

ARTICLE OPEN



Semaphorin 5A suppresses ferroptosis through activation of PI3K-AKT-mTOR signaling in rheumatoid arthritis

Qi Cheng ^{1,2}, Mo Chen¹, Mengdan Liu¹, Xin Chen^{1,2}, Lingjiang Zhu¹, Jieying Xu^{1,3}, Jing Xue¹, Huaxiang Wu ^{1,4}✉ and Yan Du ^{1,4}✉

© The Author(s) 2022, corrected publication 2023

Abnormal activation of synovial fibroblasts (SFs) plays an important role in rheumatoid arthritis (RA), the mechanism of which remains unknown. The purpose of our study is to comprehensively and systematically explore the mechanism for Semaphorin 5A-mediated abnormal SF activation in RA. Here, we found that Semaphorin 5A levels were significantly higher in synovial fluid and synovial tissue from RA patients compared with osteoarthritis patients. We further found that the mRNA level and protein abundance of Plexin-A1 was elevated in RA SFs compared with OA SFs, while Plexin-B3 expression showed no significant difference. The increased Semaphorin 5A in RA synovial fluid was mainly derived from CD68⁺ synovial macrophages, and the elevation led to increased binding between Semaphorin 5A and its receptors, thereby promoting cytokine secretion, proliferation, and migration, and decreasing apoptosis. Moreover, the effect of Semaphorin 5A on enhancing activation (cytokine secretion, cell proliferation and migration) and reducing apoptosis of SFs was significantly abolished after knockdown of Plexin-A1 and Plexin-B3 by small interfering RNA. Transcriptome sequencing and protein array detection revealed that Semaphorin 5A activated the PI3K/AKT/mTOR signaling pathway and inhibited ferroptosis. Morphologically, transmission electron microscopy results showed that Semaphorin 5A could significantly eliminate the mitochondrial diminution, membrane density increased and crest ruptured of SFs induced by ferroptosis inducer RSL3. Mechanistically, Semaphorin 5A enhanced GPX4 expression and SREBP1/SCD-1 signaling by activating the PI3K/AKT/mTOR signaling pathway, thus suppressing ferroptosis of RA SFs. In conclusion, our study provided the first evidence that elevated Semaphorin 5A in RA synovial fluid promotes SF activation by suppressing ferroptosis through the PI3K/AKT/mTOR signaling pathway.

Cell Death and Disease (2022)13:608; <https://doi.org/10.1038/s41419-022-05065-4>

INTRODUCTION

Rheumatoid arthritis (RA) is a systemic autoimmune disease with synovial inflammation as the main pathological feature, and can eventually lead to irreversible joint or organ damage [1]. Synovial macrophages (SMs) and synovial fibroblasts (SFs) play important roles in the occurrence and development of synovitis by secreting large amounts of cytokines and conducting crosstalk with various cells [2–4]. However, understanding of the synovial inflammatory microenvironment and the mechanism for synovial cell activation remains limited, and there are no effective treatment methods that directly target synovial cells.

Semaphorin 5A is a member of the Semaphorin family that has close associations with various pathophysiological phenomena including cell migration, tumor growth, and immune responses [5]. In previous studies by ourselves and Gras et al., Semaphorin 5A was confirmed to be significantly elevated in serum from patients with systemic lupus erythematosus (SLE) and RA [6–8]. The pathogenesis of the Semaphorin 5A involvement in RA has been partially revealed. Specifically, Gras et al. [7] reported that soluble Semaphorin 5A strongly induces proliferation and activation of

T cells and NK cells and promotes their secretion of a variety of pro-inflammatory factors. Xiao et al. [9] further reported that TSP1 is the key domain of Semaphorin 5A involved in fibroblast-like synovial cell proliferation and angiogenesis. SFs not only promote inflammation, angiogenesis, and bone destruction, but also exhibit tumor cell-like phenotypes, such as metabolic reprogramming, increased invasion, and migration, and decreased apoptosis [2, 10, 11]. Therefore, a comprehensive and systematic understanding of the effects of Semaphorin 5A on SFs is needed for better understanding of the disease pathophysiology in RA.

Recently, a newly discovered form of programmed cell death, ferroptosis, has attracted much attention because of its potential for application in cancer treatment strategies, and is characterized by lipid peroxidation and iron overload [12]. Li et al. [13] reported that neutrophil ferroptosis is an important driver of SLE neutropenia and contributes significantly to the disease presentation. Given the tumor cell-like phenotypes of RA synovial cells, we hypothesize that ferroptosis plays an important role in synovial cell activation. However, there are few studies on the role of ferroptosis in the pathogenesis of RA.

¹Department of Rheumatology, The Second Affiliated Hospital of Zhejiang University School of Medicine, 88 Jiefang Road, 310009 Hangzhou, China. ²Department of Clinic Medicine, The Second Affiliated Hospital of Zhejiang University School of Medicine, 88 Jiefang Road, 310009 Hangzhou, China. ³Department of Neurology, Linping District Hospital of Integrated Traditional Chinese and Western Medicine, 311199 Hangzhou, Zhejiang, China. ⁴These authors contributed equally: Huaxiang Wu, Yan Du.

✉email: wuhx8855@zju.edu.cn; duyan2014@zju.edu.cn

Edited by Professor Massimiliano Agostini

Received: 1 March 2022 Revised: 3 July 2022 Accepted: 4 July 2022

Published online: 14 July 2022

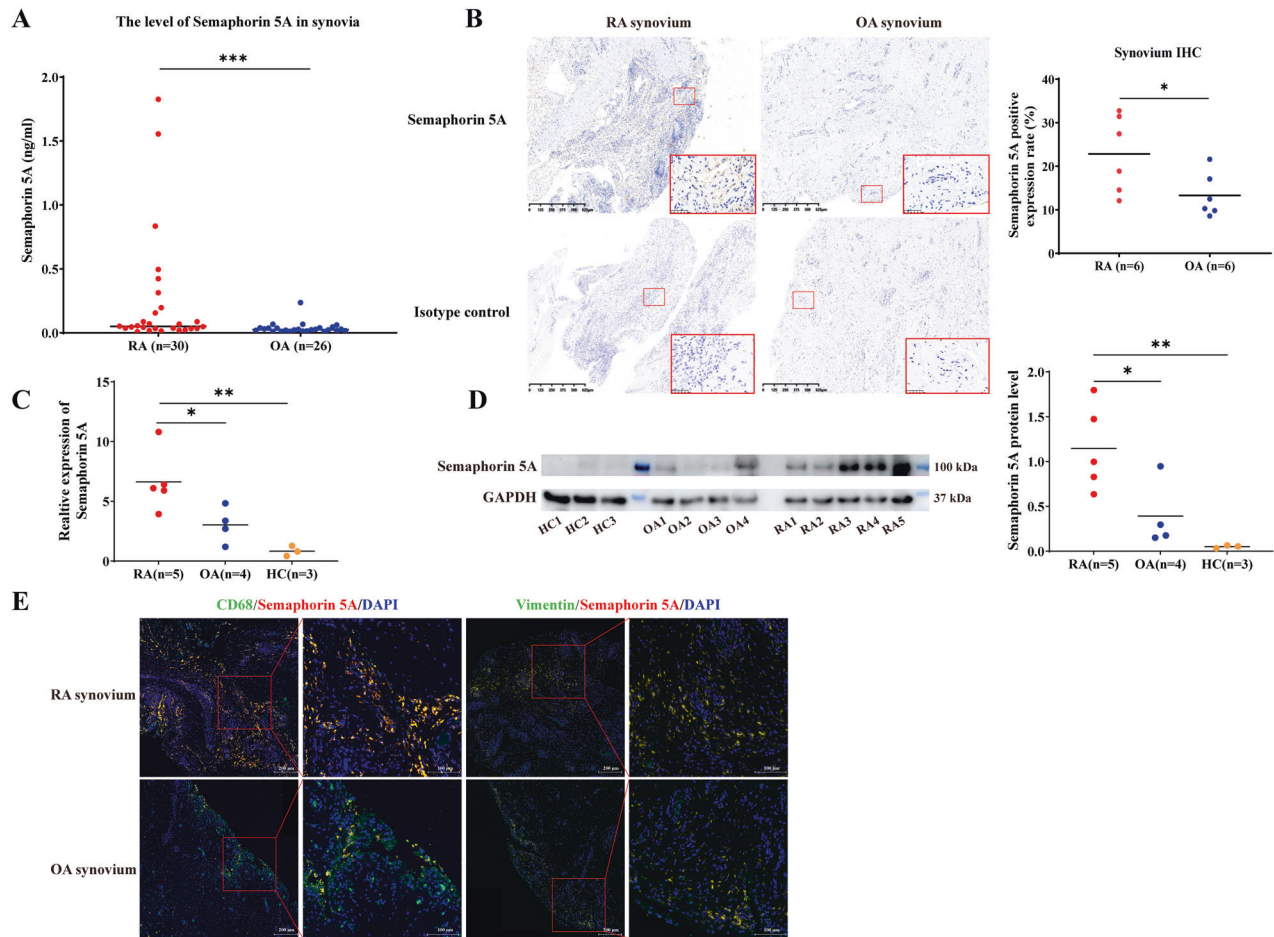


Fig. 1 Semaphorin 5A is elevated in RA, especially in SMs. **A** Levels of Semaphorin 5A in synovial fluid from 30 RA patients and 26 OA patients. **B** Immunohistochemistry of synovial tissues from RA and OA patients. The Semaphorin 5A positive expression rate was significantly higher in RA synovial tissues ($n = 6$) compared with OA synovial tissues ($n = 6$). **C**, **D** Expression levels of Semaphorin 5A in RA ($n = 5$), OA ($n = 4$) and healthy control ($n = 3$) synovial tissues detected by PCR (**C**) and western blotting (**D**). **E** Representative image of double-staining immunofluorescence of synovial tissues. Semaphorin 5A expression was higher in RA SMs (CD68⁺) compared with OA SMs ($n = 3$), but there was no difference between RA SFs (Vimentin⁺) and OA SFs ($n = 3$). * $P < 0.05$; ** $P < 0.01$. Values of $P < 0.05$ were considered significant. RA rheumatoid arthritis, OA osteoarthritis, SMs synovial macrophages, SFs synovial fibroblasts.

In the present study, we first identified SMs as the mainly origin of the increased Semaphorin 5A in RA synovial fluid and found a crosstalk in SMs and SFs which Semaphorin 5A secreted by macrophages may bind to Plexin receptors on SFs to exert its effect. Next, we specifically investigated the mechanism of Semaphorin 5A mediates the activation of SFs by high-throughput sequencing and in vitro validation and the role of Semaphorin 5A in ferroptosis of RA SFs.

RESULTS

Semaphorin 5A is elevated in RA, especially in SMs

To evaluate Semaphorin 5A expression in RA and its clinical and functional significance, we collected synovial fluid samples from 30 RA patients and 26 OA patients (Clinical and laboratory features of patients were shown in Supplementary Table 1) and examined the levels of Semaphorin 5A by ELISA. Consistent with our previous results in serum [6], the level of Semaphorin 5A in synovial fluid was significantly increased in RA compared with OA (Fig. 1A). In addition, we found a significant positive correlation between Semaphorin 5A in synovial fluid and Erythrocyte Sedimentation Rate, C-reactive protein (CRP), Rheumatoid Factor and Disease activity score of 28 joints (CRP), which are indicators of patients' disease activity (Supplementary Table 2). Because the

most basic pathological manifestation of RA is synovial inflammation, SFs and SMs are the most important effector cells. Therefore, we obtained synovial tissue specimens from RA and OA patients (Clinical and laboratory features of patients were shown in Supplementary Table 3) and detected the expression of Semaphorin 5A by immunohistochemistry, qPCR, and western blotting. Interestingly, we found that Semaphorin 5A was more strongly expressed in RA synovial tissues compared with OA synovial tissues (Fig. 1B-D). Both SFs and SMs can secrete a variety of inflammatory or effector molecules into the synovial fluid to promote the inflammation process [2]. To explore the reasons for the elevated levels of Semaphorin 5A in synovial fluid, we focused on these two major effector cells for synovial inflammation. Tissue immunofluorescence staining revealed that Semaphorin 5A expression was higher in RA SMs (CD68⁺) than in OA SMs, but there was no difference between RA SFs (Vimentin⁺) and OA SFs (Fig. 1E). These findings suggest that the difference of Semaphorin 5A in RA synovial fluid is mostly derived from SMs rather than SFs. Semaphorin 5A tends to act through binding to receptors, and Plexin-A1 and Plexin-B3 were identified as its functional receptors [14, 15]. To understand which effector cells are the targets of the elevated Semaphorin 5A in synovial fluid, we investigated the expression of the Plexin receptors on SMs and SFs using the publicly available Single-cell RNA-seq dataset (ImmPort: SDY998)

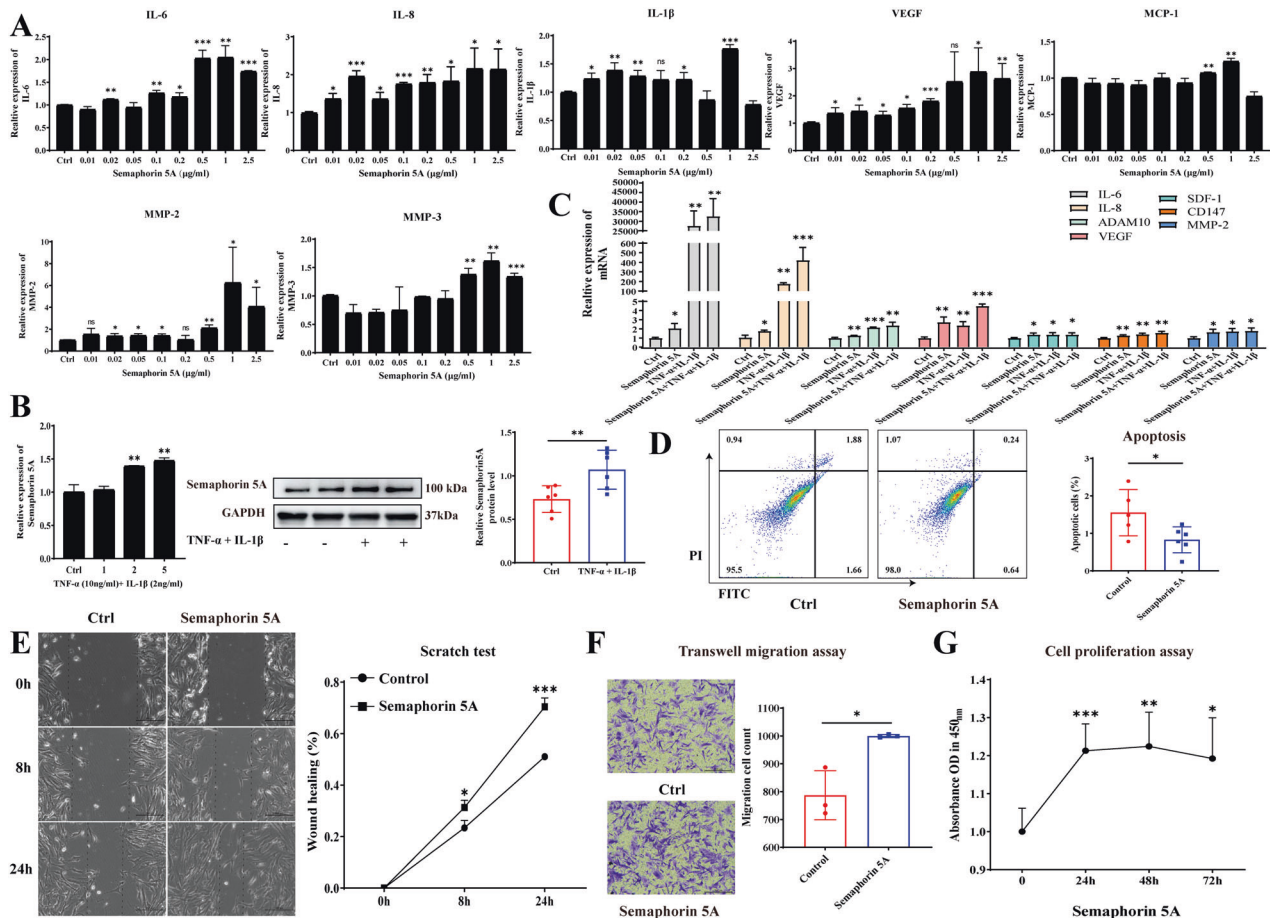


Fig. 2 Semaphorin 5A promotes cytokine secretion, cell proliferation, and cell migration and decreases apoptosis of SFs. **A** mRNA levels of various cytokines including inflammatory cytokines (IL-6, IL-8, IL-1 β), angiogenesis-related factors (VEGF, MCP-1), and matrix metalloproteinases (MMP-2, MMP-3) in SFs stimulated with different concentrations of Semaphorin 5A ($n = 3$). **B** mRNA levels and protein abundances of Semaphorin 5A in SFs activated by TNF- α (10 ng/mL) and IL-1 β (2 ng/mL) were detected by qPCR ($n = 3$) and western blotting ($n = 6$). **C** mRNA levels of IL-6, IL-8, ADAM10, VEGF, SDF-1, CD147, and MMP-2 in SFs synergistically stimulated with Semaphorin 5A (1 μ g/mL), TNF- α (10 ng/mL), and IL-1 β (2 ng/mL) ($n = 3$). **D** Apoptosis of SFs detected by flow cytometry ($n = 6$) after treatment with Semaphorin 5A (1 μ g/mL). **E** Migration of SFs detected by wound healing assays ($n = 3$) after treatment with Semaphorin 5A (1 μ g/mL). **F** Migration of SFs detected by transwell migration assays ($n = 3$) after treatment with Semaphorin 5A (1 μ g/mL). **G** Proliferation of SFs detected by a Cell Counting Kit-8 ($n = 4-5$) after treatment with Semaphorin 5A (1 μ g/mL). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$, ns, not significant. Values of $P < 0.05$ were considered significant. SFs synovial fibroblasts.

[16]. The expression of Plexin-A1 was significantly higher in SFs than in SMs, B cells, and T cells (Supplementary Fig. 1A–D). Unfortunately, Plexin-B3 was not found in this dataset. We further found that the mRNA level and protein abundance of Plexin-A1 was elevated in RA SFs compared with OA SFs, while Plexin-B3 expression showed no significant difference (Supplementary Fig. 1E–G). In addition, Semaphorin 5A and Plexin-A1 co-located on the membrane of SFs. These results suggest that Semaphorin 5A secreted by macrophages may bind to Plexin-A1 receptors on SFs to exert its effect.

Semaphorin 5A promotes activation and decreases apoptosis of SFs

Next, we examined the effects of exogenous Semaphorin 5A on MH7A. Interestingly, we found that exogenous Semaphorin 5A markedly stimulated SFs to secrete a variety of cytokines, including inflammatory cytokines (IL-6, IL-8, IL-1 β), angiogenesis-related factors (VEGF, MCP-1), matrix metalloproteinases (MMP-2, MMP-3), and others (ADAM10, SDF-1, CD147) at the optimal concentration and timing (Fig. 2A, Supplementary Fig. 2A, B). More reliably, we obtained the same results in primary human SFs isolated from RA patients (Supplementary Fig. 2C). We further

found a significant increase in the RNA level and protein abundance of Semaphorin 5A in SFs activated by TNF- α and IL-1 β (Fig. 2B). Meanwhile, Semaphorin 5A acted synergistically with TNF- α and IL-1 β to promote the secretion of cytokines from SFs (Fig. 2C). RA SFs exhibited tumor-like properties, such as increased proliferation and migration and decreased apoptosis, and Semaphorin 5A significantly promoted these characteristics (Fig. 2D–G). In addition, Semaphorin 5A was found to significantly inhibit Staurosporine-induced apoptosis (Supplementary Fig. 2D). These findings suggest that Semaphorin 5A can enhance activation and reduce apoptosis of SFs, promoting the production of an inflammatory synovial microenvironment.

Semaphorin 5A promotes activation of SFs by binding to its receptors Plexin-A1 and Plexin-B3

Like other cytokines, addition of exogenous Semaphorin 5A promoted the expression of its receptors Plexin-A1 and Plexin-B3 at both the RNA level and the protein level in SFs (Fig. 3A–C). To determine whether Semaphorin 5A really exerts its regulatory function through these receptors, we transfected siRNAs to knock-down Plexin-A1 and Plexin-B3 expression, respectively (Fig. 3D, E). As expected, the effect of Semaphorin 5A on enhancing activation

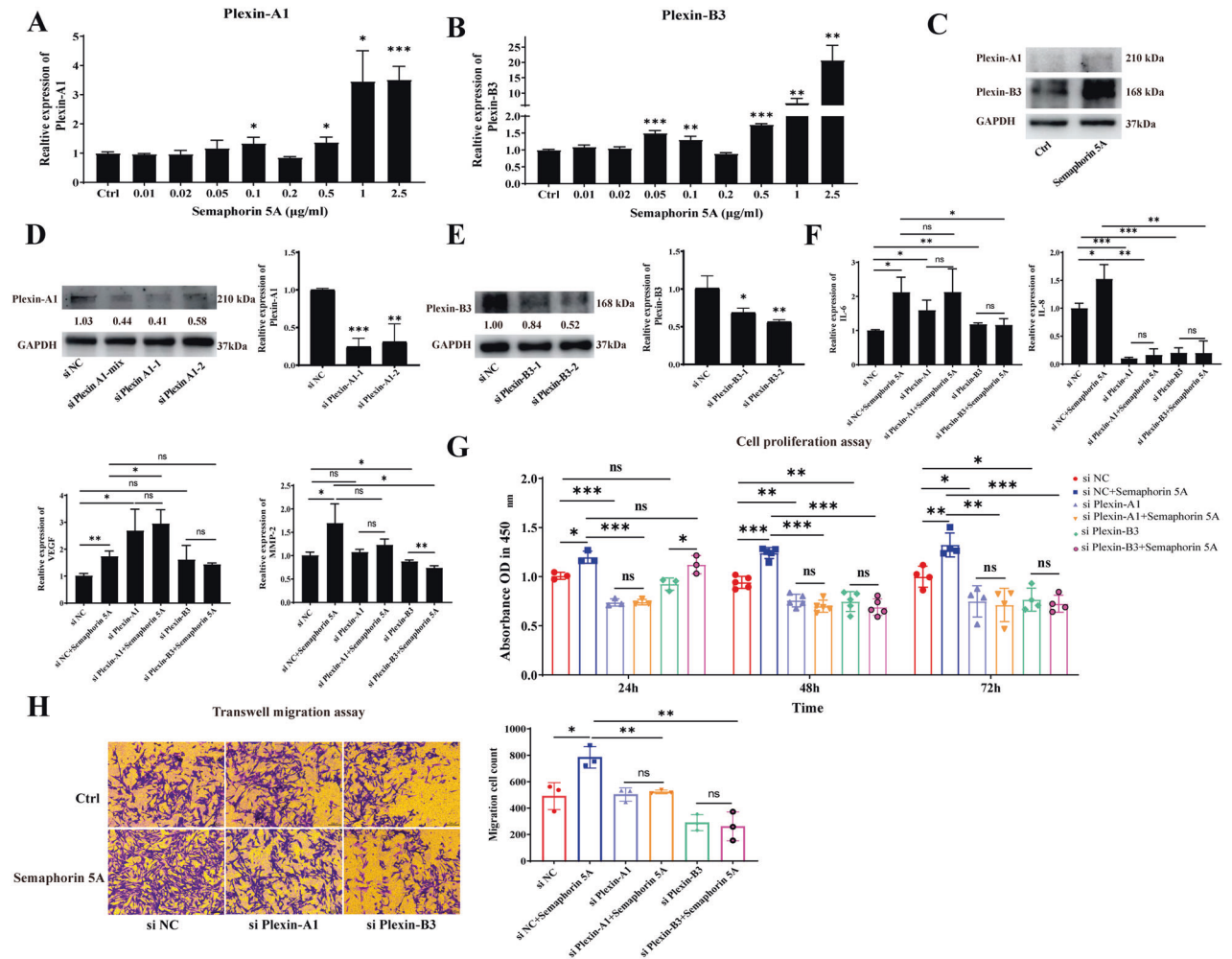


Fig. 3 Semaphorin 5A promotes activation of SFs by binding to plexin-A1 and plexin-B3. **A–C** RNA levels and protein abundances of Plexin-A1 and Plexin-B3 in SFs detected by qPCR ($n = 3$) (**A**, **B**) and western blotting ($n = 3$) (**C**). **D**, **E** After transfection of siRNAs to knock down Plexin-A1 (**D**) and Plexin-B3 (**E**), the RNA levels and protein abundances of Plexin-A1 and Plexin-B3 in SFs were significantly decreased ($n = 3$). **F** mRNA levels of IL-6, IL-8, VEGF, and MMP-2 in SFs detected by qPCR ($n = 3$) after transfection of siRNAs to knock down Plexin-A1 and Plexin-B3. **G** Proliferation of SFs detected by a Cell Counting Kit-8 ($n = 3–5$). **H** Migration of SFs detected by transwell migration assays ($n = 3$). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ns, not significant. Values of $P < 0.05$ were considered significant. SFs synovial fibroblasts, siRNA small interfering RNA.

(cytokine secretion, cell proliferation and migration) of SFs was significantly abolished after knockdown either of the two receptors (Fig. 3F–H and Supplementary Fig. 3A–F). Nevertheless, for cell apoptosis, only when Plexin-A1 and Plexin-B3 were simultaneously knockdown, the role of Semaphorin 5A could be abolished (Supplementary Fig. 3G). These suggested that Plexin-A1 and Plexin-B3 all serve as the functional receptors for Semaphorin 5A in RA SFs. Moreover, given that Plexin-B3 expression did not differ significantly between RA SFs and OA SFs, we consider that Plexin-A1 is a more specific receptor for Semaphorin 5A in RA SFs.

Semaphorin 5A can promote activation of SFs by activating the PI3K/AKT/mTOR signaling pathway

To understand how Semaphorin 5A activated SFs, transcriptome sequencing (Supplementary Fig. 4A–C) and protein array detection (Fig. 4B) were performed on SFs with and without stimulation by exogenous Semaphorin 5A. Some cytokines such as IL-1 β , IL-6, IL-8 and MMP-2 and Plexin receptors (Plexin-A1/Plexin-B3) were all up-regulated after Semaphorin 5A stimulation (Supplementary Fig. 4B). The functional and pathway enrichment analyses of differentially expressed genes and the protein microarray results all showed that Semaphorin 5A activated the PI3K/AKT/mTOR

signaling pathway (Fig. 4A, B). Subsequently, we demonstrated that Semaphorin 5A did indeed activate this pathway together with phosphorylation of the downstream molecules 4E-BP1 and BAD at different time points (Fig. 4C). These findings suggest that Semaphorin 5A has a persistent activation effect on this pathway. Moreover, we conducted rescue experiments using small-molecule inhibitors of the PI3K/AKT/mTOR signaling pathway. When we added pictilisib (PI3K inhibitor), MK-2206 (AKT inhibitor), and temsirolimus (mTOR inhibitor) to the medium, activation of the pathway and its downstream molecules by Semaphorin 5A was significantly inhibited (Fig. 4D). Meanwhile, the stimulatory effect of Semaphorin 5A on SF activation was significantly abolished, including decreased cytokine secretion (Fig. 4E), decreased cell proliferation (Fig. 4F), increased cell apoptosis (Fig. 4G), and decreased cell migration (Fig. 4H).

Semaphorin 5A reduces ferroptosis of SFs by GPX4

Our gene expression profile analysis and pathway enrichment analysis results indicated that Semaphorin 5A may affect not only the PI3K/AKT/mTOR signaling pathway, but also glutathione metabolism, apoptosis, and ferroptosis (Fig. 4A and Supplementary Fig. 4B). Through a review of the literature, we found that the final

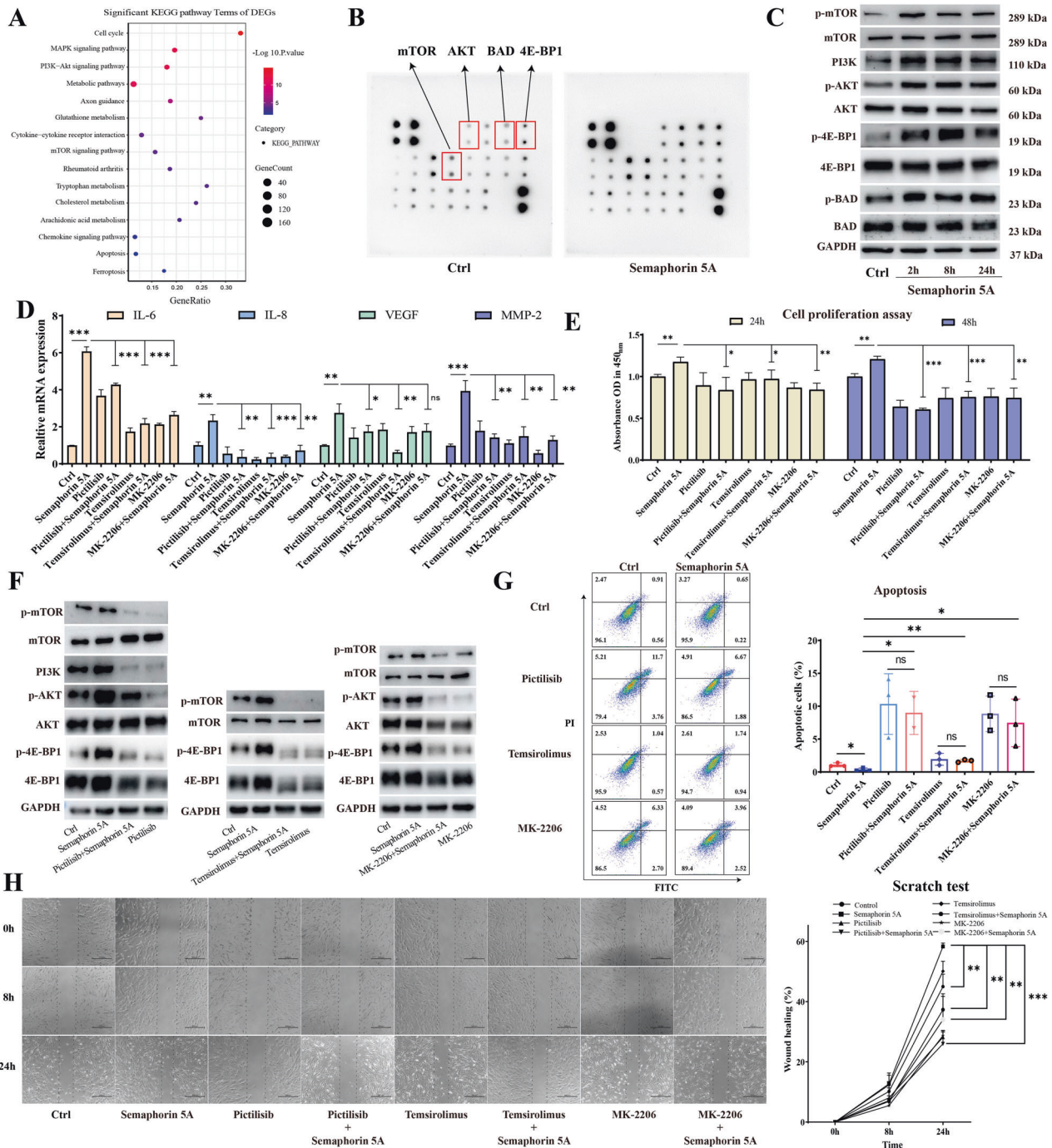


Fig. 4 Semaphorin 5A promotes activation of SFs by activating the PI3K/AKT/mTOR signaling pathway. **A** Significant KEGG pathway terms for differentially expressed genes. **B** Original image of the protein array analysis. **C** Total protein and phosphorylated protein levels for the PI3K/AKT/mTOR signaling pathway and its downstream molecules 4E-BP1 and BAD in SFs detected by western blotting after treatment with Semaphorin 5A (1 μ g/mL) at different times ($n = 3$). **D** Total protein and phosphorylated protein levels for the PI3K/AKT/mTOR signaling pathway and its downstream molecule 4E-BP1 in SFs detected by western blotting ($n = 3$). **E** mRNA levels of IL-6, IL-8, VEGF, and MMP-2 in SFs detected by qPCR after treatment with pictilisib (10 μ M), MK-2206 (5 μ M), tensirolimus (100 nM), and Semaphorin 5A (1 μ g/mL) ($n = 3$). **F** Proliferation of SFs detected by a Cell Counting Kit-8 ($n = 3-4$). **G** Apoptosis of SFs detected by flow cytometry ($n = 3$). **H** Migration of SFs detected by wound healing assays ($n = 3$). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ns not significant. Values of $P < 0.05$ were considered significant. SFs synovial fibroblasts.

outcome of PI3K/AKT/mTOR signaling pathway and glutathione metabolism is cell ferroptosis [17, 18]. Therefore, we hypothesized that Semaphorin 5A can reduce ferroptosis of SFs. Firstly, we investigated the expression of GPX4, a ferroptosis suppressor that prevents the production of lipid peroxidation reaction products [17], in synovial tissues and synovial fibroblasts of RA patients.

Unfortunately, there were no significant differences in either protein abundance or mRNA levels compared to OA patients (Supplementary Fig. 5A, B). However, we found that Semaphorin 5A significantly enhanced the RNA level and protein abundance of GPX4 in RA SFs (Fig. 5A, B). When we specifically blocked GPX4 with RSL3, we found that the enhancement of GPX4 by Semaphorin 5A

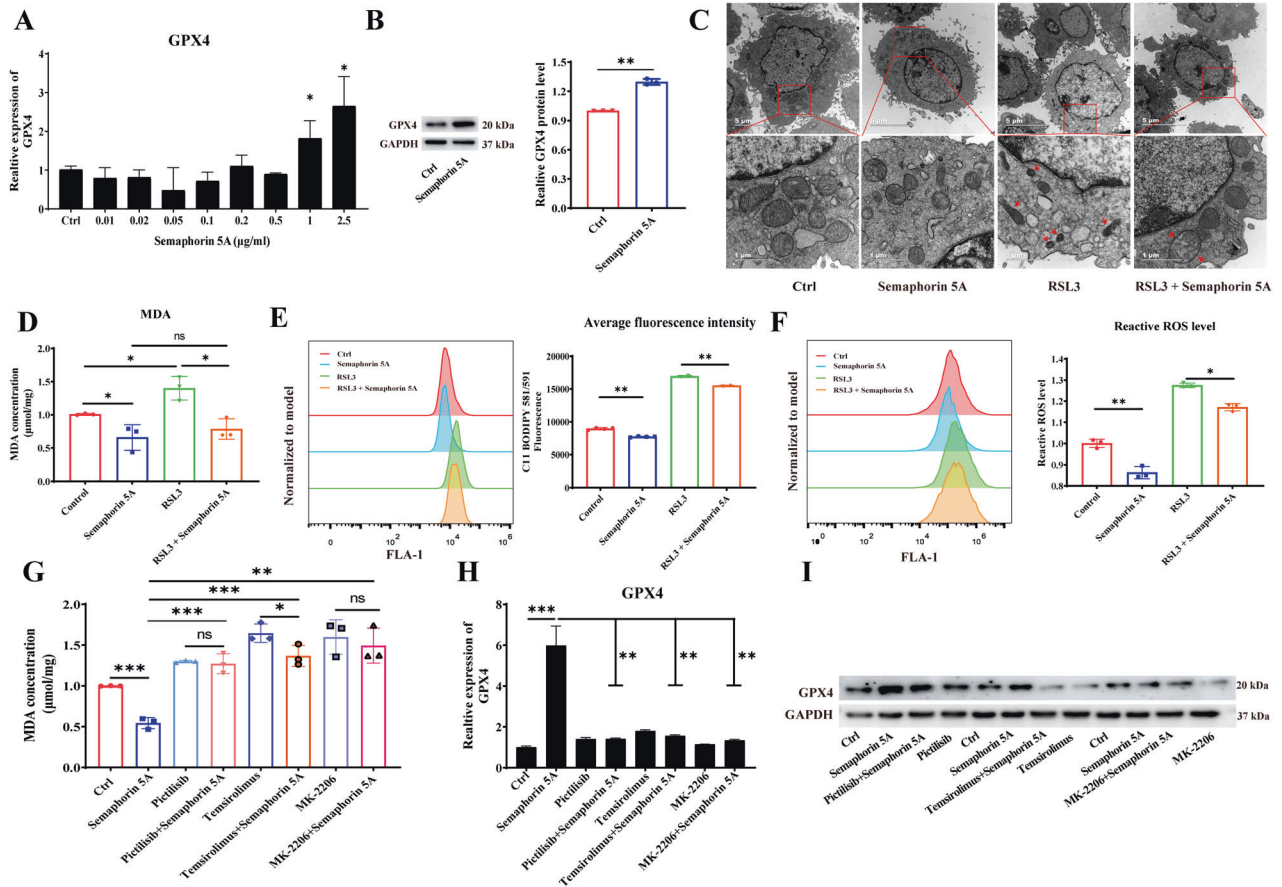


Fig. 5 Semaphorin 5A reduces ferroptosis of SFs by GPX4. **A** mRNA levels of GPX4 in SFs detected by qPCR ($n = 3$) after stimulation with different concentrations of Semaphorin 5A. **B** Protein abundances of GPX4 in SFs detected by western blotting ($n = 3$) after stimulation with Semaphorin 5A (1 $\mu\text{g/ml}$). **C** SF mitochondrial morphology observed by transmission electron microscopy ($n = 3$). **D** Intracellular MDA levels in SFs detected by a Lipid Oxidation (MDA) Assay Kit ($n = 3$). **E** Fluorescence intensity of C11-BODIPY 581/591 in SFs detected by flow cytometry ($n = 3$). **F** ROS levels in SFs detected by flow cytometry ($n = 3$). **G** Intracellular MDA levels in SFs detected by a Lipid Oxidation (MDA) Assay Kit after treatment with pictilisib (10 μM), MK-2206 (5 μM), temsirolimus (100 nM), and Semaphorin 5A (1 $\mu\text{g/ml}$) ($n = 3$). **H–I** mRNA levels and protein abundances of GPX4 in SFs detected by qPCR (**H**, $n = 3$) and western blotting (**I**, $n = 3$) after treatment with pictilisib (10 μM), MK-2206 (5 μM), temsirolimus (100 nM), and Semaphorin 5A (1 $\mu\text{g/ml}$). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ns, not significant. Values of $P < 0.05$ were considered significant. SFs synovial fibroblasts.

was attenuated (Supplementary Fig. 5C). In addition, RSL3 significantly reduced the migration and proliferation of SFs, and these effects were partially alleviated by the addition of exogenous Semaphorin 5A (Supplementary Fig. 5D, E). Next, we observed that SF mitochondria became smaller, the membrane density increased, and the cristae decreased after treatment with RSL3 alone by transmission electron microscopy (Fig. 5C). These are the typical electron microscopic characteristics of ferroptosis. Interestingly, when the cells were co-treated with Semaphorin 5A and RSL3, the changes in mitochondrial morphology were significantly improved and most of the mitochondria remained in their normal form (Fig. 5C). Changes in intracellular lipid peroxidation and ROS are other characteristics of ferroptosis. The levels of intracellular lipid peroxidation and ROS in SFs were significantly increased after treatment with RSL3 alone, but were significantly suppressed by Semaphorin 5A. Furthermore, Semaphorin 5A significantly reduced the levels of RSL3-induced lipid peroxidation and ROS (Fig. 5D–F). These findings indicate that Semaphorin 5A reduces ferroptosis of SFs by increasing the intracellular GPX4 level.

Semaphorin 5A reduces ferroptosis of SFs by activating the PI3K/AKT/mTOR/SREBP1/SCD-1 axis

Recently, several studies have confirmed a close relationship between mTORC1 signaling and ferroptosis [18, 19]. Therefore, we

hypothesized that Semaphorin 5A reduces ferroptosis of SFs by activating the PI3K/AKT/mTOR signaling pathway. First, MDA levels were measured to reflect intracellular lipid peroxidation. The intracellular MDA levels were significantly increased by the addition of pathway inhibitors alone, but were significantly reduced by Semaphorin 5A alone. We further found that the reduction in MDA levels by Semaphorin 5A was significantly abolished by the addition of the pathway inhibitors (Fig. 5G). Interestingly, the RNA level and protein abundance of GPX4 in SFs enhanced by exogenous Semaphorin 5A were significantly suppressed when PI3K/AKT/mTOR signaling was inhibited (Fig. 5H–I). These results suggest that Semaphorin 5A enhances GPX4 expression by activating the PI3K/AKT/mTOR signaling pathway, thereby suppressing ferroptosis of SFs. Yi et al. [18] reported that activation of PI3K/AKT/mTOR signaling protects cancer cells against ferroptosis through SREBP1/SCD-1-mediated lipogenesis. Therefore, we explored whether this signaling pathway was involved in the Semaphorin 5A-mediated reduction of SF ferroptosis. As expected, Semaphorin 5A significantly enhanced the RNA levels and protein abundances of SREBP1 and SCD-1 (Fig. 6A–C), and these phenomena were abolished by specific inhibitors of the PI3K/AKT/mTOR signaling pathway (Fig. 6D–F). Meanwhile, we also detected the expression of SREBP1 and SCD-1 in synovial fibroblasts to understand the basic situation of lipid synthesis

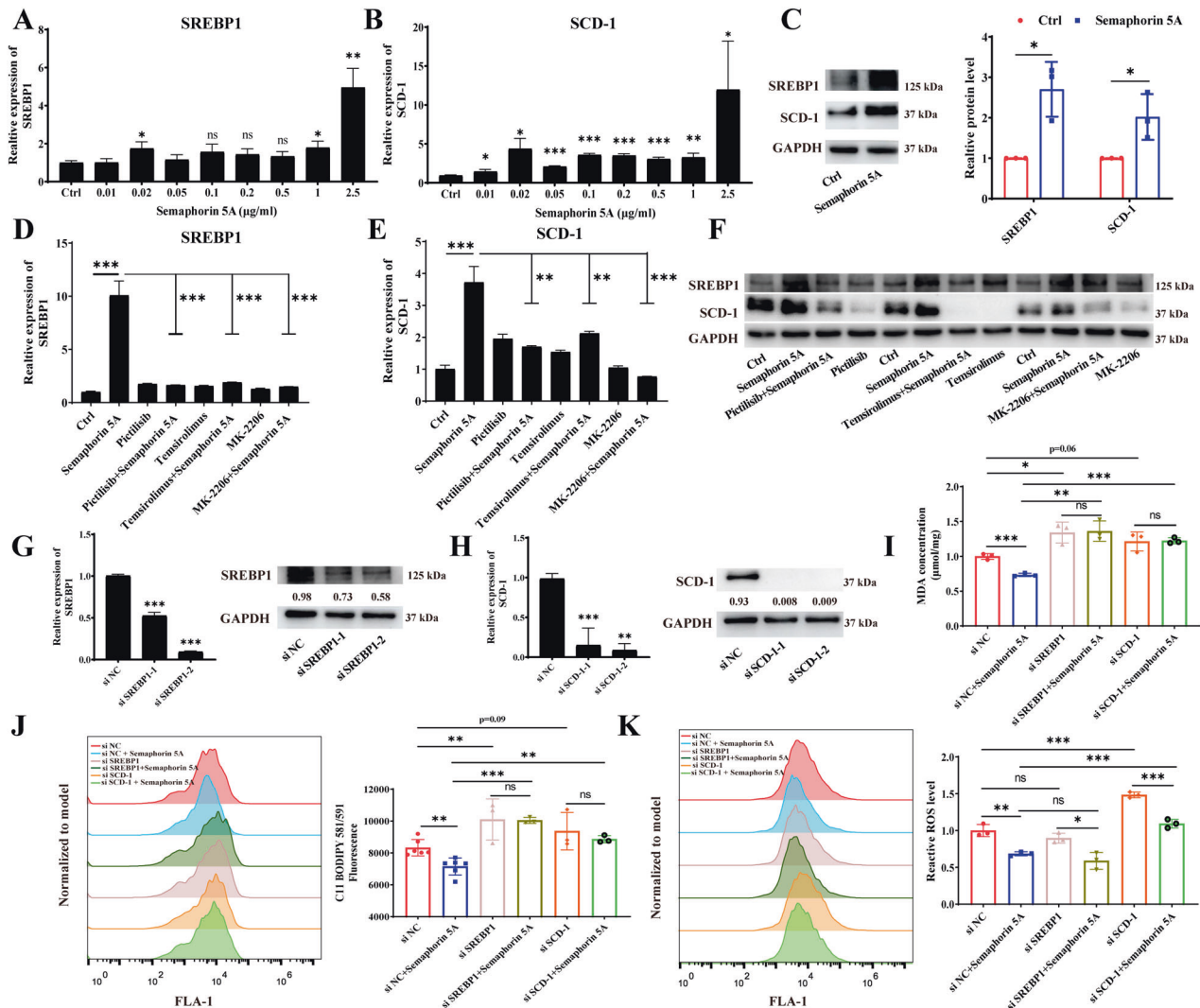


Fig. 6 Inhibitory effect of Semaphorin 5A on ferroptosis is abolished by inhibition of SREBP1/SCD-1 signaling. **A–C** mRNA levels and protein abundances of SREBP1 and SCD-1 in SFs detected by qPCR (**A**, **B**, $n = 3$) and western blotting (**C**, $n = 3$) after treatment with different concentrations of Semaphorin 5A. **D–F** mRNA levels and protein abundances of SREBP1 and SCD-1 in SFs detected by qPCR (**D**, **E**, $n = 3$) and western blotting (**F**, $n = 3$) after treatment with pictilisib (10 μM), MK-2206 (5 μM), temsirolimus (100 nM), and Semaphorin 5A (1 μg/mL). **G**, **H** RNA levels and protein abundances of SREBP1 and SCD-1 in SFs were detected by qPCR ($n = 3$) and western blotting ($n = 3$) after transfection of siRNAs to knock down SREBP1 (**G**) and SCD-1 (**H**). **I** Intracellular MDA levels of SFs detected by a Lipid Oxidation (MDA) Assay Kit ($n = 3$). **J** Fluorescence intensity of C11-BODIPY 581/591 in SFs detected by flow cytometry ($n = 3/6$). **K** ROS levels in SFs detected by flow cytometry ($n = 3$). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ns, not significant. Values of $P < 0.05$ were considered significant. SFs synovial fibroblasts.

pathway in RA patients. Compared with OA SFs, SCD-1 levels were significantly increased in RA SFs, while SREBP1 had no significant difference (Supplementary Fig. 6A–C). This is consistent with the conclusion that RA SFs underwent metabolic reprogramming, at least in the activation of lipid synthesis signals [10, 20]. Subsequently, we successfully knocked down SREBP1/SCD-1 by siRNA transfection and observed the effect on ferroptosis of SFs (Fig. 6G, H). After knockdown of SCD-1, the intracellular lipid peroxidation level and ROS level were significantly increased, while only the lipid peroxidation level was significantly enhanced after knockdown of SREBP1. We further found that the reduction in lipid peroxidation level by Semaphorin 5A was significantly abolished by knockdown of SREBP1 and SCD-1, while there was no effect on the ROS level (Fig. 6I–K). These observations further demonstrate the specificity of SREBP1/SCD-1 signaling for ferroptosis. In summary, our results confirmed that Semaphorin 5A enhances not only GPX4 expression but also SREBP1/SCD-1 signaling by activating the PI3K/AKT/mTOR signaling pathway, thereby suppressing ferroptosis of SFs.

DISCUSSION

The joint inflammation associated with RA results in an abundance of cytokines and immune cell infiltration in the interarticular space. Abnormal activation of SFs plays an important role in this process, the mechanism of which remains unknown [21]. It is of great clinical significance to identify factors that influence the abnormal activation of RA SFs and develop new therapeutic strategies.

Semaphorins play important roles in regulating various responses of the immune system. Our previous study showed that Semaphorin 5A, a member of the Semaphorin family, was elevated in the serum of RA patients [8]. Recently, Xiao et al. [9] reported that Semaphorin 5A was also elevated in the synovial membrane, synovial fluid, and SFs of RA patients. These findings are consistent with our results in synovial fluid and synovial tissue from RA patients. Meanwhile, we also found a significant positive correlation between the level of Semaphorin 5A in synovial fluid and the degree of disease activity in RA patients, suggesting that Semaphorin 5A may be involved in the disease progression and

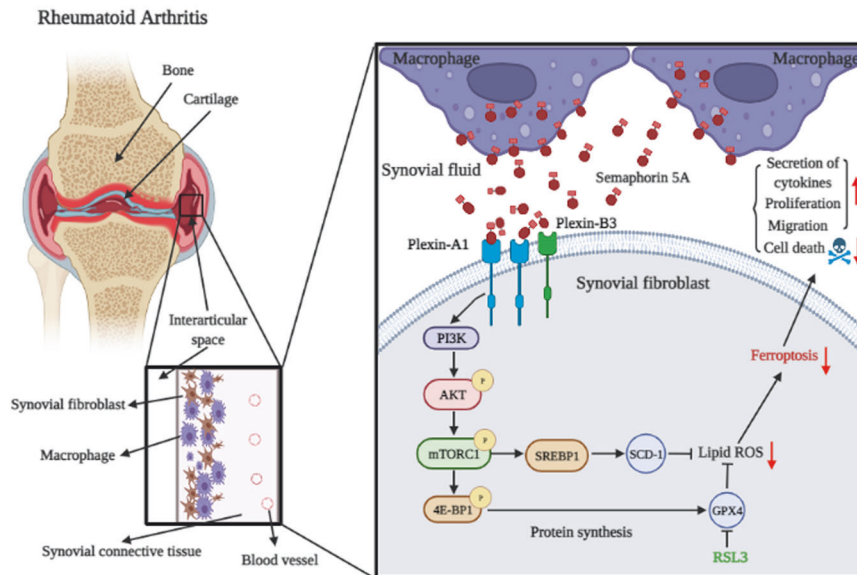


Fig. 7 Mechanism for Semaphorin 5A action on abnormal activation of RA SFs. SMs secrete a large amount of Semaphorin 5A, leading to an increased level of Semaphorin 5A in the synovial fluid. Semaphorin 5A then binds to Plexin-A1 and Plexin-B3 on SFs and activates the PI3K/AKT/mTOR signaling pathway. Activated mTORC1 signaling inhibits lipid peroxidation, ROS production, and ferroptosis by increasing the phosphorylation of 4E-BP1, leading to increased protein synthesis of GPX4, and also by activating downstream SREBP1/SCD-1 signaling. Reduced ferroptosis leads to increased survival of SFs, further promoting inflammation and immunity. SFs synovial fibroblasts, SM synovial macrophages.

activity of RA patients. However, our results showed that compared with OA, the expression of Semaphorin 5A was significantly increased in SMs from synovial tissues of RA patients, but only slightly increased in SFs without significant difference. These findings suggest that the elevated Semaphorin 5A in synovial fluid may be mainly derived from SMs, with a small part from SFs. Although a pathophysiological significance of the increased Semaphorin 5A in SFs cannot be excluded, the source of the elevated Semaphorin 5A in synovial fluid is likely to be SMs.

SFs have tumor cell-like phenotypes and are functionally altered in many ways [2, 21]. Xiao et al. [9] found that Semaphorin 5A promotes pannus formation, which reflects only a small part of the functional changes in SFs. More comprehensive and systematic studies are needed. In our study, we found that stimulation with exogenous Semaphorin 5A promoted the expression of not only angiogenic factors, but also various inflammatory cytokines and osteoclast-related factors. Joint inflammation and cartilage destruction are important causes of pain and joint deformity in RA patients. Moreover, Semaphorin 5A significantly enhanced other functions of SFs, such as cell proliferation, migration, and invasion. Importantly, Semaphorin 5A significantly reduced SF apoptosis, which is the basis for abnormal activation of RA SFs [22]. The increased survival of SFs further enhances its inflammatory, invasive, and other aspects, thereby enhancing the destructive inflammatory and immune microenvironment. Previous studies identified Plexin-A1 and Plexin-B3 as the functional receptors for Semaphorin 5A [5, 14, 15]. However, the receptor bound by Semaphorin 5A in RA SFs has not been reported in detail. Our results also indicate that SM-derived Semaphorin 5A plays an important role in abnormal activation of SFs by binding to Plexin-A1 and Plexin-B3 on SFs. Considering that the Plexin-A1 level was much higher in RA SFs compared with OA SFs, Semaphorin 5A and Plexin-A1 co-located on the membrane of SFs, we predicted that Plexin-A1 may be a more specific receptor for Semaphorin 5A in RA SFs. This is further evidence of a mutual crosstalk between SMs and SFs in the synovial microenvironment.

To determine the exact molecular mechanism, we performed transcriptome sequencing and protein array analysis. Consistent with previous studies, we found that Semaphorin 5A mainly

affected the PI3K/AKT/mTOR signaling pathway and its downstream molecule activation in SFs [23, 24]. However, we found that besides apoptosis, Semaphorin 5A activates SFs through inhibition of ferroptosis. A previous study demonstrated that GSH-based GPX4 inactivation is the most important mechanism of ferroptosis, because GPX4 is the only glutathione peroxidase in the cells for liposome peroxidase reduction [17]. We found that Semaphorin 5A inhibited ferroptosis in SFs by significantly promoting GPX4 expression. Interestingly, Semaphorin 5A still significantly inhibited ferroptosis even after treatment with RSL3, a specific inhibitor of GPX4. We further explored the relationship between the PI3K/AKT/mTOR signaling pathway and ferroptosis. Zhang et al. [19] reported that this pathway promotes GPX4 protein translation by inhibiting the downstream molecule 4E-BP1, thereby inhibiting ferroptosis. Another study by Yi et al. [18] indicated that this pathway also inhibits ferroptosis through the SREBP1-SCD-1 axis. Although these findings were obtained in tumor cells, SFs have tumor cell-like properties and we consider that there may be a similar association. As expected, Semaphorin 5A enhanced not only GPX4 expression but also SREBP1/SCD-1 signaling by activating the PI3K/AKT/mTOR signaling pathway, thereby suppressing ferroptosis of SFs (Fig. 7). These *in vitro* findings are exciting and we will be conducting animal studies in the future to verify our findings *in vivo*.

Semaphorin 5A has been described to promote angiogenesis by increasing endothelial cell proliferation and decreasing apoptosis [23]. Our study demonstrated that Semaphorin 5A not only affect apoptosis but also ferroptosis in RA. Recently, during preparation of this manuscript, Wu et al. published new evidence supporting the role of ferroptosis in RA. They found that TNF- α antagonists combined with ferroptosis inducers enhanced ferroptosis in fibroblasts and improved arthritis progression in collagen-induced arthritis mice [25]. These suggest targeting ferroptosis may be a new direction in the development of RA therapy. In addition, ferroptosis induction attenuates the interaction between fibroblasts and immune cells, which helps reduce inflammation and restore synovial homeostasis [25]. Although with the advent of biologic agents targeting cytokines such as TNF- α and IL-6, disease progression in most RA patients is controlled, some

patients do not respond well and develop resistance [1]. This is most likely due to ferroptosis resistance in RA patients. Thus, besides cell apoptosis, targeting ferroptosis may be a potential therapeutic approach and direction for the future treatment of these patients.

In the interarticular space, there are a variety of macrophages, including synovial lining layer macrophages, interstitial macrophages, and host tissue macrophages [26]. Semaphorin 5A was found to be mainly derived from CD68⁺ macrophages for the first time in our study, but the specific subgroup was not explored. A variety of emerging experimental devices and techniques, such as three-dimensional light-sheet fluorescence microscopy and single-cell sequencing technology, have provided new possibilities for dynamic origin tracing and more accurate classification of cell subsets [27]. More in-depth exploration and research are needed in the future. Meanwhile, Gras et al. [7] reported that Semaphorin 5A was cleaved by ADAM17 on the cell membrane, resulting in an elevated level of soluble Semaphorin 5A. Whether Semaphorin 5A on the surface of the SM membrane is cleaved by ADAM17 or other proteolytic enzymes also warrants further exploration. Finally, due to the apparent crosstalk between different programmed cell deaths, their effects may be different in different stages of RA [28, 29]. Therefore, although we found that Semaphorin 5A inhibited both ferroptosis and apoptosis, more studies are needed to understand the mechanisms associated with crosstalk of apoptosis and ferroptosis in RA and the role of Semaphorin 5A in the different stages of RA.

In the present study, we found that elevated Semaphorin 5A in synovial fluid of RA patients is mainly derived from CD68⁺ SMs rather than Vimentin⁺ SFs. Furthermore, Semaphorin 5A can bind to Plexin-A1 and Plexin-B3 on RA SFs to induce their activation. More importantly, we found that Semaphorin 5A promotes SF activation by inhibiting ferroptosis of SFs. Mechanistically, Semaphorin 5A enhances GPX4 expression and SREBP1/SCD-1 signaling via the PI3K/AKT/mTOR signaling pathway, thereby suppressing ferroptosis of SFs. Targeting Semaphorin 5A and ferroptosis could be an effective and valuable therapeutic strategy for patients with RA.

MATERIALS AND METHODS

Details of the cell isolation and culture, immunohistochemistry and immunofluorescence, ELISA, real-time quantitative PCR (qPCR), small interfering RNA (siRNA) transfection, cell apoptosis, cell proliferation assay, wound healing assay, transwell migration assay, RNA sequencing, protein array analysis and Western blot are provided in the Supplementary Materials.

Patient samples and informed consent

Synovial fluid and synovial tissue samples were collected from patients with clinically diagnosed RA, osteoarthritis (OA), healthy control (HC) subjects at the Second Affiliated Hospital of Zhejiang University School of Medicine from June 2020 to October 2021. Ethical approval was obtained from the Ethics Committee of the Second Affiliated Hospital of Zhejiang University School of Medicine, Hangzhou, China (approval number: 2020-306). All patients signed an informed consent form. The RA patients met the American College of Rheumatology 1987 diagnostic criteria and the American College of Rheumatology/European League Against Rheumatism 2010 diagnostic criteria for RA [30, 31]. Clinical and laboratory features in patients with RA and OA and HC subjects were shown in Supplementary Table 1 (synovia) and Table 3 (synovial tissue samples), respectively.

Transmission electron microscopy

After treatment with 1 µg/mL Semaphorin 5A (Q13591, novoprotein) and/or 150 nM RSL3 (HY-100218A; MCE), approximately $(2-6) \times 10^6$ cells were collected in a 1.5-mL eppendorf tube and fixed with 1 mL of 2.5% Glutaraldehyde Fixed Solution (PH9003; PHYGENE). After rinsing with PBS, the samples were fixed with 50 µL of 1% osmium for 1 h and then fixed/stained with 100 µL of 2% uranium acetate aqueous solution for 30 min.

Next, the samples were successively dehydrated through a series of different ethanol concentrations and 100% acetone. After embedding, ultrathin sections were cut, stained, and observed under a transmission electron microscope (Talos F200C 200kv, Thermo Scientific).

Detection of lipid peroxidation level

Malondialdehyde (MDA), a product of membrane lipid peroxidation, shows a positive correlation with ferroptosis. A Lipid Oxidation (MDA) Assay Kit (S0131S; Beyotime) was used to detect MDA levels in SFs. Briefly, the cells were lysed using Lysis Solution (P0013; Beyotime) and centrifuged. The supernatant was collected and the protein concentration was detected using a BCA Protein Assay Kit (P0010; Beyotime). After preparing the MDA working solution in accordance with the kit instructions, a sample aliquot or blank control was added for determination. The test solution was boiled for 15 min, cooled to room temperature, and centrifuged to obtain the supernatant. Finally, the absorbance was measured at 532 nm using a multimode reader (Spark Cyto, TECAN).

C11-BODIPY 581/591 is a lipid-soluble fluorescent probe used to indicate lipid peroxidation in living cells. According to the manufacturer's instructions, an appropriate volume of C11-BODIPY 581/591 (RM02821, ABclonal) was added to the cells and incubated for 30 min. After removal of the excess dye by washing with PBS, a cell pellet was obtained by 0.25% trypsin digestion and centrifugation. The cells were then resuspended in PBS containing 5% fetal bovine serum (FBS) and detected by flow cytometry (CytoFLEX LX, Beckman Coulter).

Treatment with small-molecule inhibitor

Cells were treated with small-molecule inhibitors of PI3K-AKT-mTOR signaling to detect the function of Semaphorin 5A after inhibition of this pathway. They are PI3K inhibitor, pictilisib (GDC-0941, HY-50094; MCE) 10 µM, AKT inhibitor, MK-2206 (HY-108232; MCE) 5 µM, and mTOR inhibitor, temsirolimus (CCI-779, HY-50910; MCE) 100 nM.

Detection of reactive oxygen species (ROS) level

Intracellular ROS levels were measured with a Reactive Oxygen Species Assay Kit (S0033S; Beyotime). Briefly, cells were stimulated with a ROS positive control (Rosup) or Semaphorin 5A for 24 h and then loaded with DCFH-DA as a probe. Finally, the cells were detected by flow cytometry (CytoFLEX LX, Beckman Coulter).

Statistical analysis

IBM SPSS Statistics 25 (IBM Corp.) and GraphPad Prism 8.0 (GraphPad Software Inc.) were used to analyze the data and draw scatter diagrams. For normally distributed data expressed as mean values \pm SD (standard deviation), the differences between groups were analyzed by Student's *t* test. For nonparametric data, results were expressed as median (range) values, and the differences between groups were analyzed by the Mann-Whitney *U* test. Spearman's correlation coefficient was applied to detect the correlation between two groups. *P* values less than 0.05 were considered significant. The number of independent technical repeats (*n*) are indicated in figure legends.

DATA AVAILABILITY

The data used to support the findings of this study are available from the corresponding author upon request.

MATERIALS AVAILABILITY

The materials used to support the findings of this study are available from the corresponding author upon request.

REFERENCES

- Sparks JA. Rheumatoid arthritis. *Ann Intern Med.* 2019;170:1tc1–16.
- Dakin SG, Coles M, Sherlock JP, Powrie F, Carr AJ, Buckley CD. Pathogenic stromal cells as therapeutic targets in joint inflammation. *Nat Rev Rheumatol.* 2018;14:714–26.
- Kuo D, Ding J, Cohn IS, Zhang F, Wei K, Rao DA, et al. HBEGF(+) macrophages in rheumatoid arthritis induce fibroblast invasiveness. *Sci Transl Med.* 2019;11:eaau8587.
- Take Y, Nakata K, Hashimoto J, Tsuboi H, Nishimoto N, Ochi T, et al. Specifically modified osteopontin in rheumatoid arthritis fibroblast-like synoviocytes

- supports interaction with B cells and enhances production of interleukin-6. *Arthritis Rheum.* 2009;60:3591–601.
5. Nishide M, Kumanogoh A. The role of semaphorins in immune responses and autoimmune rheumatic diseases. *Nat Rev Rheumatol.* 2018;14:19–31.
 6. Du Y, Wu X, Chen M, Wang W, Xv W, Ye L, et al. Elevated semaphorin5A in systemic lupus erythematosus is in association with disease activity and lupus nephritis. *Clin Exp Immunol.* 2017;188:234–42.
 7. Gras C, Eiz-Vesper B, Jaimes Y, Immenschuh S, Jacobs R, Witte T, et al. Secreted semaphorin 5A activates immune effector cells and is a biomarker for rheumatoid arthritis. *Arthritis Rheumatol.* 2014;66:1461–71.
 8. Du Y, Wang JW, Zhou MJ, Wu XY, Xue J, Wu HX. [The expression of Semaphorin 5A in patients with rheumatoid arthritis and its effect on osteoclastogenesis]. *Zhonghua Nei Ke za Zhi.* 2018;57:661–6.
 9. Xiao C, Lv C, Sun S, Zhao H, Ling H, Li M, et al. TSP1 is the essential domain of SEMA5A involved in pannus formation in rheumatoid arthritis. *Rheumatology (Oxford).* 2021;60:5833–42.
 10. Falconer J, Murphy AN, Young SP, Clark AR, Tiziani S, Guma M, et al. Review: synovial cell metabolism and chronic inflammation in rheumatoid arthritis. *Arthritis Rheumatol.* 2018;70:984–99.
 11. Kato M, Ospelt C, Gay RE, Gay S, Klein K. Dual role of autophagy in stress-induced cell death in rheumatoid arthritis synovial fibroblasts. *Arthritis Rheumatol.* 2014;66:40–8.
 12. Dixon SJ, Lemberg KM, Lamprecht MR, Skouta R, Zaitsev EM, Gleason CE, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell.* 2012;149:1060–72.
 13. Li P, Jiang M, Li K, Li H, Zhou Y, Xiao X, et al. Glutathione peroxidase 4-regulated neutrophil ferroptosis induces systemic autoimmunity. *Nat Immunol.* 2021;22:1107–17.
 14. Matsuoka RL, Chivatakarn O, Badea TC, Samuels IS, Cahill H, Katayama K, et al. Class 5 transmembrane semaphorins control selective Mammalian retinal lamination and function. *Neuron.* 2011;71:460–73.
 15. Artigiani S, Conrotto P, Fazzari P, Gilestro GF, Barberis D, Giordano S, et al. Plexin-B3 is a functional receptor for semaphorin 5A. *EMBO Rep.* 2004;5:710–4.
 16. Zhang F, Wei K, Slowikowski K, Fonseka CY, Rao DA, Kelly S, et al. Defining inflammatory cell states in rheumatoid arthritis joint synovial tissues by integrating single-cell transcriptomics and mass cytometry. *Nat Immunol.* 2019;20:928–42.
 17. Zheng J, Conrad M. The metabolic underpinnings of ferroptosis. *Cell Metab.* 2020;32:920–37.
 18. Yi J, Zhu J, Wu J, Thompson CB, Jiang X. Oncogenic activation of PI3K-AKT-mTOR signaling suppresses ferroptosis via SREBP-mediated lipogenesis. *Proc Natl Acad Sci USA.* 2020;117:31189–97.
 19. Zhang Y, Swanda RV, Nie L, Liu X, Wang C, Lee H, et al. mTORC1 couples cyst(e)ine availability with GPX4 protein synthesis and ferroptosis regulation. *Nat Commun.* 2021;12:1589.
 20. You S, Koh JH, Leng L, Kim WU, Bucala R. The tumor-like phenotype of rheumatoid synovium: molecular profiling and prospects for precision medicine. *Arthritis Rheumatol.* 2018;70:637–52.
 21. Nygaard G, Firestein GS. Restoring synovial homeostasis in rheumatoid arthritis by targeting fibroblast-like synoviocytes. *Nat Rev Rheumatol.* 2020;16:316–33.
 22. Bartok B, Firestein GS. Fibroblast-like synoviocytes: key effector cells in rheumatoid arthritis. *Immunological Rev.* 2010;233:233–55.
 23. Sadanandam A, Rosenbaugh EG, Singh S, Varney M, Singh RK. Semaphorin 5A promotes angiogenesis by increasing endothelial cell proliferation, migration, and decreasing apoptosis. *Microvasc Res.* 2010;79:1–9.
 24. Pan G, Zhu Z, Huang J, Yang C, Yang Y, Wang Y, et al. Semaphorin 5A promotes gastric cancer invasion/metastasis via urokinase-type plasminogen activator/phosphoinositide 3-kinase/protein kinase B. *Digestive Dis Sci.* 2013;58:2197–204.
 25. Wu J, Feng Z, Chen L, Li Y, Bian H, Geng J, et al. TNF antagonist sensitizes synovial fibroblasts to ferroptotic cell death in collagen-induced arthritis mouse models. *Nat Commun.* 2022;13:676.
 26. Udalova IA, Mantovani A, Feldmann M. Macrophage heterogeneity in the context of rheumatoid arthritis. *Nat Rev Rheumatol.* 2016;12:472–85.
 27. Culemann S, Grüneboom A, Nicolás-Ávila J, Weidner D, Lämmle KF, Rothe T, et al. Locally renewing resident synovial macrophages provide a protective barrier for the joint. *Nature.* 2019;572:670–5.
 28. Lee YS, Lee DH, Choudry HA, Bartlett DL, Lee YJ. Ferroptosis-induced endoplasmic reticulum stress: cross-talk between ferroptosis and apoptosis. *Mol Cancer Res: MCR.* 2018;16:1073–6.
 29. Zhao T, Yang Q, Xi Y, Xie Z, Shen J, Li Z, et al. Ferroptosis in rheumatoid arthritis: a potential therapeutic strategy. *Front Immunol.* 2022;13:779585.
 30. Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO 3rd, et al. 2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum.* 2010;62:2569–81.
 31. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum.* 1988;31:315–24.

ACKNOWLEDGEMENTS

The authors thank the doctors from the Department of Orthopedics, the Second Affiliated Hospital of Zhejiang University School of Medicine for helping us obtain the synovial specimens from the joints. We also thank Chenyu Yang in the Center of Cryo-Electron Microscopy (CCEM), Zhejiang University for her technical assistance in the transmission electron microscopy. We would like to thank BioRender (<https://biorender.com/>) for providing the template for the drawing mechanism diagram.

AUTHOR CONTRIBUTIONS

QC and YD performed the study concept and design. QC and MC performed the most experiments, analyzed the data, and drafted the manuscripts. QC, MC, ML, XC, Jieying Xu, Jing Xue, HW, YD contributed to the collecting of clinical samples, interpretation of data and material support. YD and HW performed supervision, project administration, and funding support. All authors read and approved the final paper.

FUNDING

This work was supported by the National Natural Science Foundation of China (No. 82071810) and the Natural Science Foundation of Zhejiang Province (No. LY20H100007).

COMPETING INTERESTS

The authors declare no competing interests.

ETHICS STATEMENT

Our study was approved by the Ethics Committee of the Second Affiliated Hospital of Zhejiang University School of Medicine (approval number: 2020-306). All participants signed a written informed consent form. The study was performed in accordance with the Declaration of Helsinki.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41419-022-05065-4>.

Correspondence and requests for materials should be addressed to Huaxiang Wu or Yan Du.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022, corrected publication 2023