### **ORIGINAL ARTICLE**



# Viscoelasticity of poly(ethylene glycol) in aqueous solutions of potassium sulfate: a comparison of quartz crystal microbalance with conventional methods

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#### Abstract

The viscoelasticity of poly(ethylene glycol) (PEG) in aqueous solutions with different concentrations of potassium sulfate ( $K_2SO_4$ ) was studied by quartz crystal microbalance with dissipation (QCM-D), after coating a rigid supported lipid bilayer of 1,2-dioleoyl-*sn*-glycero-3-phosphocholine on the silicon oxide substrate. The obtained viscoelastic properties of PEG in  $K_2SO_4$  solutions agree well with the Zimm model predictions for linear polymer chains. With increasing  $K_2SO_4$  concentration, the excluded volume exponent  $\nu$  of PEG decreased from 0.565 in water to 0.55 in 0.19 mol/L  $K_2SO_4$ , and 0.50 in 0.43 mol/L  $K_2SO_4$ . The solvent quality gradually worsened for PEG with increasing  $K_2SO_4$  concentration, which was verified by decreases in the polymer intrinsic viscosity and the corresponding Mark–Houwink exponent. The high-frequency characteristic of QCM-D makes it possible to directly study the viscoelasticity of polymer solutions in a low-viscosity solvent, which is complementary to conventional rheometers working at low frequency.

## Introduction

The dynamics of polymer chains in solutions or melts can be studied by measuring their viscoelastic properties in oscillatory shear flows at certain frequencies, which can probe the motion of whole chains or chain segments with the corresponding relaxation times [1, 2]. The typical upper frequency for conventional rheometers is approximately 100 Hz, which limits the application of such instruments in polymeric systems with sufficiently large relaxation times, usually for high-molecular-weight polymers with a large fluid viscosity. Time-temperature superposition has been employed to extend the frequency in a limited range by tuning the fluid viscosity via temperature [1]. It is, however, not applicable for aqueous polymer solutions because the viscosity of water weakly depends on temperature and the temperature range in which water acts as a good solvent for the studied polymers is limited. In some cases, a large

⊠ Yonggang Liu yonggang@ciac.ac.cn amount of inert sugars can be added to the aqueous polymer solution to slow down the dynamics of the polymer chains by increasing fluid viscosity [3–5]. On the other hand, the classical theories on polymer dynamics in solutions and melts predict various scaling behaviors at high frequencies, which contain rich information about the relaxation mechanism of the polymeric fluid [6–9]. Therefore, it is crucial to directly measure the high-frequency viscoelasticity of polymer solutions, which provides important insight into their structure and dynamics.

Instruments based on torsional resonators, which was pioneered by Mason [10], have been used to study the viscoelasticity of polymer solutions up to hundreds of kHz [11–13]. Microrheological techniques based on the measurement of the motion of tracer particles in fluids, such as quasielastic light scattering (QELS) and diffusing wave spectroscopy (DWS) [14, 15], have been developed to measure the viscoelasticity of polymer solutions up to frequencies of 100 kHz but are not applicable to dilute polymer solutions. Very recently, we reported the viscoelasticity measurement of aqueous polymer solutions at megahertz frequency by quartz crystal microbalance with dissipation (QCM-D) [16]. In the report, the perturbations arising from polymer adsorption on the crystal resonator were eliminated by coating the resonator with a rigid and inert supported lipid bilayer (SLB), and the viscoelastic properties of poly

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(ethylene glycol) (PEG) of different molecular weights in pure water were obtained from the shifts in the resonance frequency and the energy dissipation factor. The QCM-D method is proven to be a rapid and accurate way to study the viscoelasticity of aqueous polymer solutions at high frequency. This method only requires a small amount of samples, which is especially promising for costly or rare biological samples. It should be noted that biomacromolecules are often dissolved in an aqueous buffer containing salts. It is therefore necessary to study the feasibility of the QCM-D technique for polymer solutions in the presence of salts, especially in systems where the salt plays an important role in determining the conformation and dynamics of the polymer chains.

Here, we report the measurement of the viscoelasticity of aqueous PEG solutions containing different concentrations of potassium sulfate using QCM-D at high frequencies of 5-65 MHz. The effect of potassium sulfate concentration on polymer viscoelasticity is studied and compared with conventional measurements by an Ubbelohde viscometer and a rotational rheometer.

## Materials and methods

#### Materials

The lipid of 1,2-dioleoyl-sn-glycero-3-phosphocholine (DOPC) was obtained from Avanti Polar Lipids, Inc. (Alabaster, Alabam, USA) and used without further purification. PEGs with different weight average molecular weights (M<sub>w</sub>s) of 200, 400, 600, 1000, 1400, 2000, 4000, 6000, 8000, 10,000, 20,000, and 35,000 were purchased from Sigma-Aldrich and were desiccated under vacuum to a constant weight before experiments. The polydispersities  $(M_w/M_n s)$  of all PEG samples were less than 1.1, according to gel-permeation chromatography measurements. Three polyethylene oxides (PEOs) samples, synonymous with PEG, with higher molecular weights of 116,000, 278,000, and 443,000 were purchased from Polymer Laboratories, Ltd. Analytical-grade sodium chloride (NaCl) and potassium sulfate (K<sub>2</sub>SO<sub>4</sub>) were obtained from Sigma. Ultrapure water produced by a Sartorius water purification system with a resistivity of 18.2 M $\Omega$ ·cm was used for solution preparations.

#### Viscoelasticity via QCM-D

The viscoelasticity of aqueous PEG solutions is measured by QCM-D after coating the resonator surface with an inert layer, which is provided by the rupture of liposomes to form a rigid SLB on the silicon oxide substrate. More details of the experimental procedure can be found elsewhere [16].

The liposomes were prepared by the extrusion method [16]. Briefly, a DOPC solution in chloroform was transferred into a continuously rolled vial, and the organic solvent was completely removed by evaporation with a gentle stream of nitrogen gas, followed by 3 h of vacuum pumping. An aqueous solution containing 150 mM NaCl was added to the vial to dissolve the dried lipid film upon gentle vortexing. The solution was then extruded 31 times through a 100 nm polycarbonate membrane with a mini-extruder (Avanti Polar Lipids, Inc.) to produce large unilamellar vesicles (LUVs). The average diameter of the obtained liposomes was  $128 \pm 2$  nm with a narrow size distribution, according to dynamic light scattering measurements on a Malvern Nano ZS90 Zetasizer. Vesicle suspensions were kept at 4 °C and diluted to a lipid concentration of 0.1 mg/ mL prior to use.

The QCM-D experiments were carried out at  $24 \pm 0.02$  °C on a Q-Sense E4 system (Biolin Scientific, Sweden) [17]. Before mounting into the flow module, the AT-cut quartz crystal sensors with a fundamental resonance frequency of 4.95 MHz were cleaned twice with the following procedure: soaked in a 2% SDS solution for 30 min, rinsed with ultrapure water, blow-dried with nitrogen gas and exposed to an ultraviolet (UV)/ozone cleaner for 15 min [18]. The experiments were then performed in exchange mode by pumping at least 1.0 mL degassed sample solution into the 40 µL chamber at a fluid flow rate of 100 µL/min. The shifts in the resonant frequency  $\Delta f_n$  and the energy dissipation factor  $\Delta D_n$  of the sensors were monitored simultaneously at multiple harmonics (overtone numbers n = 1, 3, 5, 7, 9, 11, 13).

The measurements started by forming a rigid and inert SLB on the sensor, which was produced by the rupture of DOPC liposomes facilitated in an aqueous solution containing 150 mM NaCl. After establishing the baseline with the 150 mM NaCl solution, the DOPC vesicle solution was pumped into the chamber to form the SLB on the resonator surface, followed by again rinsing with the 150 mM NaCl solution. The chamber fluid was exchanged to pure water before the aqueous solution of PEG in pure water or in different concentrations of  $K_2SO_4$  was pumped into the chamber. Aqueous PEG solutions of different molecular weights were introduced into the chamber one by one, and a sufficient amount of water was pumped in between to rinse the SLB surface.

The relationship between the frequency shift  $\Delta f_n$  and the mass  $\Delta m$  of a thin and rigid film adsorbed onto the sensor is described by the Sauerbrey equation [19]:

$$\Delta m = -\frac{\left(\rho_q \mu_q\right)^{1/2}}{2f_0^2} \frac{\Delta f_n}{n} = -C \frac{\Delta f_n}{n} \tag{1}$$

where  $f_0$  is the fundamental resonance frequency of the sensor (4.95 MHz),  $\rho_q$  is the density of quartz (2.648 g/cm<sup>3</sup>),  $\mu_q$  is the

shear modulus of the crystal  $(2.947 \times 10^{11} \text{ g cm}^{-1} \text{ s}^{-2})$ , and *C* is the mass sensitivity constant (18.0 ng cm<sup>-2</sup> Hz<sup>-1</sup> at  $f_0 = 4.95$  MHz). For Sauerbrey films, the frequency shift  $\Delta f_n$  is proportional to the overtone number *n*, and the dissipation shift  $\Delta D_n \approx 0$ . However, a soft and viscous adlayer will dissipate energy and damp the sensor oscillation. In this case, one can obtain information about the stiffness of the adlayer from the shift in the energy dissipated per oscillation divided by the total energy stored in the oscillator.

For an oscillating quartz resonator in contact with a Newtonian fluid, the frequency shift and the dissipation shift arising from the presence of the fluid are [20-23]:

$$-\frac{\Delta f_n}{n} = n^{-1/2} f_0^{3/2} \left( \frac{\eta_l \rho_l}{\pi \mu_q \rho_q} \right)^{1/2}$$
(2)

and

$$\Delta D_n = 2n^{-1/2} f_0^{1/2} \left( \frac{\eta_l \rho_l}{\pi \mu_q \rho_q} \right)^{1/2}$$
(3)

where  $\eta_1$  and  $\rho_1$  are the fluid viscosity and density, respectively. Therefore, the QCM-D measurement of a Newtonian fluid on a nonadsorbing resonator provides the fluid viscosity if its density is accurately measured.

For a viscoelastic fluid in contact with the quartz resonator [24–26], its complex-valued viscosity is related to the frequency shift and the dissipation shift for the dynamic viscosity

$$\eta'(\omega) = -\frac{\pi \rho_q \mu_q}{\rho_l f_n} \frac{\Delta f_n \cdot \Delta \Gamma_n}{f_0^2} \tag{4}$$

and the storage viscosity

$$\eta^{\prime\prime}(\omega) = \frac{1}{2} \frac{\pi \rho_q \mu_q}{\rho_l f_n} \frac{\Delta \Gamma_n^2 - \Delta f_n^2}{f_0^2} \tag{5}$$

where  $\Delta \Gamma_n = \Delta D_n f_n/2$  is the shift of the half-bandwidth, with  $f_n = nf_0$  the frequency at each overtone *n*. Thereby, the QCM-D measurement of a viscoelastic fluid on a nonadsorbing resonator leads to its complex viscosities and shear moduli. It should be noted that the viscoelastic properties of the fluids are obtained at megahertz frequencies,  $\omega = 2\pi f_n$ .

#### Viscometry and rheometry

The viscosities of PEG solutions in pure water and different concentrations of  $K_2SO_4$  were measured at  $24 \pm 0.1$  °C by using an Ubbelohde viscometer with an efflux time of 243.0 s for pure water. Huggins and Kraemer plots were used to determine the intrinsic viscosity  $[\eta]$  of PEG. The

density of each solution was measured at  $24 \pm 0.01$  °C on a DMA 4500 M density meter (Anton Paar) with a resolution of  $5 \times 10^{-5}$  g/mL.

Linear viscoelastic measurements of PEG solutions were conducted on an ARES-G2 (TA Instruments) rheometer with a cone-and-plate geometry (with a cone radius of 25 mm and a cone angle of 2°) kept at  $24 \pm 0.1$  °C. Frequency sweep measurements were performed at small amplitudes (<10%) to measure the storage and loss moduli,  $G'(\omega)$  and  $G''(\omega)$ , as functions of the angular frequency,  $\omega$ , between 2 and 500 rad/s. Shear-rate sweep measurements were performed to measure the viscosities of PEG solutions at shear rates that ranged from 5 to 500 s<sup>-1</sup>.

### **Results and Discussion**

# Formation of a rigid supported lipid bilayer on the resonator

The formation of a SLB on a silicon oxide substrate was monitored by OCM-D, as shown in Fig. 1. After establishing the baseline with a 150 mM NaCl aqueous solution for approximately 5 min, the solution of DOPC liposome was pumped into the chamber at 7.0 min, and the observed decrease in the resonance frequency and increase in the energy dissipation factor were due to the adsorption of liposomes on the hydrophilic substrate. The frequency shifts,  $\Delta f_{\rm p}/n$ , decreased to values between -61 and -36 Hz (the value was different for each overtone) at 10.5 min, before the signals reached a plateau value of -24.5 Hz (the value was the same for all harmonics) after 14.0 min. The corresponding dissipation shifts,  $\Delta D_n$ , reached *n*-dependent maxima of approximately  $7.0 \times 10^{-6}$  before stabilizing at *n*independent values of  $0.1-0.2 \times 10^{-6}$ . Such evolutions in the shifts of both resonant frequency and energy dissipation factors indicate the rupture of adsorbed soft liposomes to form a rigid SLB on the silicon oxide substrate, which agrees well with previous experimental and theoretical studies [16-18, 27-32]. The nonfouling nature of the SLB was employed to impede the nonspecific adsorption of samples on the sensor, which allowed for the fluid viscoelastic properties to be probed by QCM-D, as shown below.

#### Viscosity of potassium sulfate solutions

After forming the DOPC SLB on the silicon oxide substrate, the adsorption behavior of  $K_2SO_4$  solutions on the SLB was subsequently studied at salt concentrations  $c_s$ between 0.1 and 0.6 mol/L. The resonance frequency and the energy dissipation factor for all 7 harmonics were monitored as the  $K_2SO_4$  solution was pumped into the chamber, followed by rinsing with water. Figure 2a shows a



**Fig. 1** QCM-D monitoring of the resonance frequency shifts  $\Delta f_n/n$  and energy dissipation shifts  $\Delta D_n$  during the formation of the DOPC SLB on a silicon oxide substrate. The frequency signals from bottom to top are for overtone numbers n = 1, 3, 5, 7, 9, 11, 13. The corresponding dissipation signal at each overtone is shown in the same color as the frequency. The black dashed line is a guide to the eyes with a value of 0 for the baseline

typical OCM-D measurement for a 0.39 mol/L K<sub>2</sub>SO<sub>4</sub> aqueous solution on the DOPC SLB. The shifts in the resonance frequency and the energy dissipation factor induced by the K<sub>2</sub>SO<sub>4</sub> solution disappeared as soon as they were rinsed away by water, indicating no irreversible adsorption. The ratio of the dissipation shift to the frequency shift,  $\Delta D_n/(-\Delta f_n/n)$ , was close to the theoretical value of  $2/f_0 = 4.04 \times 10^{-7} \text{ Hz}^{-1}$  for Newtonian fluids (Fig. 2b). The values of  $-\Delta f_n/n$  and  $\Delta D_n$  were plotted as functions of the overtone number *n*, both showing the scaling laws of  $-\Delta f_n/n \sim n^{-1/2}$  and  $\Delta D_n \sim n^{-1/2}$  (inset of Fig. 2b). One can calculate the viscosity of the  $K_2SO_4$ solution via eqs. (2) and (3) once the solution density is measured by a density meter. As shown in Fig. 3, the obtained viscosity of the K<sub>2</sub>SO<sub>4</sub> solution agrees very well with the viscosity measured by a viscometer and the data in reference [33]. Therefore, forming a rigid SLB on the quartz resonator does not prevent it being a sensor for fluid viscosity. Instead, the bilayer served as a nonadsorbing interface to prevent nonspecific adsorption between the bare silicon oxide substrate and the solutions in the chamber, and the resonator can be used to measure the viscosity of the Newtonian fluids.

# Viscoelasticity of PEG solutions in the presence of potassium sulfate

The viscoelasticity of aqueous PEG solutions in 0.19, 0.43, and 0.57 mol/L  $K_2SO_4$  was studied by QCM-D. In aqueous PEG solutions, increasing the salt concentration decreases the lower consulate temperature, and the magnitude of this decrease depends on the nature of the salt according to the



**Fig. 2** A typical QCM-D measurement for a 0.39 mol/L K<sub>2</sub>SO<sub>4</sub> aqueous solution on the DOPC SLB. **a** The resonance frequency shifts  $\Delta f_n/n$  and the energy dissipation shifts  $\Delta D_n$  for all harmonics as a function of time. The frequency signals from bottom to top are for overtone numbers n = 1, 3, 5, 7, 9, 11, 13. The corresponding dissipation signal at each overtone is shown in the same color as the frequency. **b** The ratio of the dissipation shift to the frequency shift for the 5th overtone,  $\Delta D_5/(-\Delta f_5/5)$ , as a function of time. The dashed line is the theoretical value of  $2/f_0$  for Newtonian fluids. The inset shows the dependence of the frequency shift  $-\Delta f_n/n$  and the dissipation shift  $\Delta D_n$  on the overtone number *n* for the 0.39 mol/L K<sub>2</sub>SO<sub>4</sub> aqueous solution. The solid and dashed lines are fittings to the data with a scaling exponent of -1/2

Hofmeister series [34–36]. For potassium sulfate, a strongly hydrated salt, the theta temperature of PEG decreases significantly with salt concentration. The theta temperature of PEG in 0.45 mol/L K<sub>2</sub>SO<sub>4</sub> is ~35 °C, and an extrapolation to a theta temperature of 24 °C leads to a K<sub>2</sub>SO<sub>4</sub> concentration of 0.53 mol/L [36]. Therefore, the above solvents are good, nearly theta and bad solvents for PEG/PEO. Indeed, it was found that in the 0.57 mol/L K<sub>2</sub>SO<sub>4</sub> aqueous solution, although PEG samples with  $M_w \le 35,000$  were soluble, PEO samples with the highest  $M_{\rm w}$  of 116,000, 278,000, and 443,000 were no longer soluble at a concentration of 2.0%, indicating a bad solvent condition in vicinity of the theta point. Fig. 4a shows a typical QCM-D measurement for dilute aqueous solutions of 2.0% PEG of different molecular weights in the 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> solution on the DOPC SLB. No irreversible adsorption of the PEG solution on the SLB was found in the studied molecular weight range. The ratio  $\Delta D_{\rm n}/(-\Delta f_{\rm n}/n)$  for PEG with



**Fig. 3** Concentration dependence of the density (solid squares) and viscosity obtained by QCM-D (open squares) and a viscometer (open circles) of aqueous solutions of K<sub>2</sub>SO<sub>4</sub> at 24 °C. The solid lines are fits to  $\rho = \rho_0 + a_1c_s + a_2c_s^2$ , with  $\rho_0 = 0.99730$  g/mL,  $a_1 = 0.13915$ , and  $a_2 = -0.01115$  for the density, and  $\eta = \eta_0 + b_1c_s + b_2c_s^2$ , with  $\eta_0 = 0.911$  cP,  $b_1 = 0.18759$ , and  $b_2 = 0.03515$  for the viscosity

molecular weights 200, 400, 600, and 1000 is close to the theoretical value of  $2/f_0 = 4.04 \times 10^{-7} \text{ Hz}^{-1}$  for Newtonian fluids, and it increased with molecular weight to  $1.1 \times 10^{-6} \text{ Hz}^{-1}$  for PEG-35000 (Fig. 4b). Therefore, the elastic property of PEG solutions becomes stronger for PEG of higher molecular weight.

In Fig. 5, the values of  $-\Delta f_n/n$  and  $\Delta D_n$  are plotted as functions of the normalized frequency  $\omega \tau$  for 2.0% PEG/ PEO aqueous solutions containing 0.19 mol/L K<sub>2</sub>SO<sub>4</sub>. Here, the frequency is normalized by the sample's Zimm relaxation time  $\tau$ , as given by [9]:

$$\tau = 0.325 \frac{\eta_s \overline{R}^3}{k_B T} \tag{6}$$

where  $k_{\rm B}$  is the Boltzmann constant, and  $\overline{R}$  is the end-to-end distance of polymer with  $\overline{R} = 6^{1/2} R_g$ . The radius of gyration  $R_{g}$  can be obtained from the molecular weight according to the scaling relation  $R_{\rm g} \sim M_{\rm w}^{\nu}$ . Here, we use the relation  $R_{\rm g}$  (nm) = 0.0202 ×  $M_{\rm w}^{0.58}$  for high- $M_{\rm w}$  PEG/PEO in 0.19 mol/ L K<sub>2</sub>SO<sub>4</sub> [37–39], while the relation  $R_g$  (nm) = 0.0355 ×  $M_{\rm w}^{0.50}$  obtained from light scattering and intrinsic viscosity measurements at theta condition [39, 40] was employed for all PEG/PEO samples in 0.43 and 0.57 mol/L K<sub>2</sub>SO<sub>4</sub>. The latter was also used for PEG samples with  $M_w < 2000$  in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub>, which are ideal chains with a negligible excluded volume, according to previous experiments and the coarse-grained simulation [16, 41, 42]. Both  $-\Delta f_n/n$  and  $\Delta D_{\rm n}$  increased with molecular weight in the low- $M_{\rm w}$  region but behaved differently in the high- $M_{\rm w}$  region: a peak for  $-\Delta f_n/n$  was observed, followed by a decrease to a plateau value at  $M_{\rm w} \ge 35,000$ , while  $\Delta D_{\rm n}$  continuously increased with  $M_{\rm w}$  to a plateau value at  $M_{\rm w} \ge 35,000$ . It should be



**Fig. 4** QCM-D signals for a series of aqueous solutions of 2.0% PEG of different molecular weights containing 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> exposed to the DOPC SLB. The signals from left to right are for PEG with molecular weights of 200, 400, 600, 1000, 1400, 2000, 4000, 6000, 8000, 10,000, 20,000 and 35,000. **a**, **b** are the frequency shift  $\Delta f_{3/3}$  (black curve) and the dissipation shift  $\Delta D_3$  (red curve), and their ratio  $\Delta D_3/(-\Delta f_3/3)$ , respectively. The red dashed line is the theoretical value of  $\Delta D_3/(-\Delta f_3/3) = 2/f_0$  for Newtonian fluids

noted that both the frequency shift  $-\Delta f_n/n$  and the dissipation shift  $\Delta D_n$  became lower with increases in the overtone number n. Their scaling laws with frequency  $\omega \tau$ (and overtone number n) showed a decrease in the exponents from -1/2 for low- $M_w$  PEGs with  $\omega \tau \ll 1$  to -0.76 for high- $M_{\rm w}$  ones with  $\omega \tau \gg 1$ , indicating a crossover from Newtonian to non-Newtonian behavior with increasing molecular weight. These observations are similar to previous results of PEG in pure water [16], indicating that the elastic properties of PEG in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> get stronger at either increasing  $M_{\rm w}$  or higher frequency. Fig. 6 compares the values of  $-\Delta f_n/n$  and  $\Delta D_n$  for PEG in aqueous solutions of different K<sub>2</sub>SO<sub>4</sub> concentrations to those for PEG in the pure water. It is found that these curves had the same trend, and an increase in the absolute values of  $-\Delta f_{\rm p}/n$ and  $\Delta D_n$  with increasing concentrations of K<sub>2</sub>SO<sub>4</sub> was observed. The latter is due to the increase in the viscosity and density of the K<sub>2</sub>SO<sub>4</sub> aqueous solution compared to those of pure water.

Using eqs. (4) and (5), we calculated the dynamic viscosity  $\eta'$  and storage viscosity  $\eta''$  of PEG/PEO in aqueous K<sub>2</sub>SO<sub>4</sub> solutions obtained from QCM-D. The shear moduli





**Fig. 5** Dependence of **a** the frequency shift  $-\Delta f_n/n$  and **b** the dissipation shift  $\Delta D_n$  on the normalized frequency  $\omega \tau$  for a 2.0% PEG/PEO aqueous solution containing 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> on an SLB of DOPC. The data from left to right are for PEG/PEO with molecular weights of 200, 400, 600, 1000, 1400, 2000, 4000, 6000, 8000, 10,000, 20,000, 35,000, 116,000, 278,000, and 443,000. The solid lines serve as guides to the eye with scaling exponents of -1/2 for low- $M_w$  PEG and -0.76 for high- $M_w$  PEG/PEO

attributed to the polymer can then be calculated via  $G' = \omega \eta''$  and  $G'' = \omega (\eta' - \eta_s)$ , where G' is the storage modulus, G'' is the loss modulus, and  $\eta_s$  is the solvent viscosity. In Fig. 7, we plot the reduced storage modulus  $G'M_w/(cRT)$  and reduced loss modulus  $G''M_w/(cRT)$  as functions of  $\omega \tau$  for all PEG/PEO samples in 0.19 mol/L and 0.43 mol/L K<sub>2</sub>SO<sub>4</sub>, respectively. Here, c is the polymer mass concentration, R is the gas constant, T is the absolute temperature, and  $\tau$  is the Zimm relaxation time. For comparison, we also plot the Zimm model predictions of the reduced storage and loss moduli for dilute solutions of linear polymer chains [43];

$$\frac{G'M_w}{cRT} = \frac{\omega\tau\sin[(1-1/(3\nu))\arctan(\omega\tau)]}{\left[1+(\omega\tau)^2\right]^{(1-1/(3\nu))/2}}$$
(7)

$$\frac{G''M_w}{cRT} = \frac{\omega\tau \cos[(1 - 1/(3\nu))\arctan(\omega\tau)]}{\left[1 + (\omega\tau)^2\right]^{(1 - 1/(3\nu))/2}}$$
(8)

**Fig. 6** The molecular weight dependence of **a** the frequency shift  $-\Delta f_3/3$  and **b** the dissipation shift  $\Delta D_3$  for 2.0% PEG/PEO in pure water (squares), 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> (circles), 0.43 mol/L K<sub>2</sub>SO<sub>4</sub> (upward triangles), and 0.57 mol/L K<sub>2</sub>SO<sub>4</sub> (downward triangles) on an SLB of DOPC

It can be seen that the reduced shear modulus data for all PEG/PEO samples collapsed to single curves covering the  $\omega\tau$  range between 10<sup>-2</sup> and 10<sup>4</sup>, which are very close to the Zimm model predictions with  $\nu = 0.55$  in 0.19 mol/L  $K_2SO_4$  and  $\nu = 0.50$  in 0.43 mol/L  $K_2SO_4$ . In the region of  $\omega \tau \ll 1$ , the scaling exponents were 2.0 for  $G'M_w/(cRT)$  and 1.0 for  $G''M_w/(cRT)$ . While in the region of  $\omega \tau \gg 1$ , both moduli had the same scaling exponent of  $1/(3\nu)$ , whose values were approximately 0.60 in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> and 0.67 in 0.43 mol/L K<sub>2</sub>SO<sub>4</sub>. The former and latter values are in excellent agreement with the theoretical Zimm model of linear polymer chains in a good solvent and a theta solvent, respectively. The excluded volume exponent for PEG in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub>,  $\nu = 0.55$ , was slightly smaller than the value of 0.565 for PEG in pure water [16], indicating that 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> is still a good solvent for PEG. In 0.43 mol/L K<sub>2</sub>SO<sub>4</sub>, the excluded volume exponent of  $\nu = 0.50$ indicated a state close to the theta condition. In 0.57 mol/L  $K_2SO_4$ , a bad solvent, only PEG samples with  $M_w \le 35000$ were soluble, and the obtained reduced shear modulus data were nearly the same as those for PEG in 0.43 mol/L



**Fig. 7** Reduced storage modulus  $G'M_w/(cRT)$  (solid symbols) and loss modulus  $G''M_w/(cRT)$  (open symbols) for all PEG/PEO samples in **a** 0.19 mol/L and **b** 0.43 mol/L K<sub>2</sub>SO<sub>4</sub> aqueous solutions. The solid curves are the predictions of the Zimm model with the excluded volume exponents  $\nu = 0.55$  in (**a**) and  $\nu = 0.50$  in (**b**). The dashed lines are guides to the eyes with the theoretical scaling exponents of 2.0 for  $G'M_w/(cRT)$  and 1.0 for  $G'M_w/(cRT)$  in the region of  $\omega\tau \ll 1$  and 1/  $(3\nu)$  for both moduli in the region of  $\omega\tau \gg 1$ 

 $K_2SO_4$ , agreeing with the Zimm model prediction for linear polymer chains in a theta solvent with an excluded volume exponent of  $\nu = 0.50$ .

For PEG with  $400 \le M_w \le 2000$ , in the region of  $\omega \tau \ll 1$ , the molecular weight dependence of the storage modulus  $G' \sim M_w^{6\nu-1}$  and the loss modulus  $G'' \sim M_w^{3\nu-1}$  led to the same value of  $\nu = 0.50$  in aqueous solutions containing different concentrations of K<sub>2</sub>SO<sub>4</sub> because these polymer chains were smaller than the thermal blob size and the excluded volume effect was not appreciable. For PEG with  $M_w \ge 8000$ , in the region of  $\omega \tau \gg 1$ , the frequency dependencies according to  $G'M_w/(cRT) \sim (\omega \tau)^{1/(3\nu)}$  and  $G''M_w/(cRT) \sim (\omega \tau)^{1/(3\nu)}$  led to the estimation of the excluded volume exponent  $\nu$  for each sample, and the results are plotted as a function of molecular weight in Fig. 8 for PEG in all solvents. With increasing K<sub>2</sub>SO<sub>4</sub> concentration, the excluded volume exponent in the high- $M_w$  limit decreased from 0.565 in water (a good solvent) to 0.55 in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> (a good solvent) and 0.50 in 0.43 mol/L K<sub>2</sub>SO<sub>4</sub> (a near-theta solvent). Therefore, the viscoelastic properties of PEG solutions in K<sub>2</sub>SO<sub>4</sub> at different concentrations can be obtained from the QCM-D measurement, which acts as a microrheometer working in the oscillation shear mode at high frequency.

# Comparison with conventional viscometry and rheometry

The viscosities of PEG solutions in pure water and aqueous solutions containing different concentrations of K<sub>2</sub>SO<sub>4</sub> were measured with a capillary viscometer. Fig. 9 shows the concentration dependence of the relative viscosities of PEG-35000 and PEG-600 in these solvents. The relative viscosities of PEG-600 were almost indistinguishable in solutions of different concentrations of K<sub>2</sub>SO<sub>4</sub>, and the obtained intrinsic viscosities were nearly the same in these solvents. However, the relative viscosity and intrinsic viscosity of PEG-35000 in aqueous solution decreased with increasing concentrations of K<sub>2</sub>SO<sub>4</sub>. The worsening quality of the solvent upon adding K<sub>2</sub>SO<sub>4</sub> to aqueous PEG solutions led to decreases in the intrinsic viscosity of PEG-35000 with a size above the thermal blob size, while the size of PEG-600 was smaller than the thermal blob size, and the intrinsic viscosity was independent of the solvent quality. This result agrees with the molecular weight and K<sub>2</sub>SO<sub>4</sub> concentration dependencies of the excluded volume exponent  $\nu$ , as obtained from QCM-D (Fig. 8), which took the value of 0.5 for PEG with  $M_w < 2000$  and decreased with increasing  $K_2SO_4$  concentration in the high- $M_w$  limit. The intrinsic viscosities of PEG in aqueous solutions containing different concentrations of  $K_2SO_4$  are plotted as a function of molecular weight in Fig. 10. The reference data for PEG/PEO in water and 0.45 mol/L K<sub>2</sub>SO<sub>4</sub> are also shown [39, 40, 44–47], which gave the Mark–Houwink relation  $[\eta] = KM_w^{\alpha}$  with K = 0.0308 mL/g and  $\alpha = 0.705 \pm 0.01$  in pure water, and  $[\eta]_{\theta} = K_{\theta} M_{w}^{\alpha_{\theta}}$  with  $K_{\theta} = 0.163$  mL/g and  $\alpha_{\theta}$  $= 0.50 \pm 0.01$  in 0.45 mol/L K<sub>2</sub>SO<sub>4</sub>. Our data for PEG in pure water and 0.43 mol/L K<sub>2</sub>SO<sub>4</sub> are in excellent agreement with these data. The data for PEG in 0.19 mol/L  $K_2SO_4$  were slightly lower than those for PEG in water, and the data for PEG in 0.57 mol/L K<sub>2</sub>SO<sub>4</sub> were slightly lower than those at theta condition. Fits of the data to the relation  $[\eta] = KM_w^{\alpha}$  in a limited molecular range of  $5000 \le M_w \le$ 35,000 led to  $\alpha = 0.69 \pm 0.02$  in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub> and  $\alpha =$  $0.45 \pm 0.01$  in 0.57 mol/L K<sub>2</sub>SO<sub>4</sub>. The variation of  $\alpha$  for PEG/PEO in aqueous solutions containing different concentrations of K<sub>2</sub>SO<sub>4</sub> agree well with the above results of the excluded volume exponent  $\nu$  obtained by QCM-D, by considering the relation  $\alpha = 3\nu - 1$ .



Fig. 8 The excluded volume scaling exponent  $\nu$  as a function of the molecular weight of PEG/PEO samples in water (squares) and aqueous solutions containing different amounts of K<sub>2</sub>SO<sub>4</sub>: 0.19 mol/L (circles), 0.43 mol/L (upward triangles), and 0.57 mol/L (downward triangles). The dashed lines are guides to the eyes



**Fig. 9** Concentration dependence of the relative viscosities of PEG-600 (solid symbols) and PEG-35000 (open symbols) in water (squares) and aqueous solutions containing different amounts of K<sub>2</sub>SO<sub>4</sub>: 0.19 mol/L (circles), 0.43 mol/L (upward triangles), and 0.57 mol/L (downward triangles). The lines are fits to the data according to the Huggins plot:  $\eta_r = 1 + [\eta]c + k_H[\eta]^2c^2$ . The inset shows the dependence of the intrinsic viscosity  $[\eta]$  on the K<sub>2</sub>SO<sub>4</sub> concentration  $c_s$ 

The viscoelasticity of 2% PEG/PEO aqueous solutions was also measured by a conventional rheometer at frequencies  $\omega$  between 2 and 500 rad/s. However, no reliable complex-valued viscosity can be obtained for PEG with  $M_{\rm w} \leq 35,000$  at all. For PEO samples with the highest  $M_{\rm w}$  of 116,000, 278,000, and 443,000, the dynamic viscosities can be obtained according to their agreement with the steady flow viscosities via the Cox-Merz rule [48]. However, no storage viscosity data could be obtained because the frequency range covered by the rheometer was  $\omega \ll \tau^{-1}$ . Fig. 11 shows the shear moduli of 2% PEO-116000 in water



**Fig. 10** Intrinsic viscosities of PEG/PEO in water (black solid circles) and aqueous solutions containing different amounts of K<sub>2</sub>SO<sub>4</sub>: 0.19 mol/L (red solid circles), 0.43 mol/L (green solid circles), and 0.57 mol/L (blue solid circles). The inset shows the fits to these data with exponents of 0.72, 0.69, 0.50, and 0.45. The intrinsic viscosity data in references [39, 40, 44–47] are also shown for PEG/PEO in a good solvent of water (solid squares) and in a theta solvent of 0.45 mol/L K<sub>2</sub>SO<sub>4</sub> at 35 °C (open squares). The black and green lines are fits to all data with  $M_w > 5000$ :  $[\eta] = KM_w^{\alpha}$  with K = 0.0308 mL/g and  $\alpha = 0.705$  for PEG/PEO in water, and  $[\eta]_{\theta} = K_{\theta}M_w^{\alpha 0}$  with  $K_{\theta} = 0.163$  mL/g and  $\alpha_{\theta} = 0.50$  for PEG/PEO in 0.45 mol/L K<sub>2</sub>SO<sub>4</sub>, respectively. The dashed green line is an extrapolation to the low- $M_w$  region for the fit to the theta solvent

obtained from both QCM-D and the rheometer. The viscoelastic properties of this sample were obtained in the high-frequency region via QCM-D and in the lowfrequency region by the rheometer. Both datasets are close to the Zimm model predictions of the shear moduli for dilute solutions of linear polymer chains, according to eqs. (7) and (8). As shown in the inset of Fig. 11, the dynamic viscosities  $\eta'$  of PEG/PEO solutions obtained via QCM-D agreed with the shear viscosity  $\eta$  obtained by the rheometer for low- $M_{\rm w}$  PEG but deviated significantly from that for high- $M_w$  PEG/PEO, suggesting a crossover from Newtonian to non-Newtonian behavior with increasing molecular weight. It should be noted that the dynamic viscosity  $\eta'$  of PEG/PEO solutions were obtained from QCM-D at extremely high frequency and were substantially smaller than the values obtained by the rheometer working at low frequency.

Therefore, the viscoelastic properties of aqueous PEG solutions containing different concentrations of  $K_2SO_4$  were successfully measured by QCM-D at megahertz frequency and were justified by conventional methods of viscometry and rheometry. The viscoelasticity of PEG solutions was probed by QCM-D over a broad normalized frequency range  $\omega \tau$  between 10<sup>-2</sup> and 10<sup>4</sup> by using a series of samples of different molecular weights. Newtonian behavior was observed for low- $M_w$  PEGs, while non-Newtonian behavior was found for high- $M_w$  PEGs. The high-frequency



**Fig. 11** Shear moduli of 2% PEO-116000 in water obtained from QCM-D for the loss modulus (open circles) and the storage modulus (solid circles) and from the rheometer for the loss modulus (open squares). The solid curves are the Zimm model predictions according to eqs. (7) and (8). The inset shows the dynamic viscosity  $\eta'$  of all 2.0% PEG/PEO aqueous solutions obtained from QCM-D with overtone numbers n = 1, 3, 5, 7, 9, 11, 13 (open circles from top to bottom) and by the rheometer (open squares). The steady flow viscosities obtained by the rheometer are also shown as solid diamonds

characteristic of QCM-D makes it possible to directly study the viscoelasticity of polymer solutions in a low-viscosity solvent, without the employment of time-temperature superposition. Thus the QCM-D technique is complementary to conventional rheometers working at low frequency. The QCM-D method might be useful for the viscoelasticity measurement of precious biomacromolecule samples, such as proteins, peptides, and nucleic acids. These biomacromolecules are usually studied in buffers containing different amount of salts, and their chain flexibility and conformation are controlled by the amount of added salts. It should be noted that to measure the solution viscoelasticity via QCM-D, one has to passivate the resonator by an inert layer, such as the SLB, which must be stable and nonadsorbing in the presence of the studied sample solutions.

# Conclusions

The viscoelastic properties of PEG solutions in 0.19, 0.43, and 0.57 mol/L K<sub>2</sub>SO<sub>4</sub> were studied by QCM-D after coating the resonator with a rigid SLB, which does not adsorb PEG molecules in aqueous solutions of K<sub>2</sub>SO<sub>4</sub>. The obtained results agree well with the Zimm model predictions in a good solvent and a theta solvent. The excluded volume exponent  $\nu$  for high- $M_w$  PEG decreased with increasing K<sub>2</sub>SO<sub>4</sub> concentration, i.e., from 0.565 in water to 0.55 in 0.19 mol/L K<sub>2</sub>SO<sub>4</sub>, and 0.50 at higher K<sub>2</sub>SO<sub>4</sub> concentrations, indicating the gradual worsening of solvent quality by adding  $K_2SO_4$  to aqueous solutions of PEG. The excluded volume of larger PEG chains decreased with increasing  $K_2SO_4$  concentration in aqueous solutions, as confirmed by decreases in the intrinsic viscosity as well as the Mark–Houwink exponent  $\alpha$ .

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#### **Compliance with ethical standards**

Conflict of interest The authors declare that they have no conflict of interest.

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