# **REVIEW ARTICLE** OPEN P38 kinase in gastrointestinal cancers

Thuy Phan<sup>1</sup>, Xu Hannah Zhang<sup>2</sup>, Steven Rosen  $10^{\circ}$  and Laleh G. Melstrom  $10^{1}$ 

© The Author(s) 2023

Gastrointestinal cancers are a leading cause of cancer morbidity and mortality worldwide with 4.2 million new cases and 3.2 million deaths estimated in 2020. Despite the advances in primary and adjuvant therapies, patients still develop distant metastases and require novel therapies. Mitogen-activated protein kinase (MAPK) cascades are crucial signaling pathways that regulate many cellular processes, including proliferation, differentiation, apoptosis, stress responses and cancer development. p38 Mitogen Activated Protein Kinases (p38 MAPKs) includes four isoforms: p38α (MAPK14), p38β (MAPK11), p38γ (MAPK12), and p38δ (MAPK13). p38 MAPK was first identified as a stress response protein kinase that phosphorylates different transcriptional factors. Dysregulation of p38 pathways, in particular p38γ, are associated with cancer development, metastasis, autophagy and tumor microenvironment. In this article, we provide an overview of p38 and p38γ with respect to gastrointestinal cancers. Furthermore, targeting p38γ is also discussed as a potential therapy for gastrointestinal cancers.

Cancer Gene Therapy (2023) 30:1181-1189; https://doi.org/10.1038/s41417-023-00622-1

# INTRODUCTION

Gastrointestinal (GI) cancers of the colon, stomach, liver, esophagus, and pancreas impacted an estimated 4.2 million patients (22%) and resulted in 3.2 million deaths (32%) in 2020 and together are a leading cause of cancer morbidity and mortality worldwide [1]. Current treatments for GI cancers involve multiples therapies including surgery, radiation, chemotherapy or a combination thereof. Based on the particular malignancy, radiation therapy is one of the treatments for GI cancer that can be applied before or after surgery. Chemotherapy is also combined with radiation to enhance tumor sensitization. Immunotherapy for GI cancers includes checkpoint inhibitors (PD-1, PD-L1 and CTLA-4), vaccine therapies (peptide, protein, whole tumor cells, or dendritic cell-based vaccines), cytokines (interferon-y, interleukin-2, IL-10, or GM-CSF) and adoptive T cell transfer [2]. Until now, surgical resection remains the most common treatment for patients with colorectal [3], gastric [4], and esophageal cancers [5]. Despite the advances in primary and adjuvant therapies, patients can still develop metastases and resistance to systemic therapy. Therefore, new therapeutic strategies and novel targets are urgently in need to improve the survival.

The mitogen-activated protein kinases (MAPK) pathways consist of three distinct kinases that play crucial roles in cell signaling. When exposed to extracellular and intracellular signals, MAPKKKs are activated and facilitate the direct phosphorylation of MAPKKs, leading to their activation[6]. Subsequently, MAPKKs phosphorylate and activate MAPKs [7]. MAPKs respond to a diverse range of stimuli, such as hormones, cytokines, growth factors, endogenous stress, and environmental signals [8]. Extracellular signal-regulated kinases (ERK), c-Jun NH2-terminal kinase (JNK), and p38 are the three wellknown MAPKs that play a role in carcinogenesis. Among these, the ERK pathway is predominantly triggered by growth factors like epidermal growth factor, as well as hormones and proinflammatory stimuli. On the other hand, the JNK and p38 pathways are activated by different stress-inducing stimuli, such as ultraviolet radiation, reactive oxygen species (ROS), as well as inflammatory cytokines like tumor necrosis factor (TNF)- $\alpha$  and interleukin (IL)-1 $\beta$  [9]. MAPKs are involved in regulating a range of cellular processes linked to the development of cancer, including proliferation, differentiation, apoptosis, inflammation, and immunity. Abnormal MAPK signaling may result in excessive or unregulated cell proliferation, as well as resistance to apoptosis[10].

#### **P38 PATHWAYS**

The p38 MAPK family was first identified in studies of endotoxininduced cytokine expression [11]. P38 is involved in inflammation, cell growth, cell differentiation, cell death, and the cell cycle [12]. There are four p38 isoforms including p38a (MAPK14), p38β (MAPK11), p38y (SAPK3, ERK6 or MAPK12), and p38b (MAPK13) with an overall sequence homology >60% and an identity within the kinase domains >90% [13]. The four p38 MAPK isoforms are ubiquitously expressed with different levels of expression in various tissues (Fig. 1). For example p38a is expressed in all cell types and tissues; p38ß is predominant in the brain; p38y is found in skeletal muscle and p38δ is mainly expressed in the testis, pancreas, kidney and small intestine [14]. Despite the high level of sequence homology of the four isoforms, they interact with different downstream effectors such as: MK2 (MAPK-activating protein kinase 2), PRAK (p38-related/activated protein kinase), ATF-2 (activating transcription factor-2), MEF2 (myocyte enhancement factor 2) and c-Jun and upstream MAPK kinase activators (including MKK4, MKK3 and MKK6) [15]. Phosphorylated p38 MAPK can activate a wide range of stimuli, such as transcription factors, protein kinases, cytoplasmic substrates and nuclear substrates [16]. The downstream events of these p-p38 MAPK have cell-specific consequences including regulation of RNA splicing, cytokine production, inflammatory response, apoptosis, cell-cycle arrest and cell differentiation [16]. Activated p38a has been shown to downregulate cyclins, upregulate

Check for updates

<sup>&</sup>lt;sup>1</sup>Department of Surgery, City of Hope Medical Center, Duarte, CA, USA. <sup>2</sup>Department of Hematology, City of Hope Medical Center, Duarte, CA, USA. <sup>Semail:</sup> Imelstrom@coh.org



**Fig. 1 p38 distribution and function.** The p38 MAPKs include 4 isoforms ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ ) ubiquitously expressed with different levels of expression in various organs. p38 MAPK supports cell survival via anti-apoptotic inflammatory signals interleukin-6 (IL-6) and enable DNA-repair after chemotherapy which results in drug-resistance in cancer cells. Upregulation of p38 MAPK promotes cell invasion by inducing epithelial to mesenchymal transdifferentiation (EMT), matrix metalloproteinases MMP-2 and MMP-9. Activated p38 signaling also increases cell migration via Vascular Endothelial Growth Factor (VEGF).

cyclin-dependent kinase (CDK) inhibitors, modulate the tumor suppressor p53 at the G1/S and the G2/M phases and induce apoptosis. This reduces cell proliferation in primary cells (cardiomyocytes, hepatocytes, fibroblasts, hematopoietic cells, and airway epithelium). In contrast, p38a has also been shown to support cell survival via anti-apoptotic inflammatory signals interleukin-6 (IL-6) and enable DNA-repair after chemotherapy which results in drugresistance in cancer cells [17]. Moreover, upregulation of p38 MAPK promotes cell invasion by inducing epithelial to mesenchymal transdifferentiation (EMT) [18]. Downregulation of p38 MAPK leads to suppressed expression and activity of matrix metalloproteinases MMP-2 and MMP-9. This evidence supports the role of p38 in facilitating cancer cell invasion [19]. Activated p38 signaling also increases cell migration via Vascular Endothelial Growth Factor (VEGF) expression promoting actin rearrangement [20]. The studies noted above demonstrate a duality in the role of p38 MAPK signaling as either an oncogene or tumor suppressor in many types of tumors.

In resting cells, p38 $\alpha/\beta$  are mainly found in the cytoplasm, with certain molecules undergoing phosphorylation in response to stimulation [21]. Whether phosphorylated or not, p38 $\alpha/\beta$  form a complex with either a dimer of Imp7/3 or Imp9/3, which transport them to the nuclear pores. While Imp3 remains outside, Imp7 or Imp9 accompany the p38 $\alpha/\beta$  into the nucleus. Once inside, p38 $\alpha/\beta$  disassociate from the importins and proceed to phosphorylate their substrates. Finally, they are exported back to the cytoplasm. Upon activation in myoblasts, the p38 signaling pathway phosphorylates BAF60c, facilitating the recruitment of the SWI-SNF chromatin-remodeling complex to muscle-specific regions. This, in turn, promotes the activation of gene expression [22].

Among the MAPK family, p38γ is the only isoform that possesses a short C-terminal sequence (-KETXL) capable of binding

to PDZ domains. When exposed to stress, p38γ can phosphorylate and regulate the activity of various PDZ-domain containing proteins involved in different signaling pathways. These include α1-syntrophin [23], SAP (synapse-associated protein) 90/PSD (post-synapse density) 95 [24], the scaffold protein SAP97/hDlg, and the protein tyrosine phosphatase PTPH1 [25] (Fig. 2). Interestingly, p38γ can also directly phosphorylate the transcription factor MyoD without interaction with the PDZ domain and subsequently suppress its activity [26]. In regard to tumorigenesis, p38γ binding to PTPH1 via the PDZ domain increases Ras transformation and promotes colon cancer development [25].

This review will aim to assess the role of p38 in cancers of the gastrointestinal tract.

#### P38 IN GASTROINTESTINAL CANCERS Esophageal cancer

In esophageal squamous cell carcinoma (ESCC), most publications regarding the p38 MAPK pathway in carcinogenesis have been focused on exploring the role of the p38 $\alpha$  isoform. There is limited published data on other p38 isoforms ( $\beta$ ,  $\gamma$ , and  $\delta$ ). Overall p38 MAPK expression has been found to be significantly higher in ESCC compared to normal esophageal tissue [27]. Zheng et al. found a significant association between p38 $\gamma$  expression and clinical stage, lymph nodes metastases, and tumor volume in ESCC [28]. This suggested p38 $\gamma$  may serve as a metastasis-associated gene in ESCC. Moreover, p38 $\gamma$  can promote cell motility and growth in ESCC cells in vitro. Knockdown of p38 $\gamma$  can prevent tumor progression in ESCC tumor bearing nude mice. These findings indicate that p38 $\gamma$  plays an oncogenic role in ESCC and may be targeted for therapy [28].



**Fig. 2** Schematic representation of p38γ MAPK signal transduction pathway. A wide variety of stimuli including cellular stresses, UV, proinflammatory cytokines, growth factors can activate p38γ MAPK. These lead to the initiation of a three-step MAPK phosphorylation cascade (MAPKKK, MAPKK, and MAPK). Firstly, MAPKKKs (ASK1, MLK1&2, and TAO1&2) phosphorylate the p38 MAPK-specific MAPKKs MKK6 and MKK3, respectively. These subsequently phosphorylate p38γ MAPK. The phosphorylated p38γ can activate the downstream substrates such as α-syntrophin, hDlg (human disc large), PTPH1 (protein tyrosine phosphatase H1), PSD95 (post-synapse density 95), and transcription factor MyoD, which trigger cellular responses. ASK1 (apoptosis signal-regulating kinase-1), MLKs (mixed-lineage kinases), TAO (thousand-and-one amino acid).

## GASTRIC CANCER

In the case of gastric cancer, Liu et al. demonstrated that cells pretreated with a combination of ERK1/2 and p38 inhibitors could enhance the anti-proliferative effects of 5-FU via suppressing the ERCC1 (Excision Repair Cross Complementation group 1) protein [29]. ERCC1 is upregulated in 5-FU-treated cells which may contribute to drug resistance. This study proposed that activated ERK1/2 and p38 kinases play a role in developing drug resistance via upregulating ERCC1 expression in gastric cancer [29].

Interferon gamma (IFN- $\gamma$ ) plays an important role in the innate and adaptive immune response as a cytokine with antitumor functions [30]. Zhao et al. reported that gastric cancer cells treated with IFN- $\gamma$  underwent G1/S phase cell cycle arrest and had downregulated p38 $\gamma$  expression [31]. This study suggested that IFN- $\gamma$ may inhibit gastric cancer cell proliferation via regulating p38 $\gamma$ .

#### COLORECTAL CANCER

p38 $\gamma$  is markedly upregulated in colon cancer tissues compared to surrounding colon epithelial cells [32]. p38 $\gamma$  knockdown was found to inhibit tumor progression in a colitis-associated mouse model [33]. In the same study p38 $\gamma$  activated the  $\beta$ -catenin/Wnt pathways promoting CRC development [33]. This protumor role has been corroborated by others where p38y knock down inhibited cell proliferation, migration, and induced apoptosis in colon cancer cells in vitro. Overexpression of p38y promoted CRC cell progression. Loesch et al. found that overexpressed p38y MAPK activated the transcription factor c-Jun, and then recruited p38y as a cofactor to the matrix metalloproteinase 9 (MMP-9) promoter thus enhancing cell invasion [34]. These studies indicated p38y as a potential therapeutic target for CRC.

#### PANCREATIC CANCER

In pancreatic adenocarcinoma, p38 expression was associated with a shorter survival [35]. However, Zhong et al. reported that high expression of p38 MAPK was associated with improved survival [36]. All isoforms of p38 MAPK were found in various human pancreatic cancer cells in vitro [37]. The roles of the various p38 isoforms in pancreatic cancer are controversial. Tian et al. proved that inhibition of p38 $\beta$  decreased tumor progression while inhibition of p38 $\alpha$  enhanced tumor formation in pancreatic cancer mouse models [37]. In addition, p38 $\gamma$  has been reported to promote PDAC development via KRAS signaling and aerobic glycolysis. Wang et al. indicated that

1184

KRAS mutation induced the expression and phosphorylation of p38γ, and subsequently enhanced PFKFB3 (6-Phosphofructo-2-Kinase/Fructose-2,6-Biphosphatase 3) and expression/phosphorylation of a glucose transporter GLUT2 (Glucose transporter 2). Moreover, PFKFB3 and GLUT2 depend on p38γ for aerobic glycolysis and tumor development [38]. This study proved that p38γ is crucial for the G1/S transition in cell cycle and proliferation. p38γ knockdown or p38γ inhibition in combination with a PFKFB3 inhibitor suppressed aerobic glycolysis and PDAC tumorigenesis [38]. The above data highlight the heterogenous role of p38 MAPKs isoforms and selective targeting will decide the promising effect for cancer treatment.

# LIVER CANCER

In hepatocellular carcinoma (HCC), high p38y expression was inversely related to survival [39]. In vitro, p38y knockdown suppressed proliferation and colony formation in HCC cell lines [39]. lyoda et al. showed that mutant MKK6 (a common activator of four p38 MAPK isoforms) increases p38 pathway activation, caspase-3 activity and subsequently induces apoptosis in human hepatocellular carcinoma cell lines [40]. Tomás-Lobaet al. reported that p38y shares a high sequence homology and substrate specificity with cyclin-dependent kinase (CDK)-cyclin protein which regulates cell division in cancer development [39]. In this study, p38y expression correlated with the expression of fibrosis markers ACTA2 and COL1A, that support the progression of liver cancer. p38y in human HCC biopsy samples were found to be overexpressed compared to healthy livers. The data involving HCC suggests the oncogenic role of p38y in human liver tumors and p38y as a promising target for liver cancer therapy.

Table 1 summarizes physiological function of p38 $\gamma$  and its impacts in human gastrointestinal cancers.

## **P38 IN IMMUNE REGULATION**

p38 $\alpha$  MAPK mediates the production of inflammatory cytokines in different immune cells [41]. p38 enhances transcriptional activity of NF- $\kappa$ B in primary human astrocytes via acetylation of p65 NF- $\kappa$ B [42], a key regulator of the inflammatory response, which contributes to chemoresistance through MDR1 expression in cancer cells [43]. The p38 $\alpha$  pathway can promote inflammation in several cell types. Activated myeloid p38 $\alpha$  enhances intestinal insulin-like growth factor-1 (IGF-1) production in intestinal inflammation and tumorigenesis [44]. This study showed a significant correlation between p38 $\alpha$  phosphorylation in monocytes/macrophages and IGF-1 phosphorylation in samples from ulcerative colitis patients and colon cancer patients. p38 $\alpha$  activation in dendritic cells promotes the expression of proinflammatory cytokines and chemokines and suppresses the expression of anti-inflammatory cytokine in the colon of DSS model, leading to colitis-associated tumorigenesis [45]. On the other hand, p38a also has anti-inflammatory functions in innate immune cells, which are mediated by the mitogen- and stressactivated kinases 1 and 2 (MSK1/2) resulting to the expression of anti-inflammatory genes such as IL-10, DUSP1, TTP, and IL-1ra [46].

## **P38 AND METASTASIS**

Gamma synuclein (SNCG), a neuronal protein, is overexpressed in different types of cancer. SNCG have been shown to promote TGF- $\beta$ -induced p38 MAPK phosphorylation by stabilizing MAPK kinase 3/6 (MKK3/6). The upregulation of p38 MAPK by SNCG leads to increased MMP-9 expression, which enhances cancer cell invasion. Overexpression of SNCG in liver cancer cells supports lung metastasis, which can be suppressed by the p38 MAPK inhibitor [47]. IL-1 $\beta$  induces the activation of p38 and the upregulates of MMP-2 and MMP-9 by activating AP-1-dependent transcription in gastric adenocarcinoma (GA) cells. Phospho-p38 is upregulated and correlates with the expression of IL-1 $\beta$ , MMP-2, MMP-9 and c-fos in human GA tissues and in a GA metastasis mouse model. IL-1 $\beta$  also activated JNK but it was not associated with migration and invasion in GA cells [48].

#### P38 REGULATES CELLULAR HOMEOSTASIS, AUTOPHAGY, UBIQUITINATION, AND PROTEOLYSIS Cellular homeostasis

The p38 $\alpha$  pathway plays a dual role during colorectal tumorigenesis [49]. In normal colon epithelial cells, p38 $\alpha$  maintains intestinal homeostasis and barrier function to suppress colitis-associated tumor initiation. On the other hand, p38 $\alpha$  contributes to colon tumor development by supporting proliferation and inhibiting apoptosis of transformed epithelial cells. TGF- $\beta$ 1 mediates the mRNA and protein levels of MMPs (MMP-2 and MMP-9) and their inhibitors (TIMP-2 and RECK), which plays an essential role of extracellular matrix homeostasis control in breast cancer progression. TGF- $\beta$ 1 phosphorylates p38 MAPK which can induce the expression of MMP-2 and TIMP-2, and increased migration and invasion in breast cancer cells [50].

## AUTOPHAGY

Autophagy is a conserved process that recycles damaged cellular proteins, organelles, and other cellular components to maintain energy homeostasis and to protect cells against stress [51]. p38

Type of cancer	Function	Impacts	References	
Esophageal cancer	Oncogene	p38γ promotes the cell motility and growth in vitro. Knockdown of p38γ prevents the tumor formation in mice. p38γ expression is markedly associated with clinical stage, lymph nodes metastases, and tumor volume in ECSS tissues.		
Gastric cancer	Oncogene	p38 and ERK1/2 inhibitors enhance the anti-proliferative effects of 5-FU via suppressing ERCC1 protein.	[29]	
		p38 $\gamma$ expression is down-regulated upon IFN- $\gamma$ treatment and induced G1/S phase cell cycle arrest.	[31]	
Colorectal cancer	Oncogene	Overexpressed $p38\gamma$ activates the c-Jun, recruits $p38\gamma$ to MMP-9, leads to the increasing MMP-9 expression and enhances cell invasion.	[34]	
		p38 $\gamma$ activates $\beta$ -catenin/Wnt pathways which promotes CRC development in a colitis-associated mouse model.	[33]	
		Overexpression of p38 $\gamma$ promoted CRC cell progression. p38 $\gamma$ is markedly upregulated in colon cancer tissues compared to surrounding colon epithelial cells.	[32]	
Pancreatic cancer	Oncogene	p38γ knockout or p38γ inhibitor combination with a PFKFB3 inhibitor suppressed aerobic [] glycolysis and PDAC tumorigenesis in KPC mice.		
Liver cancer	Oncogene	High p38y expression was associated with a lower survival rate in liver cancer while p38y [3 knockdown suppressed proliferation and colony formation in HCC cell lines.		

**Table 1.** Physiological function of  $p38\gamma$  and its impacts in human gastrointestinal cancers.

MAPK have been shown to positively and negatively regulate autophagy. Various stimulations (including oxidative stress, UV, inflammatory cytokines, growth factor, and chemotherapy) can activate p38 MAPK through the TAK1-MKK3/6-p38 and ASK1-MKK3/6-p38 cascades [52]. Next, activated p38 phosphorylates Atg5 leading to inhibition of autophagic membrane extension and the transformation of LCI to LCII, which inhibits the autophagy pathway [53]. p38 can regulate autophagy in response to chemotherapeutic agents. Irinotecan (IRI) induces autophagy and apoptosis through accumulation of reactive oxygen species (ROS) and activation of the JNK and p38 MAPK pathways that leads to tumor suppression in gastric cancer [54]. On the other hand, Isoliguiritigenin (ISL) promotes apoptosis and blocks autophagy through p38 activation that results in cell death and tumor suppression in pancreatic cancer [55]. The combination of ISL and Gemcitabine or 5-FU enhances the inhibition of cell viability compared to single agents.

# UBIQUITINATION

Ubiguitination, an important post-translational modification in cells, is an ATP-dependent cascade adding ubiguitin, a ubiguitously expressed protein consisting of 76 amino acids, to a substrate protein and inducing the degradation of target protein [56]. Ubiguitin can be attached via 7 lysine residues (K6, K11, K27, K29, K33, K48, and K63) or the first methionine (M1), which regulates various cellular processes including endocytosis of membrane proteins, protein degradation, and DNA repair [57-59]. Activation of protease-activated receptor 1 (PAR1), a G protein-coupled receptor (GPCR) for thrombin and inflammation, induces noncanonical p38 MAPK through autophosphorylation (independence of MKK3/MKK6) via a ubiquitin and TAB1-TAB2-dependent pathway on endosomes [60]. This study is the first one revealing the novel insight of GPCR ubiquitination in mediating the p38 pathway. On the other hand, p38 can phosphorylate Snail, which is a key regulator of epithelial-mesenchymal transition, a major step in tumor metastasis in ovarian cancer [61]. This process enhances Snail stability via suppressed DYRK2-mediated phosphorylation, which is important for GSK3β-dependent Snail phosphorylation and βTrCP-mediated Snail ubiguitination and degradation. Activated p38y and p38δ eliminate the cancer stem cell properties and tumor initiating ability of nonsmall cell lung cancer cells via the ubiquitination and degradation of stemness proteins SOX2, OCT4, Nanog, KLF4 and c-MYC through MK2-mediated phosphorylation of Hsp27, an important component of the proteasomal degradation machinery [62].

#### PROTEOLYSIS

Proteolysis is a fundamental hallmark of cancer as malignant tumors overexpress proteolytic enzymes for invasion, metastasis and angiogenesis including plasminogen activation system (PAS) and the matrix metalloproteinase family (MMPs) [63, 64]. Urokinase plasminogen activator (uPA) is overexpressed in gastric carcinoma cells by enhancing the promoter activity through p38 MAPK signaling [65]. In colon cancer, the transcription factor c-Jun is activated by p38y, and then recruits p38y into the matrix metalloproteinase 9 (MMP-9) promoter leading to MMP-9 transactivation and cell invasion [34]. In gastric cancer, activation of p38 MAPK through IL-1 increases cell invasion in vitro and promotes tumor metastasis in vivo via upregulation of MMP-2 and MMP-9 [48]. WEF, an aqueous extract of Eupatorium fortune in Chinese medicine, blocked PMA-induced p38 and JNK phosphorylation and decreased PMA-induced NF-KB activation. This results to suppress the metastatic properties such as anchorageindependent colony formation, migration and invasion, by downregulating the expression and proteolytic activity of MMP-9 in malignant metastatic cancer [66]. In addition, arsenite, an environmental carcinogen, triggers p38 MAPK activation, and subsequently induced cyclin B1 proteolysis through the ubiquitin-proteasome pathway, which contributes to G2 arrest [67]. Following arsenite exposure, DNA repair may activate p38, promote G2-arrest, cell apoptosis and genome instability.

# **P38 CONTRIBUTION TO THE TUMOR MICROENVIRONMENT**

p38 plays dual role in the tumor microenvironment. In breast cancer, tumor-derived GM-CSF induced myeloid cells ARG1 expression through p38 activation and inhibited antitumor function of T cells. ARG1 is a biomarker for protumor M2-polarized macrophages. This results in an immunosuppressive tumor microenvironment which causes resistance to adoptive T cell transfer [48]. On the other hand, AMP-activated protein kinase (AMPK) activates p38 MAPK and phosphorylates glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ). This leads to inhibition of PD-1 expression in Tregs and suppresses tumor progression [68]. In addition, CD4 + T cells with activated p38 signaling can promote pancreatic cancer progression [69].

# **TARGETING P38 GAMMA FOR CANCER TREATMENT**

Overall, there remains some controversy as to the role of  $p38\gamma$  in tumorigenesis and as a target for therapy in gastrointestinal cancers. In general,  $p38\gamma$  levels are overexpressed in many human malignancies including breast cancer [70], gliomas [71], and gastrointestinal cancers [28, 32, 38, 39]. This section will focus on targeting p38 as an oncogene in cancer therapy.

The four p38 MAPK isoforms have different sensitivity to kinase inhibitors. The p38y and p38\delta isoforms (~75% identity) are less similar in sequence compared to p38a (~60% identity) [72]. For instance, pharmacological studies showed that specific compounds (SB203580 and other pyridinyl imidazoles) can only inhibit p38a and p38β, but not p38γ and p38δ [73]. Three main p38a MAPK inhibitors have been in clinical trials: MW150 (by ADDF, in safety studies), Neflamapimod (by EIP Pharma, in efficacy studies [74]) for Alzheimer's disease and Losmapimod (by GlaxoSmithKline) for myocardial infarction [75]. PH-797804 is an oral p38 inhibitor tested for Rheumatoid Arthritis (Clinical trial NCT00383188). There are some clinical trials targeting p38 for cancer treatment. Ralimetinib (or LY2228820), a selective inhibitor of p38a and p38 $\beta$ , is being tested as monotherapy or in combination with other agents, for the treatment of ovarian cancer [76], glioblastoma, and metastatic breast cancer. This clinical trial (NCT01663857) evaluated the efficacy of ralimetinib in combination with gemcitabine and carboplatin for patients with recurrent platinum-sensitive epithelial ovarian cancer. The combination therapy resulted in the improvement of progression-free survival (PFS). p38 MAPK Inhibitor LY3007113 is being tested for advanced or metastatic cancer [77]. Talmapimod (SCIO-469), an orally active, selective, and ATP-competitive p38a inhibitor, is being tested as a monotherapy or in combination with Bortezomib for relapsed multiple myeloma (Clinical trial NCT00087867) (Table 2).

The p38 MAPK inhibitor (SB203580) significantly increased the sensitivity of colorectal cancer cell to 5-FU, a common therapy for colon cancer. The combination of SB203580 and 5-FU markedly reduced cell viability through a decrease of pro-apoptotic protein Bax expression [78]. However, long-term treatment of 5-FU results in chemoresistance. The development of multidrug resistance (MDR) related to the overexpression of ATP-binding cassette (ABC) transporters decreases drug accumulation in cancer cells and cause chemoresistance. The SW480/5-FU cells showed a significantly increased protein expression level of MDR-related proteins (P-gp, MRP1 and ABCG2). Noscapine treatment decreased the expression of these proteins in the SW480/5-FU cells, and combination with p38 MAPK inhibition enhances the sensitivity of 5-FU-resistant colon cancer cells to noscapine [79].

 Table 2.
 p38 inhibitors in clinical trials.

Inhibitor	Target	Disease	Studies		
MW150	p38α	Alzheimer	NCT05194163: safety study (phase 2)		
Neflamapimod	p38α	Alzheimer	NCT03402659: efficacy study (phase 2) [74]		
		Dementia With Lewy Bodies	NCT04001517: efficacy study (phase 2)		
Losmapimod	p38 $\alpha$ and p38 $\beta$	Myocardial infarction	NCT00910962: safety and efficacy study (phase 2) [75]		
PH-797804	p38α	Rheumatoid Arthritis	NCT00383188: safety and efficacy study (phase 2)		
Ralimetinib (LY2228820)	p38 $\alpha$ and p38 $\beta$	Ovarian cancer	NCT01663857: in combination with gemcitabine [76]		
		Advanced cancer	NCT01393990		
		Metastatic breast cancer	NCT02322853		
		Glioblastoma	NCT02364206: in combination with Temozolomide and radiotherapy		
LY3007113	p38α	Advanced cancer	NCT01463631: safety study (phase 1) [77]		
Talmapimod (SCIO-469)	p38 $\alpha$ and p38 $\beta$	Multiple myeloma	NCT00087867: monotherapy or in combination with Bortezomib (phase 2)		



#### Fig. 3 p38 in cancer hallmarks and immune response.

Until now, there is one p38y inhibitor named pirfenidone (PFD) that suppressed pro-inflammatory cytokines and tumor growth in a colitis-associated CRC model [33] and pancreatic cancer mouse model [38]. Pirfenidone has been clinically tested for patients with idiopathic pulmonary fibrosis [80]. A combined p38y and p38& inhibitor (BIRB796) reduced IFN-y [81]. A potent p38y inhibitor (F7 or PIK75) effectively suppressed tumor growth in a cutaneous T-cell lymphoma (CTCL) mouse model [82]. This small molecule p38y inhibitor has been screened through a high-throughput kinase inhibitor library. F7 or PIK75 inhibited p38y kinase activity, significantly reduced tumors burden in mice, and eliminated CD4+ malignant CTCL cells but not healthy CD4+ cells. Based on the Drugbank platform, several p38y-targeted drugs including phosphonothreonine, phosphoamino phosphonicacid-adenylate ester, CEP-1347 and KC706 are being investigated and tested [83, 84].

In the past 15 years immnunotherapy has evolved as a therapeutic strategy for several solid tumors. Different types of immunotherapies have been investigated in GI cancers, including adoptive T-cell transfer [85, 86], dendritic cell vaccines [87, 88], peptide vaccines [89, 90], and immune checkpoint inhibitors [91–94].

Recently, using CRISPR-Cas9 targeted 25 TCR-driven kinases, Gurusamy et al. found that knockout MAPK14 (p38α) can increase T cell expansion and memory. This also decreases reactive oxidative stress (ROS), and genomic stress (gH2AX) which accounts for an effective anti-tumor T cell [95]. Anti-CD19 CAR T cells expanded with the p38 inhibitor (BIRB796) markedly suppressed tumor growth and enhanced survival in tumor bearing mice. Therefore, it is interesting to combine p38 inhibitors and other immunotherapies like adoptive T cell transfer to enhance therapeutic effects in GI cancers.

#### CONCLUSION

p38 MAPKs have been explored as regulators of environmental stress and inflammation as well as mediators of homeostasis maintenance. Recent studies implicate crucial functions of p38a in the tumorigenesis and cancer development. In this review, we summarized the primary impact of p38 in gastrointestinal cancers, in which p38 is overexpressed, plays as an oncogene through regulation of various cellular processes (metastasis, autophagy, ubiquitylation and proteolysis) and facilitating tumor malignancy (Fig. 3). Future research work will clarify which isoform of p38 is more important and valuable for combination therapy in gastrointestinal cancers. An in-depth understanding of the p38 pathway and downstream mechanisms will translate into better therapeutic strategies.

#### REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71:209–49.
- Dahiya DS, Kichloo A, Singh J, Albosta M, Lekkala M. Current immunotherapy in gastrointestinal malignancies a review. J Investig Med. 2021;69:689–96.
- Kuipers EJ, Grady WM, Lieberman D, Seufferlein T, Sung JJ, Boelens PG, et al. Colorectal cancer. Nat Rev Dis Prim. 2015;1:15065.
- Sexton RE, Al Hallak MN, Diab M, Azmi AS. Gastric cancer: a comprehensive review of current and future treatment strategies. Cancer Metastasis Rev. 2020;39:1179–203.
- He S, Xu J, Liu X, Zhen Y. Advances and challenges in the treatment of esophageal cancer. Acta Pharm Sin B 2021;11:3379–92.
- Lee S, Rauch J, Kolch W. Targeting MAPK signaling in cancer: mechanisms of drug resistance and sensitivity. Int J Mol Sci. 2020;21:1102.
- Cargnello M, Roux PP. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. Microbiol Mol Biol Rev. 2011;75:50–83.
- Zhang W, Liu HT. MAPK signal pathways in the regulation of cell proliferation in mammalian cells. Cell Res. 2002;12:9–18.
- Johnson GL, Lapadat R. Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. Science 2002;298:1911–2.
- Dhillon AS, Hagan S, Rath O, Kolch W. MAP kinase signalling pathways in cancer. Oncogene 2007;26:3279–90.
- 11. Tang J, Qi X, Mercola D, Han J, Chen G. Essential role of p38gamma in K-Ras transformation independent of phosphorylation. J Biol Chem. 2005;280:23910-7.
- Ono K, Han J. The p38 signal transduction pathway: activation and function. Cell Signal. 2000;12:1–13.
- Coulthard LR, White DE, Jones DL, McDermott MF, Burchill SA. p38(MAPK): stress responses from molecular mechanisms to therapeutics. Trends Mol Med. 2009;15:369–79.
- Sanz-Ezquerro JJ, Cuenda A. p38 signalling pathway. Int J Mol Sci. 2021;22:1003.
   Feng Y, Wen J, Chang CC. p38 Mitogen-activated protein kinase and hematologic
- malignancies. Arch Pathol Lab Med. 2009;133:1850–6. 16. Cuadrado A, Nebreda AR. Mechanisms and functions of p38 MAPK signalling.
- Biochem J. 2010;429:403–17.
- 17. Wagner EF, Nebreda AR. Signal integration by JNK and p38 MAPK pathways in cancer development. Nat Rev Cancer. 2009;9:537–49.
- Bhowmick NA, Zent R, Ghiassi M, McDonnell M, Moses HL. Integrin beta 1 signaling is necessary for transforming growth factor-beta activation of p38MAPK and epithelial plasticity. J Biol Chem. 2001;276:46707–13.
- Mao L, Yuan L, Slakey LM, Jones FE, Burow ME, Hill SM. Inhibition of breast cancer cell invasion by melatonin is mediated through regulation of the p38 mitogenactivated protein kinase signaling pathway. Breast Cancer Res. 2010;12:R107.
- Rousseau S, Houle F, Landry J, Huot J. p38 MAP kinase activation by vascular endothelial growth factor mediates actin reorganization and cell migration in human endothelial cells. Oncogene 1997;15:2169–77.
- Maik-Rachline G, Lifshits L, Seger R. Nuclear P38: roles in physiological and pathological processes and regulation of nuclear translocation. Int J Mol Sci. 2020;21:6102.
- Forcales SV, Albini S, Giordani L, Malecova B, Cignolo L, Chernov A, et al. Signaldependent incorporation of MyoD-BAF60c into Brg1-based SWI/SNF chromatinremodelling complex. Embo J. 2012;31:301–16.
- Hasegawa M, Cuenda A, Spillantini MG, Thomas GM, Buée-Scherrer V, Cohen P, et al. Stress-activated protein kinase-3 interacts with the PDZ domain of alpha1syntrophin. A mechanism for specific substrate recognition. J Biol Chem. 1999;274:12626–31.
- Sabio G, Reuver S, Feijoo C, Hasegawa M, Thomas GM, Centeno F, et al. Stress- and mitogen-induced phosphorylation of the synapse-associated protein SAP90/PSD-95 by activation of SAPK3/p38gamma and ERK1/ERK2. Biochem J. 2004;380:19–30.
- Hou SW, Zhi HY, Pohl N, Loesch M, Qi XM, Li RS, et al. PTPH1 dephosphorylates and cooperates with p38gamma MAPK to increase ras oncogenesis through PDZmediated interaction. Cancer Res. 2010;70:2901–10.
- Gillespie MA, Le Grand F, Scimè A, Kuang S, von Maltzahn J, Seale V, et al. p38-{gamma}-dependent gene silencing restricts entry into the myogenic differentiation program. J Cell Biol. 2009;187:991–1005.
- Liu Q, Li W, Yang S, Liu Z. High expression of uPA related to p38MAPK in esophageal cancer indicates poor prognosis. Onco Targets Ther. 2018;11:8427–34.
- Zheng S, Yang C, Liu T, Liu Q, Dai F, Sheyhidin I, et al. Clinicopathological significance of p38β, p38γ, and p38δ and its biological roles in esophageal squamous cell carcinoma. Tumour Biol. 2016;37:7255–66.
- Liu JL, Huang WS, Lee KC, Tung SY, Chen CN, Chang SF. Effect of 5-fluorouracil on excision repair cross-complementing 1 expression and consequent cytotoxicity regulation in human gastric cancer cells. J Cell Biochem. 2018;119:8472–80.
- Ni C, Wu P, Zhu X, Ye J, Zhang Z, Chen Z, et al. IFN-γ selectively exerts proapoptotic effects on tumor-initiating label-retaining colon cancer cells. Cancer Lett. 2013;336:174–84.

- Zhao YH, Wang T, Yu GF, Zhuang DM, Zhang Z, Zhang HX, et al. Anti-proliferation effects of interferon-gamma on gastric cancer cells. Asian Pac J Cancer Prev. 2013;14:5513–8.
- Su C, Sun Q, Liu S, Wang H, Feng L, Cao Y. Targeting p38γ to inhibit human colorectal cancer cell progression. Biochem Biophys Res Commun. 2019;517:172–9.
- Yin N, Qi X, Tsai S, Lu Y, Basir Z, Oshima K, et al. p38γ MAPK is required for inflammation-associated colon tumorigenesis. Oncogene 2016;35:1039–48.
- Loesch M, Zhi HY, Hou SW, Qi XM, Li RS, Basir Z, et al. p38gamma MAPK cooperates with c-Jun in trans-activating matrix metalloproteinase 9. J Biol Chem. 2010;285:15149–58.
- Handra-Luca A, Lesty C, Hammel P, Sauvanet A, Rebours V, Martin A, et al. Biological and prognostic relevance of mitogen-activated protein kinases in pancreatic adenocarcinoma. Pancreas 2012;41:416–21.
- 36. Zhong Y, Naito Y, Cope L, Naranjo-Suarez S, Saunders T, Hong SM, et al. Functional p38 MAPK identified by biomarker profiling of pancreatic cancer restrains growth through JNK inhibition and correlates with improved survival. Clin Cancer Res. 2014;20:6200–11.
- Tian X, Traub B, Xie X, Zhou S, Henne-Bruns D, Knippschild U, et al. Opposing oncogenic functions of p38 mitogen-activated protein kinase Alpha and Beta in human pancreatic cancer cells. Anticancer Res. 2020;40:5545–56.
- Wang F, Qi XM, Wertz R, Mortensen M, Hagen C, Evans J, et al. p38γ MAPK is essential for aerobic glycolysis and pancreatic tumorigenesis. Cancer Res. 2020;80:3251–64.
- Tomás-Loba A, Manieri E, González-Terán B, Mora A, Leiva-Vega L, Santamans AM, et al. p38γ is essential for cell cycle progression and liver tumorigenesis. Nature 2019;568:557–60.
- Iyoda K, Sasaki Y, Horimoto M, Toyama T, Yakushijin T, Sakakibara M, et al. Involvement of the p38 mitogen-activated protein kinase cascade in hepatocellular carcinoma. Cancer 2003;97:3017–26.
- 41. Canovas B, Nebreda AR. Diversity and versatility of p38 kinase signalling in health and disease. Nat Rev Mol Cell Biol. 2021;22:346–66.
- Saha RN, Jana M, Pahan K. MAPK p38 regulates transcriptional activity of NFkappaB in primary human astrocytes via acetylation of p65. J Immunol. 2007;179:7101–9.
- Bentires-Alj M, Barbu V, Fillet M, Chariot A, Relic B, Jacobs N, et al. NF-kappaB transcription factor induces drug resistance through MDR1 expression in cancer cells. Oncogene 2003;22:90–7.
- Youssif C, Cubillos-Rojas M, Comalada M, Llonch E, Perna C, Djouder N, et al. Myeloid p38α signaling promotes intestinal IGF-1 production and inflammationassociated tumorigenesis. EMBO Mol Med. 2018;10:e8403.
- 45. Zheng T, Zhang B, Chen C, Ma J, Meng D, Huang J, et al. Protein kinase p38α signaling in dendritic cells regulates colon inflammation and tumorigenesis. Proc Natl Acad Sci USA. 2018;115:E12313–e12322.
- 46. Reyskens KM, Arthur JS. Emerging roles of the mitogen and stress activated kinases MSK1 and MSK2. Front Cell Dev Biol. 2016;4:56.
- Liu J, Shao T, Zhang J, Liu Q, Hua H, Zhang H, et al. Gamma synuclein promotes cancer metastasis through the MKK3/6-p38MAPK cascade. Int J Biol Sci. 2022;18:3167–77.
- 48. Huang Q, Lan F, Wang X, Yu Y, Ouyang X, Zheng F, et al. IL-1β-induced activation of p38 promotes metastasis in gastric adenocarcinoma via upregulation of AP-1/ c-fos, MMP2 and MMP9. Mol Cancer. 2014;13:18.
- 49. Gupta J, del Barco Barrantes I, Igea A, Sakellariou S, Pateras Ioannis S, Gorgoulis, et al. Dual function of p38α MAPK in colon cancer: suppression of colitisassociated tumor initiation but requirement for cancer cell survival. Cancer Cell. 2014;25:484–500.
- 50. Gomes LR, Terra LF, Wailemann RA, Labriola L, Sogayar MC. TGF-β1 modulates the homeostasis between MMPs and MMP inhibitors through p38 MAPK and ERK1/2 in highly invasive breast cancer cells. BMC Cancer. 2012;12:26.
- Hansen M, Rubinsztein DC, Walker DW. Autophagy as a promoter of longevity: insights from model organisms. Nat Rev Mol Cell Biol. 2018;19:579–93.
- 52. Chen C, Gao H, Su X. Autophagy-related signaling pathways are involved in cancer (Review). Exp Ther Med. 2021;22:710.
- Keil E, Höcker R, Schuster M, Essmann F, Ueffing N, Hoffman B, et al. Phosphorylation of Atg5 by the Gadd45β-MEKK4-p38 pathway inhibits autophagy. Cell Death Differ. 2013;20:321–32.
- 54. Zhu Q, Guo Y, Chen S, Fu D, Li Y, Li Z, et al. Irinotecan induces autophagydependent apoptosis and positively regulates ROS-related JNK- and P38-MAPK pathways in gastric cancer cells. Onco Targets Ther. 2020;13:2807–17.
- Zhang Z, Chen WQ, Zhang SQ, Bai JX, Liu B, Yung KK, et al. Isoliquiritigenin inhibits pancreatic cancer progression through blockade of p38 MAPK-regulated autophagy. Phytomedicine 2022;106:154406.
- 56. Damgaard RB. The ubiquitin system: from cell signalling to disease biology and new therapeutic opportunities. Cell Death Differ. 2021;28:423–6.
- 57. Foot N, Henshall T, Kumar S. Ubiquitination and the regulation of membrane proteins. Physiol Rev. 2017;97:253–81.

- Sun T, Liu Z, Yang Q. The role of ubiquitination and deubiquitination in cancer metabolism. Mol Cancer. 2020;19:146.
  - 59. Yu J, Qin B, Lou Z. Ubiquitin and ubiquitin-like molecules in DNA double strand break repair. Cell Biosci. 2020;10:13.
  - Grimsey NJ, Aguilar B, Smith TH, Le P, Soohoo AL, Puthenveedu MA, et al. Ubiquitin plays an atypical role in GPCR-induced p38 MAP kinase activation on endosomes. J Cell Biol. 2015;210:1117–31.
  - Ryu KJ, Park SM, Park SH, Kim IK, Han H, Kim HJ, et al. p38 stabilizes snail by suppressing DYRK2-mediated phosphorylation that is required for GSK3β-βTrCPinduced snail degradation. Cancer Res. 2019;79:4135–48.
  - Fang Y, Wang J, Wang G, Zhou C, Wang P, Zhao S, et al. Inactivation of p38 MAPK contributes to stem cell-like properties of non-small cell lung cancer. Oncotarget 2017;8:26702–17.
  - Wyganowska-Świątkowska M, Tarnowski M, Murtagh D, Skrzypczak-Jankun E, Jankun J. Proteolysis is the most fundamental property of malignancy and its inhibition may be used therapeutically (Review). Int J Mol Med. 2019;43:15–25.
  - 64. Vizovisek M, Ristanovic D, Menghini S, Christiansen MG, Schuerle S. The tumor proteolytic landscape: a challenging frontier in cancer diagnosis and therapy. Int J Mol Sci. 2021;22:2514.
  - Shin BA, Yoo HG, Kim HS, Kim MH, Hwang YS, Chay KO, et al. P38 MAPK pathway is involved in the urokinase plasminogen activator expression in human gastric SNU-638 cells. Oncol Rep. 2003;10:1467–71.
  - Kim A, Im M, Yim NH, Ma JY. Reduction of metastatic and angiogenic potency of malignant cancer by Eupatorium fortunei via suppression of MMP-9 activity and VEGF production. Sci Rep. 2014;4:6994.
  - 67. Li JP, Yang JL. Cyclin B1 proteolysis via p38 MAPK signaling participates in G2 checkpoint elicited by arsenite. J Cell Physiol. 2007;212:481–8.
  - Pokhrel RH, Acharya S, Ahn JH, Gu Y, Pandit M, Kim JO, et al. AMPK promotes antitumor immunity by downregulating PD-1 in regulatory T cells via the HMGCR/p38 signaling pathway. Mol Cancer. 2021;20:133.
  - Alam MS, Gaida MM, Bergmann F, Lasitschka F, Giese T, Giese NA, et al. Selective inhibition of the p38 alternative activation pathway in infiltrating T cells inhibits pancreatic cancer progression. Nat Med. 2015;21:1337–43.
  - Meng F, Zhang H, Liu G, Kreike B, Chen W, Sethi S, et al. p38γ mitogen-activated protein kinase contributes to oncogenic properties maintenance and resistance to poly (ADP-ribose)-polymerase-1 inhibition in breast cancer. Neoplasia 2011;13:472–82.
  - 71. Yang K, Liu Z, Liu J, Liu X, Chen X, et al. p38γ overexpression in gliomas and its role in proliferation and apoptosis. Sci Rep. 2013;3:2089.
  - 72. Risco A, del Fresno C, Mambol A, Alsina-Beauchamp D, MacKenzie KF, Yang HT, et al. p38γ and p38δ kinases regulate the Toll-like receptor 4 (TLR4)-induced cytokine production by controlling ERK1/2 protein kinase pathway activation. Proc Natl Acad Sci USA. 2012;109:11200–5.
  - Bain J, Plater L, Elliott M, Shpiro N, Hastie CJ, McLauchlan H, et al. The selectivity of protein kinase inhibitors: a further update. Biochem J. 2007;408:297–315.
  - 74. Prins ND, Harrison JE, Chu HM, Blackburn K, Alam JJ, Scheltens P. A phase 2 double-blind placebo-controlled 24-week treatment clinical study of the p38 alpha kinase inhibitor neflamapimod in mild Alzheimer's disease. Alzheimers Res Ther. 2021;13:106.
  - Newby LK, Marber MS, Melloni C, Sarov-Blat L, Aberle LH, Aylward PE, et al. Losmapimod, a novel p38 mitogen-activated protein kinase inhibitor, in non-STsegment elevation myocardial infarction: a randomised phase 2 trial. Lancet 2014;384:1187–95.
  - 76. Vergote I, Heitz F, Buderath P, Powell M, Sehouli J, Lee CM, et al. A randomized, double-blind, placebo-controlled phase 1b/2 study of ralimetinib, a p38 MAPK inhibitor, plus gemcitabine and carboplatin versus gemcitabine and carboplatin for women with recurrent platinum-sensitive ovarian cancer. Gynecol Oncol. 2020;156:23–31.
  - Goldman JW, Rosen LS, Tolcher AW, Papadopoulos K, Beeram M, Shi P, et al. Phase 1 and pharmacokinetic study of LY3007113, a p38 MAPK inhibitor, in patients with advanced cancer. Invest N. Drugs. 2018;36:629–37.
  - Yang SY, Miah A, Sales KM, Fuller B, Seifalian AM, Winslet M. Inhibition of the p38 MAPK pathway sensitises human colon cancer cells to 5-fluorouracil treatment. Int J Oncol. 2011;38:1695–702.
  - Han Z, Meng L, Huang X, Tan J, Liu W, Chen W, et al. Inhibition of p38 MAPK increases the sensitivity of 5-fluorouracil-resistant SW480 human colon cancer cells to noscapine. Oncol Lett. 2022;23:52.
  - King TE Jr., Bradford WZ, Castro-Bernardini S, Fagan EA, Glaspole I, Glassberg MK, et al. A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis. N. Engl J Med. 2014;370:2083–92.
  - Yamaguchi R, Kawata J, Yamamoto T, Ishimaru Y, Sakamoto A, Ono T, et al. Mechanism of interferon-gamma production by monocytes stimulated with myeloperoxidase and neutrophil extracellular traps. Blood Cells Mol Dis. 2015;55:127–33.
  - Zhang XH, Nam S, Wu J, Chen CH, Liu X, Li H, et al. Multi-kinase inhibitor with antip38y activity in cutaneous T-cell lymphoma. J Invest Dermatol. 2018;138:2377–87.

- Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. Nucleic Acids Res. 2018;46:D1074–D1082.
- Xu W, Liu R, Dai Y, Hong S, Dong H, Wang H. The role of p38γ in cancer: from review to outlook. Int J Biol Sci. 2021;17:4036–46.
- Zhang C, Wang Z, Yang Z, Wang M, Li S, Li Y, et al. Phase I escalating-dose trial of CAR-T therapy targeting CEA(+) metastatic colorectal cancers. Mol Ther. 2017;25:1248–58.
- Feng K, Liu Y, Guo Y, Qiu J, Wu Z, Dai H, et al. Phase I study of chimeric antigen receptor modified T cells in treating HER2-positive advanced biliary tract cancers and pancreatic cancers. Protein Cell. 2018;9:838–47.
- Shindo Y, Hazama S, Maeda Y, Matsui H, Iida M, Suzuki N, et al. Adoptive immunotherapy with MUC1-mRNA transfected dendritic cells and cytotoxic lymphocytes plus gemcitabine for unresectable pancreatic cancer. J Transl Med. 2014;12:175.
- Maeda Y, Yoshimura K, Matsui H, Shindo Y, Tamesa T, Tokumitsu Y, et al. Dendritic cells transfected with heat-shock protein 70 messenger RNA for patients with hepatitis C virus-related hepatocellular carcinoma: a phase 1 dose escalation clinical trial. Cancer Immunol Immunother. 2015;64:1047–56.
- Hazama S, Nakamura Y, Tanaka H, Hirakawa K, Tahara K, Shimizu R, et al. A phase II study of five peptides combination with oxaliplatin-based chemotherapy as a first-line therapy for advanced colorectal cancer (FXV study). J Transl Med. 2014;12:108.
- Miyazawa M, Katsuda M, Maguchi H, Katanuma A, Ishii H, Ozaka M, et al. Phase II clinical trial using novel peptide cocktail vaccine as a postoperative adjuvant treatment for surgically resected pancreatic cancer patients. Int J Cancer. 2017;140:973–82.
- Muro K, Chung HC, Shankaran V, Geva R, Catenacci D, Gupta S, et al. Pembrolizumab for patients with PD-L1-positive advanced gastric cancer (KEYNOTE-012): a multicentre, open-label, phase 1b trial. Lancet Oncol. 2016;17:717–26.
- El-Khoueiry AB, Sangro B, Yau T, Crocenzi TS, Kudo M, Hsu C, et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): an openlabel, non-comparative, phase 1/2 dose escalation and expansion trial. Lancet 2017;389:2492–502.
- Overman MJ, McDermott R, Leach JL, Lonardi S, Lenz HJ, Morse MA, et al. Nivolumab in patients with metastatic DNA mismatch repair-deficient or microsatellite instability-high colorectal cancer (CheckMate 142): an open-label, multicentre, phase 2 study. Lancet Oncol. 2017;18:1182–91.
- 94. Kang YK, Boku N, Satoh T, Ryu MH, Chao Y, Kato K, et al. Nivolumab in patients with advanced gastric or gastro-oesophageal junction cancer refractory to, or intolerant of, at least two previous chemotherapy regimens (ONO-4538-12, ATTRACTION-2): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017;390:2461–71.
- Gurusamy D, Henning AN, Yamamoto TN, Yu Z, Zacharakis N, Krishna S, et al. Multi-phenotype CRISPR-Cas9 screen identifies p38 kinase as a target for adoptive immunotherapies. Cancer Cell. 2020;37:818–833.e9.

## **AUTHOR CONTRIBUTIONS**

TP contributed to the design of the article, table, figures and prepared the manuscript. LGM provided supervision, prepared, and finalized the manuscript for submission. XHZ and SR contributed the idea about p38 in cancers. All the authors contributed to the manuscript preparation and approved the submitted version.

## FUNDING

Open access funding provided by SCELC, Statewide California Electronic Library Consortium.

## **COMPETING INTERESTS**

The authors declare no competing interests.

### ADDITIONAL INFORMATION

**Correspondence** and requests for materials should be addressed to Laleh G. Melstrom.

Reprints and permission information is available at http://www.nature.com/ reprints

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http:// creativecommons.org/licenses/by/4.0/.

#### © The Author(s) 2023

1189