



# Cumulative inactivation of *Nell-1* in *Wnt1* expressing cell lineages results in craniofacial skeletal hypoplasia and postnatal hydrocephalus

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## Abstract

Upregulation of *Nell-1* has been associated with craniosynostosis (CS) in humans, and validated in a mouse transgenic *Nell-1* overexpression model. Global *Nell-1* inactivation in mice by *N*-ethyl-*N*-nitrosourea (ENU) mutagenesis results in neonatal lethality with skeletal abnormalities including cleidocranial dysplasia (CCD)-like calvarial bone defects. This study further defines the role of *Nell-1* in craniofacial skeletogenesis by investigating specific inactivation of *Nell-1* in *Wnt1* expressing cell lineages due to the importance of cranial neural crest cells (CNCCs) in craniofacial tissue development. *Nell-1*<sup>fllox/fllox</sup>; *Wnt1*-Cre (*Nell-1*<sup>Wnt1</sup> KO) mice were generated for comprehensive analysis, while the relevant reporter mice were created for CNCC lineage tracing. *Nell-1*<sup>Wnt1</sup> KO mice were born alive, but revealed significant frontonasal and mandibular bone defects with complete penetrance. Immunostaining demonstrated that the affected craniofacial bones exhibited decreased osteogenic and *Wnt*/ $\beta$ -catenin markers (Osteocalcin and active- $\beta$ -catenin). *Nell-1*-deficient CNCCs demonstrated a significant reduction in cell proliferation and osteogenic differentiation. Active- $\beta$ -catenin levels were significantly low in *Nell-1*-deficient CNCCs, but were rescued along with osteogenic capacity to a level close to that of wild-type (WT) cells via exogenous *Nell-1* protein. Surprisingly, 5.4% of young adult *Nell-1*<sup>Wnt1</sup> KO mice developed hydrocephalus with premature ossification of the intrasphenoidal synchondrosis and widened frontal, sagittal, and coronal sutures. Furthermore, the epithelial cells of the choroid plexus and ependymal cells exhibited degenerative changes with misplaced expression of their respective markers, transthyretin and vimentin, as well as dysregulated *Pit-2* expression in hydrocephalic *Nell-1*<sup>Wnt1</sup> KO mice. *Nell-1*<sup>Wnt1</sup> KO embryos at E9.5, 14.5, 17.5, and newborn mice did not exhibit hydrocephalic phenotypes grossly and/or histologically. Collectively, *Nell-1* is a pivotal modulator of CNCCs that is essential for normal development and growth of the cranial vault and base, and mandibles partially via activating the *Wnt*/ $\beta$ -catenin pathway. *Nell-1* may also be critically involved in regulating cerebrospinal fluid homeostasis and in the pathogenesis of postnatal hydrocephalus.

## Introduction

The *Nell-1* gene was originally cloned from a chick embryonic cDNA library as a binding complex to a nuclear protein and was found to be highly conserved among mice and humans [1, 2]. Clinically, human *NELL-1* has been linked to non-syndromic unilateral coronal craniosynostosis (CS) due to its overexpression at premature fusing and fused coronal sutures [3]. Global gain and loss of *Nell-1* mouse models have demonstrated *Nell-1*'s major function in the development of skeletal tissues, particularly for the craniofacial skeleton [4, 5] and for the growth and maintenance of the appendicular bones [6, 7]. Currently, the data suggested that *Nell-1* promotes osteogenesis primarily by

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augmenting the Wnt/ $\beta$ -catenin signaling pathway through interactions with its binding partner,  $\beta$ 1, and/or its newly identified receptor, contactin-associated protein-like 4 (Ctnnap4), in osteoblasts [6, 8]. The manifestations of CS in Nell-1 overexpression transgenic (CMV-Nell-1) neonatal mice and cleidocranial dysplasia (CCD) like calvarial defects in N-ethyl-N-nitrosourea (ENU)-induced Nell-1 deficient (END) neonatal mice have clearly revealed the prominent role of Nell-1 in craniofacial skeletal development [4, 9]. Unfortunately, the drawbacks of super-physiological overexpression of Nell-1 driven by CMV and the neonatal lethality of END mouse models have precluded the study of Nell-1's effects in more defined conditions and at postnatal stages [4, 5]. Advances in mouse genetic manipulation for highly-specific cell/tissue inactivation of target genes and reliable *in vivo* reporter systems enabled further delineation of Nell-1's role in craniofacial conditions [10, 11]. Due to the great importance of cranial neural crest cells (CNCCs) in craniofacial tissue development [12, 13], this study further defines the functionalities of Nell-1 in the craniofacial skeleton originating from CNCCs.

Notably, a variety of craniofacial tissues including the nerves, ganglia, connective tissues, cartilage, and bones are CNCC derivatives [14, 15]. It has been reported that CNCC precursors arise at the lateral margin of the neural fold at the boundary between the surface and neural ectoderm, and migrate ventrolaterally as the cranial ganglia and skeletogenic neural crest cells and populate the branchial arches [14, 16]. Multiple experimental approaches have validated that CNCCs are derived and emigrated from Wnt1-expressing precursor cells in the dorsal central nervous system (CNS) during the early stage of embryogenesis (E8.5–11.5) [16–19]. Consequently, upon confirmation of Wnt1 as a faithful cell lineage marker of CNCCs, it becomes possible to investigate the dynamic involvement of CNCCs in the development and growth of the craniofacial tissues [18, 20]. In particular, studies on the tissue origins of the mouse skull vault and mandibular skeleton using Wnt1-Cre and ROSA26R reporter mouse lines have provided evidence that CNCCs contribute significantly to the development of the craniofacial tissues [14, 21]. It is worth noting that Wnt1-Cre has been widely used for specific gene knockouts in CNCC lineages and has produced tremendous amounts of valuable data in neuroscience and other relevant fields [16, 22], although other CNCC lineage mouse lines are also available [23, 24].

To further define the role and molecular mechanisms of Nell-1 in CNCCs during craniofacial skeletal development and postnatal growth, we have conditionally inactivated the Nell-1 gene using the Cre/loxP recombination system [11]. Here we show that specific inactivation of the Nell-1 gene in Wnt1 expressing cell lineages results in frontonasal and

mandibular bone (MB) hypoplasia and postnatal hydrocephalus with premature ossification of the synchondroses.

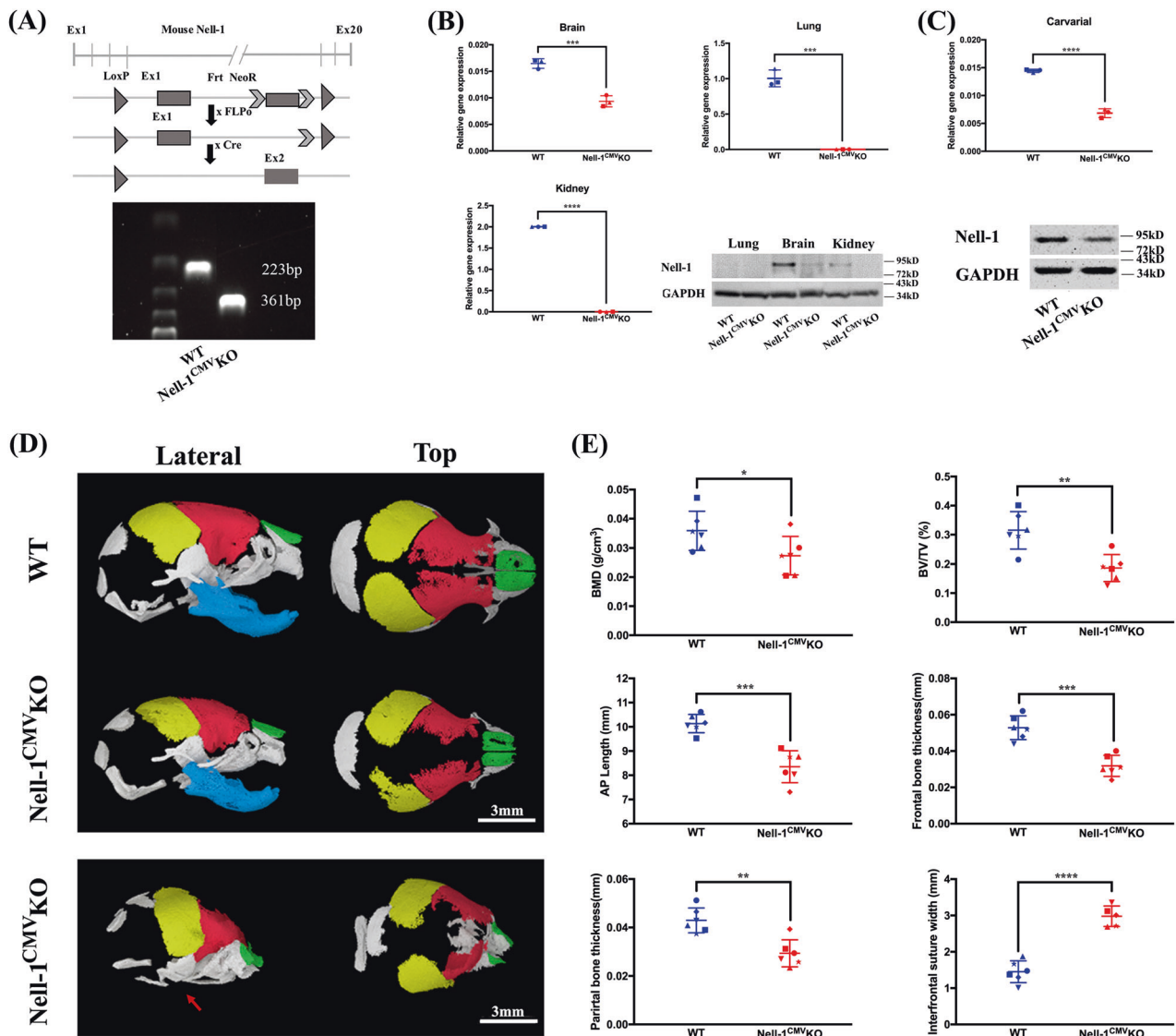
## Materials and methods

### Animals

All animals were cared for according to the institutional guidelines set at the University of California, Los Angeles. Nell-1<sup>flox/flox</sup> mice were generated by flanking Exon 1 containing ATG site with loxP (Strain: B6.129Sv/Pas-Nell1tm from EMMA-European Mouse Mutant Archive) and then excising the FRT-flanked Neo cassette with FLPo-10 delete mice (Stock# 011065 from Jackson Laboratory) in C57BL/6J background (Fig. 1a). Nell-1<sup>flox/flox</sup> mice were mated with either CMV-Cre (Stock# 006054) or Wnt1-Cre (Stock# 022501) mice to obtain Nell-1<sup>flox/flox</sup>;CMV-Cre (Nell-1<sup>CMV</sup>KO) or Nell-1<sup>Wnt1</sup>KO mice as well as Cre negative littermates (WT), respectively. Moreover, R26R (Stock# 003474) and R26R-tdTomato (Stock# 007909) mice were used to yield Nell-1<sup>flox/flox</sup>; Wnt1-Cre; R26R (Nell-1<sup>Wnt1-R26R</sup>KO) or Nell-1<sup>flox/flox</sup>; Wnt1-Cre; R26R-tdTomato (Nell-1<sup>Wnt1-tdTomato</sup> KO) mice for cell lineage validation purpose. Mating was carried out overnight and females were examined for the presence of vaginal plugs and defined as 0.5 dpc (days post coitum). Animal groups were of mixed gender unless otherwise stated. The mutant and R26R mouse genotypes were identified by PCR using primers listed in Supplementary Table S1. Wnt1 expressing cell lineage validation was done either by X-gal staining (see details in Supplementary Material and Methods) or by fluorescent microscopy. Newborn mice (P0) were sacrificed for CNCCs culture, histology, and micro-CT analysis. In addition, mice (P21–48) with hydrocephalus were collected for histology and micro-CT analysis along with age- and gender-matched control littermates.

### Cell culture

Primary calvarial cells were isolated from neonatal mouse calvarial bones following the established protocol reported in previous studies [4, 25]. CNCCs were isolated from the frontal and nasal bones of cranial vault only and expanded in growth medium containing DMEM + 10% heat-inactivated fetal bovine serum (FBS) + 100 U/ml penicillin/streptomycin (P/S) at 37 °C, 5% CO<sub>2</sub>. Passage 2 cells were used for all *in vitro* experiments of this study. Cells were treated with Dickkopf-related protein 1 (DKK-1) or PNU 74654 at the final concentration of 500 ng/ml and 10  $\mu$ M, respectively, 1 h prior to adding Nell-1 protein at 1000 ng/ml [6, 7, 26]. The osteogenic induction was performed in osteogenic differentiation (OD) medium containing 100 nmol/L dexamethasone, 10 mmol/L  $\beta$ -glycerophosphate,



**Fig. 1** Generation of Nell-1 conditional mutant mice. **a** Schematic diagrams of the genetic construct for floxed Nell-1 generation, the breeding strategy for Nell-1<sup>CMV</sup>KO mice, and genotyping (lower row); **b** Nell-1 mRNA and protein levels vary among different organs in Nell-1<sup>CMV</sup>KO newborn mice (P0); **c** Nell-1<sup>CMV</sup>KO newborn mice exhibit significantly decreased Nell-1 in the calvarial bones; **d** 3D reconstructions of the craniofacial skeleton of Nell-1<sup>CMV</sup>KO newborn

mice demonstrate visible hypoplasia and cranial suture widening. Extreme deficits were also observed in the mandibular bones of Nell-1<sup>CMV</sup>KO newborn mice, but less frequently (arrow in lower row); **e** Quantification of the craniofacial bones as measured by BV/TV, BMD, bone thickness, and suture width confirm their hypoplastic changes. \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.005$ ; \*\*\*\* $P < 0.001$ ;  $n = 3$  for **b** and **c**;  $n = 6$  for **e**

and 0.05 mmol/L L-ascorbic acid-2-phosphate in growth medium.

### Cell proliferation and apoptosis

To determine the effect of Nell-1 inactivation on the proliferation of CNCCs, cells from Nell-1<sup>Wnt1</sup>KO and WT mice were examined either by the Chick-iT EdU Alexa Fluor<sup>TM</sup> 488 Image Kit (Thermo Fisher Scientific, Waltham, MA, USA) and fluorescence microscopy (OLYMPUS BX51, Japan) for direct visualization or by cell cycle analysis for quantifying the percentage of S (DNA synthesis replication)

phase cells using flow cytometry [27]. For apoptosis analyses, the Annexin V and propidium iodide (PI) Kit (BD Pharmingen, San Jose, CA, USA) was used for cell staining, followed by flow cytometric analysis for quantifying the percentage of cells at early and late apoptosis by the same protocol as described previously [28].

### Alkaline phosphatase (ALP) and mineralization assays

Assays for CNCCs osteogenic differentiation were adapted from our previous publications using the Leukocytes ALP

Staining Kit (86R-1KT, Sigma-Aldrich Corp., St. Louis, MO, USA). Alizarin Red S (ARS) staining was carried out with 2% Alizarin Red stains (CM-0058, Lifeline, Frederick, MD, USA). The staining intensity of ALP and ARS was measured and quantified using Image-Pro Plus 6.0 (Media Cybernetics, Rockville, MD, USA).

### Western blot and Real-time quantitative PCR (RT-qPCR)

Western blot analysis was performed as previously described [6, 29]. Nell-1 antibody (1:500) (GeneTex, Irvine, CA, USA), active- $\beta$ -catenin (1:1000) (Cell Signaling, USA), and GAPDH (1:1000) (Cell Signaling, USA) primary antibodies were used at different dilutions. Real-time PCR was performed using the QuantStudio<sup>TM</sup> 3 real-time PCR System instrument (Applied Biosystems, Foster City, CA, USA) as described previously [6]. The primer sequences are summarized in Supplementary Table S2. All data are representative of three experimental sets of cells or the three mice tissue specimens with PCR duplicates and are presented as the fold change.

### High-resolution micro-CT analysis

High-resolution micro-CT scanning was performed on mice crania (Skyscan 1176, Bruker-microCT, Kontich, Belgium) of neonatal stage for the most samples, and postnatal stage when hydrocephaly became apparent for some samples, respectively. The images were scanned with the setting of 10  $\mu$ m pixel size, 50 kV Voltage, 500  $\mu$ A current, 0.5 mm Al filter, 1000 ms exposure time, 0.500 rotation step, and 3 frame averaging. All resulting images were reconstructed by NRecon (Version 1.7.1.0, Bruker-microCT, Kontich, Belgium) as described previously [6]. The detailed settings are described in Supplementary Material and Methods. Quantitative data are presented as the mean  $\pm$  SD and were analyzed using the two-tailed Student's *t* test with  $p < 0.05$  considered significant.

### Histological analysis and immunohistochemistry

All craniofacial samples of embryonic, neonatal, and postnatal stages were fixed in 4% paraformaldehyde (Sigma-Aldrich, St. Louis, MO, USA) for 24–48 h, then processed for paraffin embedding. Hematoxylin and Eosin (H&E) staining (H&E stain kit, abcam, Cambridge, England) was used per standard protocols, while immunohistochemistry and its semi-quantification were performed as described previously [4, 9]. The working concentration of primary antibodies was set at 2–5  $\mu$ g/ml in general and the primary antibodies are listed in Supplementary Table S3. Photomicrographs were acquired using Olympus BX 51 and IX 71 microscopes equipped with Cell Sense digital imaging system (Olympus, Japan).

### Statistical analysis

All quantifiable data were presented as mean  $\pm$  SD. The gross assessment was performed with 15 mice of each mutant and wild type group, while the detailed micro-CT and histological analyses were performed with 3 to 6 mice of each genotype for the various parameters. All in vitro experiments were run in triplicate. An unpaired two-tailed Student's *t* test was used to analyze the data. A value of  $p < 0.05$  was considered to indicate a statistically significant difference. The statistical analyses were performed in consultation with the UCLA Statistical Biomathematical Consulting Clinic.

## Results

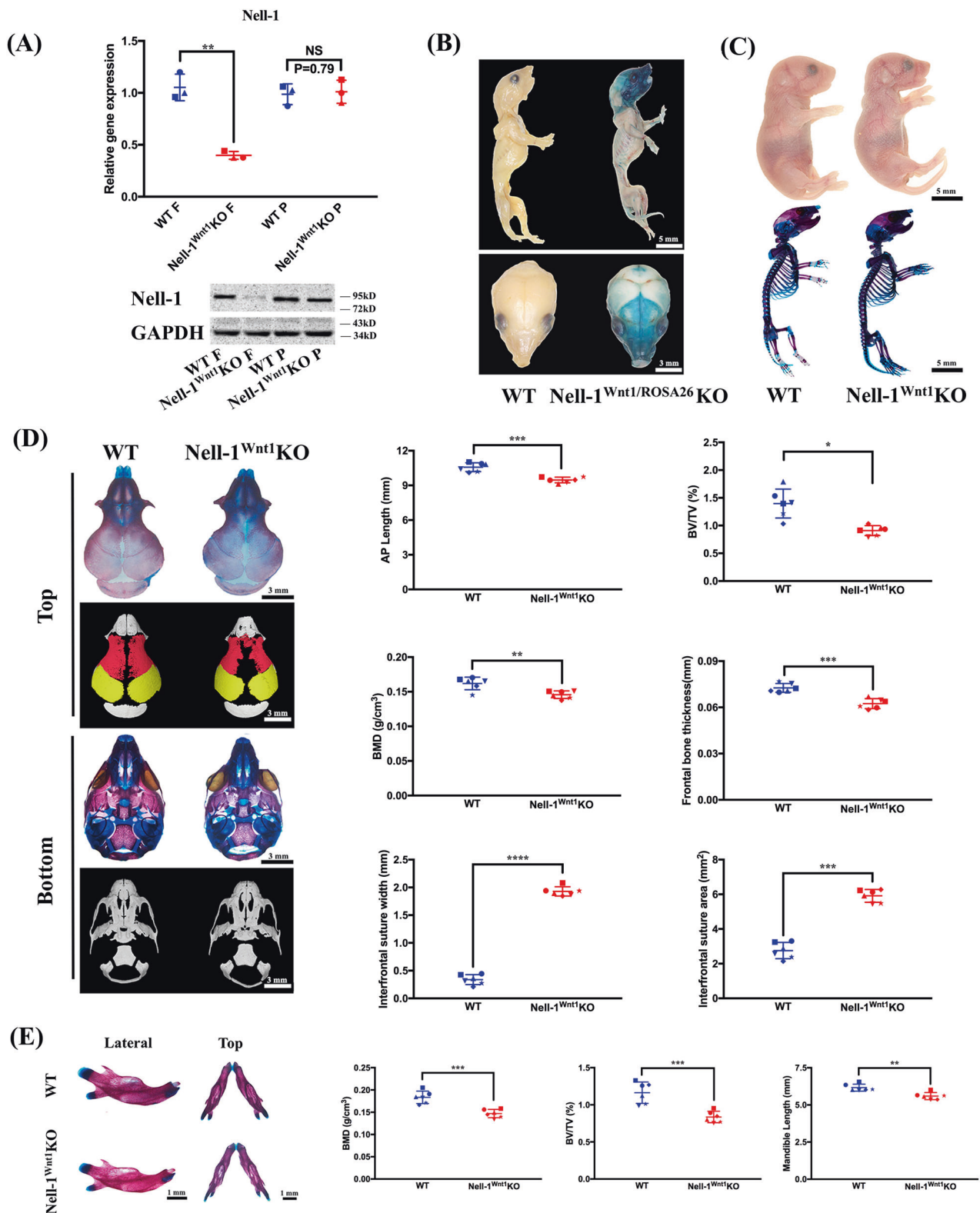
### Nell-1 is validated to be a key factor for craniofacial anomalies in new conditional Nell-1 knockout mutants

Upon successful rederivation of the Nell-1<sup>flx/flx</sup> mouse line, the complete deletion of the Neo cassette was achieved with FLPo-10 deleter mice to eliminate the possible genetic influences to the resultant floxed mutant mice [30]. CMV-Cre [31] and Wnt1-Cre [32] transgenic mice were then used to obtain Nell-1<sup>CMV</sup>KO mice for the initial screening or Nell-1<sup>Wnt1</sup>KO mice for comprehensive analyses. Genotyping and Nell-1 gene and protein expression were performed to confirm the knockout in the respective tissues (Fig. 1a–c). Due to the neonatal lethality seen in ENU-induced Nell-1 deficient (END) mice [5, 9], we first selected CMV-Cre as a global knockout strategy to evaluate the newly generated floxed Nell-1 mice. Surprisingly, the majority of Nell-1<sup>CMV</sup>KO mice were born alive but demonstrated various degrees of hypoplastic craniofacial skeletal defects as characterized by micro-CT scanning (Fig. 1d, e). Notably, in the most severe form, newborn Nell-1<sup>CMV</sup>KO mice were found still-born and exhibited extreme craniofacial malformations with grossly deficient or absent mandibles, severe frontal and parietal bone defects, and widened anterior fontanels as characterized by Micro-CT analysis (Fig. 1d, bottom row). Thus, the phenotypical changes in Nell-1<sup>CMV</sup>KO newborns once again indicate the pivotal role of Nell-1 in craniofacial skeletal development, and validates the floxed Nell-1 mouse model.

### Nell-1 inactivation in CNCCs induces specific craniofacial bone defects with high and consistent penetrance

Given the importance of CNCCs to the development and growth of the craniofacial tissues, Nell-1<sup>Wnt1</sup>KO mice were used for comprehensive analyses. To confirm and trace the





Nell-1 knockout in Wnt1 lineages, the R26R mouse [33] was utilized to make Nell-1<sup>Wnt1-R26R</sup> KO mice for validation in certain experiments. All Nell-1<sup>Wnt1</sup>KO and Nell-1<sup>Wnt1-R26R</sup> KO mice were born alive without obvious visible

abnormalities. Genotyping, Nell-1 gene and protein expression analyses, and X-gal staining were performed to confirm the CNCCs-derived tissue specific knockouts (Fig. 2a, b). Grossly, there were no visible abnormalities of the

◀ **Fig. 2** *Nell-1* inactivation in CNCCs results in frontonasal bone hypoplasia and micrognathia in *Nell-1<sup>Wnt1</sup>KO* newborn mice. **a** As a result of *Nell-1* inactivation in CNCCs, *Nell-1* mRNA, and protein expression decreased only in the frontonasal bones (*Nell-1<sup>Wnt1</sup>KO* (F) and not in the parietal bones (*Nell-1<sup>Wnt1</sup>KO* P); **b** The craniofacial tissues stained positive for X-gal, with the frontonasal and mandibular bones being particularly targeted in *Nell-1<sup>Wnt1-R26R</sup>KO* mice; **c** gross appearance and skeletal staining showed no visible defects of the appendicular and axial bones in neonatal *Nell-1<sup>Wnt1</sup>KO* mice; **d** detailed analyses of the cranial vault of newborn mice exhibited significant hypoplasia of the frontonasal bones, while the synchondroses of the cranial base were unaffected as demonstrated by skeletal staining, high resolution micro-CT imaging, and quantification as measured by bone volume (BV/TV), bone mineral density (BMD), frontal bone thickness and defect area, the suture width as well as the anteroposterior length of calvarial vault; **e** *Nell-1<sup>Wnt1</sup>KO* mice exhibited significantly smaller and shorter mandibles (micrognathia) with decreased BV/TV and BMD. NS no significance; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.005$ ; \*\*\*\* $P < 0.001$ ;  $n = 3$  for **a**;  $n = 6$  for **d** and **e**)

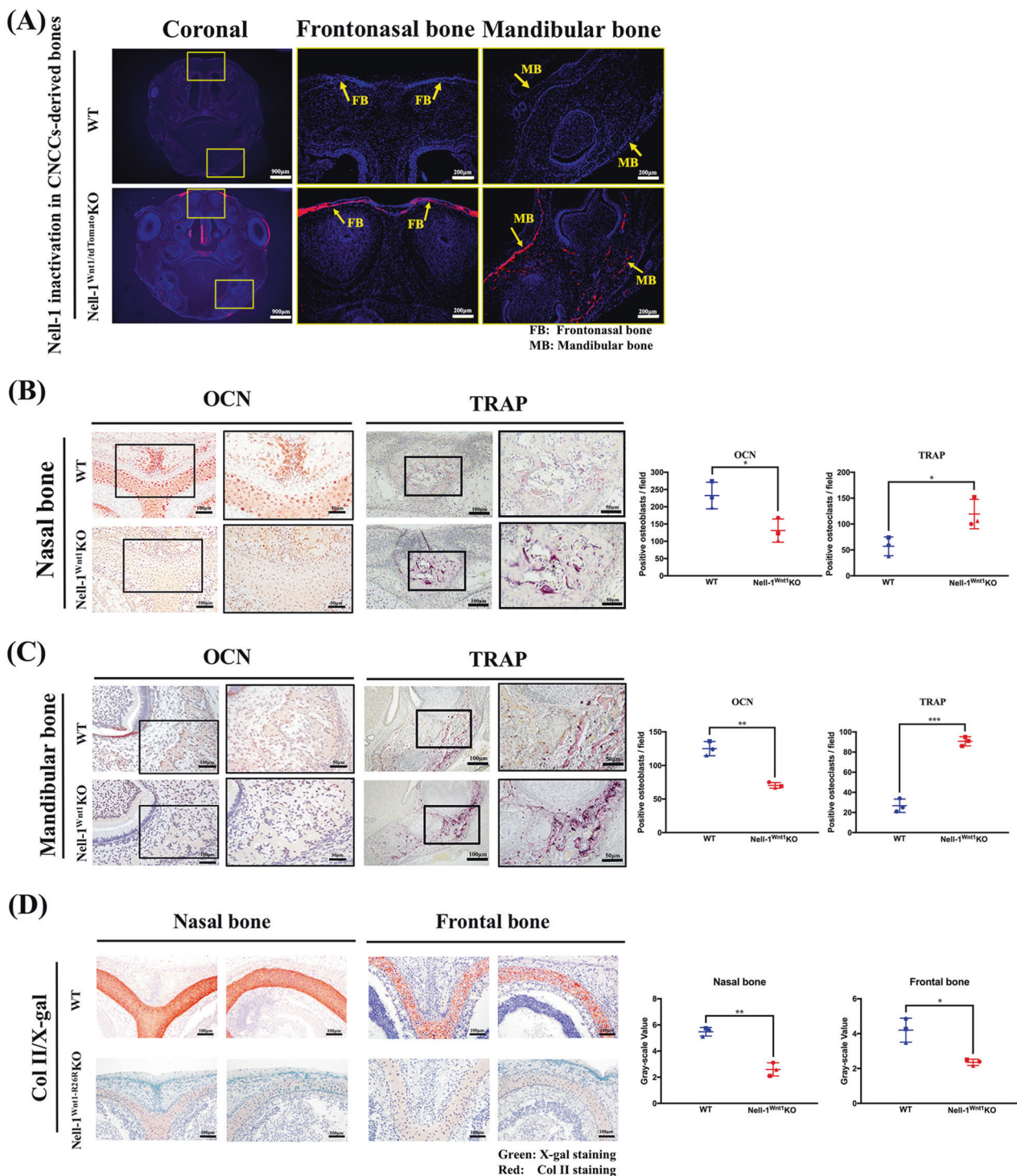
appendicular and axial bones in *Nell-1<sup>Wnt1-R26R</sup>KO* newborns as compared with WT littermates (Fig. 2b, c). Detailed analyses of the craniofacial bones (P0) were performed with skeletal staining and micro-CT analyses (Fig. 2d). *Nell-1<sup>Wnt1</sup>KO* mice had significantly reduced mineralization and volume of the frontonasal bones (FB), while the parietal bones and cranial base remained unaffected compared to WT littermates. Notably, the size, bone volume over tissue volume (BV/TV), and bone mineral density (BMD) of the mandible of *Nell-1<sup>Wnt1</sup>KO* mice (P0) were also significantly reduced (Fig. 2e). However, there is no significant difference in the comparative micro CT examination of the affected cranial vault vs. MB in *Nell-1<sup>Wnt1</sup>KO* mice in terms of severity of skeletal hypoplasia (Supplementary Fig. 1).

Expectedly, the frontonasal bones (FB) and MB were specifically targeted for *Nell-1* inactivation in *Nell-1<sup>Wnt1-tdTomato</sup>KO* mice (P0) (Fig. 3a). Osteocalcin (OCN) immunostaining clearly revealed decreased osteogenic properties in both the frontonasal and MB of *Nell-1<sup>Wnt1</sup>KO* mice (P0) (Fig. 3b, c). Interestingly, tartrate-resistant acid phosphatase (TRAP) staining exhibited more positive cells in both the frontonasal and MB of *Nell-1<sup>Wnt1</sup>KO* mice (Fig. 3b, c). Histomorphometric analysis demonstrated a significant decrease in the number of osteoblasts and an increase in the number of osteoclasts in affected bones (Fig. 3b, c). In sharp contrast to WT littermates, *Nell-1<sup>Wnt1-R26R</sup>KO* mice had significantly less Collagen type II (Col2) in the X-gal positive-stained cartilaginous wall and septum of the nasal cavity under the FB (P0) (Fig. 3d). Overall, specific *Nell-1* inactivation in CNCCs consistently resulted in frontonasal skeletal hypoplasia and micrognathia in newborn mice with complete penetrance.

## ***Nell-1<sup>Wnt1</sup>KO* mice develop postnatal hydrocephalus at significantly higher rates**

In contrast to the neonatal lethality of END mice [5], the majority of *Nell-1<sup>Wnt1</sup>KO* mice are born full-term, fertile, and live to maturity without visible clinical manifestations different from WT littermates. However, among *Nell-1<sup>Wnt1</sup>KO* mice, 5.4% of young adult mice ranging from 21 to 48 days old developed classical domed head phenotypes of hydrocephalus (Fig. 4a, b), a rate significantly higher than the <0.01% prevalence of spontaneous hydrocephalus in C57BL/6J mice (JAX Notes@ <https://www.jax.org/news-and-insights/1991/april/spontaneous-hydrocephalus-in-inbred-strains-of-mice>) (\*\* $P < 0.01$ ). Notably, hydrocephalic mice were characterized by smaller and less mineralized FB with widened frontal, sagittal, and coronal sutures on the calvarial vault along with various degrees of premature ossification of intrasphenoidal synchondrosis (ISS) and/or sphenoid-occipital synchondrosis (SOS) (Fig. 4c, d). At the tissue level, profound dilation of the brain ventricles along with compressed and thinner brain cortex layers were observed, likely secondary to cerebrospinal fluid (CSF) accumulation (Fig. 4e). Histology and OCN immunohistochemistry of the cranial base supported premature fusion/ossification of the synchondroses (Fig. 4f, g and Supplementary Fig. 2A). The epithelial cells of the choroid plexus (CP) were enlarged with rich cytoplasm, and the ependymal cells were flattened with focal loss of apical structures (Fig. 4h). Vimentin, a marker of ependymal cells [34], was undetectable in the flattened ependymal cells and/or mis-expressed in some of the enlarged CP epithelial cells, while the expression of prealbumin/transferrin (TTR), a specific marker of CP epithelial cells [35], was significantly lower or even undetectable in hydrocephalic *Nell-1<sup>Wnt1</sup>KO* mice (Fig. 4i and Supplementary Fig. 2B). Next, a type III Na<sup>+</sup>-dependent inorganic phosphate transporter, Pit-2, was explored due to the fact that it was one of the phosphate transporters that *Nell-1* used to increase pre-osteoblast mineralization [36] and its causative role in hydrocephalus in Pit-2 deficient mice [37]. The intracellular protein level of Pit-2 was significantly reduced in CP epithelial cells of the fourth ventricle, but slightly elevated in both swollen CP epithelial cells and flattened ependymal cells of the lateral ventricle in hydrocephalic *Nell-1<sup>Wnt1</sup>KO* mice as compared with those cells in WT mice (Fig. 4j and Supplementary Fig. 2C). These results may reflect the functional properties and distinct pathological changes of CP epithelial and ependymal cells at different locations of brain ventricular system under hydrocephalic condition in *Nell-1<sup>Wnt1</sup>KO* mice.

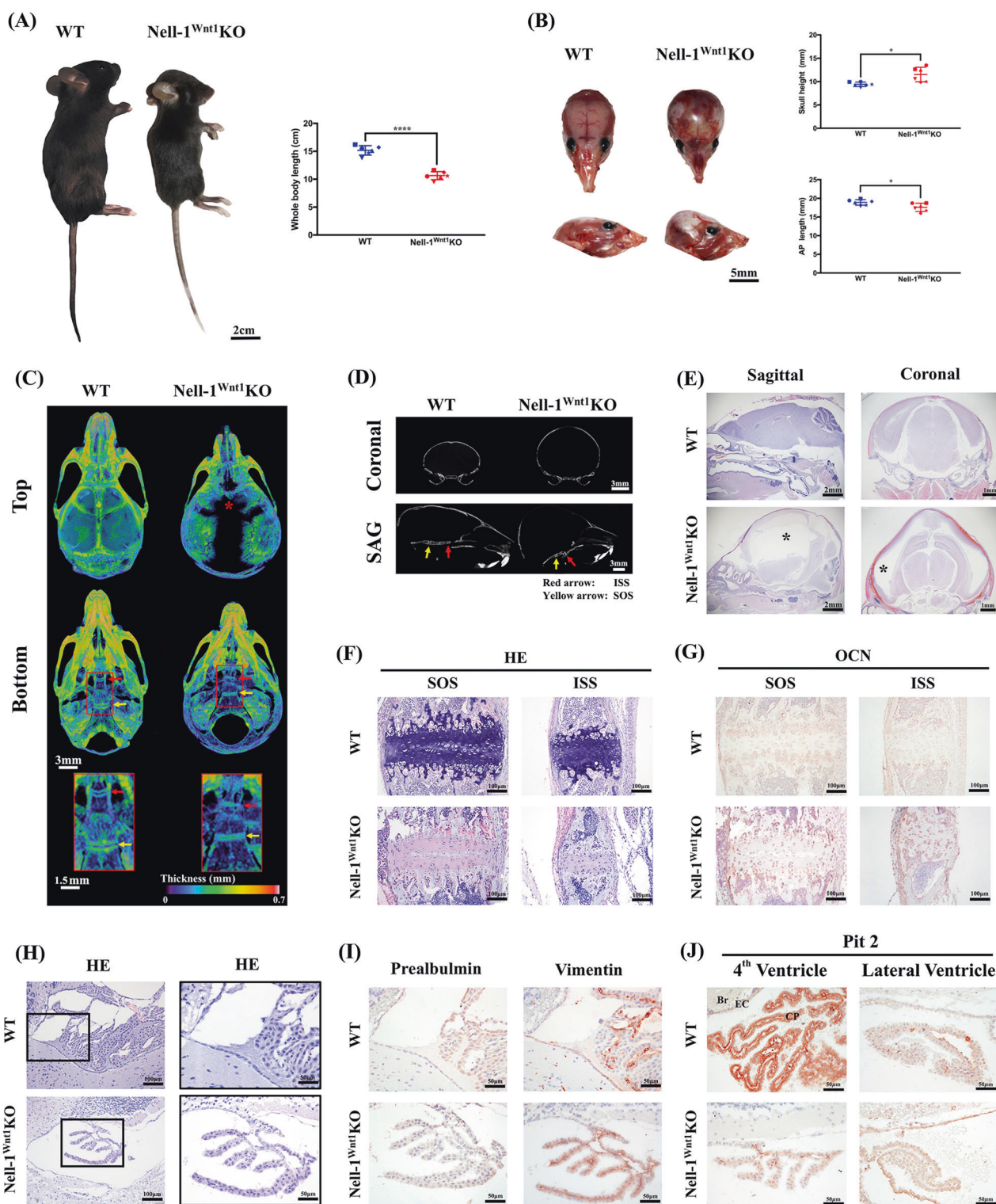
To determine if hydrocephalus occurred as a congenital condition, the embryos at E9.5, 14.5, and 17.5 were



**Fig. 3** Nell-1<sup>Wnt1</sup>KO newborn mice demonstrate defective osteochondrogenesis in CNCC-derived bones. **a** Cell lineage tracking in Nell-1<sup>Wnt1-tdTomato</sup>KO mice clearly showed red fluorescence (yellow arrows) in the frontonasal bones (FB) and mandibular bones (MB) which are derived from Wnt1 expressing CNCC lineages; **b, c** More OCN positive cells were observed in the osteogenic front of the calvarial bone plate and mandibular bones in WT samples than in Nell-1<sup>Wnt1</sup>KO samples. In contrast to OCN staining, TRAP positive staining

showed a drastic increase in both the frontonasal and mandibular bones of Nell-1<sup>Wnt1</sup>KO mice. The distinct correlation of OCN and TRAP staining in these bones was further revealed by quantitative analyses between WT and Nell-1<sup>Wnt1</sup>KO mice; **d** The maturation of CNCC-derived cartilaginous tissues was severely inhibited in Nell-1<sup>Wnt1-R26R</sup>KO mice with evidence of extremely low levels of Collagen type II matrix. \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.005$ ;  $n = 3$  for **b-d**





subsequently collected and analyzed although we did not observe noticeable hydrocephalic manifestations in any newborn mice. *Nell-1<sup>Wnt1-R26R</sup>* KO embryos were stained positive by X-gal to display areas of *Nell-1* inactivation and to assist lineage tracing (Supplementary Fig. 3A). Grossly, there were no visible malformations of the craniofacial

tissues in either *Wnt1* expressing cell lineage and other origins in *Nell-1<sup>Wnt1-R26R</sup>* KO embryos. At the tissue level, the embryonic and neonatal *Nell-1<sup>Wnt1</sup>* KO mice did not demonstrate any pathological changes of the brain ventricles, although *Nell-1* inactivation did occur in CSF producing cells from *Nell-1<sup>Wnt1-tdTomato</sup>* KO mice



◀ **Fig. 4** Nell-1<sup>Wnt1</sup>KO young adult mice develop postnatal hydrocephalus. **a** The gross appearance of the whole body and **b** skulls of young adult WT and affected Nell-1<sup>Wnt1</sup>KO mice (P32) showed classic dome-shaped heads in KO specimens. The skull length (Anteroposterior length, AP), skull height, and body length were measured, showing significant features of hydrocephalus; **c** Micro-CT imaging with color-coded bone density heat-maps showed distinct decreases in the size and mineral density of the frontonasal bones, defects of the frontal bones (\*), widened cranial sutures, premature ossification of the intraphenoidal synchondrosis (ISS, red arrow) and sphenoid-occipital synchondrosis (SOS, yellow arrow) in hydrocephalic Nell-1<sup>Wnt1</sup>KO mice compared with WT controls; **d** 2D coronal and sagittal X-ray images demonstrated an enlarged hydrocephalic skull profile and premature ossification/fusion of the synchondrosis in the cranial base (arrows) in Nell-1<sup>Wnt1</sup>KO mice; **e**) Histologically, the typical morphological changes of the dilated brain ventricles (\*) and the compressed brain cortex in hydrocephalus were revealed from the sagittal and coronal views. **f** At the cellular level, the composition of the cartilaginous matrix corroborates the ossified synchondrosis observed in hydrocephalic Nell-1<sup>Wnt1</sup>KO mice as compared to WT controls; **g**) stronger OCN immunostaining was observed in the hydrocephalic samples over WT controls; **h** The swollen epithelial cells of the choroid plexus were readily observed in hydrocephalic Nell-1<sup>Wnt1</sup>KO specimens; **i** Immunohistochemistry of the markers of epithelial cell of the choroid plexus, prealbumin, and the ependymal cells, vimentin, were performed respectively. The misplaced expression pattern of vimentin was observed in hydrocephalic Nell-1<sup>Wnt1</sup>KO specimens; **j**) Immunohistochemistry of Pit-2 exhibited dysregulation among CP epithelial and ependymal cells of the dilated brain ventricle and the fourth ventricle in hydrocephalic Nell-1<sup>Wnt1</sup>KO specimens. \**P* < 0.05; \*\*\**P* < 0.001; *n* = 6 for **a** and **b**; *n* = 3 for **i** and **j**

(Supplementary Fig. 3B, C). Micro-CT imaging exhibited obvious delayed development of the MB and cranial vault in E17.5 Nell-1<sup>Wnt1</sup>KO embryos (Supplementary Fig. 3D). Immunostaining of OCN and TRAP staining clearly demonstrated that inactivation of Nell-1 in CNCCs has a significant impact on both osteoblastic and osteoclastic activities, resulting in delayed formation and mineralization of the MB even at E17.5 (Supplementary Fig. 3E).

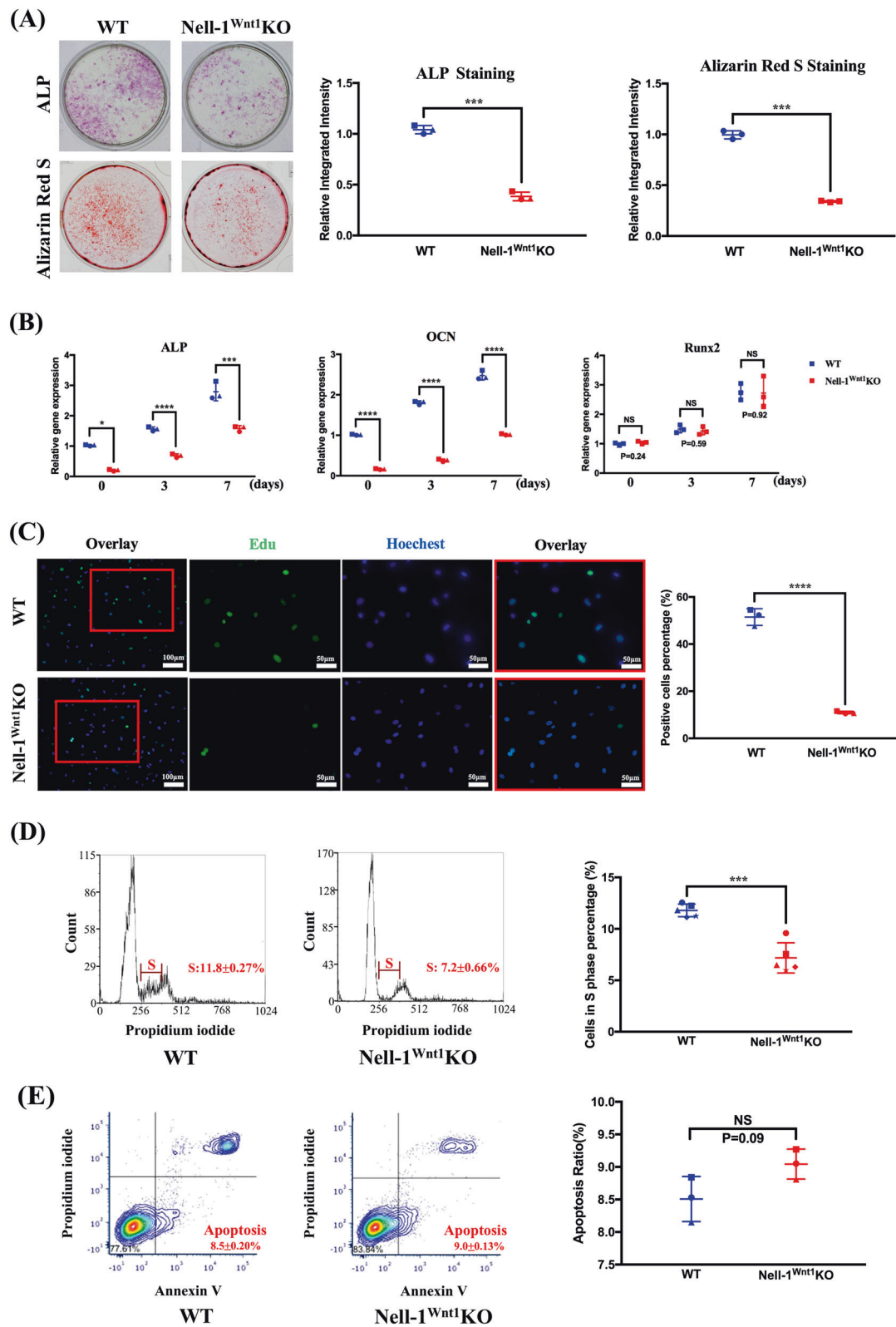
### Nell-1 is required for the full osteogenic differentiation potential of CNCCs

Clearly, the osteogenic potential of primary calvarial cells was significantly diminished in Nell-1 KO cells as evidenced by decreased expression of the osteogenic marker genes, OCN, Osteopontin (OPN), and Runx2 (Supplementary Fig. 4A), lower ALP at day 5, and reduced intensity of mineralization at day 21 using Alizarin red staining (Supplementary Fig. 4B). The later stage inactivation of Nell-1 by adenoviral CMV-Cre delivery to Nell-1<sup>flx/flx</sup> calvarial cells did not affect mineralization as compared with the earlier stage (Day 9) inactivation (Supplementary Fig. 5). Similarly, the CNCCs isolated from the FB of Nell-1<sup>Wnt1</sup>KO mice also exhibited significant reduction of ALP activity at day 7, and reduction of mineralization at day 21 compared with CNCCs from WT mice (Fig. 5a). The quality control

of isolated CNCCs from non-cranial neural crest calvarial cells was monitored at two steps: (1) during physical separation, and (2) during Nell-1 gene expression profiling (Fig. 2a). The gene expression of the major osteogenic markers, ALP and OCN, but not Runx2, was significantly downregulated in CNCCs of Nell-1<sup>Wnt1</sup>KO mice (Fig. 5b), but not in the parietal bone-derived calvarial cells from same batch of Nell-1<sup>Wnt1</sup>KO mice as compared with cells from WT mice (Supplementary Fig. 6). The EdU cell proliferation assay revealed that Nell-1<sup>Wnt1</sup>KO CNCCs had significantly less proliferative potential (Fig. 5c), which was supported with cell cycle analysis showing much lower DNA synthesis replication phase (S phase) CNCCs in Nell-1<sup>Wnt1</sup>KO mice ( $7.2 \pm 0.66\%$ ) than in WT mice ( $11.8 \pm 0.27\%$ ) (Fig. 5d). In contrast, the apoptotic cells were slightly greater in Nell-1<sup>Wnt1</sup>KO CNCCs ( $9.0 \pm 0.13\%$ ) over those of the WT ( $8.5 \pm 0.20\%$ ) (Fig. 5e), while the gating strategies for flow cytometry analyses were strictly enforced (Supplementary Fig. 7). Thus, Nell-1 significantly modulates the proliferation and osteogenic potential of CNCCs, which correlate with the abnormal craniofacial skeletal phenotypes observed in Nell-1<sup>Wnt1</sup>KO mice.

### Wnt/ $\beta$ -catenin signaling is downregulated in Nell-1 deficient CNCCs and its craniofacial bone derivatives

Previously, Nell-1 has been reported as a novel agonist of the canonical Wnt signaling pathway due to its pro-osteogenic properties [29]. The critical impact of the Wnt signaling pathway has also been identified in the development and growth of the craniofacial skeletal tissues [38]. Therefore, it is meaningful to investigate the changes of Wnt/ $\beta$ -catenin signaling in the context of CNCC specific Nell-1 KO mice for craniofacial malformations. Apparently, the inactivation of Nell-1 is responsible for the lower basal level of active  $\beta$ -catenin and the gene expressions of Wnt/ $\beta$ -catenin downstream targets, C-myc and Cyclin D, in CNCCs from Nell-1<sup>Wnt1</sup>KO newborns (Fig. 6a). CNCCs from Nell-1<sup>Wnt1</sup>KO mice exhibited significantly lower expression patterns of Wnt signaling molecules and reduced osteogenic markers, ALP and OCN, compared with those from WT mice (Fig. 6b). Immunostaining of active  $\beta$ -catenin from CNCCs-derived bone tissue sections further demonstrated the significantly defective Wnt canonical signaling pathway in Nell-1<sup>Wnt1</sup>KO mice (P0) (Fig. 6c). Significantly, recombinant Nell-1 protein can partially rescue not only the defective Wnt signaling molecules, but also osteogenic differentiation to a level close to that in WT CNCCs at 1000 ng/ml that was empirically determined with primary CNCCs (Fig. 6b and Supplementary Fig. 8). Nevertheless, Nell-1 protein can also partially rescue osteogenic deficits and elevate Wnt downstream targets of gene expression in WT CNCCs that were treated with Wnt signaling inhibitor



[39, 40], DKK-1 that interferes interaction of Wnt ligands and receptors by binding to *lrp5/6* and Kremen proteins, but not in WT CNCCs being treated with PNU 74654, which prevents Tcf from binding to  $\beta$ -catenin (Fig. 6d–f). As a

result, Nell-1 is a pivotal component of the canonical Wnt pathway in regulating CNCC osteogenesis. However, it remains unclear whether Nell-1 uses distinct mechanisms from Wnt ligands in achieving its regulatory role.

◀ **Fig. 5** Nell-1 inactivation reduces CNCCs osteogenic capacity and cell proliferation. The CNCCs isolated from Nell-1<sup>Wnt1</sup>KO newborn mice demonstrated significantly impaired osteogenic capacity as demonstrated by **a** ALP staining and mineralization, and **b** gene expression of the osteogenic markers ALP, OCN, and Runx2; **c** in vitro EdU labeling exhibited significantly reduced positive cells from Nell-1<sup>Wnt1</sup>KO mice, and **d** flow cytometry cell cycle analyses further confirmed the lower proliferative cell populations in Nell-1<sup>Wnt1</sup>KO mice. Specifically, CNCCs in the DNA synthesis phase (S phase) of WT mice ( $11.8 \pm 0.27\%$ ) were significantly more than that of Nell-1<sup>Wnt1</sup>KO ( $7.2 \pm 0.66\%$ ). **e** While the rates of early apoptosis of CNCCs measured by flow cytometry from Nell-1<sup>Wnt1</sup>KO mice ( $9.0 \pm 0.13\%$ ) was slightly higher than that of WT mice ( $8.5 \pm 0.20\%$ ), no statistically significant difference was found. NS no significance; \* $P < 0.05$ ; \*\*\* $P < 0.005$ ; \*\*\*\* $P < 0.001$

## Discussion

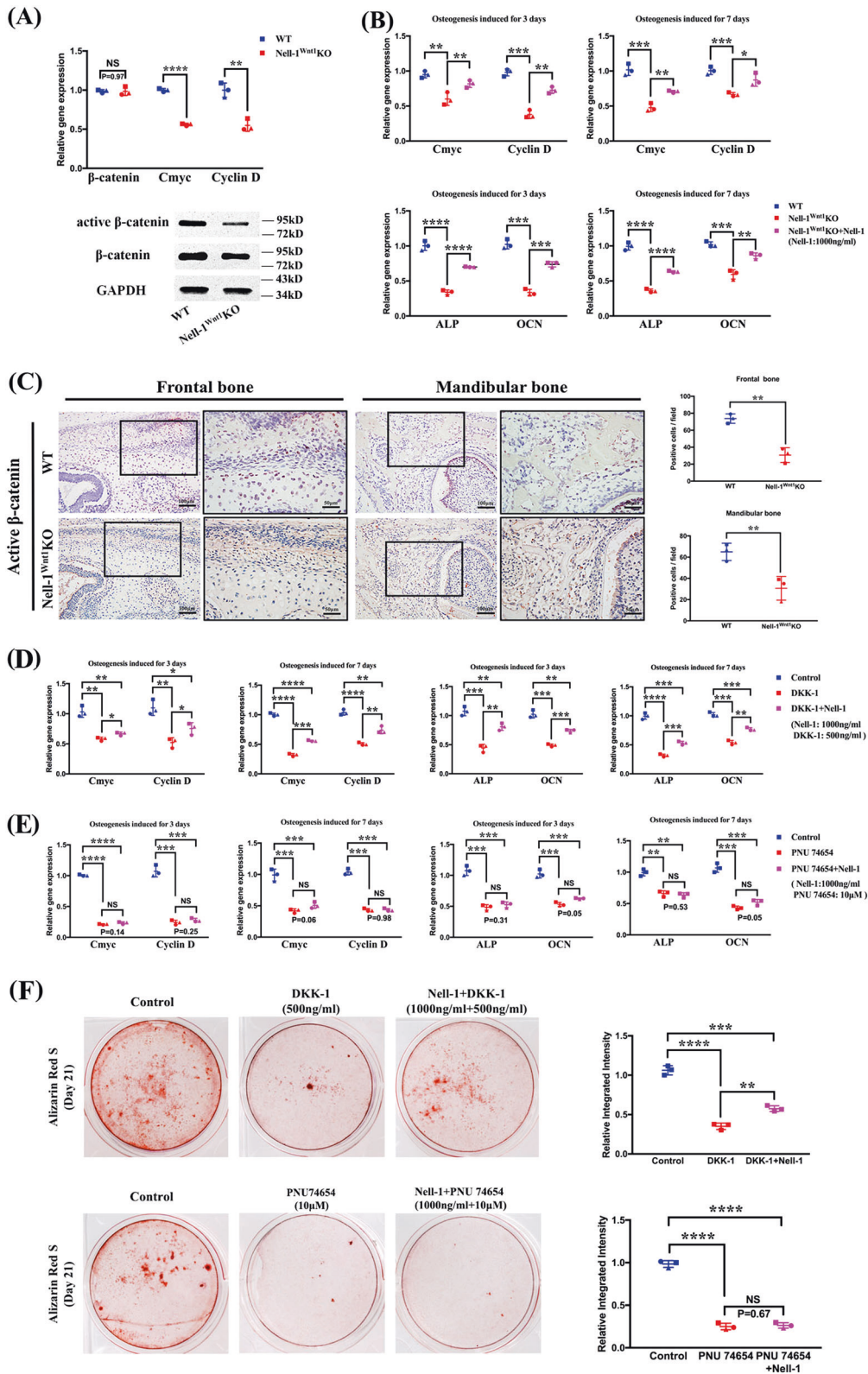
With defined conditional knockout of Nell-1 in Wnt1 expressing cells, the CNCC-derived craniofacial skeletons of the frontonasal and MB exhibited lower BMD and reduced bone volume in embryonic and neonatal Nell-1<sup>Wnt1</sup>KO mice at complete penetrance. The similar craniofacial skeletal hypoplasia phenotype was also identified in the majority of Nell-1<sup>CMV</sup>KO mice except for some severe cases. This further implicated the prominent role of Nell-1 in craniofacial skeletal tissues. Surprisingly, a relatively high prevalence of hydrocephalus was identified in Nell-1<sup>Wnt1</sup>KO mice at postnatal 30–40 days. This is the first report demonstrating the association of Nell-1 with hydrocephalus, which has shown to be frequently associated with abnormalities of the choroid plexus and ependymal cells as well as corticogenesis in murine models [22, 41–43]. This new finding may indicate possible functional impact by Nell-1 on choroid plexus epithelial cells and ependymal cells of the central nervous system in addition to its prominent pro-osteochondral properties, thereby warranting further investigation. The global deletion of Nell-1 in END mice resulted in neonatal lethality with severe cartilage and bone deformities including CCD-like phenotypes in craniofacial tissues [5, 9]. In contrast, the majority of Nell-1<sup>CMV</sup>KO mice survived at birth with various degrees of craniofacial skeletal hypoplasia. The phenotypic discrepancies between END and Nell-1<sup>CMV</sup>KO mice, as well as among Nell-1<sup>CMV</sup>KO mice themselves, may be attributable to different degrees of inactivation/diminishment of the Nell-1 molecule because Cre activation and Cre-loxP recombination are highly influenced by promoter activity of the CMV driver mouse [5, 11, 44, 45]. Consequently, in comparison to the CMV-Cre mouse line, the Wnt1-Cre line has been shown to be a more reliable tool to study CNCC specific knockouts [16, 22, 32].

The hypoplastic skeletal phenotype in CNCC derivatives- frontonasal and MB, was readily noticeable at late embryonic stages and became more significant in neonatal

Nell-1<sup>Wnt1</sup>KO mice as measured by micro-CT and histological analyses. Similar to previous findings with primary calvarial cells from END mice [9, 46], the osteogenic differentiation and proliferation of CNCCs were impaired by the cell specific knockout of Nell-1. Significantly, the application of recombinant Nell-1 can compensate for the loss of osteogenic differentiation of Nell-1<sup>Wnt1</sup>KO CNCCs. Thus, Nell-1 is an essential player for the full osteogenic potential of CNCCs and for normal bone formation during craniofacial skeletal development. Mechanistically, Nell-1, as a direct downstream target of Runx2 [47], has been identified to involve multiple major signaling pathways including MAPK (ERK/JNK) [46, 48, 49], Wnt/ $\beta$ -catenin [6, 29], Integrin  $\beta$ 1/FAK [50, 51], and IHH [52, 53] due to its osteogenic role in various experimental settings [54, 55]. Since the inactivation of  $\beta$ -catenin in Wnt1 expressing cell lineages resulted in severe craniofacial skeletal defects [16], the influence of Nell-1 inactivation on Wnt/ $\beta$ -catenin signaling during osteogenic differentiation of CNCCs was chosen as a primary focus of the mechanistic study. Nell-1<sup>Wnt1</sup>KO CNCCs exhibited significantly reduced gene expression of the downstream targets of Wnt/ $\beta$ -catenin. The application of recombinant Nell-1 can compensate the reduction of these genes' expression not only in Nell-1<sup>Wnt1</sup>KO CNCCs, but also in WT CNCCs treated with Wnt inhibitors that act at two different points of the Wnt signaling pathway [39, 40]. Clearly, our data indicate that the major point/site of Nell-1 in activating Wnt/ $\beta$ -catenin signaling would be downstream of Wnt receptor/co-receptors in CNCCs. This is in agreement with the latest finding of Cntnap4, a cell surface receptor of Nell-1, to be required for Nell-1's osteogenic effects in primary calvarial cells and MC3T3-E1 cells. Specifically, Wnt/ $\beta$ -catenin signaling activation and osteoblastic differentiation by Nell-1 stimulation were attenuated in MC3T3-E1 cells with Cntnap4 knockdown [8]. Collectively, these data indicate that Nell-1 is not only a pivotal component of the canonical Wnt/ $\beta$ -catenin signaling pathway, but also a novel agonist of Wnt signaling [6, 29] through interacting with its specific receptor and/or other potential binding partners including integrin  $\beta$ 1 in modulating the osteogenic potential of post-migratory CNCCs during craniofacial skeletal development. Apparently, a more complex mechanism is expected as a result from possible crosstalk among the aforementioned signaling pathways [55, 56]. Alternatively, Nell-1 may also employ mechanisms involving  $\beta$ -catenin irrelevant to Wnt signaling as suggested in the  $\beta$ -catenin<sup>Wnt1</sup> KO mice model [16, 57].

Interestingly, Nell-1 inactivation in Wnt1 expressing cell lineages was associated with postnatal hydrocephalus for the first time. As defined by the International Hydrocephalus Working Group, hydrocephalus is "an active distension of the ventricular system resulting from inadequate





passage of CSF from its point of production within the cerebral ventricles to its point of absorption into the systemic circulation” [58]. Thus, the inadequate production and absorption of CSF or the abnormal passage of CSF

circulation may result in hydrocephalus. In agreement with CNCCs being a multipotent mesenchymal stem cell population for neural and craniofacial tissues [59], the choroid plexus where CSF was produced and the ependymal cells

◀ **Fig. 6** Nell-1 inactivation significantly alters Wnt/ $\beta$ -catenin signaling in CNCCs and cranial bone derivatives of newborn mice. **a** Gene expression of Wnt/ $\beta$ -catenin signaling molecules showed a significant decrease in the  $\beta$ -catenin downstream targets, C-myc and Cyclin D, but  $\beta$ -catenin itself remained unchanged in CNCCs from Nell-1<sup>Wnt1</sup>KO mice as evidenced by qPCR analysis (upper row). However, active  $\beta$ -catenin protein was significantly reduced in CNCCs from Nell-1<sup>Wnt1</sup>KO mice as evidenced by Western blot analysis (lower row). **b** A similar trend of reduced gene expression of osteogenic markers along with downstream targets of  $\beta$ -catenin was observed during osteogenic induction of CNCCs from both WT and Nell-1<sup>Wnt1</sup>KO mice. Notably, administration of exogenous Nell-1 can partially compensate for the defective osteogenic capacity of CNCCs from Nell-1<sup>Wnt1</sup>KO mice. **c** The reduced amounts of active  $\beta$ -catenin protein was also confirmed in the frontal and mandibular bones of Nell-1<sup>Wnt1</sup>KO mice, indicating the essential role of Nell-1 in Wnt/ $\beta$ -catenin signaling for osteogenesis. **d** DKK-1, an inhibitor of the Wnt/ $\beta$ -catenin signaling pathway that interferes with Wnt ligand binding to its receptors, also resulted in reduced gene expression of osteogenic markers (ALP and OCN), and  $\beta$ -catenin downstream targets (C-myc and Cyclin D). However, exogenous Nell-1 can partially rescue the loss of osteogenic differentiation by increasing Wnt/ $\beta$ -catenin signaling. **e** Gene expressions of osteogenic markers (ALP and OCN), and the  $\beta$ -catenin downstream targets (C-myc and Cyclin D) were severely blocked with PNU74654, an inhibitor of the Wnt/ $\beta$ -catenin signaling pathway that prevents Tcf binding to  $\beta$ -catenin. Administration of exogenous Nell-1 had no compensatory effects to this inhibition in Wnt/ $\beta$ -catenin signaling and osteogenic differentiation in CNCCs. **f** Moreover, recombinant Nell-1 protein is capable of rescuing the terminal osteogenic differentiation of CNCCs treated with DKK-1 to a certain extent. However, recombinant Nell-1 protein cannot rescue the terminal osteogenic differentiation of CNCCs treated with PNU74654. NS no significance; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.005$ ; \*\*\*\* $P < 0.001$

which are the ciliated lining cells of the brain ventricles in charge of directing CSF flow were reported to be derived from Wnt1-expressing cell populations of the dorsal midline [42, 60, 61] and experimentally verified using Nell-1<sup>Wnt1-tdTomato</sup> KO mouse in this study. This led us to examine whether these structures were affected in Nell-1<sup>Wnt1</sup>KO mice that may account for the development of hydrocephalus. Likely, the structural abnormalities and misplaced expression of vimentin and pre-albumin/transferrin in the degenerative choroid plexus epithelial cells and ependymal cells are part of the pathogenesis of hydrocephalus in postnatal Nell-1<sup>Wnt1</sup>KO mice. The high prevalence of premature ISS fusion in hydrocephalic Nell-1<sup>Wnt1</sup>KO mice may implicate the considerable impacts of skeletal deformities or abnormal mechanical forces on the developing brain to hydrocephalus development as suggested in other relevant studies [62, 63]. The seemingly paradoxical results of premature ossification of the ISS with overall cranial skeletal hypoplasia in hydrocephalic Nell-1<sup>Wnt1</sup>KO mice need to be further investigated in future studies. Furthermore, our data on the significant alterations of Pit expression in hydrocephalic Nell-1<sup>Wnt1</sup>KO mice may implicate that the dysregulation of phosphate import from the CSF is a part of critical mechanisms in the pathogenesis of hydrocephalus [37, 64]. Coincidentally, there were

several conditional gene inactivation models using Wnt1-Cre resulting in hydrocephalus including Huntingtin protein [22], Pax3 transcription factor [65], and RFX3 [41]. Likely, this is due to the cumulative insults by inactivation of these functional molecules to the subcommissural organ, ependymal cells, and epithelial cells of the CP that originate from early Wnt1 expressing neuroepithelial cells [20, 24]. Nevertheless, L1 cell adhesion molecule (L1cam) was recognized as a highly affected gene for X-linked congenital hydrocephalus in humans [66], and its underlying mechanism was proposed as the loss of L1cam function affecting radial migration in corticogenesis in a murine model [43]. Therefore, the exact mechanism of the development of hydrocephalus may be more complex than what has been reported. It is significant that Nell-1 becomes a newly identified factor in contributing to the pathogenesis of postnatal hydrocephalus. The dual role of Nell-1 in osteochondral and neural tissue development and growth as evidenced in this study may provide a new research avenue to the growing field of neuroskeletal biology [3, 54, 67–70].

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## Compliance with ethical standards

**Conflict of interest** XZ, KT, and CS are inventors of Nell-1 related patents. XZ, KT, and CS are founders and/or previous board members of Bone Biologics Inc./Bone Biologics Corp., which sublicenses Nell-1 patents from the UC Regents, which also hold equity in the company. XZ, KT, and CS = Xinli Zhang, Kang Ting, Chia Soo

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## References

1. Matsubashi S, Noji S, Koyama E, Myokai F, Ohuchi H, Taniguchi S, et al. New gene, nel, encoding a M(r) 93 K protein with EGF-like repeats is strongly expressed in neural tissues of early stage chick embryos. *Dev Dyn*. 1995;203:212–22.
2. Watanabe TK, Katagiri T, Suzuki M, Shimizu F, Fujiwara T, Kanemoto N, et al. Cloning and characterization of two novel human cDNAs (NELL1 and NELL2) encoding proteins with six EGF-like repeats. *Genomics*. 1996;38:273–6.

3. Ting K, Vastardis H, Mulliken JB, Soo C, Tieu A, Do H, et al. Human NELL-1 expressed in unilateral coronal synostosis. *J Bone Miner Res: Off J Am Soc Bone Miner Res.* 1999;14:80–9.
4. Zhang X, Kuroda S, Carpenter D, Nishimura I, Soo C, Moats R, et al. Craniosynostosis in transgenic mice overexpressing Nell-1. *J Clin Investig.* 2002;110:861–70.
5. Desai J, Shannon ME, Johnson MD, Ruff DW, Hughes LA, Kerley MK, et al. Nell1-deficient mice have reduced expression of extracellular matrix proteins causing cranial and vertebral defects. *Hum Mol Genet.* 2006;15:1329–41.
6. James AW, Shen J, Zhang X, Asatrian G, Goyal R, Kwak JH, et al. NELL-1 in the treatment of osteoporotic bone loss. *Nat Commun.* 2015;6:7362. Epub 2015/06/18
7. Qi H, Kim JK, Ha P, Chen X, Chen E, Chen Y, et al. Inactivation of Nell-1 in chondrocytes significantly impedes appendicular skeletogenesis. *J Bone Miner Res: Off J Am Soc Bone Miner Res.* 2018;34:533–46.
8. Li C, Zheng Z, Ha P, Chen X, Jiang W, Sun S, et al. Neurexin superfamily cell membrane receptor contactin-associated protein like-4 (Cntnap4) is involved in neural EGFL-Like 1 (Nell-1)-responsive osteogenesis. *J Bone Miner Res: Off J Am Soc Bone Miner Res.* 2018;33:1813–25.
9. Zhang X, Ting K, Pathmanathan D, Ko T, Chen W, Chen F, et al. Calvarial cleidocraniodysplasia-like defects with ENU-induced Nell-1 deficiency. *J Craniofac Surg.* 2012;23:61–6.
10. Soriano P. Generalized lacZ expression with the ROSA26 Cre reporter strain. *Nat Genet.* 1999;21:70–1.
11. Gu H, Marth JD, Orban PC, Mossmann H, Rajewsky K. Deletion of a DNA polymerase beta gene segment in T cells using cell type-specific gene targeting. *Science.* 1994;265:103–6.
12. Bronner-Fraser M. Origins and developmental potential of the neural crest. *Exp Cell Res.* 1995;218:405–17.
13. Szabo-Rogers HL, Smithers LE, Yakob W, Liu KJ. New directions in craniofacial morphogenesis. *Dev Biol.* 2010;341:84–94.
14. Chai Y, Jiang X, Ito Y, Bringas P Jr., Han J, Rowitch DH, et al. Fate of the mammalian cranial neural crest during tooth and mandibular morphogenesis. *Development.* 2000;127:1671–9.
15. Douarin NL, Kalcheim C. *The Neural Crest.* Cambridge University Press; Cambridge, United Kingdom; 1999.
16. Brault V, Moore R, Kutsch S, Ishibashi M, Rowitch DH, McMahon AP, et al. Inactivation of the beta-catenin gene by Wnt1-Cre-mediated deletion results in dramatic brain malformation and failure of craniofacial development. *Development.* 2001;128:1253–64.
17. McMahon AP, Bradley A. The Wnt-1 (int-1) proto-oncogene is required for development of a large region of the mouse brain. *Cell.* 1990;62:1073–85.
18. Echelard Y, Vassileva G, McMahon AP. Cis-acting regulatory sequences governing Wnt-1 expression in the developing mouse CNS. *Development.* 1994;120:2213–24.
19. Roelink H, Nusse R. Expression of two members of the Wnt family during mouse development-restricted temporal and spatial patterns in the developing neural tube. *Genes Dev.* 1991;5:381–8.
20. Hsu W, Miranda AJ, Yu HM. Manipulating gene activity in Wnt1-expressing precursors of neural epithelial and neural crest cells. *Dev Dyn.* 2010;239:338–45.
21. Jiang X, Iseki S, Maxson RE, Sucov HM, Morriss-Kay GM. Tissue origins and interactions in the mammalian skull vault. *Dev Biol.* 2002;241:106–16.
22. Dietrich P, Shanmugasundaram R, Shuyu E, Dragatsis I. Congenital hydrocephalus associated with abnormal subcommissural organ in mice lacking huntingtin in Wnt1 cell lineages. *Hum Mol Genet.* 2009;18:142–50.
23. Yamauchi Y, Abe K, Mantani A, Hitoshi Y, Suzuki M, Osuzu F, et al. A novel transgenic technique that allows specific marking of the neural crest cell lineage in mice. *Dev Biol.* 1999;212:191–203.
24. Murdoch B, DelConte C, Garcia-Castro MI. Pax7 lineage contributions to the mammalian neural crest. *PLoS ONE.* 2012;7:e41089.
25. Wong GL, Cohn DV. Target cells in bone for parathormone and calcitonin are different: enrichment for each cell type by sequential digestion of mouse calvaria and selective adhesion to polymeric surfaces. *Proc Natl Acad Sci USA.* 1975;72:3167–71.
26. Leal LF, Bueno AC, Gomes DC, Abduch R, de Castro M, Antonini SR. Inhibition of the Tcf/beta-catenin complex increases apoptosis and impairs adrenocortical tumor cell proliferation and adrenal steroidogenesis. *Oncotarget.* 2015;6:43016–32.
27. Jacobberger JW, Sramkoski RM, Stefan T, Woost PG. Multi-parameter cell cycle analysis. *Methods Mol Biol.* 2018;1678:203–47.
28. Wang C, Tanjaya J, Shen J, Lee S, Bisht B, Pan HC, et al. Peroxisome proliferator-activated receptor-gamma knockdown impairs bone morphogenetic protein-2-induced critical-size bone defect repair. *Am J Pathol.* 2019;189:648–64.
29. Shen J, James AW, Zhang X, Pang S, Zara JN, Asatrian G, et al. Novel Wnt regulator NEL-like molecule-1 antagonizes adipogenesis and augments osteogenesis induced by bone morphogenetic protein 2. *Am J Pathol.* 2016;186:419–34.
30. Wu Y, Wang C, Sun H, LeRoith D, Yakar S. High-efficient FLPo deleter mice in C57BL/6J background. *PLoS ONE.* 2009;4:e8054.
31. Schwenk F, Baron U, Rajewsky K. A cre-transgenic mouse strain for the ubiquitous deletion of loxP-flanked gene segments including deletion in germ cells. *Nucleic Acids Res.* 1995;23:5080–1.
32. Lewis AE, Vasudevan HN, O'Neill AK, Soriano P, Bush JO. The widely used Wnt1-Cre transgene causes developmental phenotypes by ectopic activation of Wnt signaling. *Dev Biol.* 2013;379:229–34.
33. Stoller JZ, Degenhardt KR, Huang L, Zhou DD, Lu MM, Epstein JA. Cre reporter mouse expressing a nuclear localized fusion of GFP and beta-galactosidase reveals new derivatives of Pax3-expressing precursors. *Genesis.* 2008;46:200–4.
34. Yamazaki Y, Hirai Y, Miyake K, Shimada T. Targeted gene transfer into ependymal cells through intraventricular injection of AAV1 vector and long-term enzyme replacement via the CSF. *Sci Rep.* 2014;4:5506.
35. Baehr C, Reichel V, Fricker G. Choroid plexus epithelial monolayers—a cell culture model from porcine brain. *Cerebrospinal Fluid Res.* 2006;3:13.
36. Cowan CM, Zhang X, James AW, Kim TM, Sun N, Wu B, et al. NELL-1 increases pre-osteoblast mineralization using both phosphate transporter Pit1 and Pit2. *Biochem Biophys Res Commun.* 2012;422:351–7.
37. Wallingford MC, Chia JJ, Leaf EM, Borgeia S, Chavkin NW, Sawangmake C, et al. SLC20A2 deficiency in mice leads to elevated phosphate levels in cerebrospinal fluid and glymphatic pathway-associated arteriolar calcification, and recapitulates human idiopathic basal ganglia calcification. *Brain Pathol.* 2017;27:64–76.
38. Ikeya M, Lee SM, Johnson JE, McMahon AP, Takada S. Wnt signalling required for expansion of neural crest and CNS progenitors. *Nature.* 1997;389:966–70.
39. Li J, Sarosi I, Cattle RC, Pretorius J, Asuncion F, Grisanti M, et al. Dkk1-mediated inhibition of Wnt signaling in bone results in osteopenia. *Bone.* 2006;39:754–66.
40. Yan M, Li G, An J. Discovery of small molecule inhibitors of the Wnt/beta-catenin signaling pathway by targeting beta-catenin/Tcf4 interactions. *Exp Biol Med (Maywood).* 2017;242:1185–97.
41. Baas D, Meiniel A, Benadiba C, Bonnafé E, Meiniel O, Reith W, et al. A deficiency in RFX3 causes hydrocephalus associated with abnormal differentiation of ependymal cells. *Eur J Neurosci.* 2006;24:1020–30.



42. Louvi A, Wassef M. Ectopic engrailed 1 expression in the dorsal midline causes cell death, abnormal differentiation of circumventricular organs and errors in axonal pathfinding. *Development*. 2000;127:4061–71.
43. Itoh K, Fushiki S. The role of L1cam in murine corticogenesis, and the pathogenesis of hydrocephalus. *Pathol Int*. 2015;65:58–66.
44. Loser P, Jennings GS, Strauss M, Sandig V. Reactivation of the previously silenced cytomegalovirus major immediate-early promoter in the mouse liver: involvement of NFkappaB. *J Virol*. 1998;72:180–90.
45. Schmidt-Supprian M, Rajewsky K. Vagaries of conditional gene targeting. *Nat Immunol*. 2007;8:665–8.
46. Zhang X, Ting K, Bessette CM, Culiati CT, Sung SJ, Lee H, et al. Nell-1, a key functional mediator of Runx2, partially rescues calvarial defects in Runx2(+/-) mice. *J Bone Miner Res: Off J Am Soc Bone Miner Res*. 2011;26:777–91.
47. Truong T, Zhang X, Pathmanathan D, Soo C, Ting K. Craniosynostosis-associated gene nell-1 is regulated byrunx2. *J Bone Miner Res: Off J Am Soc Bone Miner Res*. 2007;22:7–18.
48. Chen F, Walder B, James AW, Soofer DE, Soo C, Ting K, et al. NELL-1-dependent mineralisation of Saos-2 human osteosarcoma cells is mediated via c-Jun N-terminal kinase pathway activation. *Int Orthop*. 2012;36:2181–7.
49. Bokui N, Otani T, Igarashi K, Kaku J, Oda M, Nagaoka T, et al. Involvement of MAPK signaling molecules and Runx2 in the NELL1-induced osteoblastic differentiation. *FEBS Lett*. 2008;582:365–71.
50. Hasebe A, Nakamura Y, Tashima H, Takahashi K, Iijima M, Yoshimoto N, et al. The C-terminal region of NELL1 mediates osteoblastic cell adhesion through integrin alpha3beta1. *FEBS Lett*. 2012;586:2500–6.
51. Shen J, James AW, Chung J, Lee K, Zhang JB, Ho S, et al. NELL-1 promotes cell adhesion and differentiation via Integrinbeta1. *J Cell Biochem*. 2012;113:3620–8.
52. James AW, Pan A, Chiang M, Zara JN, Zhang X, Ting K, et al. A new function of Nell-1 protein in repressing adipogenic differentiation. *Biochem Biophys Res Commun*. 2011;411:126–31.
53. Li CS, Zhang X, Peault B, Jiang J, Ting K, Soo C, et al. Accelerated chondrogenic differentiation of human perivascular stem cells with NELL-1. *Tissue Eng Part A*. 2016;22:272–85.
54. Zhang X, Zara J, Siu RK, Ting K, Soo C. The role of NELL-1, a growth factor associated with craniosynostosis, in promoting bone regeneration. *J Dent Res*. 2010;89:865–78.
55. Pakvasa M, Alverdy A, Mostafa S, Wang E, Fu L, Li A, et al. Neural EGF-like protein 1 (NELL-1): Signaling crosstalk in mesenchymal stem cells and applications in regenerative medicine. *Genes Dis*. 2017;4:127–37.
56. Wang Y, Li YP, Paulson C, Shao JZ, Zhang X, Wu M, et al. Wnt and the Wnt signaling pathway in bone development and disease. *Front Biosci (Landmark Ed)*. 2014;19:379–407.
57. Aberle H, Schwartz H, Kemler R. Cadherin-catenin complex: protein interactions and their implications for cadherin function. *J Cell Biochem*. 1996;61:514–23.
58. Tully HM, Dobyns WB. Infantile hydrocephalus: a review of epidemiology, classification and causes. *Eur J Med Genet*. 2014;57:359–68.
59. Takashima Y, Era T, Nakao K, Kondo S, Kasuga M, Smith AG, et al. Neuroepithelial cells supply an initial transient wave of MSC differentiation. *Cell*. 2007;129:1377–88.
60. Awatramani R, Soriano P, Rodriguez C, Mai JJ, Dymecki SM. Cryptic boundaries in roof plate and choroid plexus identified by intersectional gene activation. *Nat Genet*. 2003;35:70–5.
61. Fernandez-Llebrez P, Grondona JM, Perez J, Lopez-Aranda MF, Estivill-Torrus G, Llebrez-Zayas PF, et al. Msx1-deficient mice fail to form prosomere 1 derivatives, subcommissural organ, and posterior commissure and develop hydrocephalus. *J Neuropathol Exp Neurol*. 2004;63:574–86.
62. Cinalli G, Sainte-Rose C, Kollar EM, Zerah M, Brunelle F, Chumas P, et al. Hydrocephalus and craniosynostosis. *J Neurosurg*. 1998;88:209–14.
63. Kanavaki A, Jenny B, Hanquinet S, Chiari I malformation associated with premature unilateral closure of the posterior intraoccipital synchondrosis in a preterm infant. *J Neurosurg Pediatr*. 2013;11:658–60.
64. Jensen N, Autzen JK, Pedersen L. Slc20a2 is critical for maintaining a physiologic inorganic phosphate level in cerebrospinal fluid. *Neurogenetics*. 2016;17:125–30.
65. Zhou HM, Conway SJ. Restricted Pax3 deletion within the neural tube results in congenital hydrocephalus. *J Dev Biol*. 2016;4:1–17.
66. Adle-Biassette H, Saugier-Verber P, Fallet-Bianco C, Delezoide AL, Razavi F, Drouot N, et al. Neuropathological review of 138 cases genetically tested for X-linked hydrocephalus: evidence for closely related clinical entities of unknown molecular bases. *Acta Neuropathol*. 2013;126:427–42.
67. Zhang X, Cowan CM, Jiang X, Soo C, Miao S, Carpenter D, et al. Nell-1 induces acrania-like cranioskeletal deformities during mouse embryonic development. *Lab Invest*. 2006;86(Jul):633–44.
68. Patel MS, Elefteriou F. The new field of neuroskeletal biology. *Calcif tissue Int*. 2007;80:337–47.
69. Spencer GJ, Hitchcock IS, Genever PG. Emerging neuroskeletal signalling pathways: a review. *FEBS Lett*. 2004;559:6–12.
70. Nakamura R, Nakamoto C, Obama H, Durward E, Nakamoto M. Structure-function analysis of Nel, a thrombospondin-1-like glycoprotein involved in neural development and functions. *J Biol Chem*. 2012;287:3282–91.

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