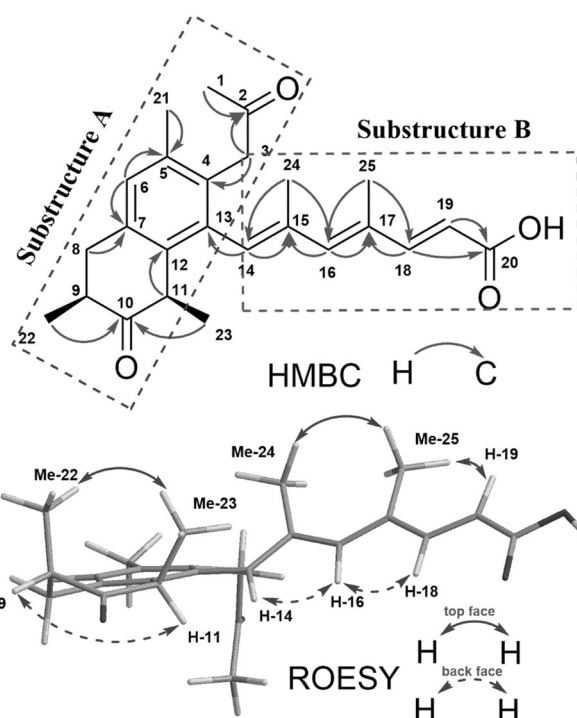


Fig. 2 Selective HMBC and ROESY correlations of **1**

and C-16 (δ_{C} 142.5), from H-16 (δ_{H} 3.82, d, $J = 4.0$ Hz) to C-14 (δ_{C} 132.0) and C-18 (δ_{C} 151.6), from Me-25 (δ_{H} 3.09, s) to C-16, C-17 (δ_{C} 142.5), and C-18 implied a 4,6-dimethylhepta-2,4,6-trienoic acid motif (substructure A) in the structure of **1**. Two specific doublet methyl groups and one singlet methyl group in ^1H NMR and the HMBC correlations from H-9 (δ_{H} 2.43, m) to Me-22 (δ_{H} 1.23, d, $J = 6.5$ Hz), H-11 (δ_{H} 3.48, m) to Me-23, H₂-8 (δ_{H} 2.95, m) to C-7 (δ_{C} 136.3), C-12 (δ_{C} 136.9), Me-22 and Me-23 to C-10 (δ_{C} 217.9) and H-11 to C-12 and C-7 indicated a decalin derivative motif (substructure B) in **1** like in preussilide **C**. The HMBC correlation between H-14 and C-13 suggested the connection point between substructure A and substructure B located at C-13. The singlet methyl (δ_{H} 2.08) and two germinal coupled protons (δ_{H} 3.82, d, $J = 18.0$ Hz; 3.88, d, $J = 18.0$ Hz) showed HMBC correlations to the ketone carbonyl carbon (δ_{C} 208.8), and this indicated a methyl ketone unit in **1** located at C-4. Thus, the planar structure of **1** was determined as shown in Fig. 2.

The relative configuration of **1** was determined based on vicinal coupling constants and ROESY correlations. The coupling constant $J_{18,19} = 15.5$ Hz indicated an *E*-configuration of this double bond. The trisubstituted double bonds $\Delta^{14,15}$ and $\Delta^{16,17}$ were assigned as *E*-configurations based on the ROESY correlations between H-16 and H-14, H-18 as well as the ROESY correlations between Me-25 and Me-24, H-19. The ROESY correlations between the H-9 and H-11 indicated Me-22 and Me-23 were co-facial, and H-11 and H-9 were on the same face of the decalin motif. Thus, the relative configuration of the decalin moiety was determined

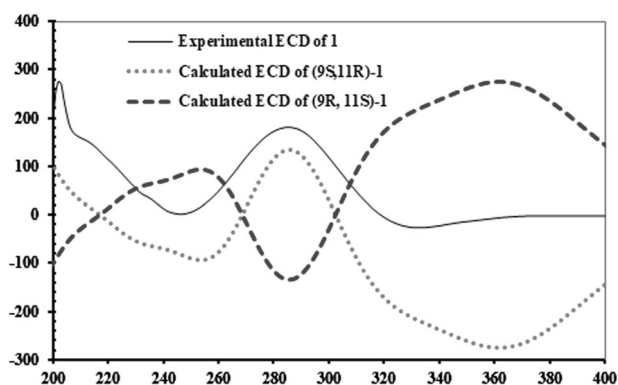


Fig. 3 Experimental and calculated ECD spectra of **1**

as shown in Fig. 1, which was consistent with the configuration of the revised preussilide A¹⁰.

Unfortunately, the curvulaide A (**1**) failed to crystallize in different solvent. To determine the absolute configuration of curvulaide (**1**), the theoretical calculation of ECD spectra was performed with the Gaussian 09 program package, which was conducted in MeOH using time-dependent density functional theory at B3LYP/6–311G** (Fig. 3), and led us to determine the absolute configuration of **1** (9S, 11R) by comparing experimental and calculated ECD spectra.

The anti-anaerobic bacteria activity was evaluated against periodontal pathogen *Porphyromonas gingivalis* by broth microdilution method in a 96-well plate¹¹. Compound **1** exhibited moderate activity with the MIC value of 62.5 μ M (tinidazole was added as the positive control with MIC value of 7.8 μ M). The cytotoxic activity of **1** was assessed against human hepatoma cell line BEL7402 and the drug-resistant cell line BEL7402/5-Fu¹², and showed effective cytotoxicity with IC₅₀ values of 19.85 μ M and 12.46 μ M, respectively¹³ [the 5-Fu was used as the positive control (BEL7402, IC₅₀: 14 μ M; BEL7402/5-FU, IC₅₀: 1630 μ M)].

In conclusion, a new bicyclic polyketide, curvulaide A (**1**), with a 4,6-dimethylohepta-2,4,6-trienoic acid substructure and decalin derivative motif, was produced by solid-state fermentation with *Curvularia* sp. IFB-Z10 using rice as substrate. Compound **1** showed moderate anti-anaerobic bacteria activity and cytotoxic activities. Therefore, the isolation of the new bicyclic polyketide **1** suggested that more unique bioactive compounds can be isolated from *Curvularia* sp. IFB-Z10, and these novel compounds could serve as lead compounds for new drug discovery.

Curvulaide A (1): pale-yellow resin, $[\alpha]_D^{23} + 10.3$ (c, 0.1, MeOH); UV (MeOH) λ_{\max} (log ϵ) 207 nm (4.56) 280 nm (4.02); ¹H and ¹³C NMR (CD₃OD, 600 MHz), see Table 1;

negative ESIMS m/z 393.2 [M – H][–]; positive ESIMS m/z 417.2 [M + Na]⁺; HRESIMS m/z 417.2035 [M + Na]⁺ (calcd for C₂₅H₃₀O₄Na, 417.2036).

Acknowledgements This work was financially supported by the NSFC grants (81703388, 81741156), the Shanghai Sailing Program (17YF1403700), the Fundamental Research Funds for the Central Universities (222201714023, 22221818014), and Talent Program of School of Biotechnology in East China University of Science and Technology.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Minotti G, et al. Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. *Pharmacol Rev.* 2004;56:185–229.
- Blunt JW, et al. Marine natural products. *Nat Prod Rep.* 2018; 34:235–94.
- Noumeur SR, et al. Preussilides A–F, bicyclic polyketides from the endophytic fungus *Preussia similis* with antiproliferative activity. *J Nat Prod.* 2017;80:1531–40.
- Shiono Y, et al. Antarones A and B, two polyketides from an endophytic *Penicillium antarcticum*. *Z Nat.* 2008;63b:909–14.
- Jung HJ, et al. Embellistatin, a microtubule polymerization inhibitor, inhibits angiogenesis both *in vitro* and *in vivo*. *Biochem Biophys Res Commun.* 2007;353:376–80.
- Breinholt J, et al. Hamigerone and dihydrohamigerone: two acetate-derived, antifungal metabolites from *Hamigera avellanea*. *Acta Chem Scand.* 1997;51:1241–4.
- Han WB, et al. Curvulamane, a new antibacterial alkaloid incorporating two undescribed units from a *Curvularia* species. *Org Lett.* 2014;16:5366–9.
- Han WB, et al. Curindolizine, an anti-inflammatory agent assembled via Michael addition of pyrrole alkaloids inside fungal cells. *Org Lett.* 2016;18:1816–9.
- Dong JW, et al. Production of a new tetracyclic triterpene sulfate metabolite sambacide by solid-state cultivated *Fusarium sambucinum* B10.2 using potato as substrate. *Bioresour Technol.* 2016; 218:1266–70.
- Pescitelli G, Di BL. Revision of the absolute configuration of preussilides A–F established by the exciton chirality method. *J Nat Prod.* 2017;80:2855–9.
- Zhang Y, et al. Antibacterial and antibiofilm activities of eugenol from essential oil of *Syzygium aromaticum* (L.) Merr. & L. M. Perry (clove) leaf against periodontal pathogen *Porphyromonas gingivalis*. *Microb Pathog.* 2017;113:396–402.
- Huang HY, et al. Reversal effect of 2',4'-dihydroxy-6'-methoxy-3',5'-dimethylchalcone on multi-drug resistance in resistant human hepatocellular carcinoma cell line BEL-7402/5-FU. *Phytochemistry.* 2011;18:1086–92.
- Reed LJ. A simple method estimating fifty percent endpoint. *Am J Epidemiol.* 1938;27:493–7.