



## CORRESPONDENCE OPEN

## Targeting IL-22 and IL-22R protects against experimental osteoarthritis

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Osteoarthritis (OA) is characterized by cartilage degradation, pain, and synovitis.<sup>1</sup> Joint inflammation driven by cytokines has been demonstrated to cause cartilage degradation and pain.<sup>2</sup> However, approaches to neutralize cytokines, such as IL-1 and TNF- $\alpha$ , known to be involved in OA have shown poor clinical efficacy.<sup>3</sup> There is an unmet clinical need to find better anti-inflammatory and pain targets for OA therapy and to elucidate the role of other cytokines in OA pathogenesis. Previous studies have shown that IL-22 and its receptor IL-22R play central roles in inflammation and diseases such as psoriasis, ulcerative colitis, graft-versus-host disease, certain infections and tumors, as well as in liver and pancreas damage.<sup>4,5</sup> The role of IL-22/IL-22R and the potential for therapeutic targeting of both proteins in OA remain largely unknown, which we sought to investigate.

We first examined human OA tissues to investigate whether IL-22/IL-22R expression levels change in disease. IL-22 was increased in the synovial fluid (SF) but not in the sera of OA patients compared to non-OA patients (Fig. 1a, b). Protein and mRNA expression of IL-22R was elevated in human chondrocytes isolated from OA patients (Fig. 1c–e). However, IL-22 protein and mRNA expression was only increased in fibroblast-like synoviocytes (FLS) isolated from OA patients (Fig. 1f–h). The increased concentration of IL-22 in the SF but not in the sera of OA patients seems to suggest that this cytokine plays a local role in OA joints with tissue-specific expression. It seems plausible that IL-22 produced by FLS in OA joints is important for disease progression. FLS-produced IL-22 may act mainly on IL-22R on chondrocytes, as indicated by the elevated expression levels of the receptor in human OA chondrocytes. Although IL-22/IL-22R have been reported to be increased in inflamed OA synovium and linked with increased protease expression,<sup>6,7</sup> further studies investigating the precise downstream signaling of IL-22/IL-22R in chondrocytes need to be conducted.

Having observed the tissue-specific increase in IL-22 in chondrocytes and IL-22R in FLS from OA patients, we next investigated whether these results had an in vivo relevance during

disease pathogenesis. We tested this hypothesis first by successfully generating inducible IL-22R chondrocyte-specific KO mice (IL-22R<sup>Acan Cre-ERT2</sup>) (Supplementary Fig. S1a, b). IL-22R<sup>Acan Cre-ERT2</sup> mice displayed decreased cartilage degradation, synovitis, osteophyte maturity and pain compared to IL-22R<sup>fl/fl</sup> control mice post experimental OA (surgical destabilization of the medial meniscus (DMM)) (Fig. 1i–k and Supplementary Fig. S1c–e). We also successfully generated inducible IL-22 FLS-specific KO mice (IL-22<sup>Col1a2 Cre-ERT2</sup>) (Supplementary Fig. S2a, b). IL-22<sup>Col1a2 Cre-ERT2</sup> mice demonstrated reduced disease outcomes and pain compared to IL-22<sup>fl/fl</sup> control mice post DMM surgery (Supplementary Fig. S2c–h). To our knowledge, this is the first set of in vivo data that show, using tissue-specific KO mice, the pathogenic role of IL-22/IL-22R in both OA disease progression and OA-related pain. IL-22 in RA joints has both beneficial<sup>8</sup> and pathogenic<sup>9</sup> roles; together with our results, this may suggest that IL-22 is part of divergent inflammatory responses orchestrated by different joint cells in OA compared to RA.

Next, we wanted to investigate whether therapeutically neutralizing IL-22 and IL-22R may attenuate OA in vivo. WT mice treated with an IL-22R neutralizing antibody demonstrated decreased cartilage degradation, synovitis, osteophyte maturity and pain compared to IgG1-treated control mice post DMM surgery (Fig. 1l, m and Supplementary Fig. S3a–c). Similarly, WT mice treated with an anti-IL-22 antibody displayed reduced disease outcomes (Supplementary Fig. S4a–f). Our in vivo studies also showed the possible potential of using anti-IL-22/IL-22R antibodies to treat OA and its related pain. A number of basic studies and clinical trials have shown the benefits of targeting IL-22/IL-22R in systemic immune diseases.<sup>4</sup> Our study indicates a further benefit of administering anti-IL-22/IL-22R in the joint to avoid any adverse systemic effects.<sup>10</sup>

Together, our data reveal the cell-specific pathogenic role of IL-22 (FLS specific) and IL-22R (chondrocyte specific) in OA. Targeting both IL-22 and IL-22R seems to be a plausible treatment option for OA and pain.

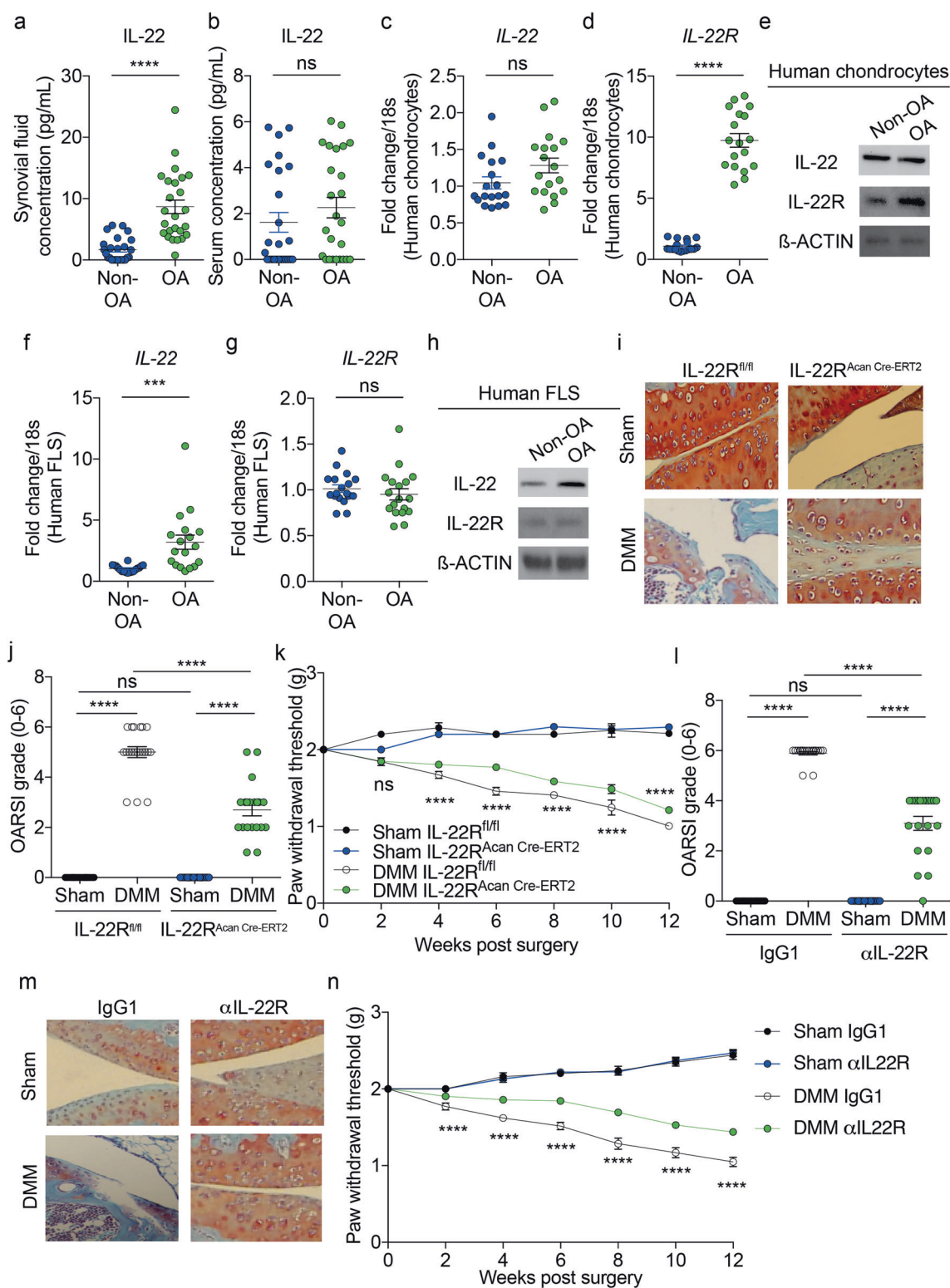
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**Fig. 1** Targeting IL-22 signaling protects against experimental osteoarthritis. IL-22 concentration (pg/mL) in **a** SF and **b** serum of non-OA and OA patients ( $n = 25$ ). **c** IL-22 mRNA, **d** IL-22R mRNA and **e** IL-22 and IL-22R protein expression in isolated human chondrocytes from non-OA and OA patients ( $n = 18$ ). **f** IL-22 mRNA, **g** IL-22R mRNA and **h** IL-22 and IL-22R protein expression in isolated FLS from non-OA and OA patients ( $n = 18$ ). **i**, **j** OARSJ scoring of cartilage and **k** von Frey pain assessment of sham- or DMM-operated IL-22R<sup>fl/fl</sup> control mice and IL-22R<sup>Acn Cre-ERT2</sup> mice (12 weeks post surgery end timepoint) ( $n = 20$ ). **l**, **m** OARSJ scoring of cartilage and **n** von Frey pain assessment from sham- or DMM-operated WT mice treated i.a. with either IgG1 (control; 50  $\mu$ g per mouse; 3 times per week for 12 weeks post surgery) or  $\alpha$ IL-22R (50  $\mu$ g per mouse; 3 times per week for 12 weeks post surgery) ( $n = 20$ ). All RT-qPCR gene expression levels were normalized to the endogenous level of 18S in the respective groups. Data are expressed as the mean  $\pm$  SEM with two-tailed *t*-test or two-way analysis of variance followed by the Tukey-Kramer test or repeated measures 2-way ANOVA with Bonferroni's post hoc tests. *n* indicates the number of human specimens or mice per group. NS nonsignificant. \*\*\* $p < 0.001$  or \*\*\*\* $p < 0.0001$  is represented in all figures

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## AUTHOR CONTRIBUTIONS

Design and experimentation: C.Y., J.L., and P.K.S.; supervision: Y.Y., Q.J., and P.K.S.; and manuscript writing: P.K.S.

## ADDITIONAL INFORMATION

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**Competing interests:** The authors declare no competing interests.

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